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## 4 **Biochar granulation and particle size influence hydrological** 5 **performance of green roof substrates**

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18 **Abstract**

19 Green roofs are increasingly being implemented in cities to improve stormwater management  
20 and provide additional ecosystem services. Biochar, a carbon-rich material derived from  
21 pyrolyzed biomass, has emerged as a promising substrate additive to improve hydrological  
22 performance of green roofs; however, unprocessed biochar is susceptible to erosion loss. Biochar  
23 granulation and particle size modification have been suggested as effective approaches to  
24 mitigate erosion losses but research on the effects of biochar post-processing on green roof  
25 substrate properties and hydrological performance remains scarce. We investigated how biochar  
26 granulation and particle size influences surface and pore characteristics and hydraulic properties  
27 of green roof substrates. Granulated biochars exhibited uniform surface morphology and pore  
28 structure, resulting in consistent hydrological responses across particle size ranges, likely due to  
29 the homogeneous structure formed during granulation. In contrast, unprocessed biochars showed  
30 a significant increase in Brunauer–Emmett–Teller (BET) specific surface area and pore volume  
31 with decreasing particle size. The amendment of granulated biochars improved substrate field  
32 capacity, indicating enhanced stormwater retention. In contrast, the addition of fine (0.5–1 mm)  
33 and intermediate-sized (2–2.8 mm) unprocessed biochars increased plant available water,  
34 potentially enhancing plant performance. Although reduced saturated hydraulic conductivity  
35 associated with fine unprocessed biochars may improve green roof detention capacity, fine  
36 biochar particle size is expected to increase erosion losses and periodic waterlogging. We  
37 conclude that the application of either granulated biochar or intermediate-sized unprocessed  
38 biochar provide a balanced enhancement of stormwater retention and detention capacities, as  
39 well as improved plant available water in green roof systems.

40

41 **Keywords:** biochar post-processing, surface and pore characteristics, soil water characteristic  
42 curve, hydraulic properties, stormwater management, green infrastructure  
43

## 44 **1. Introduction**

45           Rapid urbanization has led to a proliferation of impervious surfaces in cities, increasing  
46 the risks of urban flooding (Chen et al., 2015; Rentschler, 2023). Such flooding can have  
47 profound adverse socioeconomic and environmental impacts, including damage to property and  
48 infrastructure, contamination of water resources, and loss of life (Hammond et al., 2015).  
49 Climate change is predicted to amplify the frequency and intensity of extreme precipitation  
50 globally (Madsen et al., 2014; Donat et al., 2016), further exacerbating the hazards associated  
51 with urban flooding. It is projected that nearly two-thirds of global population will live in urban  
52 areas by 2050 (Ritchie & Roser, 2022), which will pose severe stresses on urban drainage  
53 systems and increase the risks of system failure, particularly during extreme precipitation events  
54 (Yazdanfar & Sharma, 2015).

55           Green roofs, which are rooftops with vegetation planted in engineered growing media,  
56 have been increasingly implemented in cities to improve stormwater management (Zheng, 2021;  
57 Alim et al., 2022). In general, green roofs can mitigate flood risks by attenuating total discharge  
58 volume and reducing and delaying peak flows (Li & Babcock, 2014; Palermo et al., 2019).  
59 Substrate and vegetation play important roles in hydrological performance on green roofs.  
60 Permeable substrates with large pore sizes often show low water retention and detention  
61 capacities (Stovin et al., 2015). Vegetation generally enhances green roof water retention and  
62 reduces discharge water volume through evapotranspiration (Stovin et al., 2015), water uptake  
63 (Wolf & Lundholm, 2008; Nagase & Dunnett, 2012), and rainwater interception by plant  
64 aboveground structures (Tan & Wang, 2023). However, green roofs are highly exposed to  
65 intense storms, severe and periodic drought, and high wind (Oberndorfer et al., 2007), presenting  
66 challenging conditions for substrate and vegetation performance. For example, substrates are

67 susceptible to particle loss through water and wind erosion (Liao et al., 2022), potentially  
68 reducing hydrological performance. In addition, difficult plant growth conditions may result in  
69 vegetation degradation, reducing stormwater and ecological benefits of green roof systems. With  
70 the intensification of extreme precipitation events under climate change, enhancing hydrological  
71 performance of substrates and vegetation is essential for improving stormwater management of  
72 green roofs and alleviating pressure on urban stormwater infrastructure (Kourtis & Tsihrintzis,  
73 2021).

74         Hydraulic conductivity (K) and the soil-water characteristic curve (SWCC) are two  
75 hydrological properties of substrates that are fundamental determinants of stormwater retention  
76 and plant performance on green roofs. K determines the rate at which water flows through the  
77 substrate, which affects the detention capacity of roof systems (Peng et al., 2019). The SWCC  
78 describes the substrate's ability to retain water as a function of matric suction, and is used to  
79 determine key parameters, including water retention capacity (WRC), permanent wilting point,  
80 and plant available water (De-Ville et al., 2017). The hydrological properties and performance of  
81 substrates are typically influenced by their physical characteristics, particularly pore size  
82 distribution, particle size distribution, and particle hydrophobicity (De-Ville et al., 2017; Hill et  
83 al., 2019). Optimizing substrate properties is essential to balance stormwater retention and  
84 detention while enhancing plant available water to support vegetation performance. Accurate and  
85 high-resolution measurements of hydraulic properties are critical as they provide key parameters  
86 for reliable hydrological modeling and large-scale simulations of green roof performance.

87         Biochar, a carbon-rich, porous material produced from pyrolyzed biomass, has been  
88 suggested as an ideal substrate additive for green roofs to enhance hydrological and vegetation  
89 performance (Cao et al., 2014; Liao et al., 2023; Lee & Kwon, 2024). Biochar amendment can

90 improve water retention and detention capacity (Huang et al., 2020; Gan et al., 2021; Tan &  
91 Wang, 2023), enhance plant available water content (Cao et al., 2014; Petreje et al., 2025), and  
92 reduce saturated (Huang et al., 2020; Gan et al., 2021) and unsaturated (Hussain & Ravi, 2021)  
93 hydraulic conductivity of substrates, primarily due to large specific surface area and high  
94 porosity of biochar and increased water flow tortuosity. In addition, biochar often exhibits a  
95 bimodal pore structure with pores at the nano- to micro-scale (Lu & Zong, 2018), which can  
96 influence water retention and infiltration processes in substrates. However, unprocessed biochar  
97 is lightweight and thus susceptible to wind and water erosion, particularly on highly exposed  
98 roof systems, potentially resulting in biochar loss and environmental pollution (Liao et al.,  
99 2022c).

100 Biochar granulation and particle size modification are important post-processing  
101 approaches suggested to enhance biochar physicochemical properties (Thomas, 2021) while  
102 mitigating biochar erosion loss (Liao et al., 2022c). We recently found that addition of  
103 granulated biochar to green roof substrates increased WRC by 26%, though this improvement  
104 was less pronounced than unprocessed biochar amendment due to lower porosity and  
105 hydrophobicity (Liao et al., 2022a). Granulated biochar has also been shown to reduce erosion  
106 losses of both biochar and non-biochar particles on green roof substrates (Liao et al., 2022c).  
107 However, no research has been done examining hydraulic properties of green roof substrates  
108 amended with granulated versus unprocessed biochars or the underlying mechanisms involved.

109 Application of relatively large biochar particles can potentially reduce biochar erosion  
110 losses due to increased particle mass, while enhancing hydraulic properties of the substrates.  
111 Coarse biochar particles may also better increase substrate WRC than fine biochar particles due  
112 to their higher intra-porosity and hydrophilicity (Wang et al., 2019; Edeh & Mašek, 2021). In

113 contrast, small biochar particles generally have large specific surface area and high intra-  
114 porosity, thereby more effectively enhancing substrate WRC compared to large biochar particles  
115 (Ibrahim et al., 2017). Moreover, the addition of fine biochar particles can increase water flow  
116 tortuosity by clogging substrate pores, further improving WRC (Esmaeelnejad et al., 2017).  
117 However, fine biochar particles may reduce air-filled porosity and water infiltration, resulting in  
118 waterlogging of green roofs during and after intense rainfall events that requires additional  
119 stormwater control measures (Werdin et al., 2021). Studies have also shown that the effects of  
120 biochar particle size on hydraulic properties vary with soil particle size distribution (Razzaghi et  
121 al., 2020). Green roof substrates are engineered media consisting of materials in various particle  
122 sizes, which generally have different textures and hydraulic properties than natural soils (Liao et  
123 al., 2023), highlighting the importance of designing appropriate biochar for green roof  
124 applications. Research on the effects of biochar particle size on the hydraulic properties of green  
125 roof substrates is lacking, in particular, accurate and high-resolution measurements of hydraulic  
126 properties, including K and SWCC.

127         In the present study we investigated the effects of biochar type (granulated vs.  
128 unprocessed biochars) and particle size (0.25–6.3 mm) on K and SWCC of green roof substrate  
129 using accurate, high-resolution KSAT and HYPROP devices, respectively. To understand the  
130 underlying mechanisms of biochar effects on substrate hydraulic properties, we also examined  
131 specific surface area, pore structure and characteristics, and chemical functional groups of  
132 biochars. The following hypotheses were tested: (1) unprocessed biochar will better increase  
133 WRC and decrease K of substrates compared to granulated biochar due to higher macroporosity;  
134 (2) small biochar particles will result in increased WRC and reduced K compared to large  
135 biochar particles due to large specific surface areas; (3) intermediate-sized biochar particles will

136 better balance water retention and detention of substrates; (4) all biochar-substrate mixtures will  
137 show a biphasic or multiphasic SWCC.

