

14 **Abstract**

15 *Bactrocera dorsalis* (Hendel) is the most economically important invasive fruit fly in sub-Saharan
16 Africa, yet long-term empirical evidence linking its seasonal dynamics to environmental drivers
17 across altitudinal gradients remains limited. We analysed 279 site-month observations of male trap
18 catches (flies per trap per day, FTD) collected between October 2004 and February 2012 at six
19 sites spanning 526–1,650 m above sea level along the Uluguru Mountain transect in Morogoro,
20 Tanzania. Climatic variables (temperature, rainfall, relative humidity) were obtained from ERA5
21 reanalysis, and host fruit phenology was characterised for key commercial species. Abundance
22 declined sharply with altitude, with mean FTD decreasing from 87.1 at the lowland site (SUA, 526
23 m) to 0.07 at the highest site (Nyandira, 1,650 m), where 55% of months recorded zero catches.
24 Seasonal peaks occurred during the warm–wet period (November–April), but both their magnitude
25 and duration declined with altitude, with the high season shortening from seven months at SUA to
26 three months at Nyandira. Seasonal patterns also became less predictable at higher elevations, with
27 the coefficient of variation of peak abundance increasing from 0.54 to 1.38. Relative humidity
28 showed the strongest association with FTD (Spearman $\rho = 0.202$, $p = 0.0007$), while temperature
29 showed no significant overall relationship. Host fruit availability further shaped population
30 dynamics, with ripening of jew plum, mango, and soursop associated with increased abundance,
31 whereas citrus and loquat had minimal influence. Overall, *B. dorsalis* abundance along the gradient
32 is primarily structured by altitude and host availability, with climate acting as a secondary, context-
33 dependent influence. These findings provide a clearer ecological basis for predicting pest pressure
34 and improving management strategies across heterogeneous landscapes in East Africa.

35 **Keywords:** *Bactrocera dorsalis*, altitudinal gradient, seasonal dynamics, host phenology,
36 population abundance, relative humidity, climatic drivers, tropical agroecosystems

37

38 **Introduction**

39 Tephritid fruit flies (Diptera: Tephritidae) rank among the most economically damaging insect
40 pests of horticultural crops worldwide, inflicting severe pre- and post-harvest losses and
41 constraining international trade in fresh produce [1,2]. Within this family, the oriental fruit fly,
42 *Bactrocera dorsalis* (Hendel), stands out as a destructive invasive species whose broad host range
43 encompasses more than 400 fruit and vegetable species [3–5]. Originally native to continental
44 South and South-East Asia, *B. dorsalis* was first detected on the African continent along the
45 Kenyan coast in 2003 [6]. The African populations were initially described as a separate species,
46 *Bactrocera invadens* Drew, Tsuruta & White [7], but integrative taxonomic analyses combining
47 morphological, molecular, and cytogenetic evidence subsequently demonstrated conspecificity,
48 leading to formal synonymisation under *B. dorsalis* [8,9].

49 Following its arrival in East Africa, *B. dorsalis* spread rapidly, and by 2020 it had been recorded
50 in more than 35 sub-Saharan countries including Indian Ocean Island nations [10]. This invasion
51 success has been attributed to a combination of favourable tropical climates, the availability of
52 diverse cultivated and wild host plants, and the species' competitive superiority over indigenous
53 *Ceratitis* species [11,12]. In areas where *B. dorsalis* has established, crop losses in mango, citrus,
54 guava, and other commercially important fruits routinely range from 30% to 80% if populations
55 are left unmanaged [13–15], causing significant economic losses for smallholder farmers [16,17].

56 Across Africa, the spatial and temporal distribution of *B. dorsalis* is highly heterogeneous and
57 strongly seasonal. Spatially, the species attains its highest densities in low- to mid-altitude
58 agroecological zones characterised by warm temperatures and year-round host availability [1,18].

59 Along altitudinal gradients in East Africa, population densities decline with increasing elevation,
60 reflecting the combined effects of cooler temperatures, reduced host diversity, and changes in
61 landscape composition [19–21]. Temporally, populations peak during warm, humid periods that
62 coincide with the main host fruiting seasons, declining during cooler or drier months when
63 developmental rates slow and host resources become scarce [13,22].

64 Predictive distribution and ecological niche-modelling studies indicate that large areas of Africa
65 and other tropical and subtropical regions remain climatically suitable for the establishment of *B.*
66 *dorsalis* [23], and phenology-based models suggest that warming temperatures may promote
67 seasonal expansion and year-round persistence, increasing invasion risk in temperate fruit-growing
68 regions under future climate scenarios [24,25]. Understanding how environmental drivers interact
69 to shape population dynamics across heterogeneous landscapes is therefore essential for
70 developing climate-sensitive surveillance and evidence-based integrated pest management (IPM)
71 strategies.

72 Previous studies on the ecology of *B. dorsalis* in Tanzania documented its host utilisation,
73 seasonality, competitive interactions with native fruit flies, and progressive dominance along an
74 altitudinal transect in Morogoro [13,19,20,26]. However, these investigations were generally
75 limited in temporal scope, typically spanning one to three years, and did not examine long-term
76 population dynamics in relation to host fruit phenology, landscape-level host availability, and the
77 full altitudinal gradient simultaneously. Long-term datasets are critical for distinguishing
78 persistent ecological signals from short-term variability and for strengthening the evidence base
79 on which surveillance and management decisions rest [1,10].

80 The present study addresses this gap by analysing *B. dorsalis* population dynamics over a seven-
81 year period (October 2004 to February 2012) across six trapping sites spanning 526 to 1,650 m

82 above sea level along the eastern slopes of the Uluguru Mountains, Morogoro Region, Tanzania.
83 We hypothesised that: (i) abundance declines with increasing altitude, reflecting thermal
84 constraints and reduced host availability; (ii) seasonal population peaks are stronger and longer at
85 lower elevations; (iii) moisture-related climatic variables, particularly relative humidity, are more
86 strongly associated with abundance than temperature alone; and (iv) the timing and availability of
87 preferred host fruits contribute substantially to local population increases. By integrating long-
88 term trapping data with ERA5 climate reanalysis products, host fruit phenology records, and both
89 parametric and machine learning analytical approaches, this study aims to provide a
90 comprehensive ecological characterisation of *B. dorsalis* population dynamics along a tropical
91 altitudinal gradient and to inform targeted, landscape-specific management strategies.

92 **Methodology**

93 **Study area and agroecological zones**

94 Monitoring of *B. dorsalis* was carried out in Morogoro Region, eastern–central Tanzania, along
95 an altitudinal gradient on the eastern slopes of the Uluguru Mountains. Sites were selected to
96 represent four agroecological zones following the classification systems described for the Eastern
97 Arc Mountains [27,28]. This zonation reflects distinct changes in cropping systems, host plant
98 composition, and climatic regimes along the transect.

99 **Fig 1.** 3D elevation surface around the six monitoring localities (SRTM DEM), highlighting the
100 lowland–highland gradient.

101 A preliminary analysis revealed a strong negative correlation between altitude and temperature
102 (Spearman’s $\rho = -0.79$), consistent with the well-documented adiabatic lapse rate in tropical

103 mountains. To reduce collinearity in subsequent analyses, agroecological zone was retained as a
104 categorical landscape variable while preserving the ecological meaning of the altitudinal gradient
105 [29].

106 The lowland agro-mosaic (below ~600 m a.s.l.) encompassed two sites: SUA (526 m) and Mlali
107 (581 m), characterised by warm agricultural landscapes with diverse tropical fruit production,
108 including citrus (*Citrus sinensis* (L.) Osbeck, *C. limon* (L.) Osbeck, *C. reticulata* Blanco), mango
109 (*Mangifera indica* L.), guava (*Psidium guajava* L.), jew plum (*Spondias cytherea* Sonn.), and
110 soursop (*Annona muricata* L.). The mid-elevation transition zone contained one site, Kibundi (843
111 m), in a mixed smallholder landscape dominated by banana (*Musa* spp.), avocado (*Persea*
112 *americana* Mill.), maize, and beans. The sub-montane farming zone included two sites: Langali
113 (1,105 m) and Visada (1,302 m), where staple crops predominated and tropical fruit trees were
114 relatively sparse. Finally, the montane agroforestry zone was represented by Nyandira (1,650 m),
115 a cooler landscape dominated by temperate fruit crops such as apple (*Malus domestica* Borkh.),
116 peach (*Prunus persica* (L.) Batsch), and plum (*Prunus salicina* Lindl.), often intercropped with
117 coffee (*Coffea canephora* Pierre ex A. Froehner).

118

119 **Sampling design and Trap deployment**

120 Six monitoring sites, spanning approximately 500 to 1,650 m a.s.l., were established along the
121 gradient. Monitoring commenced in October 2004 at the two altitudinal extremes, SUA (lowland)
122 and Nyandira (montane), with routine monitoring at SUA beginning in October 2005 following an
123 initial pilot phase. In October 2008, the network was expanded to include Mlali, Kibundi, Langali,
124 and Visada, and all six sites were monitored continuously until February 2012.

125 Adult male *B. dorsalis* were monitored using modified McPhail traps baited with methyl eugenol
126 (ME; Scentry Cie, Billings, MT, USA), a highly effective male-specific parapheromone widely
127 used in tephritid surveillance programmes [31,32]. Methyl eugenol attracts *B. dorsalis* from the
128 surrounding landscape, thereby enabling sampling at a broader spatial scale. Traps were fitted with
129 dichlorvos (DDVP) strips as a killing agent and suspended approximately 2 m above ground level
130 within the host tree canopy. Across all sites, inter-trap spacing was maintained at a minimum of
131 1,000 m to ensure spatial independence of catches.

132 At SUA, traps were initially placed in mango (cv. Dodo), citrus (cv. Sweet Seedling), and guava
133 trees, later expanded to five traps with additional placements in mango (cv. Sindano Nyeusi) and
134 citrus (cv. Tahiti lime). At Nyandira, traps were installed in peach and plum trees. At the four
135 intermediate sites, three replicate traps per site were placed predominantly in mango trees. After
136 each sampling event, traps were randomly repositioned within the sampling area to minimise
137 location bias. Lure plugs and DDVP strips were replaced at approximately monthly intervals to
138 maintain trapping efficiency.

139 Sampling procedures followed established protocols for tephritid monitoring in East Africa [13,19].
140 Collected specimens were transported to the SUA Entomology Laboratory, where they were sorted
141 and identified using standard morphological keys [2,34] and confirmed with the integrative
142 diagnostic tools of Virgilio et al. [33]. Specimens previously identified as *Bactrocera invadens*
143 were treated as *B. dorsalis* following formal synonymisation based on integrative taxonomic
144 evidence [8,9].

145 **Fruit phenology data**

146 Phenological data for major host fruit species were obtained from the systematic weekly
147 observations of Shechambo [30], conducted at the SUA Horticultural Unit. Representative host
148 species selected for analysis included mango, citrus, guava, soursop, loquat (*Eriobotrya japonica*
149 (Thunb.) Lindl.), jew plum, and strawberry guava (*Psidium cattleianum* Sabine). These species
150 were chosen on the basis of their documented high incidence and infestation rates for *B. dorsalis*
151 in the study area [19,20,26].

152 Phenological stages recorded included flowering, fruit set, immature fruit, mature fruit, and ripe
153 fruit. For subsequent analyses, periods of host fruit availability (i.e., presence of mature and ripe
154 fruit) were coded as binary indicators for each host species and calendar month (S1 Fig).

155 **Weather data collection**

156 Monthly climatic variables; mean air temperature (°C), total rainfall (mm), and mean relative
157 humidity (%) were compiled for each monitoring site. At SUA, primary data were obtained from
158 the Tanzania Meteorological Authority (TMA) and supplemented with on-site temperature records
159 from iButton data loggers. At Nyandira, temperature was recorded with iButton loggers, while
160 rainfall data were derived from gridded climate products.

161 To ensure temporal continuity across all sites and fill observational gaps, additional climatic data
162 were extracted from the ERA5 global reanalysis [36], accessed via Google Earth Engine [37]. Site-
163 level baseline climate was further characterised using bioclimatic variables (bio01–bio19) from
164 WorldClim version 2.1 at 30-arc-second resolution [35]. ERA5 and WorldClim data supplemented,
165 but did not replace, direct observations where these were available.

166 For analytical consistency, daily records were aggregated into monthly means (temperature,
167 relative humidity) and monthly totals (rainfall). These three climatic variables were selected on
168 the basis of their established influence on the development, survival, reproduction, and dispersal
169 of *B. dorsalis* [14,38].

170 **Abundance index and temporal aggregation**

171 Fruit fly abundance was expressed as flies per trap per day (FTD), the standard index
172 recommended for tephritid population monitoring [31]. For each inspection interval i , FTD was
173 calculated as:

$$174 \text{FTD}_i = C_i / (T_i \times D_i)$$

175 where C_i is the total number of *B. dorsalis* captured, T_i is the number of traps deployed, and D_i is
176 the number of days since the previous inspection. Monthly mean FTD for a given calendar month
177 m was then calculated as a trap-day weighted mean, i.e., the total flies attributed to month m
178 divided by the total trap-days in that month. When inspection intervals spanned two calendar
179 months, catches were allocated proportionally based on the number of days in each month,
180 assuming a constant daily catch rate. Monthly uncertainty was summarised as the weighted
181 standard error of inspection-interval FTD values, with asymmetric clipping at zero to prevent
182 negative lower bounds.

183 **Data analysis**

184 Fruit fly abundance was expressed as flies per trap per day (FTD) and log-transformed using
185 $\log_{10}(1 + x)$ prior to analysis [39]. Seasonal variation was assessed using the Kruskal–Wallis test
186 [40]. Associations between FTD and climatic variables were quantified using Spearman’s rank

187 correlation [41], chosen for its robustness to the skewed, non-linear relationships characteristic of
188 count data.

189 To model climate–abundance relationships while accounting for overdispersion, generalised linear
190 models (GLMs) with a negative binomial error distribution and log link function were fitted
191 [42,43]. Two models were compared: a climate-only model and a full model incorporating locality
192 as a proxy for agroecological zone. Within-site temporal correlation was addressed using
193 generalised estimating equations (GEE) with an exchangeable correlation structure [44]. Model
194 effects were expressed as incidence rate ratios (IRR) with 95% Wald confidence intervals, and
195 model selection was guided by Akaike’s Information Criterion.

196 To capture non-linear relationships and interactions among predictors, three ensemble machine
197 learning algorithms were applied: LightGBM with Tweedie loss ($p = 1.9$) [45], Random Forest
198 [46], and Gradient Boosting [47]. The Tweedie loss was selected because it accommodates the
199 semi-continuous, zero-inflated distribution of FTD. Model performance was evaluated using year-
200 blocked GroupKFold cross-validation to prevent temporal leakage [47]. Predictor importance was
201 quantified using SHapley Additive exPlanations (SHAP) [48], which assigns each feature an
202 importance value grounded in cooperative game theory. From the SHAP output, mean absolute
203 SHAP values provided global predictor rankings, dependence plots revealed non-linear predictor–
204 response relationships, and interaction analyses examined the altitude–mango ripening interaction.
205 A predictor evidence summary integrated SHAP importance with univariate statistical tests
206 (Spearman’s ρ for continuous, Mann–Whitney U for binary, and Kruskal–Wallis H for categorical
207 predictors), with p -values corrected using the Benjamini–Hochberg false discovery rate procedure
208 [49].

209 All analyses were performed in Python 3.13.5 using pandas [50], NumPy [51], scikit-learn [47],
210 LightGBM [45], SHAP [48], statsmodels [52], and SciPy [53]. Statistical significance was
211 assessed at $\alpha = 0.05$ throughout.

212 **Results**

213 **Temporal dynamics and altitudinal gradient in abundance**

214 Monthly FTD varied markedly across the six sites, reflecting a pronounced altitudinal gradient in
215 *B. dorsalis* abundance (Fig 2). At the lowland SUA site (526 m a.s.l.), FTD fluctuated between
216 approximately 10 and 250 flies trap⁻¹ day⁻¹ throughout the study period, with recurrent peaks
217 during the warm, wet months (November–April) and troughs during the cool, dry season (May–
218 October). The adjacent lowland site of Mlali (581 m) showed a similar seasonal pattern but at
219 lower absolute densities (mean FTD = 38.9). At the mid-elevation site of Kibundi (843 m), FTD
220 was further reduced (mean = 22.3), and seasonal peaks rarely exceeded 100 flies trap⁻¹ day⁻¹. At
221 Langali (1,105 m), representing the sub-montane transition zone, mean FTD declined to 2.2, with
222 only sporadic increases above 5 flies trap⁻¹ day⁻¹. At the two highest sites, Visada (1,302 m) and
223 Nyandira (1,650 m), abundance was extremely low (mean FTD = 0.19 and 0.07, respectively),
224 with Nyandira recording zero catches in 55% of all months monitored (Fig 1).

225 **Fig 2.** Monthly temporal dynamics of *B. dorsalis* FTD at six sites along the Mount Uluguru
226 transect, Morogoro Region, Tanzania, 2004–2012. Sites are arranged from lowest (SUA, 526 m,
227 top) to highest elevation (Nyandira, 1,650 m, bottom). Points show monthly mean FTD \pm SEM.

228 **Seasonal phenology across the gradient**

229 Seasonal boxplots of monthly FTD at each site (Fig 3) showed that the lowland SUA site exhibited
230 the most pronounced and statistically significant monthly variation (Kruskal–Wallis, $p < 0.001$),
231 with the highest median FTD in January (114.9) and the lowest in September (18.7). At the
232 montane Nyandira site, monthly variation was also significant ($p = 0.037$), driven by elevated FTD
233 in January–March and near-zero values from May to November. In contrast, at the intermediate
234 sites of Mlali, Kibundi, and Langali, Kruskal–Wallis tests were not significant ($p = 0.114, 0.085,$
235 and 0.195 , respectively), indicating weaker seasonal structure relative to interannual variability.
236 Similarly, no significant monthly variation was detected at Visada ($p = 0.117$).

237 **Fig 3.** Seasonal phenology of *B. dorsalis* across the altitudinal gradient. (A) Monthly FTD boxplots
238 by site with Kruskal–Wallis test p-values.

239 Seasonal predictability metrics (Fig 4) described three aspects of how the population changes
240 across the gradient. First, seasonal amplitude (the difference between the highest and lowest
241 monthly FTD) decreased sharply with altitude, from 96.2 at SUA to 0.2 at Nyandira (Fig 4A).
242 Second, the coefficient of variation (CV) of peak-month FTD increased with altitude, from 0.54
243 at SUA to 1.38 at Nyandira (Fig 4B), showing that seasonal peaks became less predictable at higher
244 elevations. Third, the duration of the high season (months when FTD was above the site’s annual
245 average) ranged from seven months at SUA to only three months at Nyandira (Fig 4C).

246 **Fig 4.** Seasonal predictability metrics: (A) amplitude (peak–trough FTD) versus altitude; (B)
247 coefficient of variation of peak-month FTD; (C) duration of the high season at each site.

248 **Seasonal effects from negative binomial GLMs**

249 The negative binomial models (Fig 5) confirmed the seasonal pattern observed above. At SUA
250 (526 m), FTD was significantly lower than in January from April to December, with the largest
251 reductions in September (about -80%) and October (-70%). At Mlali and Kibundi, FTD decreased
252 by 60–90% from April to November. At the three highest sites, FTD was also lower than in January
253 in most months, although overall levels were already very low.

254 **Fig 5.** Seasonal effects on *B. dorsalis* FTD from negative binomial GLMs. (A) Percentage change
255 in FTD for each month relative to January (reference) at all six sites. Green bars indicate significant
256 departures ($p < 0.05$); beige bars are non-significant.

257 Annotated seasonal bar charts (Fig 6) showed that population peaks occurred consistently during
258 the early warm–wet season (January at most sites), with minor secondary peaks at Langali and
259 Visada in September. The contrast between peak and low periods increased sharply with altitude,
260 indicating that populations at higher elevations are concentrated into a much shorter seasonal
261 window.

262 **Fig 6.** Monthly mean FTD (\pm SEM) at each site with peak and trough months annotated. Red
263 shading indicates months above the site-specific annual mean.

264 **Effects of climatic variables on abundance**

265 Bivariate relationships between FTD and climate variables (Fig 7) showed that relative humidity
266 had the strongest positive association with FTD (Spearman $\rho = 0.202$, $p = 0.0007$), followed by
267 rainfall ($\rho = 0.119$, $p = 0.046$). Temperature showed no significant overall relationship ($\rho = -0.084$,
268 $p = 0.162$).

269 The correlation matrices (Fig 8) showed that rainfall and relative humidity were strongly correlated
270 with each other ($\rho = 0.75$, $p < 0.001$), indicating that they represent similar seasonal moisture
271 conditions.

272 **Fig 7.** Effects of climatic variables on *B. dorsalis* FTD: Bivariate scatter plots of FTD against
273 temperature, rainfall, and relative humidity with LOESS trend lines and Spearman ρ values. Points
274 are coloured by site.

275 **Fig 8.** Effects of climatic variables on *B. dorsalis* FTD: Multivariate Pearson (left) and Spearman
276 (right) correlation matrices across all sites combined.

277 Site-specific correlations (S2 Fig) showed that these relationships were strongest at the lowland
278 sites. The association between FTD and relative humidity was highest at SUA ($\rho = 0.55$, $p < 0.001$),
279 Mlali ($\rho = 0.50$, $p < 0.01$), and Kibundi ($\rho = 0.55$, $p < 0.001$). Rainfall also showed significant
280 positive relationships with FTD at these sites (SUA: $\rho = 0.47$, $p < 0.001$; Mlali: $\rho = 0.42$, $p < 0.01$;
281 Kibundi: $\rho = 0.50$, $p < 0.01$). In contrast, no significant relationships were observed at the higher-
282 elevation sites (Langali, Visada, Nyandira), where overall abundance was low.

283 **Host fruit phenology and abundance**

284 The SHAP analysis of host fruit ripening (Fig 9) showed that not all host fruits contributed equally
285 to *B. dorsalis* abundance. Ripening of jew plum (*Spondias cytherea*) had the strongest positive
286 effect on FTD (SHAP difference = +1.221). Soursop (*Annona muricata*) and mango (*Mangifera*
287 *indica*) also increased FTD, but to a lesser extent (+0.275 and +0.159, respectively). In contrast,
288 citrus and loquat had little to no effect, with slightly negative contributions (-0.022 and -0.055),
289 indicating that their presence did not meaningfully increase local populations.

290 **Fig 9.** Host phenology effects on *B. dorsalis* abundance. SHAP difference (ripe minus not-ripe)
291 for five host fruits, indicating the contribution of each host's ripening to predicted FTD.

292 The altitude–mango interaction (Fig 10) showed that the effect of altitude on FTD depended on
293 host availability. At lower elevations, where mango was present and ripening between January and
294 March, altitude contributed positively to FTD (mean SHAP = +3.07 at SUA and +2.17 at Mlali).
295 At mid-elevation (Langali), the effect was close to zero (−0.22), while at higher elevations (Visada
296 and Nyandira), where mango was absent, altitude had a strong negative effect (−1.92 and −8.43,
297 respectively).

298 **Fig 10.** Host phenology effects on *B. dorsalis* abundance. SHAP value for altitude stratified by
299 mango ripening status, showing the altitude–mango interaction.

300 Overall, the negative effect of altitude became stronger above about 1,100 m, especially where key
301 host fruits such as mango were no longer present. This pattern was consistent across seasons,
302 although small differences were observed between months when mango was ripe and when it was
303 not. The phenological calendar (S1 Fig) showed that mango ripening (December–May) at the
304 lower sites coincided with the main period of *B. dorsalis* activity.

305 **Key drivers of *B. dorsalis* abundance**

306 Across all analyses, including the machine learning model used to assess predictor importance, a
307 consistent pattern emerged in the factors shaping *B. dorsalis* abundance along the altitudinal
308 gradient (Fig 11). Altitude was the primary factor, with abundance declining sharply at higher
309 elevations. This decline was accompanied by weaker seasonality, reduced predictability, and
310 shorter periods of population activity.

311 **Fig 11.** Machine learning predictor importance. (A) Mean absolute SHAP values ranking all 20
312 predictors by their contribution to FTD prediction.

313 Host fruit availability further shaped these patterns (Fig 12). The presence and ripening of key host
314 fruits, particularly jew plum and mango, were associated with higher FTD, while sites lacking
315 these hosts—especially at higher elevations—supported only low and sporadic populations.

316 **Fig 12.** Machine learning predictor importance. Predictor evidence summary integrating SHAP
317 importance (x-axis, lollipop length) with the type and FDR-adjusted significance of univariate
318 statistical tests (point colour and size).

319 Climatic conditions also influenced abundance, with relative humidity showing the strongest and
320 most consistent relationship with FTD. However, these effects were most evident at lower
321 elevations, where populations were well established, and were weak or absent at higher elevations.
322 Together, these results show that *B. dorsalis* abundance along the gradient is primarily structured
323 by altitude and host availability, with climate acting as a secondary, context-dependent influence.

324 **Discussion**

325 This long-term study demonstrates that *B. dorsalis* populations exhibit pronounced seasonal
326 dynamics, with consistent peaks during the warm–wet season and declines during the cool-dry
327 period. Such rainfall-driven seasonality, which regulates host availability, is widely reported
328 across tropical systems, including Tanzania [13], Kenya [38], Benin [18], Cameroon [22], and
329 Asia [54,55], indicating a broadly conserved temporal structure. More broadly, the ecology and
330 spread of *B. dorsalis* across Africa reflect interacting effects of climate, host availability, and
331 landscape structure [10]. However, the long-term data further show that seasonal dynamics are not

332 uniform across environments. Along the altitudinal gradient, population peaks became
333 progressively shorter, weaker, and less predictable at higher elevations. This pattern indicates that
334 while lowland populations follow relatively stable climatic and phenological cycles, highland
335 populations are more constrained and temporally variable. Our study has refined seasonal
336 understanding by resolving the timing and reliability of peak and low population periods across
337 years, which is important for anticipating periods of highest risk.

338 Our results showed a clear decline in abundance with increasing elevation, demonstrating the
339 important role of altitude in structuring populations. This widespread pattern indicates that altitude
340 imposes fundamental constraints on population persistence. While lower temperatures at higher
341 elevations are known to slow development and reduce reproductive performance [14,56,57], the
342 present study shows that these effects are compounded by host limitation. The marked decline in
343 abundance above mid-elevations coincided with the absence of key hosts, and interaction analyses
344 showed that altitude effects were strongly mediated by host availability, consistent with
345 observations in Cameroon [22]. Similar patterns have been documented along the Morogoro
346 transect in Tanzania [19,20], across East Africa [21], and in southern Africa [64], as well as across
347 ecological zones in India [69]. Species distribution modelling studies further show that climatic
348 suitability, including temperature gradients associated with elevation, strongly determines the
349 geographic range of *B. dorsalis* [58]. In particular, phenology-based models incorporating
350 environmental gradients have reported how temperature and moisture jointly shape its potential
351 distribution across regions [23].