138

## 139 **2. Materials and methods**

### 140 ***2.1. Biochar and substrate preparation and characterization***

141 The two biochars used in this study were produced from conifer feedstock. Unprocessed  
142 biochar was produced from conifer sawmill wastes with mixed particle sizes through slow  
143 pyrolysis at 625 °C and a residence time of ~20 minutes (Titan Clean Energy Projects Co.,  
144 Canada). Granulated biochar was produced from similar raw biochar through a drum granulation  
145 process with ground unprocessed biochar and a proprietary binder. Unprocessed biochar was  
146 sieved into particle size ranges of 0.25–0.5 mm, 0.5–1 mm, 1–2 mm, 2–2.8 mm, 2.8–4 mm,  
147 while granulated biochar was sieved into 1–2mm, 2–2.8 mm, 2.8–4mm, and 4–6.3 mm. The  
148 overlapping 1–4 mm particle size range was precisely controlled across both biochar types to  
149 investigate biochar granulation effects. Unprocessed biochar was thermally treated at 100 °C for  
150 24 hours to reduce phytotoxic compounds. A commercial green roof substrate (Gro-Bark, Inc.,  
151 Canada) that follows FLL (Forschungsgesellschaft Landschaftsentwicklung Landschaftsbau)  
152 guidelines was used in this study. The substrate is classified as well-graded sand and consists of  
153 70% porous aggregates, 25% composted organic matter, and 5% fine sand. Unprocessed and  
154 granulated biochars in each particle size class were applied to the substrate at a dosage of 25 t/ha  
155 (equivalent to ~4.5% w/w). Detailed information on biochar and substrate preparation,  
156 characterization, and testing methods are documented in [Liao et al. \(2022a\)](#).

157 Scanning electron microscope (SEM) images were taken to qualitatively assess surface  
158 structure and morphological features of unprocessed and granulated biochars using a Hitachi

159 Scanning Electron Microscope SU3500 (Hitachi Ltd., Japan). Representative biochar samples  
160 were mounted on stubs with carbon tape and were sputter-coated with gold-palladium using a  
161 sputter coater (Bal-Tec SCD 050, Bal-Tec, USA) to enhance conductivity. The surface  
162 functional groups of biochar were characterized using Fourier transform infrared spectroscopy  
163 (FTIR) with a Bruker Tensor 27 FTIR Spectrometer (Bruker, USA). Spectra were acquired in the  
164 range of 4000 to 400  $\text{cm}^{-1}$  at a resolution of 4  $\text{cm}^{-1}$ , with 32 scans per sample. The samples were  
165 prepared as compressed pellets from a homogenized mixture of 0.2 g potassium bromide (KBr)  
166 and 0.25% of biochar ground with a mortar and pestle. Biochar was oven-dried at 65 °C for 12  
167 hours prior to the measurements to remove moisture.

168 Nitrogen adsorption-desorption isotherms were determined at 77 K using a NOVA 1200e  
169 surface area analyzer (Quantachrome Instruments, USA). Biochar specific surface area was  
170 measured through the Brunauer–Emmett–Teller (BET) method with a relative pressure ( $P/P_0$ )  
171 range of 0.01–0.3. The total pore volume of biochar was determined using a single nitrogen  
172 adsorption point at  $P/P_0 = 0.98$  (Sigmund et al., 2017). Biochar micropore volume was obtained  
173 through the Dubinin-Radushkevich (DR) method and the mesopore volume was calculated as the  
174 difference between total pore volume and micropore volume (Leng, 2021). The mesopore and  
175 micropore diameter distribution of biochar was determined by the Barrett–Joyner–Halenda  
176 (BJH) and Density Functional Theory (DFT) methods (Bardestani et al., 2019; Leng, 2021).  
177 Biochar was degassed at 150 °C for 20 hours prior to the measurements. The data were analyzed  
178 in NOVAVin software (Quantachrome Instruments, USA) to obtain the specific surface area,  
179 pore volume, and pore size distribution of biochar. Duplicate measurements were conducted for  
180 unprocessed and granulated biochars in each particle size range.

181

## 182 2.2. Hydraulic property measurements

### 183 2.2.1. *Soil-water characteristic curve and unsaturated hydraulic conductivity*

184 The soil-water characteristic curve (SWCC) and unsaturated hydraulic conductivity  
185 ( $K_{\text{unsat}}$ ) of the substrate and biochar-substrate mixtures were determined using a HYPROP device  
186 (METER Group, Germany). The HYPROP system measures the soil water retention and release  
187 processes using an evaporation method (Peters & Durner, 2008; Peters, 2013). The substrate and  
188 biochar-substrate mixtures were packed into the HYPROP sample ring (volume = 250 cm<sup>3</sup>,  
189 height = 5 cm, inside diameter = 8 cm). The samples were saturated in deionized, degassed water  
190 from bottom to top in a container for 24 hours prior to measurements. The samples were  
191 mounted on the HYPROP instrument with two tensiometers positioned at heights of 1.75 cm and  
192 3.75 cm to measure matric suction, while a connected HYPROP balance simultaneously  
193 recorded mass changes throughout the evaporation process. Samples were measured at room  
194 temperature (20 to 25 °C, with an average temperature of 23°C). The HYPROP measurements  
195 were terminated when cavitation occurred in both tensiometers. The dry mass of the samples was  
196 determined by oven-drying at 105 °C for 24 hours. Each sample measurement lasted an average  
197 of 24 days, ranging from a minimum of 11 days (pure substrate) to a maximum of 32 days  
198 (biochar-amended substrates). Duplicate measurements were conducted for each substrate and  
199 biochar-substrate mixture.

200 The HYPROP-FIT software (METER Group, USA) was used to analyze data from the  
201 evaporation experiments and to fit  $K_{\text{unsat}}$  following the HYPROP-FIT manual (Pertassek, et al.,  
202 2015). The data were fitted with the Peters–Durner–Iden (PDI) expression (Peters, 2013; Iden &  
203 Durner, 2014) of the bimodal constrained van Genuchten (VGbi-PDI) model (Van Genuchten,  
204 1980; Haghverdi et al., 2020), which accounts for bimodal pore structure. Model fits were

205 assessed using the root mean square error (RMSE) and sample-sized adjusted Akaike  
 206 Information Criterion (AICa) values. The PDI variant was employed to capture the full SWCC,  
 207 including the dry-end, due to its improved accuracy and reliable model performance even  
 208 without direct dry-end measurements (Haghverdi et al., 2020). The general form of the PDI  
 209 retention model is expressed as the sum of capillary and adsorptive water retention (Peters, 2013;  
 210 Iden & Durner, 2014):

$$211 \quad \theta(h) = (\theta_s - \theta_r)S_{cap}(h) + \theta_r S_{ad}(h) \quad (1)$$

212 where  $\theta$  is the volumetric water content ( $\text{m}^3/\text{m}^3$ ),  $h$  is the matric suction (cm),  $\theta_s$  is the saturated  
 213 water content ( $\text{m}^3/\text{m}^3$ ),  $\theta_r$  is the maximum water content for water adsorption ( $\text{m}^3/\text{m}^3$ ), and  $S_{cap}$   
 214 and  $S_{ad}$  are the saturation of capillary and adsorptive water retention ( $\text{m}^3/\text{m}^3$ ), respectively.

215 The capillary scaling function  $S_{cap}$  of the PDI variant is calculated as:

$$216 \quad S_{cap}(h) = \frac{\Gamma(h) - \Gamma_0}{1 - \Gamma_0} \quad (2)$$

217 where  $\Gamma_0$  is the basic function at  $h = h_0$ . The adsorptive scaling function is calculated as:

$$218 \quad S_{ad}(x) = 1 + \frac{1}{x_a - x_0} \left\{ x - x_a + b \ln \left[ 1 + \exp \left( \frac{x_a - x}{b} \right) \right] \right\} \quad (3)$$

219 where  $x = \text{pF} = \log_{10}(h, \text{cm})$ ,  $x_a = \log_{10}(h_a, \text{cm})$ , with  $h_a$  being the suction at air entry for the  
 220 adsorptive retention, and  $x_0 = \log_{10}(h_0, \text{cm})$ , with  $h_0$  being the suction where the water content is  
 221 0. The value of  $h_0 = 10^{6.8}$  cm was used to represent the section at oven dryness for 105 °C. The  
 222 smoothing parameter  $b$  is dependent on the capillary scaling function  $S_{cap}$  and is given by:

$$223 \quad b = 0.1 + \frac{0.2}{n^2} \left\{ 1 - \exp \left[ - \left( \frac{\theta_r}{\theta_s - \theta_r} \right)^2 \right] \right\} \quad (4)$$

224 where  $n$  is an empirical curve shape parameter. The bimodal constrained van Genuchten model is  
 225 a weighted sum of the unimodal van Genuchten functions, expressed as:

$$226 \quad \Gamma(h) = \sum_{i=1}^2 w_i \left[ \frac{1}{1 + (\alpha_i h)^{n_i}} \right]^{m_i} = \sum_{i=1}^2 (w_i \Gamma_i) \quad (5)$$

227 where  $\Gamma$  is the effective saturation ranging from 0 to 1,  $h$  is the matric suction (cm),  $w_i$  is the  
 228 weighting factor that  $\sum w_i = 1$ , and  $\alpha_i$ ,  $n_i$ , and  $m_i$  are empirical curve shape parameters, with  
 229  $m_i = 1 - n_i^{-1}$ .