352 Climatic influences on abundance were dominated by moisture-related variables, with relative
353 humidity and rainfall showing stronger associations than temperature. This pattern is consistent
354 with established mechanisms, where adequate moisture improves survival, pupation conditions,

355 and host fruit quality [14,38]. In Benin, infestation levels reflect combined effects of abiotic
356 conditions and host factors [15], while broader studies demonstrate that climate operates in
357 conjunction with landscape and season to regulate population dynamics [64]. Thermal biology
358 studies have reported the role of temperature in shaping seasonal abundance [65]. Overall, these
359 findings indicate that moisture acts as a proximate driver of seasonal population fluctuations within
360 suitable environments, whereas temperature and host distribution define broader spatial limits.

361 Host fruit availability emerged as a key driver of population dynamics, with a small number of
362 hosts contributing disproportionately to population increases. The strong influence of jew plum
363 highlights the importance of non-commercial hosts in sustaining populations between major
364 cropping periods, consistent with the recognised role of *Spondias* species as preferred hosts [3].
365 Similar phenological linkages have been reported elsewhere, including in Uganda, where
366 population peaks track mango fruiting cycles [59], and in Senegal, where fruiting stages influence
367 oviposition behaviour [60]. Host suitability also varies with fruit ripening stage, influencing
368 infestation potential [61], while seasonal host availability and diversity further regulate population
369 dynamics [62].

370 Mango remains a primary host in East Africa [1,26], although its influence is partly mediated by
371 favourable climatic conditions at low elevations. In contrast, citrus contributes little to population
372 growth under natural conditions. Host prioritisation studies further confirm that reproductive
373 success varies significantly among host plants [63]. Importantly, host availability interacts with
374 altitude to shape population patterns. The absence of key hosts at higher elevations amplifies the
375 decline in abundance, reinforcing the need to consider host distribution alongside climatic
376 gradients. By isolating the role of key host fruits, this study clarifies how resource availability
377 drives population build-up, which is important for targeting host-based management strategies.

378 The observed patterns provide a basis for improving surveillance and management strategies.
379 Altitude offers a practical framework for risk stratification, with lowland areas experiencing
380 consistently higher and more predictable pest pressure. In these environments, climate-informed
381 forecasting—particularly using rainfall and humidity indicators—can improve the timing of
382 interventions such as bait sprays and male annihilation [18,66]. Similar approaches have proven
383 effective in Sri Lanka using time-series models [55]. At higher elevations, reduced predictability
384 of population peaks suggests that fixed seasonal calendars are less effective, and adaptive
385 monitoring based on trap thresholds is more appropriate. Integrating host phenology with climate-
386 based forecasts could further enhance early warning systems. In island systems, invasion dynamics
387 and niche interactions further illustrate the role of landscape structure [67].

388 This study is based on male trap captures, which do not directly reflect female oviposition or
389 infestation levels. Incorporating fruit sampling would improve inference on crop damage. In
390 addition, unequal sampling periods and site-specific phenological data may limit comparability
391 across the gradient. Finally, unmeasured biotic factors such as parasitism, competition, and
392 management practices may also influence population dynamics [11,68].

393 **Conclusion**

394 Long-term monitoring revealed consistent seasonal dynamics and strong altitudinal structuring of
395 *B. dorsalis* populations. Lowland populations exhibited predictable peaks, whereas highland
396 populations remained low and irregular. These patterns reflect interacting effects of climate, host
397 phenology, and landscape context, underscoring the importance of long-term data for developing
398 predictive and adaptive pest management strategies. These findings support the use of long-term,
399 climate-informed monitoring systems to improve the timing and targeting of *B. dorsalis*

400 management. In lowland areas, where populations are consistently high and seasonally predictable,
401 interventions should be synchronised with rainfall patterns and host fruit phenology to maximise
402 effectiveness. In contrast, highland systems require adaptive, threshold-based monitoring due to
403 lower and less predictable population dynamics. Integrating host phenology, climatic indicators,
404 and spatial risk stratification into early warning systems will enhance decision-making and support
405 more efficient, landscape-level pest management strategies.

406 **Author's contributions**

407 Maulid W. Mwatawala (MWM): Conceptualization, Methodology, Investigation, Data Collection,
408 Writing-original draft.

409 Marc De Meyer (MDM): Methodology, Resources, Data Collection, Writing-review & editing.

410 Jackline Bakengesa (JB) and Mwajuma Zinga (MW): Writing-review & editing.

411 Joseph O Ruboha (JOR): Formal analysis, Writing-review & editing

412 **Ethics**

413 The research was conducted from 2004 and 2012, before the establishment of formal ethical review
414 boards. The study involved only insect sampling and environmental observations and did not
415 involve human participants or vertebrate animals. All procedures complied with institutional
416 guidelines and accepted scientific practices applicable at the time of the study.

417 **Acknowledgements**

418 The study was carried out in several phases between 2004 and 2012 and was supported by multiple
419 funding sources, including the Belgian Development Cooperation through the Framework
420 Programme with the Royal Museum for Central Africa (Projects F13 and S1_TNZ_IPM), the joint

421 Food and Agriculture Organization / International Atomic Energy Agency (FAO/IAEA)
422 Programme on Nuclear Techniques in Food and Agriculture through the Coordinated Research
423 Project “*Resolution of Cryptic Species Complexes of Tephritid Pests to Overcome Constraints to*
424 *the Sterile Insect Technique (SIT) Application and International Trade*”, the International
425 Foundation of Science (Project C3956), and the Belgian Science Policy (Project MO/37/017).

426 **Declaration**

427 Artificial intelligence (AI) tools were used solely for grammar checking, spelling correction, and
428 improving language clarity. All analyses, interpretations, and scientific conclusions were
429 conducted entirely by the authors.

430 **Conflict of interest**

431 The authors declare that there is NO conflict of interest regarding the publication of this
432 Manuscript.

433 **References**

- 434 1. Ekesi S, De Meyer M, Mohamed SA, Virgilio M, Borgemeister C. Taxonomy, Ecology,
435 and Management of Native and Exotic Fruit Fly Species in Africa. *Annu Rev Entomol.*
436 2016;61: 219–238. doi:10.1146/annurev-ento-010715-023603
- 437 2. White I, Elson-Harris M. *Fruit Flies of Economic Significance: Their Identification and*
438 *Bionomics*. Wallingford: CAB International; 1992. doi:10.1079/9780851987903.0000
- 439 3. Allwood AJ, Chinajariyawong A, Drew RAI, Hamacek EL, Hancock DL, Hengsawad C,
440 et al. Host plant records for fruit flies (Diptera: Tephritidae) in Southeast Asia. *Raffles Bull*
441 *Zool.* 1999;47(7): 1–102.
- 442 4. Clarke AR, Armstrong KF, Carmichael AE, Milne JR, Raghu S, Roderick GK, et al.
443 Invasive phytophagous pests arising through a recent tropical evolutionary radiation: the
444 *Bactrocera dorsalis* complex of fruit flies. *Annu Rev Entomol.* 2005;50: 293–319.
445 doi:10.1146/ANNUREV.ENTO.50.071803.130428
- 446 5. Leblanc L, Hossain MA, Khan SA, San Jose M, Rubinoff D. A Preliminary Survey of the
447 Fruit Flies (Diptera: Tephritidae: Dacinae) of Bangladesh. *Proc Hawaiian Entomol Soc.*
448 2013;45: 51–58.
- 449 6. Lux SA, Copeland RS, White IM, Manrakhan A, Billah MK. A New Invasive Fruit Fly
450 Species from the *Bactrocera dorsalis* (Hendel) Group Detected in East Africa. *Int J Trop*
451 *Insect Sci.* 2003;23(4): 355–361. doi:10.1017/S174275840001242X
- 452 7. Drew RAI, Tsuruta K, White IM. A new species of pest fruit fly (Diptera: Tephritidae:
453 Dacinae) from Sri Lanka and Africa. *Afr Entomol.* 2005;13(1): 149–154.
- 454 8. Schutze MK, Mahmood K, Pavasovic A, Bo W, Newman J, Clarke AR, et al. One and the
455 same: Integrative taxonomic evidence that *Bactrocera invadens* (Diptera: Tephritidae) is

- 456 the same species as the Oriental fruit fly *Bactrocera dorsalis*. *Syst Entomol.* 2015;40(2):
457 472–486. doi:10.1111/SYEN.12114
- 458 9. Schutze MK, Aketarawong N, Amornsak W, Armstrong KF, Augustinos AA, Barr N, et
459 al. Synonymization of key pest species within the *Bactrocera dorsalis* species complex
460 (Diptera: Tephritidae). *Syst Entomol.* 2015;40(2): 456–471. doi:10.1111/SYEN.12113
- 461 10. Mutamiswa R, Nyamukondiwa C, Chikowore G, Chidawanyika F. Overview of oriental
462 fruit fly, *Bactrocera dorsalis* (Hendel) (Diptera: Tephritidae) in Africa: From invasion, bio-
463 ecology to sustainable management. *Crop Prot.* 2021;141: 105492.
464 doi:10.1016/J.CROPRO.2020.105492
- 465 11. Ekesi S, Billah MK, Nderitu PW, Lux SA, Rwomushana I. Evidence for competitive
466 displacement of *Ceratitis cosyra* by *Bactrocera invadens*. *Bull Entomol Res.* 2009;99(2):
467 153–161.
- 468 12. Rwomushana I, Ekesi S, Ogol CKPO, Gordon I. Mechanisms contributing to the
469 competitive success of the invasive fruit fly *Bactrocera invadens* over the indigenous
470 mango fruit fly, *Ceratitis cosyra*. *Entomol Exp Appl.* 2009;133(1): 27–37.
471 doi:10.1111/J.1570-7458.2009.00897.X
- 472 13. Mwatawala MW, De Meyer M, Makundi RH, Maerere AP. Seasonality and host utilization
473 of the invasive fruit fly, *Bactrocera invadens* (Dipt., Tephritidae) in central Tanzania. *J*
474 *Appl Entomol.* 2006;130(9–10): 530–537. doi:10.1111/J.1439-0418.2006.01099.X
- 475 14. Rwomushana I, Ekesi S, Ogol CKPO, Gordon I. Effect of temperature on development and
476 survival of immature stages of *Bactrocera invadens* (Diptera: Tephritidae). *J Appl Entomol.*
477 2008;132(9–10): 832–839. doi:10.1111/J.1439-0418.2008.01318.X

- 478 15. Vayssières JF, Korie S, Ayegnon D. Correlation of fruit fly (Diptera Tephritidae)
479 infestation of major mango cultivars in Borgou (Benin) with abiotic and biotic factors and
480 assessment of damage. *Crop Prot.* 2009;28(6): 477–488.
481 doi:10.1016/J.CROPRO.2009.01.010
- 482 16. Badii KB, Billah MK, Afreh Nuamah K, Obeng Ofori D, Nyarko G. Review of the pest
483 status, economic impact and management of fruit-infesting flies (Diptera: Tephritidae) in
484 Africa. *Afr J Agric Res.* 2015;10(12): 1488–1498. doi:10.5897/AJAR2014.9278
- 485 17. Muriithi BW, Diiro GM, Affognon H, Ekesi S. Economic impact of integrated pest
486 management strategies for the suppression of mango-infesting fruit fly species in Africa.
487 In: Ekesi S, Mohamed SA, De Meyer M, editors. *Fruit Fly Research and Development in*
488 *Africa.* Cham: Springer; 2016. pp. 755–770. doi:10.1007/978-3-319-43226-7_33
- 489 18. Vayssières JF, De Meyer M, Ouagoussounon I, Sinzogan A, Adandonon A, Korie S, et al.
490 Seasonal Abundance of Mango Fruit Flies (Diptera: Tephritidae) and Ecological
491 Implications for Their Management in Mango and Cashew Orchards in Benin. *J Econ*
492 *Entomol.* 2015;108(5): 2213–2230. doi:10.1093/jee/tov143
- 493 19. Geurts K, Mwatawala M, Meyer M De. Indigenous and Invasive Fruit Fly Diversity along
494 an Altitudinal Transect in Eastern Central Tanzania. *J Insect Sci.* 2012;12: 12.
495 doi:10.1673/031.012.1201
- 496 20. Geurts K, Mwatawala MW, De Meyer M. Dominance of an invasive fruit fly species,
497 *Bactrocera invadens*, along an altitudinal transect in Morogoro, Eastern Central Tanzania.
498 *Bull Entomol Res.* 2014;104(3): 288–294. doi:10.1017/S0007485313000722
- 499 21. Odanga JJ, Mohamed S, Mwalusepo S, Olubayo F, Nyankanga R, Khamis F, et al. Spatial
500 Distribution of *Bactrocera dorsalis* and *Thaumatotibia leucotreta* in Smallholder Avocado

- 501 Orchards along Altitudinal Gradient of Taita Hills and Mount Kilimanjaro. *Insects*.
502 2018;9(2): 71. doi:10.3390/INSECTS9020071
- 503 22. Nanga Nanga S, Hanna R, Fotso Kuate A, Fiaboe KKM, Nchoutnji I, Ndjab M, et al.
504 Tephritid Fruit Fly Species Composition, Seasonality, and Fruit Infestations in Two
505 Central African Agro-Ecological Zones. *Insects*. 2022;13(11): 1045.
506 doi:10.3390/insects13111045
- 507 23. De Villiers M, Hattingh V, Kriticos DJ, Brunel S, Vayssières JF, Sinzogan A, et al. The
508 potential distribution of *Bactrocera dorsalis*: Considering phenology and irrigation patterns.
509 *Bull Entomol Res*. 2016;106(1): 19–33. doi:10.1017/S0007485315000693
- 510 24. Liu H, Zhang DJ, Xu YJ, Wang L, Cheng DF, Qi YX, et al. Invasion, expansion, and
511 control of *Bactrocera dorsalis* (Hendel) in China. *J Integr Agric*. 2019;18(4): 771–787.
512 doi:10.1016/S2095-3119(18)62015-5
- 513 25. Stephens AEA, Kriticos DJ, Leriche A. The current and future potential geographical
514 distribution of the oriental fruit fly, *Bactrocera dorsalis* (Diptera: Tephritidae). *Bull*
515 *Entomol Res*. 2007;97(4): 369–378. doi:10.1017/S0007485307005044
- 516 26. Mwatawala MW, De Meyer M, Makundi RH, Maerere AP. Host range and distribution of
517 fruit-infesting pestiferous fruit flies (Diptera, Tephritidae) in selected areas of Central
518 Tanzania. *Bull Entomol Res*. 2009;99(6): 629–641. doi:10.1017/S0007485309006695
- 519 27. Burgess ND, Butynski TM, Cordeiro NJ, Doggart NH, Fjeldså J, Howell KM, et al. The
520 biological importance of the Eastern Arc Mountains of Tanzania and Kenya. *Biol Conserv*.
521 2007;134(2): 209–231. doi:10.1016/j.biocon.2006.08.015
- 522 28. Lovett J, Wasser SK, editors. *Biogeography and Ecology of the Rain Forests of Eastern*
523 *Africa*. Cambridge: Cambridge University Press; 1993.

- 524 29. Zuur AF, Ieno EN, Walker NJ, Saveliev AA, Smith GM. Mixed Effects Models and
525 Extensions in Ecology with R. New York: Springer; 2009. doi:10.1007/978-0-387-87458-
526 6
- 527 30. Shechambo L. Effects of host availability on seasonal abundance of the fruit fly *Bactrocera*
528 *invadens* Drew, Tsuruta & White in Morogoro [dissertation]. Morogoro: Sokoine
529 University of Agriculture; 2008.
- 530 31. FAO/IAEA. Trapping guidelines for area-wide fruit fly programmes. Enkerlin WR, Reyes-
531 Flores J, editors. 2nd ed. Vienna: IAEA; 2018.
- 532 32. Tan KH, Nishida R. Methyl Eugenol: Its Occurrence, Distribution, and Role in Nature,
533 Especially in Relation to Insect Behavior and Pollination. *J Insect Sci.* 2012;12: 56.
534 doi:10.1673/031.012.5601
- 535 33. Virgilio M, Jordaens K, Breman FC, Backeljau T, de Meyer M. Identifying Insects with
536 Incomplete DNA Barcode Libraries, African Fruit Flies (Diptera: Tephritidae) as a Test
537 Case. *PLoS ONE.* 2012;7(2): e31581. doi:10.1371/JOURNAL.PONE.0031581
- 538 34. Virgilio M, White I, De Meyer M. A set of multi-entry identification keys to African
539 frugivorous flies (Diptera, Tephritidae). *ZooKeys.* 2014;428: 97–108.
540 doi:10.3897/zookeys.428.7366
- 541 35. Fick SE, Hijmans RJ. WorldClim 2: new 1-km spatial resolution climate surfaces for global
542 land areas. *Int J Climatol.* 2017;37(12): 4302–4315. doi:10.1002/JOC.5086
- 543 36. Hersbach H, Bell B, Berrisford P, Hirahara S, Horányi A, Muñoz-Sabater J, et al. The
544 ERA5 global reanalysis. *Q J R Meteorol Soc.* 2020;146(730): 1999–2049.
545 doi:10.1002/QJ.3803

- 546 37. Gorelick N, Hancher M, Dixon M, Ilyushchenko S, Thau D, Moore R. Google Earth
547 Engine: Planetary-scale geospatial analysis for everyone. *Remote Sens Environ.* 2017;202:
548 18–27. doi:10.1016/J.RSE.2017.06.031
- 549 38. Ekesi S, Nderitu PW, Rwomushana I. Field infestation, life history and demographic
550 parameters of the fruit fly *Bactrocera invadens* (Diptera: Tephritidae) in Africa. *Bull*
551 *Entomol Res.* 2006;96(4): 379–386.
- 552 39. Zar JH. *Biostatistical Analysis*. 5th ed. Upper Saddle River: Pearson Prentice Hall; 2010.
- 553 40. Kruskal WH, Wallis WA. Use of Ranks in One-Criterion Variance Analysis. *J Am Stat*
554 *Assoc.* 1952;47(260): 583–621. doi:10.1080/01621459.1952.10483441
- 555 41. Spearman C. The Proof and Measurement of Association between Two Things. *Am J*
556 *Psychol.* 1904;15(1): 72. doi:10.2307/1412159
- 557 42. Cameron AC, Trivedi PK. *Regression Analysis of Count Data*. 2nd ed. Cambridge:
558 Cambridge University Press; 2013. doi:10.1017/CBO9781139013567
- 559 43. Hilbe JM. *Negative Binomial Regression*. 2nd ed. Cambridge: Cambridge University Press;
560 2011. doi:10.1017/CBO9780511973420
- 561 44. Liang KY, Zeger SL. Longitudinal data analysis using generalized linear models.
562 *Biometrika.* 1986;73(1): 13–22. doi:10.1093/BIOMET/73.1.13
- 563 45. Ke G, Meng Q, Finley T, Wang T, Chen W, Ma W, et al. LightGBM: A Highly Efficient
564 Gradient Boosting Decision Tree. In: Guyon I, Von Luxburg U, Bengio S, Wallach H,
565 Fergus R, Vishwanathan S, et al., editors. *Advances in Neural Information Processing*
566 *Systems*. Vol. 30. 2017. pp. 3146–3154.
- 567 46. Breiman L. Random forests. *Mach Learn.* 2001;45(1): 5–32.
568 doi:10.1023/A:1010933404324

- 569 47. Pedregosa F, Varoquaux G, Gramfort A, Michel V, Thirion B, Grisel O, et al. Scikit-learn:
570 Machine Learning in Python. *J Mach Learn Res.* 2011;12: 2825–2830.
- 571 48. Lundberg SM, Lee SI. A Unified Approach to Interpreting Model Predictions. In: Guyon
572 I, Von Luxburg U, Bengio S, Wallach H, Fergus R, Vishwanathan S, et al., editors.
573 *Advances in Neural Information Processing Systems*. Vol. 30. 2017. pp. 4766–4775.
- 574 49. Benjamini Y, Hochberg Y. Controlling the False Discovery Rate: A Practical and Powerful
575 Approach to Multiple Testing. *J R Stat Soc Ser B.* 1995;57(1): 289–300.
576 doi:10.1111/J.2517-6161.1995.TB02031.X
- 577 50. McKinney W. Data Structures for Statistical Computing in Python. *Proc 9th Python Sci*
578 *Conf.* 2010;445(1): 56–61. doi:10.25080/majora-92bf1922-00a
- 579 51. Harris CR, Millman KJ, van der Walt SJ, Gommers R, Virtanen P, Cournapeau D, et al.
580 *Array programming with NumPy.* *Nature.* 2020;585(7825): 357–362. doi:10.1038/s41586-
581 020-2649-2
- 582 52. Seabold S, Perktold J. *Statsmodels: Econometric and Statistical Modeling with Python.*
583 *Proc 9th Python Sci Conf.* 2010;7(1): 92–96. doi:10.25080/majora-92bf1922-011
- 584 53. Virtanen P, Gommers R, Oliphant TE, Haberland M, Reddy T, Cournapeau D, et al. *SciPy*
585 *1.0: fundamental algorithms for scientific computing in Python.* *Nat Methods.* 2020;17(3):
586 261–272. doi:10.1038/s41592-019-0686-2
- 587 54. Dong Z, He Y, Ren Y, Wang G, Chu D. Seasonal and Year-Round Distributions of
588 *Bactrocera dorsalis* (Hendel) and Its Risk to Temperate Fruits under Climate Change.
589 *Insects.* 2022;13(6): 550. doi:10.3390/INSECTS13060550
- 590 55. Wijekoon WMCD, Ganehiarachchi GASM, Wegiriya HCE, Vidanage SP. Seasonal
591 forecasting of *Bactrocera dorsalis* Hendel, 1912 (Diptera: Tephritidae) in bioclimatic zones

- 592 of Sri Lanka using the SARIMA model. CABI Agric Biosci. 2024;5(1): 40.
593 doi:10.1186/S43170-024-00241-2
- 594 56. Michel A, Dongmo K, Fiaboe KKM, Kekeunou S, Nanga SN, Kuate AF, et al.
595 Temperature-based phenology model to predict the development, survival, and
596 reproduction of the oriental fruit fly *Bactrocera dorsalis*. J Therm Biol. 2021;97: 102877.
597 doi:10.1016/J.JTHERBIO.2021.102877
- 598 57. Salum JK, Mwatawala MW, Kusolwa PM, Meyer MD. Demographic parameters of the
599 two main fruit fly (Diptera: Tephritidae) species attacking mango in Central Tanzania. J
600 Appl Entomol. 2014;138(6): 441–448. doi:10.1111/JEN.12044
- 601 58. De Meyer M, Robertson MP, Mansell MW, Ekesi S, Tsuruta K, Mwaiko W, et al.
602 Ecological niche and potential geographic distribution of the invasive fruit fly *Bactrocera*
603 *invadens* (Diptera, Tephritidae). Bull Entomol Res. 2010;100(1): 35–48.
604 doi:10.1017/S0007485309006713
- 605 59. Mayamba A, Nankinga CK, Isabirye B, Akol AM. Seasonal population fluctuations of
606 *Bactrocera invadens* (Diptera: Tephritidae) in relation to mango phenology in the Lake
607 Victoria Crescent, Uganda. Fruits. 2014;69(6): 473–480. doi:10.1051/FRUITS/2014033
- 608 60. Diatta P, Rey JY, Vayssières JF, Diarra K, Coly EV, Lechaudel M, et al. Fruit phenology
609 of citrus, mangos and papayas influences egg-laying preferences of *Bactrocera invadens*
610 (Diptera: Tephritidae). Fruits. 2013;68(6): 507–516. doi:10.1051/FRUITS/2013093
- 611 61. Cugala D, Ekesi S, Ambasse D, Adamu RS, Mohamed SA. Assessment of ripening stages
612 of Cavendish dwarf bananas as host or non-host to *Bactrocera invadens*. J Appl Entomol.
613 2014;138(6): 449–457. doi:10.1111/JEN.12045

- 614 62. Ndiaye O, Vayssières JF, Yves Rey J, Ndiaye S, Diedhiou PM, Ba CT, et al. Seasonality
615 and range of fruit fly (Diptera Tephritidae) host plants in orchards in Niayes and the Thies
616 Plateau (Senegal). *Fruits*. 2012;67(5): 311–331. doi:10.1051/FRUITS/2012024
- 617 63. Dominiak BC, Hoskins JL. Managing Oriental fruit fly, *Bactrocera dorsalis* (Hendel):
618 prioritising host plants using the host reproduction number. *Int J Trop Insect Sci*.
619 2025;45(3): 1249–1275. doi:10.1007/S42690-025-01514-7
- 620 64. Manrakhan A, Verykouki E, Serfontein L, Goldshtein E, Beck RR, Kriticos DJ, et al.
621 Temporal and spatial patterns of *Bactrocera dorsalis* (Diptera: Tephritidae) populations in
622 its southern limits of distribution. *Bull Entomol Res*. 2025;115(5): 644–659.
623 doi:10.1017/S0007485325100230
- 624 65. Motswagole R, Gotcha N, Nyamukondiwa C. Thermal Biology and Seasonal Population
625 Abundance of *Bactrocera dorsalis* Hendel (Diptera: Tephritidae): Implications on Pest
626 Management. *Int J Insect Sci*. 2019;11: 1179543319863417.
627 doi:10.1177/1179543319863417
- 628 66. Mwatawala MW, Mziray H, Malebo H, De Meyer M. Guiding farmers' choice for an
629 integrated pest management program against the invasive *Bactrocera dorsalis* Hendel
630 (Diptera: Tephritidae) in mango orchards in Tanzania. *Crop Prot*. 2015;76: 103–107.
631 doi:10.1016/j.cropro.2015.07.001
- 632 67. Mze Hassani I, Raveloson-Ravaomanarivo LH, Delatte H, Chiroleu F, Allibert A, Nouhou
633 S, et al. Invasion by *Bactrocera dorsalis* and niche partitioning among tephritid species in
634 Comoros. *Bull Entomol Res*. 2016;106(6): 749–758. doi:10.1017/S0007485316000456

- 635 68. Duyck PF, David P, Quilici S. A review of relationships between interspecific competition
636 and invasions in fruit flies (Diptera: Tephritidae). *Ecol Entomol.* 2004;29(5): 511–520.
637 doi:10.1111/J.0307-6946.2004.00638.X
- 638 69. Satarkar VR, Krishnamurthy SV, Faleiro JR, Verghese A. Spatial distribution of major
639 *Bactrocera* fruit flies attracted to methyl eugenol in different ecological zones of Goa, India.
640 *Int J Trop Insect Sci.* 2009;29(4): 195–201. doi:10.1017/S174275840999035X

641 **Supporting Information**

- 642 **S1 Fig.** Phenological stages of six key host fruit species and *B. dorsalis* seasonal activity at all
643 sites along the Uluguru transect. Source: adapted from Shechambo [30].
- 644 **S2 Fig.** Multivariate Spearman correlation matrices between FTD and weather variables at each
645 site separately (n = 38–81 per site).