230 The unsaturated hydraulic conductivity ( $K_{\text{unsat}}$ ) is calculated as:

$$231 \quad K = K_s[(1 - \omega)K_{rel}^{cap}(S_{cap}) + \omega K_{rel}^{film}(S_{ad})] \quad (6)$$

232 where  $K$  is the unsaturated hydraulic conductivity,  $K_s$  is the saturated conductivity,  $\omega$  is a  
 233 weighting factor that  $\sum w_i = 1$ ,  $K_{rel}^{cap}$  is the relative conductivity associated with capillary flow,  
 234 and  $K_{rel}^{film}$  is the relative conductivity associated with film flow, as shown below:

$$235 \quad K_{rel}^{cap} = \left(\sum_{i=1}^2 w_i S_{cap_i}\right)^\tau \left[1 - \frac{\sum_{i=1}^2 w_i \alpha_i (1 - \Gamma_i^{1/m_i})^{m_i}}{\sum_{i=1}^2 w_i \alpha_i (1 - \Gamma_0^{1/m_i})^{m_i}}\right]^2 \quad (7)$$

$$236 \quad K_{rel}^{film} = \left(\frac{h_0}{h_a}\right)^{a(1-S_{ad})} \quad (8)$$

237 where  $\tau$  is the tortuosity and connectivity parameter and the parameter  $a$  is the slope on the log-  
 238 log scale, which is set to  $-1.5$  (Peters, 2013).

239 The field capacity and permanent wilting point of the samples were determined as water  
 240 retention at pF = 2.5 ( $\sim 33$  kPa) and pF = 4.2 ( $\sim 1500$  kPa), respectively (Cassel & Nielsen,  
 241 1986). Plant available water content was calculated as the difference between field capacity and  
 242 permanent wilting point (Cassel & Nielsen, 1986).

243

## 244 **2.2.2. Pore size distribution of substrates and biochar-substrate mixtures**

245 The pore size distribution of the substrate and biochar-substrate mixtures was derived  
 246 from the sample water retention data at matric potentials of approximately  $-3$ ,  $-6$ , and  $-33$  kPa,  
 247 which correspond to pore diameters of 100, 50, and 9  $\mu\text{m}$ , respectively (Edeh & Mašek, 2021).

248 Pore volume fractions were estimated from variations in volumetric water content, using

249 designated pore diameter ranges to define class boundaries. Macro-porosity, meso-porosity, and  
250 micro-porosity corresponded to the volume fractions of pores with radii greater than 100  $\mu\text{m}$ ,  
251 between 50 and 100  $\mu\text{m}$ , and less than 50  $\mu\text{m}$ , respectively (Edeh & Mašek, 2021).

252

### 253 **2.2.3. Saturated hydraulic conductivity**

254 Saturated hydraulic conductivity ( $K_{\text{sat}}$ ) was measured using a KSAT device (METER  
255 Group, USA) with a 2-cm constant head and 1-second measurement intervals. The  $K_{\text{sat}}$  was  
256 determined using the Darcy's law:

$$257 \quad K_{\text{sat}} = \frac{Q}{A} \times \frac{L}{H} \quad (9)$$

258 where  $Q$  is the steady-state flow rate from a Mariotte flask,  $A$  is the cross-sectional area of the  
259 substrate sample,  $L$  is the length of the substrate sample, and  $H$  is the hydraulic head difference  
260 between inlet and outlet level.  $K_{\text{sat}}$  of the samples was normalized to 10 °C by accounting for the  
261 temperature dependence of water viscosity (Gao & Shao, 2015). The substrate and biochar-  
262 substrate mixtures were saturated with deionized water for 24 hours prior to the measurements.  
263 Triplicate samples were analyzed for each substrate and biochar-substrate mixture, with each  
264 sample measured ten times.

265

### 266 **2.3. Statistical analysis**

267 The effects of biochar processing method and particle size on surface area and pore  
268 characteristics of biochars, as well as the hydraulic properties of the substrate and biochar-  
269 substrate mixtures, were analyzed using one-way analysis of variance (ANOVA). Two-way  
270 ANOVA was performed for the subset of factorial treatments (granulated and unprocessed  
271 biochars in particle size ranges of 1–2 mm, 2–2.8 mm, and 2.8–4 mm) to examine the biochar

272 type by particle size interactions. Post-hoc Tukey's HSD tests were conducted for ANOVA  
273 results that were significant. *t*-tests were performed to determine whether each biochar treatment  
274 was statistically different from the pure substrate. *P*-values were adjusted using the false  
275 discovery rate procedure for multiple comparisons (Benjamini & Hochberg, 1995). All statistical  
276 analyses and data visualization were conducted in R version 4.4.1 (R Core Team, 2024).

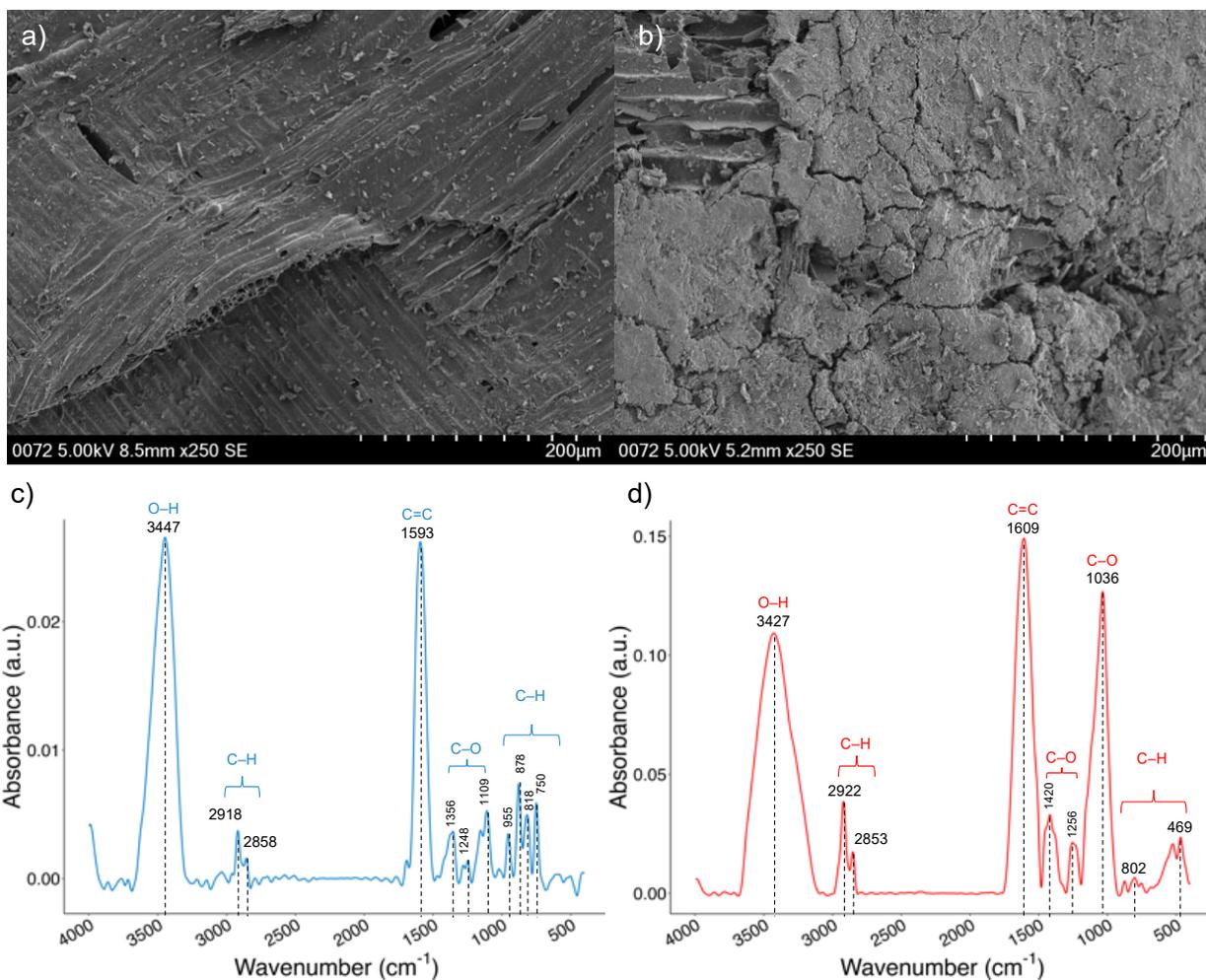
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### 278 **3. Results**

#### 279 ***3.1. Biochar physicochemical properties***

##### 280 *3.1.1. Biochar pore structure and functional groups*

281 The SEM images showed that unprocessed biochar was well-structured, with the pores  
282 corresponding to the wood cells of the conifer feedstock (Fig. 1a). In contrast, the pore structure  
283 of the granulated biochar collapsed during the granulation process (Fig. 1b). In addition,  
284 granulated biochar exhibited a non-uniform, fractured surface morphology and was composed of  
285 biochar particles of varying sizes (Fig. 1b).



286

287 **Fig. 1.** Scanning electron microscope (SEM) images for unprocessed (a) and granulated (b)  
 288 biochars at the magnification of  $250\times$ , and Fourier-transform infrared spectroscopy (FTIR)  
 289 spectra of unprocessed (c) and granulated (d) biochars. Dashed lines indicate peaks of FTIR  
 290 spectra. Unprocessed and granulated biochars are shown in blue and red colors, respectively.