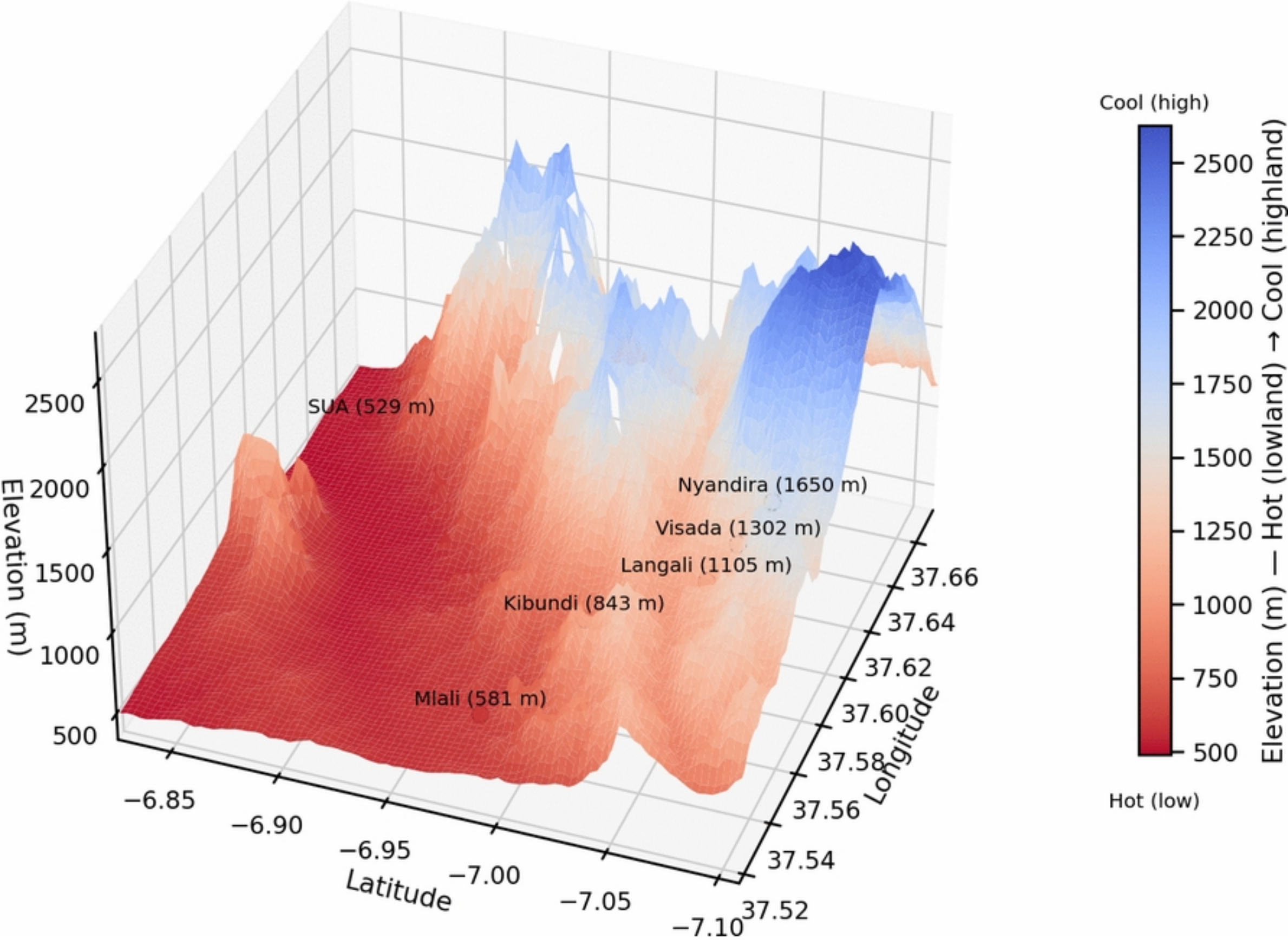


Figure1

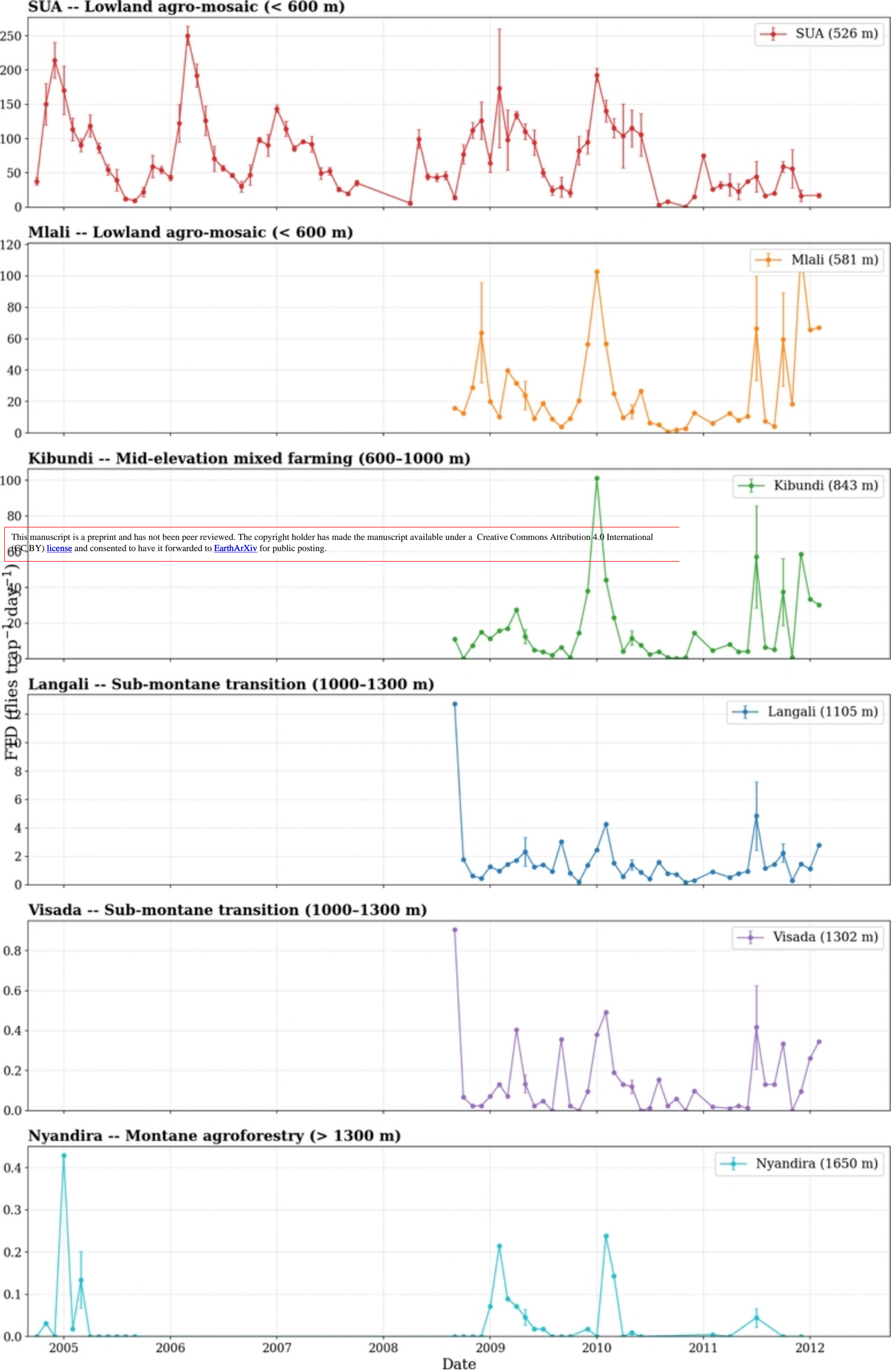


Figure2

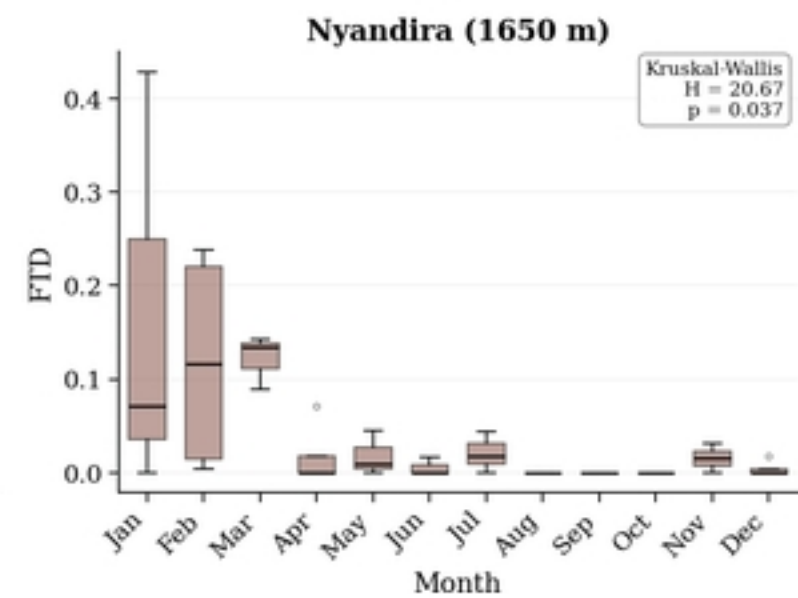
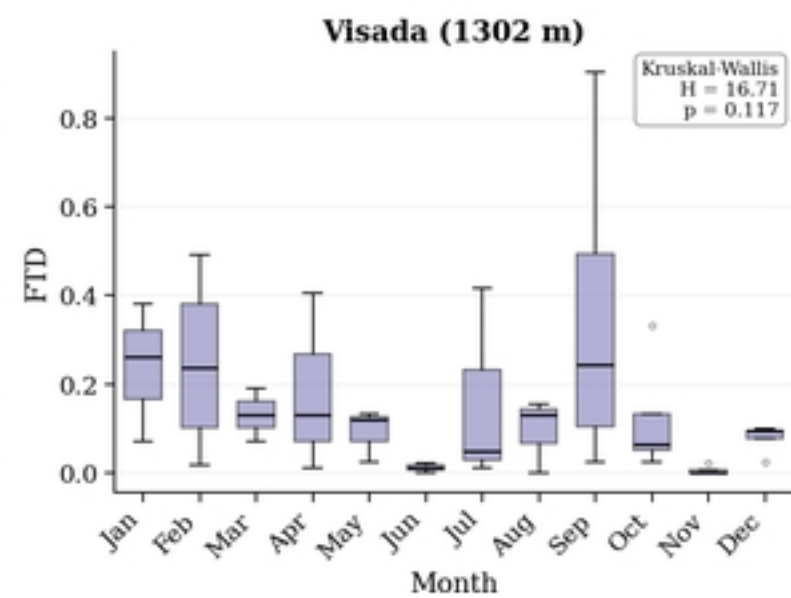
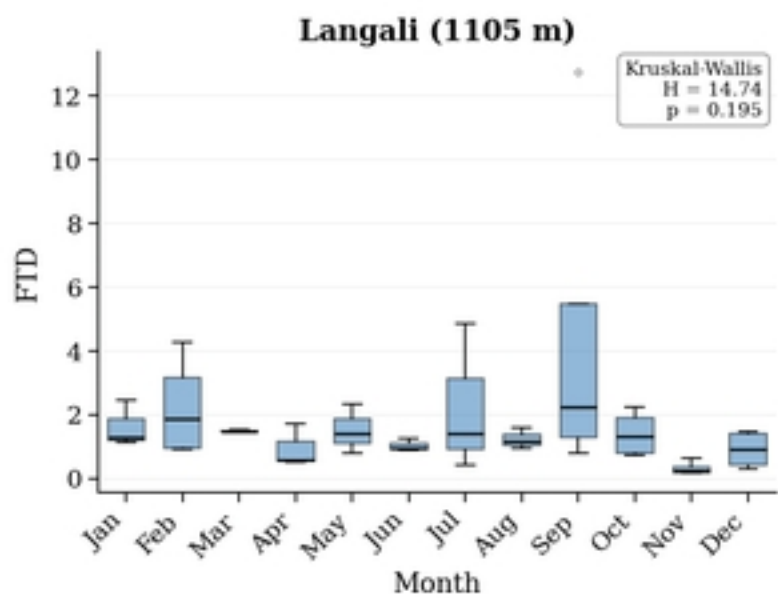
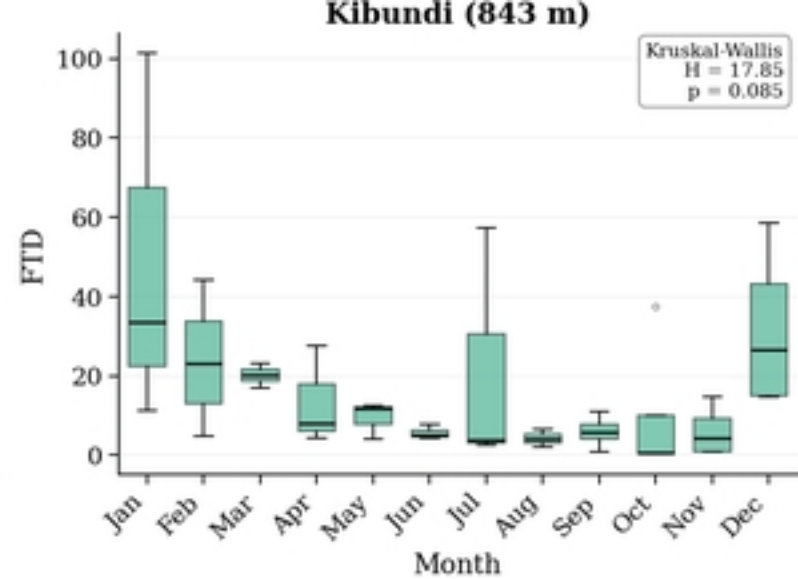
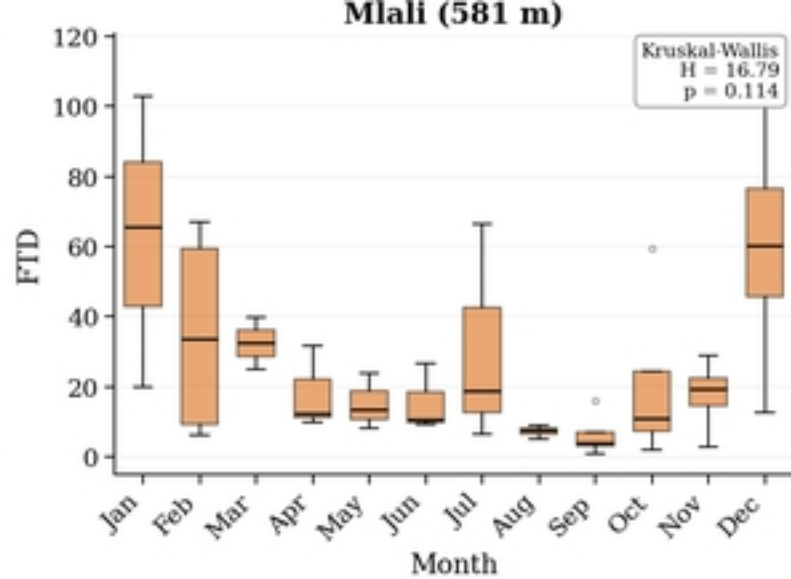
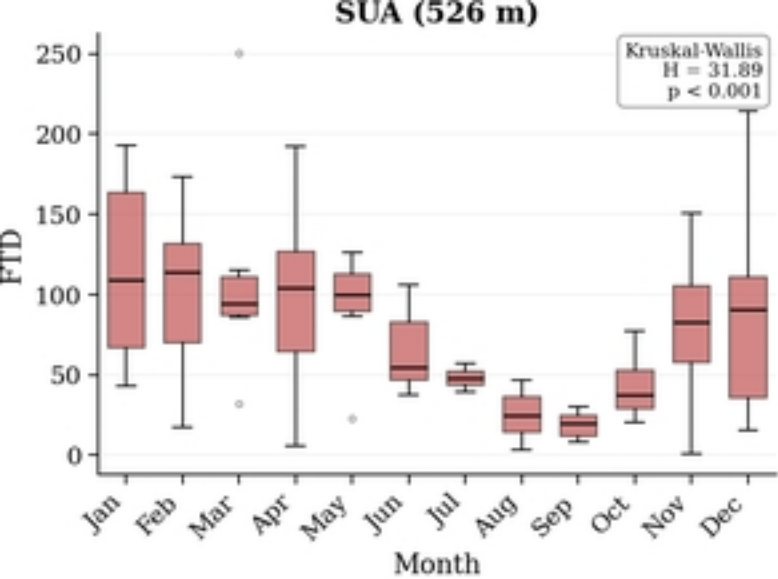


Figure3

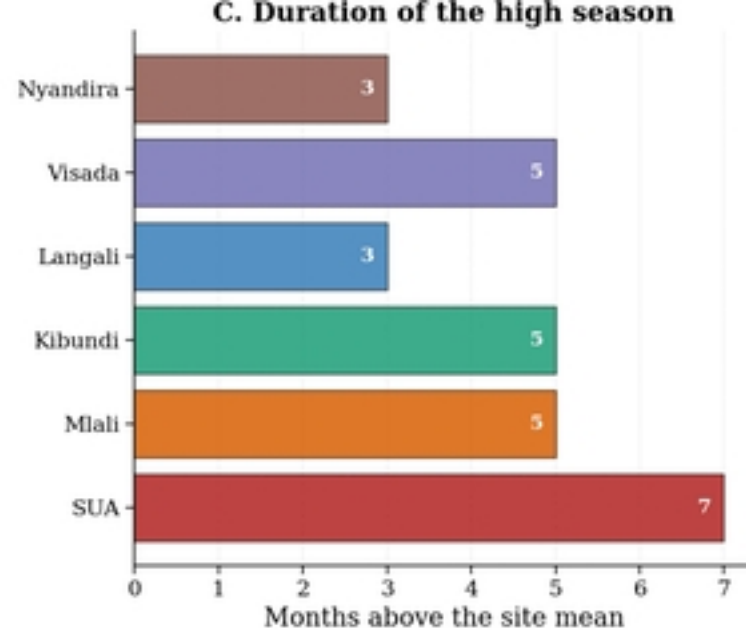
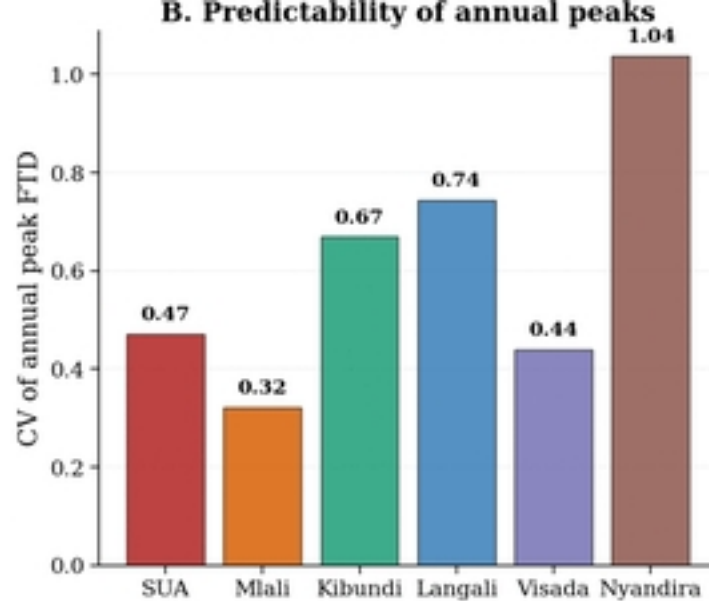
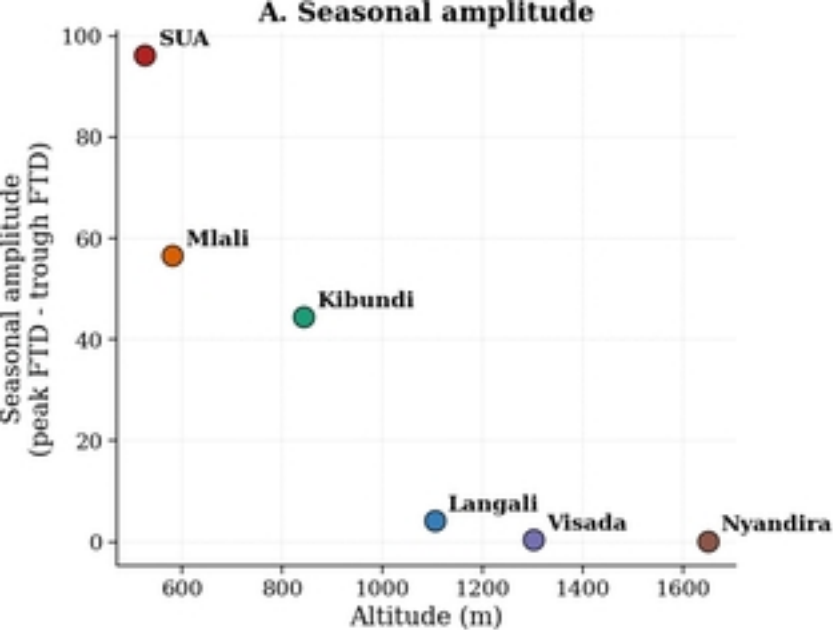


Figure4

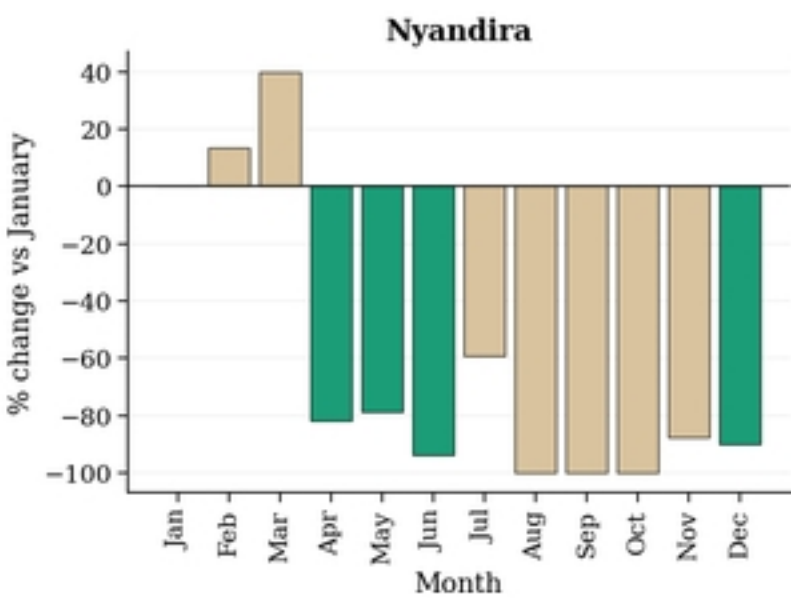
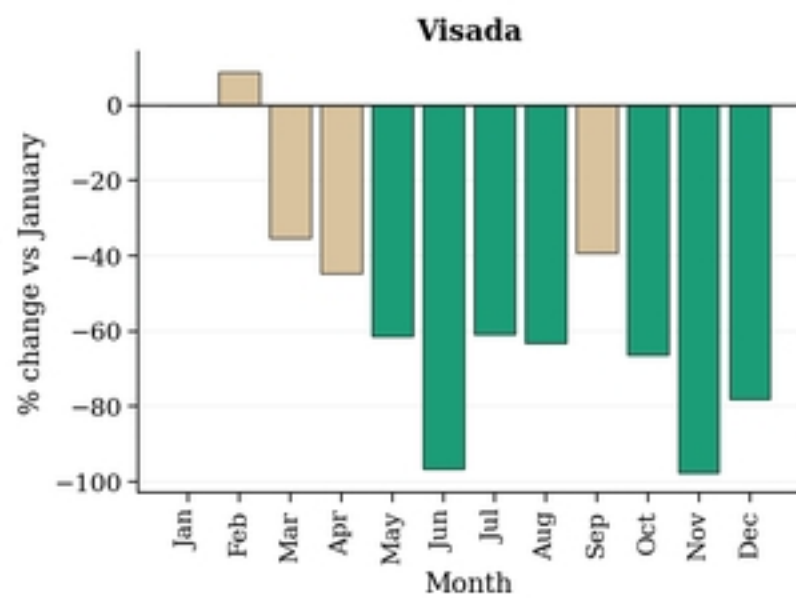
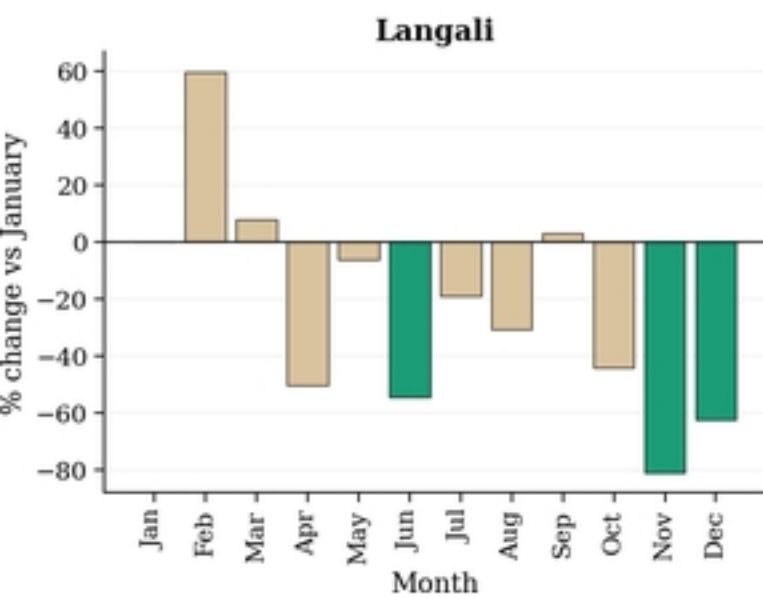
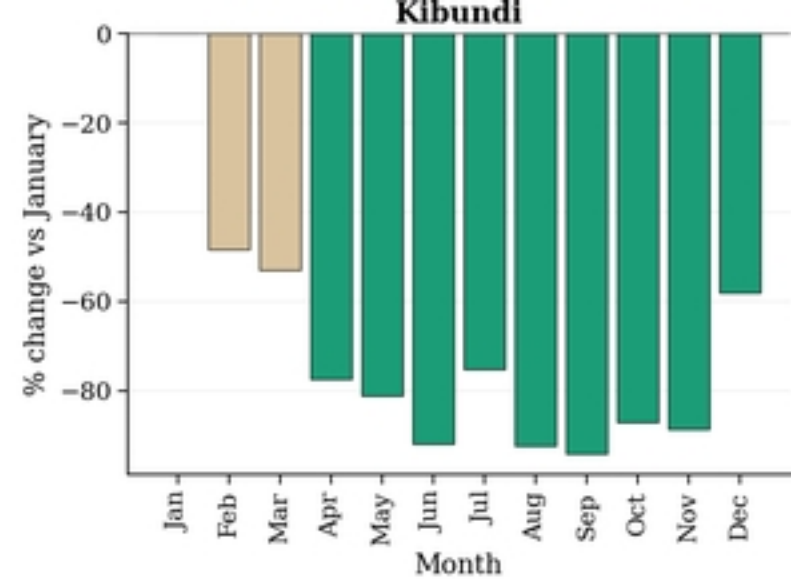
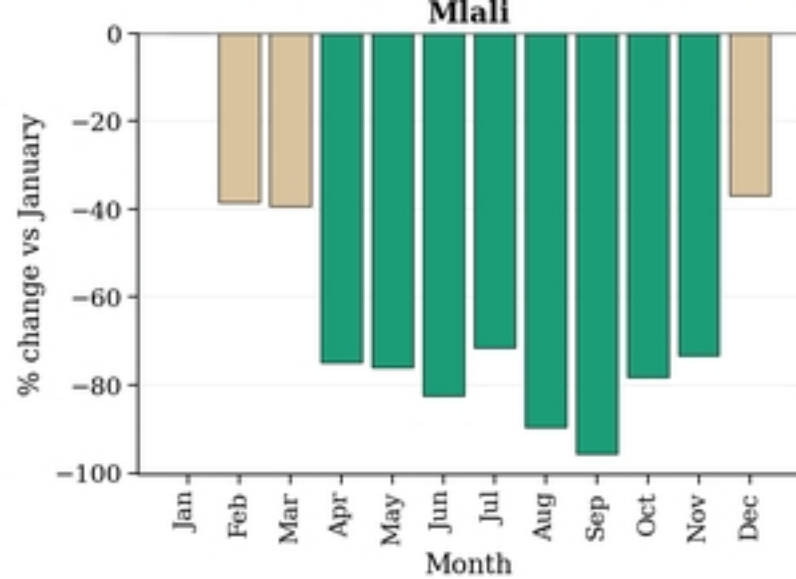
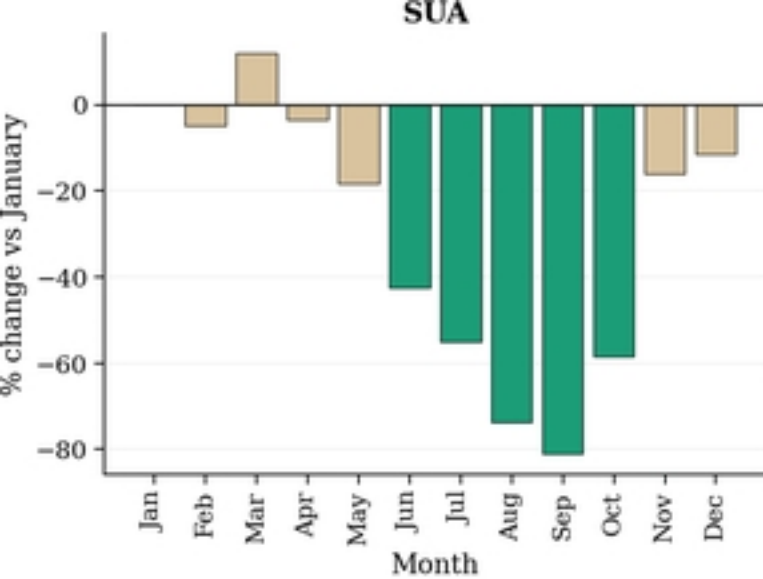