291

292 The FTIR spectra of unprocessed (Fig. 1c) and granulated (Fig. 1d) biochars revealed  
 293 characteristic absorbance bands associated with organic and oxygen-containing functional  
 294 groups. In the unprocessed biochar, a broad absorption band at  $3447\text{ cm}^{-1}$  was attributed to O–H  
 295 stretching vibrations of hydroxyl groups. A prominent peak at  $1593\text{ cm}^{-1}$  was assigned to  
 296 aromatic C=C stretching. Additionally, weak absorption bands at  $2918\text{ cm}^{-1}$  and  $2858\text{ cm}^{-1}$  were  
 297 related to C–H bonds. The FTIR spectrum of the granulated biochar exhibited similar strong

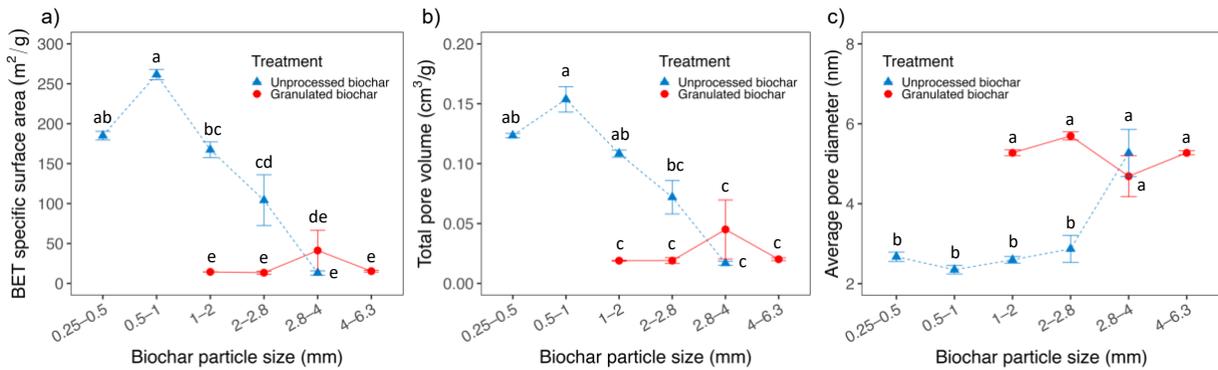
298 bands at  $3427\text{ cm}^{-1}$  and  $1609\text{ cm}^{-1}$ , corresponding to O–H and aromatic C=C stretching  
299 vibrations, respectively. The C–H stretching peaks were also present at  $2922\text{ cm}^{-1}$  and  $2853\text{ cm}^{-1}$ .  
300 Notably, a distinct and sharp peak at  $1036\text{ cm}^{-1}$  appeared in the granulated biochar, which  
301 corresponds to C–O stretching vibrations. Several weak bands related to C–O and C–H  
302 stretching vibrations were also observed in both unprocessed and granulated biochars, indicating  
303 the presence of residual oxygenated functional groups and aliphatic linkages.

304

### 305 *3.1.2. Biochar specific surface area and pore characteristics*

306 Unprocessed biochars had a greater mean BET specific surface area ( $95 \pm 30\text{ m}^2/\text{g}$  [mean  
307  $\pm$  SE]) than granulated biochars ( $23 \pm 9\text{ m}^2/\text{g}$ ) for the particle size ranges where these overlapped  
308 (i.e., 1–2 mm, 2–2.8 mm, 2.8–4 mm) ( $p < 0.05$ ) (Table 1). There were also strong interactive  
309 effects (Table 1): BET specific surface areas of unprocessed biochars increased with decreasing  
310 biochar particle sizes ( $p < 0.05$ ), with the unprocessed biochar at 0.5–1 mm ( $262 \pm 6\text{ m}^2/\text{g}$ ) and  
311 2.8–4 mm ( $13 \pm 3\text{ m}^2/\text{g}$ ) showing the highest and lowest BET specific surface areas, respectively,  
312 while there was no particle size effect with granulated biochar (Fig. 2a; Table 2). Likewise,  
313 unprocessed biochars ( $0.066 \pm 0.017\text{ cm}^3/\text{g}$ ) had a higher total pore volume than granulated  
314 biochars for the overlapping particle sizes ( $0.028 \pm 0.008\text{ cm}^3/\text{g}$ ), with significant interactive  
315 effects of biochar type and particle size ( $p < 0.05$ ) (Fig. 2b; Table 1). The total pore volume of  
316 unprocessed biochars generally declined with biochar particle size (Fig. 2b), with the  
317 unprocessed biochars at 0.5–1 mm ( $0.154 \pm 0.011\text{ cm}^3/\text{g}$ ) and 2.8–4 mm ( $0.017 \pm 0.002\text{ cm}^3/\text{g}$ )  
318 showing the highest and lowest total pore volumes, respectively (Fig. 2b; Table 2). However, no  
319 particle size effect on BET specific surface area or total pore volume was observed for the  
320 granulated biochars. Similar patterns were observed for the biochar micropore volume (Fig. S1

321 and Table 2). Overall, unprocessed biochars (86.4%) had a higher percentage of micropores than  
 322 granulated biochars (60.7%), while granulated biochars presented higher percentage of mesopore  
 323 (29.3%) than unprocessed biochars (13.6%) (Fig. S1 and Table S1).

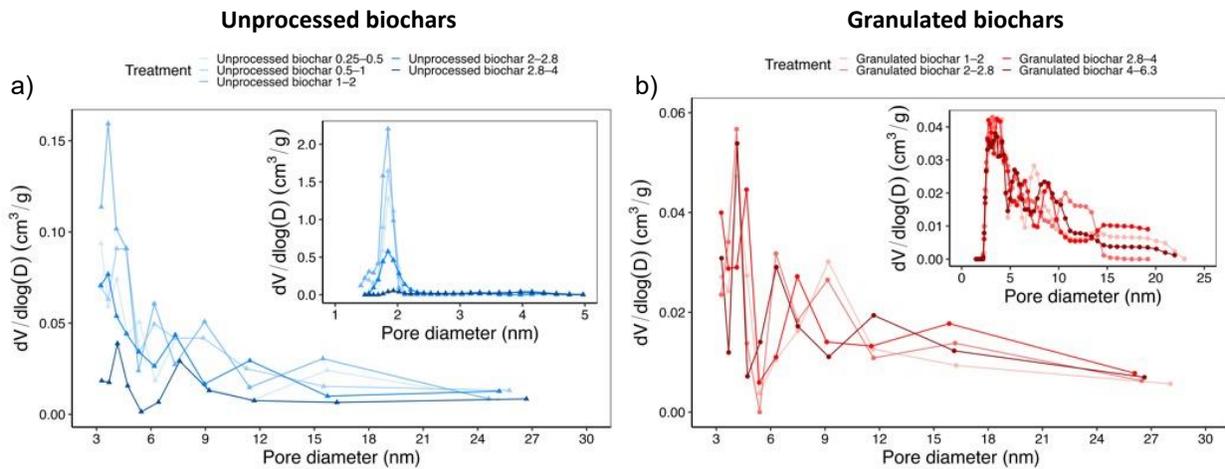


324 **Fig. 2.** BET specific surface area (a), total pore volume (b), and average pore diameter (c) of  
 325 unprocessed and granulated biochars across different particle size ranges. Means are plotted  $\pm$  1  
 326 SE based on duplicate measurements for each biochar. Dots with different lowercase letters  
 327 differ significantly at  $p < 0.05$  according to Tukey's HSD test. Unprocessed and granulated  
 328 biochars are shown in blue and red colors, respectively.

330

331 Biochar pore size distributions were also estimated from gas sorption data. Granulated  
 332 biochars had a larger estimated average pore diameter (5.22 nm) than unprocessed biochars (3.58  
 333 nm) for the overlapping particle sizes, with biochar type and biochar particle size showing  
 334 interactive effects ( $p < 0.05$ ) (Table 1). Unprocessed biochars presented significant particle size  
 335 effects, with the 2.8–4 mm fraction showing the largest average pore diameter of  $5.27 \pm 0.59$  nm,  
 336 while no differences were observed among the other particle size ranges (Fig. 2c). Additionally,  
 337 granulated biochars exhibited a larger mean DFT pore diameter ( $2.92 \pm 0.08$  nm), indicative of  
 338 microporous structure, compared to unprocessed biochars ( $1.90 \pm 0.03$  nm) (Fig. S2; Table S1).  
 339 No particle size effect on average pore diameter or DFT pore diameter was found for the  
 340 granulated biochars.

341 Consistent with the pore volume and diameter results, the pore size distribution indicated  
 342 that unprocessed biochars generally presented greater pore volumes across 3–30 nm ranges (Fig.  
 343 3). Moreover, unprocessed biochars exhibited a high concentration of small pores around ~2 nm  
 344 in diameter (Fig. 3a). In contrast, granulated biochars showed a highly variable pore size  
 345 distribution with relatively low pore volumes across the 2–20 nm range (Fig. 3b).

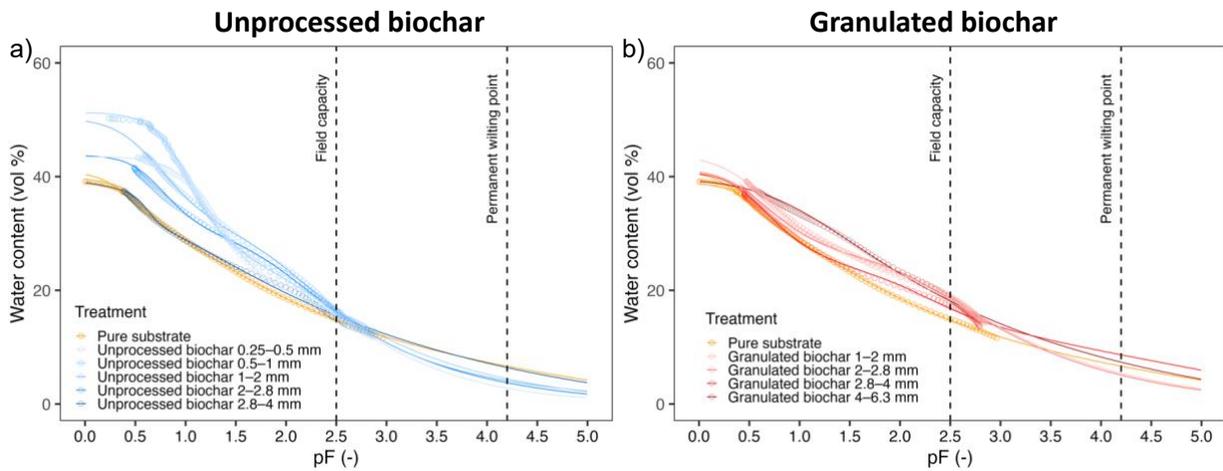


346  
 347 **Fig. 3.** Pore size distribution of unprocessed (a) and granulated (b) biochars across different  
 348 particle size ranges. Mesopore diameter distributions, determined by the Barrett–Joyner–Halenda  
 349 (BJH) method, are shown in the main panels, while micropore diameter distributions, derived  
 350 from Density Functional Theory (DFT), are presented in the embedded insets.

351  
 352 **3.2. Soil-water characteristic curves and hydraulic properties**

353 Biochar post-processing and particle size affected water retention and release behavior of  
 354 the substrates, highlighting their important roles in influencing hydrological performance and  
 355 stormwater management of green roofs (Fig. 4). The water retention and release processes of  
 356 biochar-amendment substrates were best described by the PDI variant of the bimodal constrained  
 357 van Genuchten (VGbi-PDI) model, indicating the presence of a bimodal pore structure in the  
 358 biochar-substrate mixtures. The water retention of substrates increased from 0 pF to 2.5 pF with  
 359 the addition of unprocessed and granulated biochars across all particle size ranges, except for the

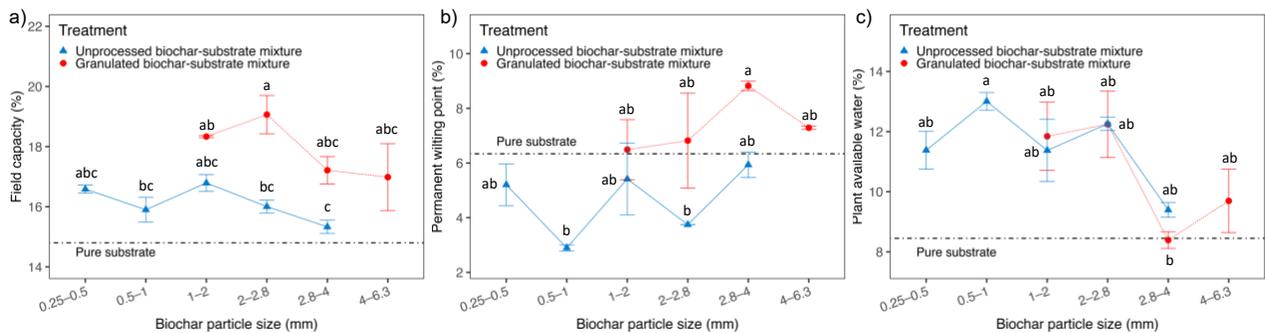
360 2.8–4 mm biochars (Fig. 4). The increase was more pronounced with unprocessed biochars. In  
 361 addition, substrates amended with unprocessed biochars in the 0.25–2.8 mm particle size range  
 362 showed sharper declines in water content between 0.5 pF and 2.5 pF compared to those amended  
 363 with granulated biochars (Fig. 4). The fitted parameter values for the VGbi-PDI models were  
 364 describe in Table S2. The SWCC in kPa was shown in Fig. S3.



365  
 366 **Fig. 4.** Soil-water characteristic curves for the pure substrate (a–b) and the substrates amended  
 367 with unprocessed (a) and granulated (b) biochars in different particle size ranges. Open symbols  
 368 represent the measurement data from the HYPROP device. Lines represent the fitted PDI variant  
 369 of the bimodal constrained van Genuchten model for one representative replicate. Dashed lines  
 370 indicate field capacity at 2.5 pF and permanent wilting point at 4.2 pF. Blue and red colors  
 371 indicate unprocessed and granulated biochars, respectively, with the darker colors presenting  
 372 larger particle size ranges. Orange color indicates the pure substrate.

373  
 374 The substrates with granulated biochar addition (18%) showed significantly higher mean  
 375 field capacity compared to those amended with unprocessed biochars (16%) and those without  
 376 biochar (15%) (Fig. 5a and Table 3). In addition, the granulated biochar-amended substrates  
 377 (7.4%) had a higher permanent wilting point than the unprocessed biochar-amended substrates  
 378 (5.0%) ( $p < 0.05$ ) (Fig. 5b and Table 3). However, biochar post-processing showed no effect on  
 379 plant available water of substrates (Fig. 5c and Table 3). Biochar particle size effects on field  
 380 capacity and plant available water were observed within the overlapping size ranges, with the

381 biochars in the 1–2 mm and 2–2.8 mm categories better increasing mean field capacity and plant  
 382 available water than the 2.8–4 mm biochar ( $p < 0.05$ ) (Fig. 5 and Table 3). In addition, the  
 383 substrate with the amendment of unprocessed biochar in 0.5–1 mm had higher plant available  
 384 water than that with the granulated biochar in 2.8–4 mm (Fig. 5c). The substrates amended with  
 385 0.25–0.5 mm and 1–2 mm unprocessed biochars and 1–2 mm and 2–2.8 mm granulated biochars  
 386 had higher field capacity than the pure substrate ( $p < 0.05$ ). Moreover, the addition of 0.5–1 mm  
 387 and 2–2.8 mm unprocessed biochars increased the substrate plant available water compared to  
 388 the control ( $p < 0.05$ ).

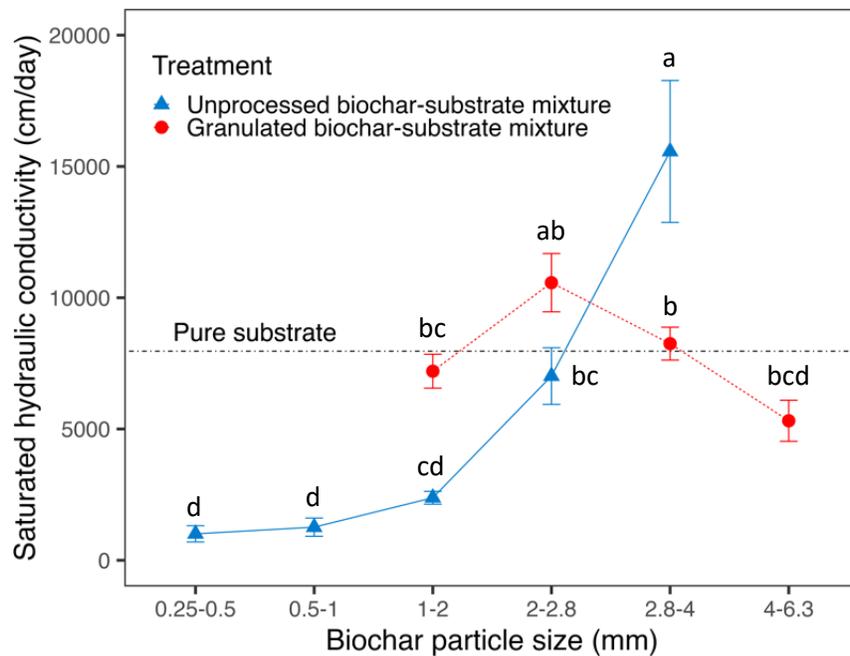


390 **Fig. 5.** Field capacity (a), permanent wilting point (b), and plant available water (c) for the pure  
 391 substrate and the substrates amended with unprocessed and granulated biochars. Blue and red  
 392 labels represent unprocessed and granulated biochars, respectively. Dashed lines indicate the  
 393 pure substrate. Means are plotted  $\pm 1$  SE for each biochar treatment and the pure substrate. Dots  
 394 with different lowercase letters differ significantly at  $p < 0.05$  according to Tukey's HSD test.

### 396 3.3. Hydraulic conductivity

397 Unprocessed and granulated biochars showed strong particle size effects on saturated  
 398 hydraulic conductivity ( $K_{sat}$ ) (Tables 3 and S3). The  $K_{sat}$  of the biochar-substrate mixtures  
 399 increased with the increasing unprocessed biochar particle sizes, with the biochar in the 2.8–4  
 400 mm category showing the highest  $K_{sat}$  (15570 cm/day) (Fig. 6). The addition of granulated  
 401 biochar in the 2–2.8 mm size range resulted in a greater increase in  $K_{sat}$  compared to the 4–  
 402 6.3 mm granulated biochar (Fig. 6). Biochar type and particle sizes showed interactive effects on

403  $K_{sat}$ , indicating that biochar particle size effect differed between unprocessed and granulated  
 404 biochars (Table 3). Overall, biochars in the 1–2 mm particle size range showed a lower  $K_{sat}$  than  
 405 those in the 2–2.8 mm and 2.8–4 mm ranges (Table 3). In addition, the substrates amended with  
 406 the unprocessed biochars in 0.25–0.5 mm, 0.5–1 mm, and 1–2 mm particle size ranges had lower  
 407  $K_{sat}$  than the pure substrate ( $p < 0.05$ ). No difference was observed in substrate  $K_{sat}$  for the pure  
 408 substrate or the substrates amended with the unprocessed biochars and granulated biochars ( $p >$   
 409 0.05) (Table 3).