Figure5

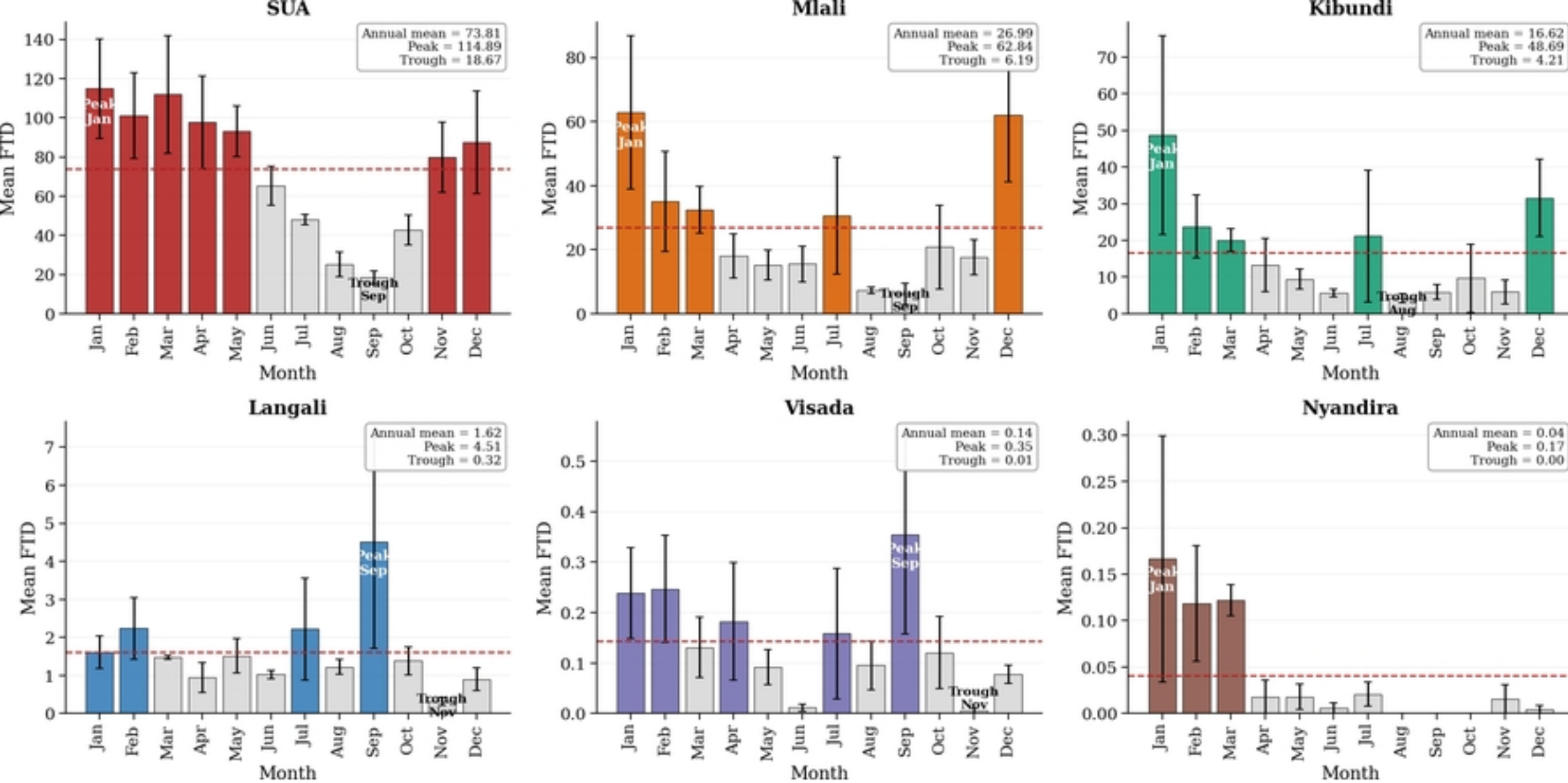


Figure6

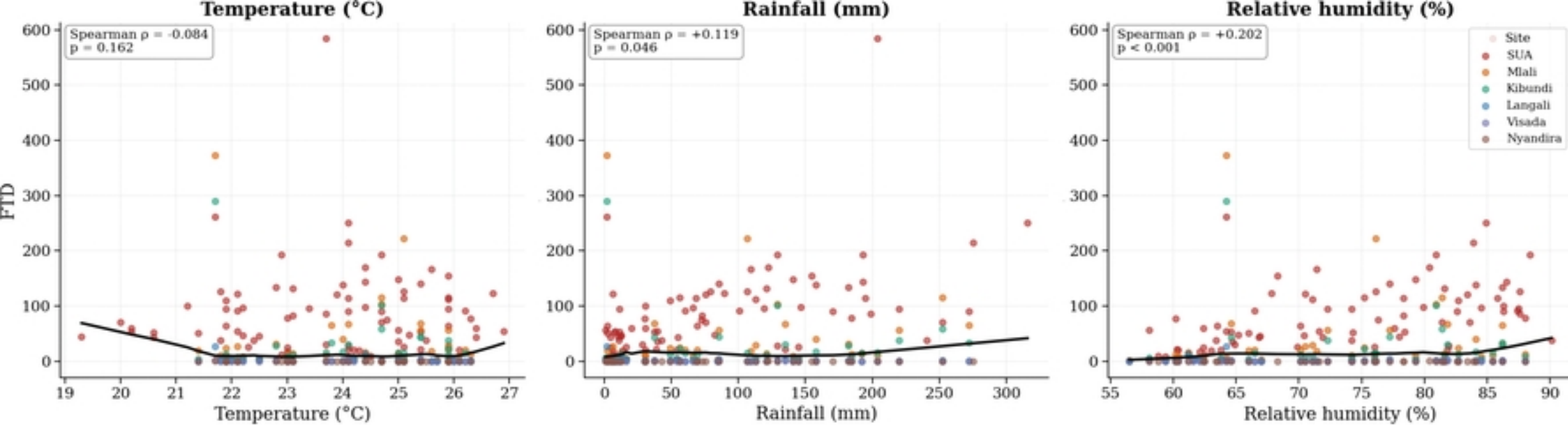


Figure7

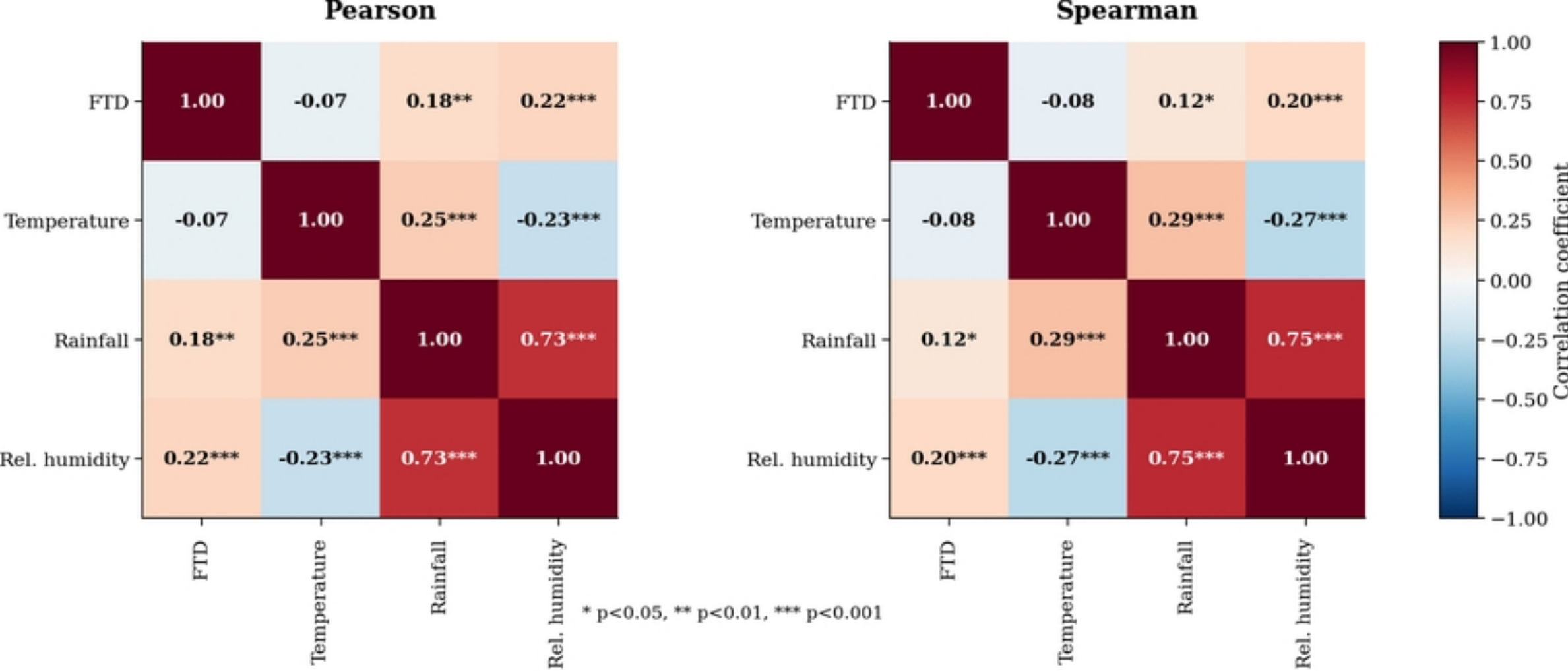


Figure8

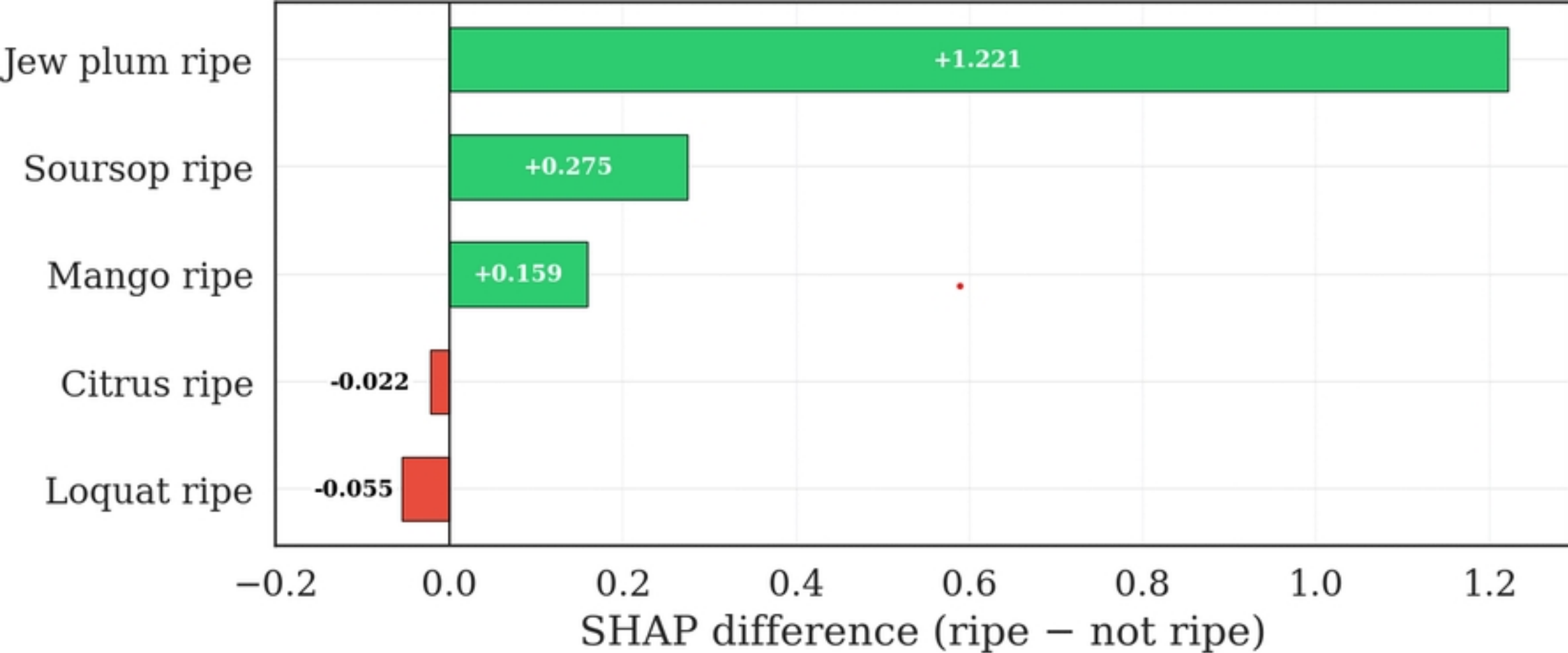