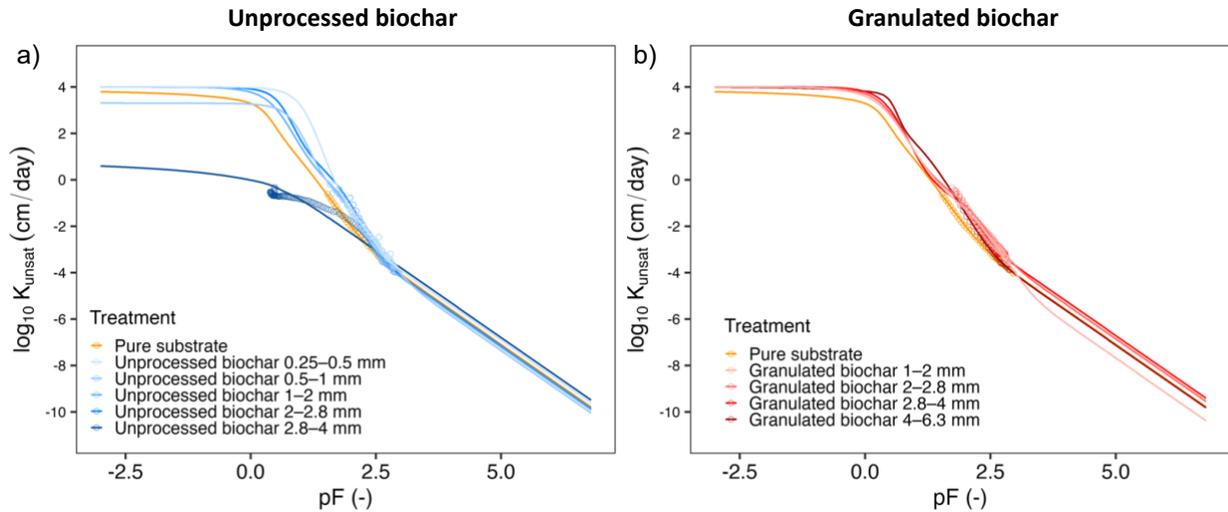
410

411 **Fig. 6.** Saturated hydraulic conductivity ( $K_{sat}$ ) for the pure substrate and the substrates amended  
 412 with unprocessed and granulated biochars in different particle size ranges. Blue and red labels  
 413 represent unprocessed and granulated biochars, respectively. Dashed lines indicate the pure  
 414 substrate. Means are plotted  $\pm 1$  SE for each biochar treatment and the pure substrate. Dots with  
 415 different lowercase letters differ significantly at  $p < 0.05$  according to Tukey's HSD test.

416

417 The addition of biochars slightly increased the substrate unsaturated hydraulic  
 418 conductivity ( $K_{unsat}$ ) for  $pF < 2.5$ , except for the unprocessed biochar in 0.5–1 mm and 2.8–4 mm

419 (Fig. 7). However, the effects were not significant. In addition, no biochar effect on  $K_{\text{unsat}}$  were  
 420 observed for  $pF > 2.5$ .



421  
 422 **Fig. 7.** Unsaturated hydraulic conductivity curves for the pure substrate (a–b) and the substrates  
 423 amended with unprocessed (a) and granulated (b) biochars in different particle size ranges. Open  
 424 symbols represent measurement data obtained from the HYPROP device, while solid lines  
 425 indicate model fits for one representative replicate. Blue and red colors indicate unprocessed and  
 426 granulated biochars, respectively, with the darker colors presenting larger particle size ranges.  
 427 Orange color indicates the pure substrate.

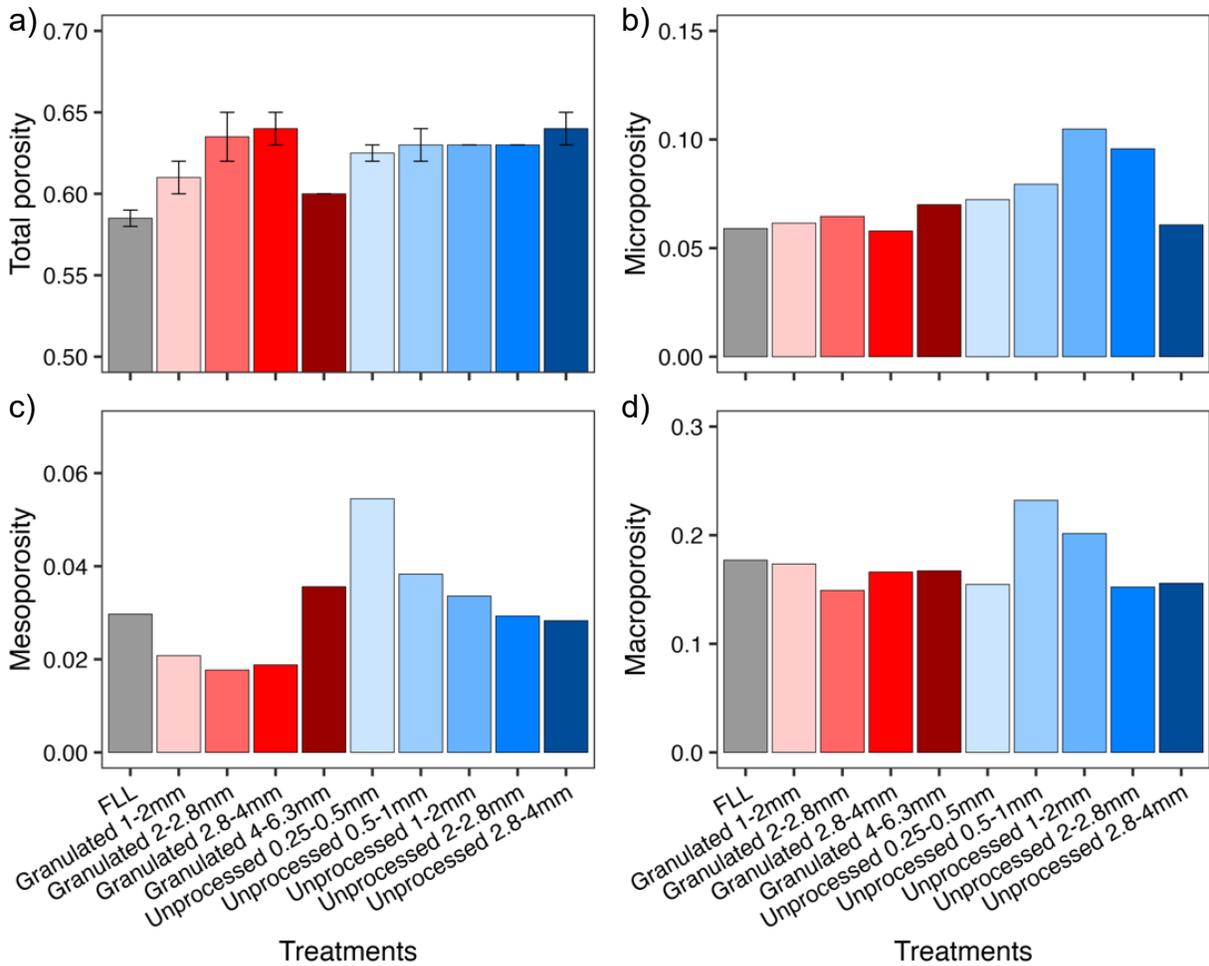
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### 429 3.4. Pore size distribution of the substrate and biochar-substrate mixtures

430 Biochar applications altered the pore size distribution and total porosity of the green roof  
 431 substrates. The addition of unprocessed and granulated biochars increased the total porosity of  
 432 the pure substrate by 8.3% and 7.4%, respectively (Fig. 8a). The amendment of unprocessed and  
 433 granulated biochars increased the substrate microporosity by 40% and 8%, respectively,  
 434 compared to the pure substrate (Fig. 8b). On average, substrates with the unprocessed biochars  
 435 (3.7%) had higher proportion of mesopores than those amended with the granulated biochars  
 436 (2.3%) and the pure substrate (3.0%) (Fig. 8c). The substrate with intermediate-sized and small-  
 437 sized granulated biochars reduced the substrate mesoporosity (Fig. 8c). In contrast, the addition

438 of granulated biochars reduced the substrate macroporosity to 16%, whereas unprocessed  
 439 biochars had a negligible effect on substrate microporosity (Fig. 8d).

440 Substrates amended with unprocessed biochars of small (0.25–0.5 mm) and large (2–  
 441 2.8 mm and 2.8–4 mm) particle sizes exhibited lower macroporosity compared to those with  
 442 intermediate particle sizes (0.5–1 mm and 1–2 mm) (Fig. 8d). In contrast, mesoporosity generally  
 443 increased with decreasing unprocessed biochar particle size (Fig. 8c). No consistent pattern in  
 444 pore size distribution was observed for the substrates amended with granulated biochars.



445  
 446 **Fig. 8.** Pore size distribution and total porosity of the pure substrate and the substrate amended  
 447 with unprocessed and granular biochars. Pore size distribution was derived from HYPROP  
 448 measurements for one representative replicate. Mean total porosity is plotted  $\pm$  1 SE and is  
 449 shown based on duplicate measurements for each biochar treatment and the pure substrate.

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## **4. Discussion**

### **4.1. Biochar post-processing affects biochar surface and pore characteristics**

Our results showed that unprocessed biochars present well-structured pores. In contrast, granulated biochars exhibit non-uniform and fractured surface morphology, along with reduced porosity and lower BET specific surface area. This is likely attributable to the collapse of physical macropore structure during processing (Fig. 1ab) and the presence of binding agents during the granulation process, which likely occluded pores and decreased the overall porosity of the granulated biochar (Liao et al., 2022a). In addition, granulated biochars exhibited more uniform and consistent BET specific surface area and pore structure across different particle sizes compared to unprocessed biochars. Minimal particle size effects were observed for granular biochars, likely due to the uniformity of biochar granulation process, which preserves the biochar's internal structure regardless of particle size (Bowden-Green & Briens, 2016; Briens & Bowden-Green, 2019). In contrast, the BET specific surface area and pore characteristics of unprocessed biochars varied strongly with biochar particle size. As particle size decreased, the BET specific surface area and total pore volume increased, possibly due to the greater exposure of inner pores (Fahmi et al., 2018). These trends are consistent with the previous findings in natural soils (Jaafar et al., 2015; Fahmi et al., 2018). The enhanced BET specific surface area and total pore volume in smaller biochar particles may enhance water and nutrient retention on green roofs (Liao et al., 2022b). Our results also showed that biochar addition increased the total porosity of green roof substrates, particularly the proportion of micropores, due to the large BET specific surface area and pore volume of biochars.

473 Unprocessed biochars had smaller average pore diameters than granulated biochars,  
474 presenting higher microporosity. This can be attributed to the intrinsic microporous structure of  
475 the conifer feedstock (Lu & Zong, 2018). The high concentration of micropores (< 2 nm)  
476 presented in the unprocessed biochars can contribute to high biochar water retention capacity  
477 (Liao et al., 2022a). In contrast, granulated biochars showed highly variable pore size  
478 distributions and relatively low pore volume across micropore and mesopore scales, which may  
479 result from collapsed pore structures and non-uniform surface morphologies introduced during  
480 the granulation process (Bowden-Green & Briens, 2016), as evidenced by the SEM images (Fig.  
481 1ab). The alteration of pore characteristics during biochar granulation has been shown to reduce  
482 biochar water retention capacity compared to unprocessed biochar (Liao et al., 2022a).

483 The FTIR spectra revealed that granulated biochar exhibited stronger hydrophilic  
484 character than unprocessed biochar, as indicated by the distinct C–O stretching band at 1036  
485  $\text{cm}^{-1}$  and the pronounced O–H and C=C peaks (Fan et al., 2022). The distinct FTIR spectra of  
486 granulated biochars may reflect either oxidation during processing or the chemistry of the  
487 binding agent. These oxygen-containing functional groups enhance water affinity (Fan et al.,  
488 2022) and pollutant adsorption capacity (Fahmi et al., 2018; Dai et al., 2021), suggesting that  
489 granulated biochars may improve water retention and contaminant removal performance. The  
490 presence of distinct functional groups, particularly the C–O peak, in the granulated biochar is  
491 likely due to the proprietary organic binding agents used during granulation processes (Hu et al.,  
492 2015; Dos Santos Júnior et al., 2025), although oxidation during processing may also contribute.  
493 In contrast, the relatively weak C–O signal in unprocessed biochar implies reduced surface  
494 polarity and lower hydrophilicity (Fan et al., 2022). This is consistent with previous study

495 indicating the moderate hydrophobicity of wood-derived granulated biochar (Briens & Bowden-  
496 Green, 2019).

497

#### 498 **4.2. Biochar improves substrate hydraulic properties**

499 Consistent with the effects of post-processing on biochar surface and pore characteristics,  
500 the addition of granulated biochar had more uniform effects on substrate hydraulic properties  
501 across particle size ranges compared to unprocessed biochars, consistent with the more  
502 homogeneous structure generated during the granulation process. The application of granulated  
503 biochar increased substrate field capacity, potentially enhancing stormwater retention on green  
504 roofs. This is likely due to the large BET specific surface area, high total pore volume, and  
505 hydrophilicity of the granulated biochar (as evidenced by the presence of oxygen-containing  
506 functional groups). These findings are consistent with the previous study that reported increased  
507 substrate water retention capacity following granular biochar amendment (Liao et al., 2022a).  
508 Likewise, a recent review indicated that biochar amendment improves hydraulic properties  
509 primarily in sandy soils, particularly under laboratory conditions where uniform homogenization  
510 of biochar-soil mixtures is ensured (Rabbi et al., 2021). The pronounced biochar effects on  
511 hydraulic properties observed in our study are likely enhanced by the coarse-textured nature of  
512 the green roof substrates, which are similar to coarse sandy soils. However, granulated biochar-  
513 substrate mixtures exhibited higher permanent wilting points than those amended with  
514 unprocessed biochar, suggesting that a larger proportion of the retained water was tightly bound  
515 within biochar intrapores and therefore less available to plants. Consequently, our results also  
516 showed that granulated biochar amendment had no effect on substrate plant available water. This  
517 finding contrasts with previous studies that reported increased plant available water and extended

518 days to permanent wilting point following unprocessed biochar application (Cao et al., 2014;  
519 Petreje et al., 2025).

520 Our results indicate that the addition of fine biochar particles more effectively improved  
521 substrate field capacity compared to coarse biochar particles, possibly due to the increased BET  
522 specific surface area (Fig. 1) and micropores (Fig. S1) associated with smaller particle sizes, as  
523 well as the clogging of inter-pores by fine biochar particles (Esmaeelnejad et al., 2017). The  
524 amendment of 0.5–1 mm and 2–2.8 mm unprocessed biochars significantly increased the  
525 substrate plant available water, likely due to the greater amount of mesopores (Fig. 8), which are  
526 typically associated with plant available water storage (Edeh & Mašek, 2021). Higher plant  
527 available water can enhance plant performance on green roof substrates (Petreje et al., 2025),  
528 particularly for drought-sensitive species or during prolonged drought periods. In contrast,  
529 although the substrate amended with 0.25–0.5 mm unprocessed biochar exhibited a relatively  
530 high proportion of mesopores, it did not improve plant available water, highlighting the critical  
531 role of biochar intrapores in regulating water retention and release dynamics that influence plant  
532 available water (Edeh & Mašek, 2021). This finding contradicts prior studies that reported higher  
533 soil plant available water with the addition of smaller-sized biochar (Alghamdi et al., 2020;  
534 Werdin et al., 2021), possibly due to differences in the matrix substrate and the specific biochar  
535 particle size ranges used.

536 All biochar-amended substrates were best described by the PDI variant of the bimodal  
537 constrained van Genuchten model using the HYPROP-FIT software, indicating the presence of a  
538 bimodal pore structure. This likely resulted from the combination of intrapores within the  
539 biochar particles and interpores formed between biochar and substrate particles. Forms of the van  
540 Genuchten model (Edeh & Mašek, 2021; Zhang et al., 2021) or the bimodal Durner model

541 (Trifunovic et al., 2018; Zanutel et al., 2024) have most often been fit to characteristic water  
542 release curves of biochar-amended soils. However, the highly porous structure of biochar,  
543 particularly granulated biochar composed of varying particle sizes and binding agents, can  
544 generate a complex pore network comprising intrapores at a range of spatial scales as well as  
545 interpores formed between particles (Zhao et al., 2023). A recent modeling study of soil  
546 hydrological processes across nearly 1,000 soil samples showed that soils often exhibit a bimodal  
547 pore size distribution due to particle aggregation (Shi et al., 2026), suggesting a strong tendency  
548 toward multimodal pore size distributions in biochar-amended substrates. As such, trimodal or  
549 multi-modal models may more accurately capture the pore architecture and hydraulic behavior of  
550 biochar-amended substrate (Zhao et al., 2023), particularly those containing granulated biochars.

551 Our results showed that  $K_{\text{sat}}$  decreased with decreasing unprocessed biochar particle size,  
552 particularly in the 0.25–0.5 mm, 0.5–1 mm, and 1–2 mm size ranges, likely due to the clogging  
553 of substrate interpores by fine biochar particles, which reduces effective pore space and increases  
554 water flow tortuosity (Edeh & Mašek, 2021). This suggests that fine unprocessed biochar  
555 particles may enhance the detention capacity of green roof substrates by reducing water flow  
556 rates and delaying peak discharge during rainfall events, particularly under saturated conditions.  
557 However, the reduced infiltration rates associated with the amendment of fine unprocessed  
558 biochar particles, especially those in the 0.25–0.5 mm and 0.5–1 mm ranges, may increase the  
559 risk of waterlogging during and after intense precipitation events, necessitating additional  
560 stormwater control measures (Werdin et al., 2021). The waterlogging associated with small  
561 biochar particles may also enhance methane emissions from green roofs (Halim et al., 2022). In  
562 contrast, the largest size class of unprocessed biochar (2.8–4 mm) most effectively increased  
563  $K_{\text{sat}}$ , likely due to the increased interpores between particles formed by the elongated-shaped

564 unprocessed biochar (Liao & Thomas, 2019). However, a reduction in  $K_{\text{unsat}}$  was observed with  
565 the addition of large unprocessed biochars, suggesting reduced water infiltration rates under  
566 unsaturated conditions, potentially due to increased water retention associated with their  
567 hydrophilic surfaces and higher intraporosity (Edeh & Mašek, 2021). In general, intermediate  
568 biochar particle sizes can provide a balanced infiltration rate, thereby improving green roof  
569 stormwater retention and detention capacity.

570

### 571 **4.3. Importance and future directions**

572 This study is the first to investigate the effects of biochar granulation on the hydraulic  
573 properties of green roof substrates, providing critical insights for optimizing green roof substrate  
574 design to improve stormwater management. The stable and uniform surface and pore  
575 characteristics and hydrological properties of granulated biochar, combined with the strong  
576 erosion resistance (Liao et al., 2022), highlight its high potential to improve the  
577 multifunctionality of green roofs, including improved stormwater management, water quality  
578 (Liao et al., 2022b), and plant performance (Liao et al., 2022a). Although the application of  
579 unprocessed biochars in 0.5–1 mm and 2–2.8 mm particle size ranges increases plant available  
580 water, intermediate-sized biochar (i.e., 2–2.8 mm) is recommended for green roofs to minimize  
581 human exposure to fine dusts and to reduce erosion losses and risks of periodic waterlogging  
582 (Tang et al., 2023). Granulation of fine biochar particles offers an effective solution for utilizing  
583 biochar powders generated during production, which are often unsuitable for field applications  
584 due to their high erodibility while enhancing the profitability of biochar production by enabling  
585 the industrial-scale use of low-value biochar powders. In addition, our study provides essential  
586 parameters for hydrological modeling, enabling a better mechanistic understanding of biochar

587 effects on hydrological performance and stormwater management under different conditions,  
588 including future climate scenarios. When combined with geospatial analysis and stormwater  
589 modeling tools, such as Storm Water Management Model (SWMM), these findings support the  
590 evaluation and optimization of stormwater management strategies across large spatial and  
591 temporal scales.