Figure9

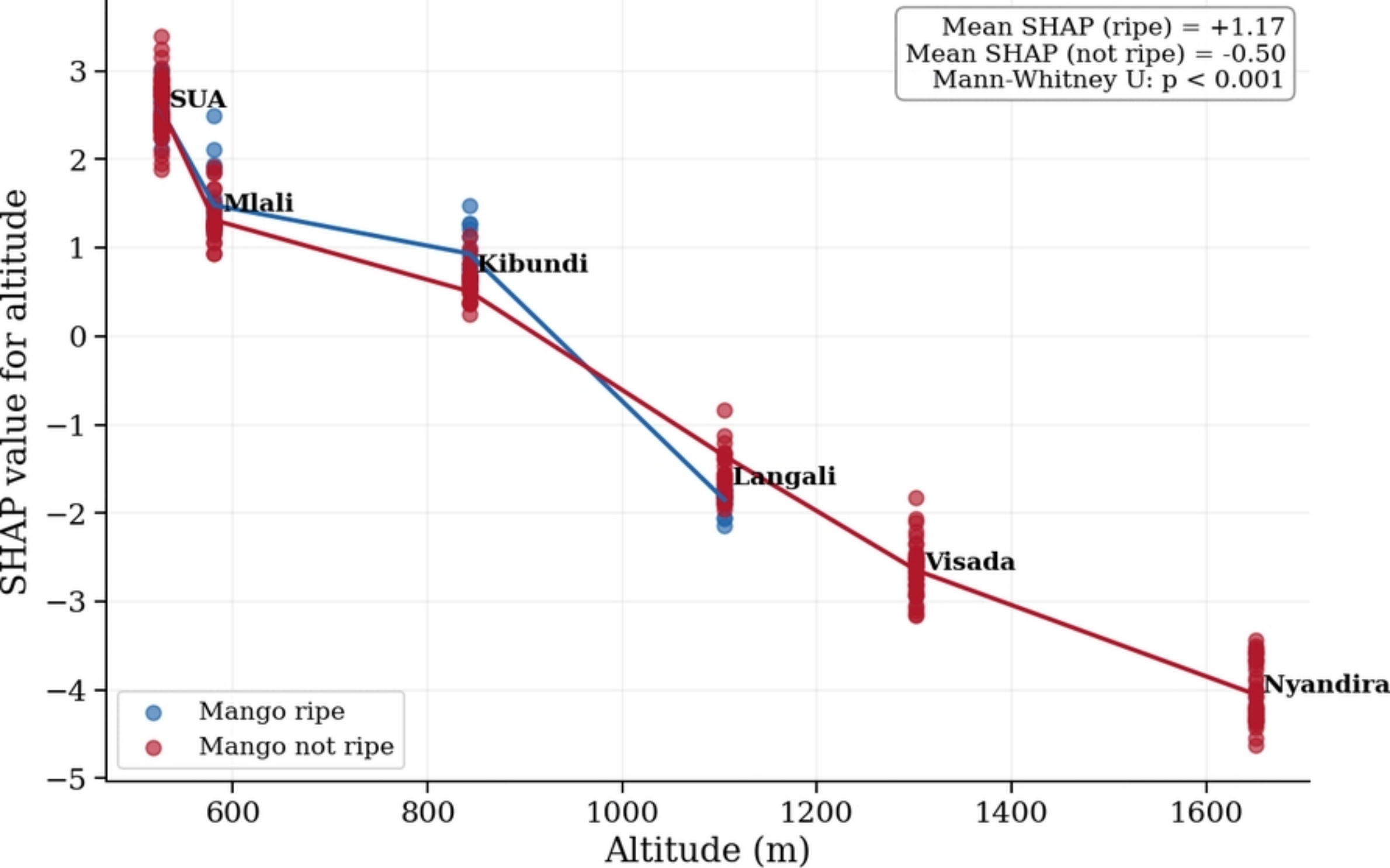


Figure10

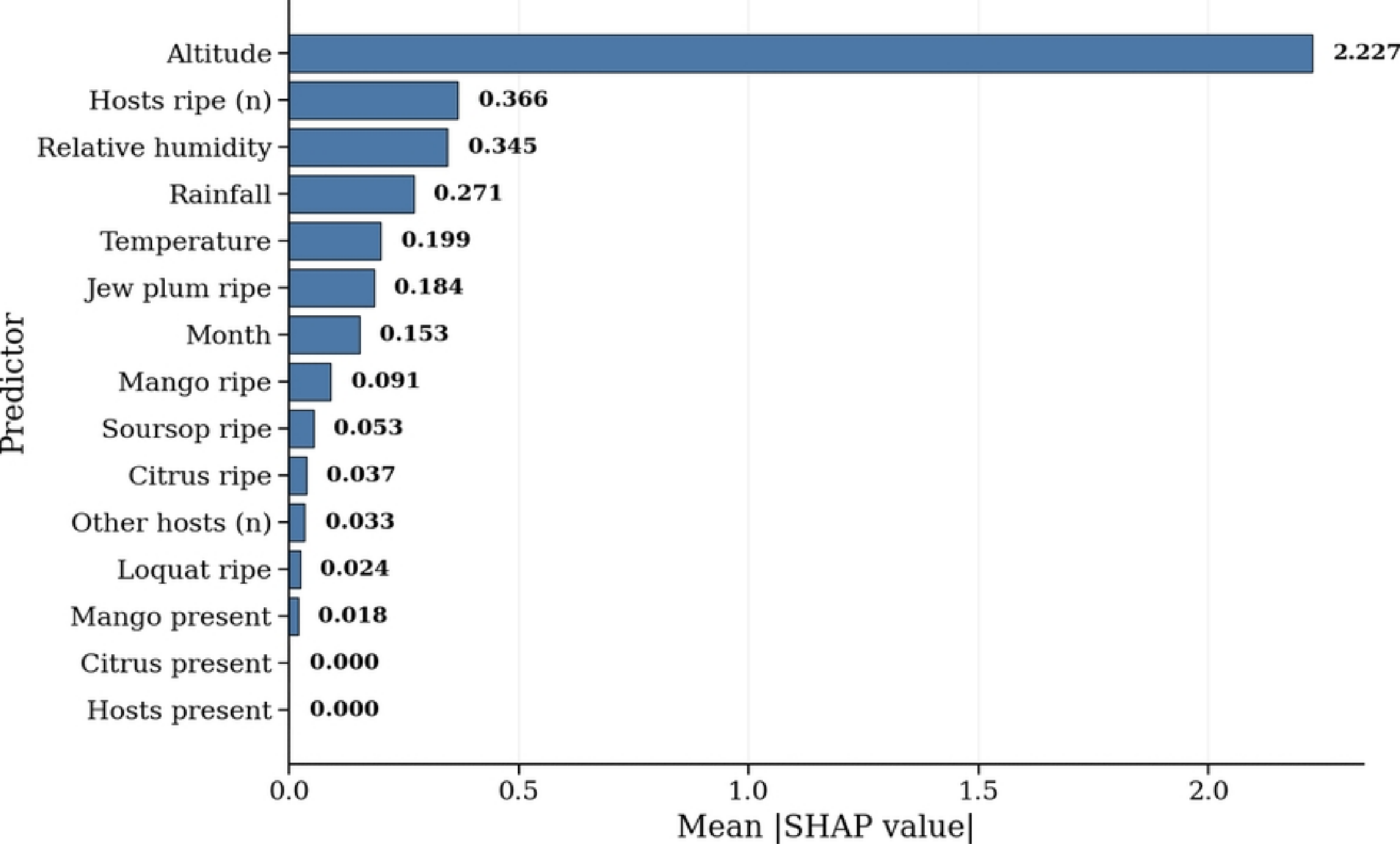


Figure 11

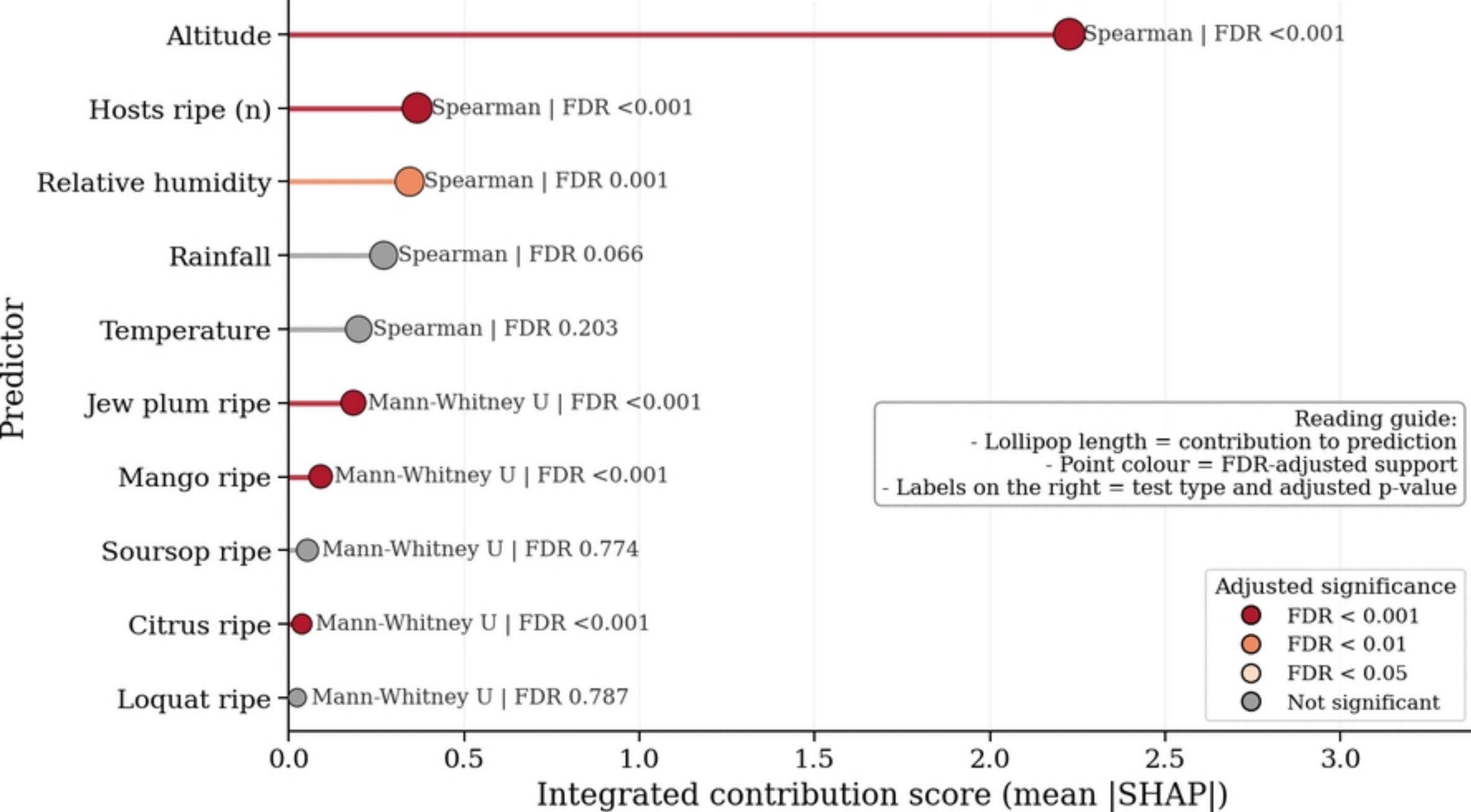


Figure12