592         It should be emphasized that the present study presents a short-term experimental data on  
593 freshly prepared biochar-substrate mixtures. The hydraulic properties of green roof substrates are  
594 likely to evolve over time due to changes in substrate physicochemical characteristics, including  
595 shifts in particle and pore size distributions, organic matter accumulation or depletion, and plant-  
596 biochar-soil interactions, such as aggregation and root exudate production (De-Ville et al., 2017;  
597 Bouzouidja et al., 2018; De-Ville et al., 2018; Yang & Davidson, 2021). Oxidation and “ageing”  
598 of biochar are also likely to alter hydrological properties (Wang et al., 2020). Future research  
599 should investigate the long-term hydrological performance of biochar-amended green roof  
600 substrates, accounting for time-dependent changes in substrate properties. In addition, future  
601 studies should evaluate the hydrological performance of granulated and unprocessed biochars  
602 under field conditions. Biochar application rate also plays a critical role in influencing the  
603 hydraulic properties of green roof substrates (Werdin et al., 2021). In this study, biochars were  
604 applied at a rate of 4.5% w/w (equivalent to ~25 t/ha), targeting optimal plant performance (Gale  
605 & Thomas, 2019). Future research should explore the optimal application rates that balance  
606 enhanced hydrological performance with improved vegetation growth and discharge water  
607 quality. Additional studies on alternative binding agents are also needed to optimize the  
608 properties of granulated biochar (Thomas, 2021), with the goal of improving hydrological  
609 performance and enhancing plant available water.

610

## 611 **5. Conclusions**

612 Our study shows that biochar granulation enhances the uniformity of surface and pore  
613 characteristics, resulting in more consistent effects on hydraulic properties across particle size  
614 ranges. In contrast, the BET specific surface area, total pore volume, and microporosity of  
615 unprocessed biochars increased with decreasing particle sizes. The addition of granulated biochar  
616 increased substrate field capacity, potentially enhancing stormwater retention on green roofs.  
617 However, a substantial portion of the retained water was tightly bound within biochar intrapores  
618 and thus not readily available to plants, though it should also be noted that the permanent wilting  
619 point metric has important limitations (e.g., [Garg et al. \(2020\)](#)). In contrast, the amendment of  
620 unprocessed biochars in the 0.5–1 mm and 2–2.8 mm size ranges enhanced plant available water,  
621 which is crucial for supporting vegetation performance on green roofs, particularly under  
622 prolonged drought conditions. Our results also show that the addition of fine unprocessed  
623 biochars reduced saturated hydraulic conductivity, indicating enhanced detention capacity  
624 through decreased infiltration rates and delayed peak flows. However, the use of fine biochar  
625 particles (e.g., < 1 mm) may increase the risks of human exposure to dust, erosion losses, and  
626 periodic waterlogging. Overall, granulated biochar and intermediate-sized unprocessed biochar  
627 may provide balanced stormwater retention and detention capacities, as well as enhanced plant  
628 available water on green roofs. Our study provides critical insights and key modeling parameters  
629 on the effects of biochar post-processing, especially granulation and particle size modification,  
630 on hydraulic properties and hydrological processes of green roof substrates. Granulated biochar,  
631 with demonstrated benefits for water quality improvement, enhanced plant performance, and

632 erosion control, holds strong potential for urban green infrastructure applications, thereby  
633 contributing to long-term urban sustainability.

634

## 635 **Declarations**

### 636 **Competing interests**

637 The authors declare no competing interests.

638

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647

### 648 **Author contributions**

649 Conceptualization: W.L., J.D. and S.C.T. Methodology: W.L., J.D. and S.C.T. Investigation:  
650 W.L. Formal analysis: W.L. Visualization: W.L. Writing – Original Draft: W.L. Funding  
651 acquisition: J.D. and S.C.T. Writing – Review & Editing: all authors.

652

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882 **Tables**

883

884 **Table 1.** Two-way ANOVA outputs for the surface area and pore characteristics of factorial treatment (granulated and unprocessed  
 885 biochars in 1-2 mm, 2-2.8 mm, and 2.8-4 mm particle size ranges). PM and PS indicate biochar processing method (granulated vs.  
 886 unprocessed biochars) and particle size, respectively. Asterisks indicate significance of two-way ANOVA: (\*),  $P < 0.1$ ; \*,  $P < 0.05$ ;  
 887 \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$ .

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Source of variation	df	Sum of Squares	Mean Square	F	P
<b>BET specific surface area</b>					
PM	1	15496	15496	26.30	<b>0.002 **</b>
PS	2	8123	4061	6.89	<b>0.028 *</b>
PM × PS	2	16933	8467	14.37	<b>0.005 **</b>
Error	6	3535	589		
<b>Total pore volume</b>					
PM	1	0.0043	0.0043	15.92	<b>0.007 **</b>
PS	2	0.0022	0.0011	3.94	0.081 (*)
PM × PS	2	0.0072	0.0036	13.27	<b>0.006 **</b>
Error	6	0.0016	0.0003		
<b>Average pore diameter</b>					
PM	1	8.0803	8.0803	32.35	<b>0.001 **</b>
PS	2	2.2648	1.1324	4.53	0.063 (*)
PM × PS	2	7.4281	3.7141	14.87	<b>0.005 **</b>
Error	6	1.4987	0.2498		
<b>Micropore volume</b>					
PM	1	0.00593	0.00593	33.40	<b>0.001 **</b>
PS	2	0.00287	0.00144	8.09	<b>0.020 *</b>
PM × PS	2	0.00607	0.00303	17.07	<b>0.003 **</b>
Error	6	0.00107	0.00018		
<b>Mesopore volume</b>					

PM	1	0.00012	0.00012	3.52	0.110
PS	2	0.00008	0.00004	1.18	0.370
PM × PS	2	0.00005	0.00003	0.75	0.513
Error	6	0.00021	0.00004		
<b>Percentage of micropore</b>					
PM	1	1987.8	1987.8	191.25	< <b>0.001</b> ***
PS	2	409.9	205.0	19.72	<b>0.002</b> **
PM × PS	2	496.6	248.3	23.89	<b>0.001</b> **
Error	6	62.4	10.4		
<b>Percentage of mesopore</b>					
PM	1	1987.8	1987.8	191.25	< <b>0.001</b> ***
PS	2	409.9	205.0	19.72	<b>0.002</b> **
PM × PS	2	496.6	248.3	23.89	<b>0.001</b> **
Error	6	62.4	10.4		

890 **Table 2.** One-way ANOVA outputs for the biochar particle size effects on surface area and pore characteristics for unprocessed and  
 891 granulated biochars. Asterisks indicate significance of two-way ANOVA: (\*),  $P < 0.1$ ; \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.001$ .  
 892

Source of variation	Biochar particle size			
	Unprocessed biochar		Granulated biochar	
	F(4, 5)	<i>P</i>	F(3, 4)	<i>P</i>
BET specific surface area	36.703	< <b>0.001</b> ***	1.104	0.4451
Total pore volume	42.785	< <b>0.001</b> ***	1.074	0.4545
Average pore diameter	14.476	<b>0.006</b> **	2.448	0.2036
Micropore volume	36.963	< <b>0.001</b> ***	1.096	0.4476
Mesopore volume	0.873	0.5394	1.044	0.4643
Percentage of micropore	21.575	<b>0.002</b> **	2.055	0.2489
Percentage of mesopore	21.575	<b>0.002</b> **	2.055	0.2489

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**Table 3.** Two-way ANOVA outputs for the hydraulic properties of factorial treatment (granulated and unprocessed biochars in 1-2 mm, 2-2.8 mm, and 2.8-4 mm particle size ranges). PM and PS indicate biochar processing method (granulated vs. unprocessed biochars) and particle size, respectively. Asterisks indicate significance of two-way ANOVA: (\*),  $P < 0.1$ ; (\*),  $P < 0.05$ ; (\*\*),  $P < 0.01$ ; (\*\*\*),  $P < 0.001$ .

Source of variation	df	Sum of Squares	Mean Square	F	P
<b>Field capacity</b>					
PM	1	14.04	14.04	53.69	<0.001 ***
PS	2	4.35	2.17	8.31	0.019 *
PM × PS	2	1.26	0.63	2.42	0.170
Error	6	1.57	0.26		
<b>Permanent wilting point</b>					
PM	1	16.52	16.52	7.98	0.030 *
PS	2	9.18	4.59	2.22	0.190
PM × PS	2	2.44	1.22	0.59	0.584
Error	6	12.43	2.07		
<b>Plant available water</b>					
PM	1	0.10	0.10	0.08	0.786
PS	2	25.45	12.72	10.13	0.012 *
PM × PS	2	1.13	0.57	0.45	0.657
Error	6	7.53	1.26		
<b>Saturated hydraulic conductivity</b>					
PM	1	558977	558977	0.11	0.751
PS	2	152832303	76416151	14.46	< 0.001 ***
PM × PS	2	133634277	66817138	12.64	< 0.01 **
Error	12	63422172	5285181		