

1 **Impacts of land-use and land-cover change on stream hydrochemistry**  
2 **in the Cerrado and Amazon biomes**

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1 Abstract – Studies on the impacts of land-use and land-cover change on stream  
2 hydrochemistry in active deforestation zones of the Amazon agricultural frontier are  
3 limited and have often used low-temporal-resolution datasets. Moreover, these impacts  
4 are not concurrently assessed in well-established agricultural areas and new  
5 deforestation hotspots. We aimed to identify these impacts using an experimental setup  
6 to collect high-temporal-resolution hydrological and hydrochemical data in two pairs of  
7 low-order streams in catchments under contrasting land use and land cover (native  
8 vegetation vs. pasture) in the Amazon and Cerrado biomes. Our results indicate that the  
9 conversion of natural landscapes to pastures increases carbon and nutrient fluxes via  
10 streamflow in both biomes. These changes were the greatest in total inorganic carbon in  
11 the Amazon and in potassium in the Cerrado, representing a 5.0- and 5.5-fold increase  
12 in the fluxes of each biome, respectively. We found that stormflow, which is often  
13 neglected in studies on stream hydrochemistry in the tropics, plays a substantial role in  
14 the carbon and nutrient fluxes, especially in the Amazon biome, as its contributions to  
15 hydrochemical fluxes are mostly greater than the volumetric contribution to the total  
16 streamflow. These findings demonstrate that assessments of the impacts of deforestation  
17 in the Amazon and Cerrado biomes should also take into account rapid hydrological  
18 pathways; however, this can only be achieved through collection of high-temporal-  
19 resolution data.

20 Keywords: carbon, nutrients, agricultural frontier, rainforest, savanna, deforestation.

## 21 1. Introduction

22 It has been widely acknowledged that surface conditions of terrestrial ecosystems have  
23 strong synergies with hydrological processes (Cuo et al., 2013; Neill et al., 2008; Recha  
24 et al., 2012; Rodriguez et al., 2010). These processes are often influenced by land-use  
25 practices, which, in turn, can change catchment responses, such as stream  
26 hydrochemistry (Crossman et al., 2014; El-Khoury et al., 2015; Oni et al., 2014; Öztürk et  
27 al., 2013; Salemi et al., 2013; Vogt et al., 2015). Because of large-scale environmental  
28 impacts resulting from the conversion of native habitats into agricultural frontiers  
29 (Schiesari et al., 2013), it is fundamental to comprehend how land-use and land-cover  
30 (LULC) change influences hydrochemical processes in pristine catchments undergoing

1 anthropogenic changes (Jordan et al., 1997; Neill et al., 2013). Therefore, studies have  
2 often focused on regions under intensive forest degradation due to agricultural expansion,  
3 such as the Brazilian Amazon, to assess the impacts of LULC change on stream  
4 hydrochemistry (Dias et al., 2015; Figueiredo et al., 2010b; Germer et al., 2009; Neill et  
5 al., 2011; Recha et al., 2013; Williams and Melack, 1997).

6 The Amazonian agricultural frontier (AAF), also known as the arc of deforestation,  
7 extends from the eastern to the southwestern edge of the Brazilian Amazon, comprising  
8 a wide area along the Amazon–Cerrado ecotone (Do Vale et al., 2015; Durieux, 2003;  
9 Silva et al., 2013). Deforestation in this region has taken place due to agricultural  
10 expansion during recent decades, and represents most of the deforestation of the AAF  
11 (Brannstrom et al., 2008; Fearnside, 2001; Riskin et al., 2013; Tollefson, 2015). This  
12 ongoing change threatens the services provided by native ecosystems, such as the water  
13 quantity and quality that sustain aquatic biodiversity and mitigates eutrophication of water  
14 bodies (Coe et al., 2013; Davidson et al., 2012; Neary, 2016; Penaluna et al., 2017).  
15 However, despite the important contribution of several research initiatives (e.g., Andreae  
16 et al., 2015; Lahsen and Nobre, 2007; Satinsky et al., 2014), an understanding of the  
17 influence of LULC change on water resources in the Brazilian Amazon region remains  
18 limited. Furthermore, the Cerrado biome, where most of the AAF deforestation has  
19 occurred (Klink and Machado, 2005), is often not integrated in studies regarding Amazon  
20 deforestation; consequently, it is one of the lesser-studied regions in terms of the  
21 environmental effects of LULC change resulting from agricultural expansion (Hunke et  
22 al., 2015a; Jepson et al., 2010; Oliveira et al., 2015) despite being a biodiversity hotspot  
23 for conservation comprised of dry forests, woodland savannas and grasslands (Spera et  
24 al., 2016; Strassburg et al., 2017). The conversion of native vegetation to crops and  
25 pastures has removed ca. 50% of the original 2 million km<sup>2</sup> in the Cerrado, which is  
26 greater than the forest loss in the Amazon biome (Klink and Machado, 2005; Lambin et  
27 al., 2013).

28 The negative impacts on water quality due to LULC change are reported to be a result of  
29 interrelated processes (i.e., changes in vegetation, soil and hydrology) that negatively  
30 disturbs its land capability, which is the ability of the land to sustain its use (Valle et al.,  
31 2014; Valle Junior et al., 2015). On the AAF, soil and hydrological changes have been

1 linked to forest clearing and conversion to pastures (Neill et al., 2008; Zimmermann et al.,  
2 2006). Indeed, LULC change on the AAF has been primarily driven by the expansion of  
3 pastures (Armenteras et al., 2013; Schierhorn et al., 2016). After some years, these  
4 pastures are often either replaced by cash crop systems (Barona et al., 2010; Cohn et  
5 al., 2016) or abandoned due to decreased grass productivity, ultimately reaching  
6 advanced stages of degradation (Davidson et al., 2012). Variations in nutrient input into  
7 rivers caused by LULC change on the AAF deserve particular attention because of their  
8 potential impact on both biogeochemistry and aquatic ecosystem functioning (Neill et al.,  
9 2011). Even though rain and dry forests account for ca. 60% of the net primary production  
10 of global terrestrial ecosystems (Grace et al., 2006; Potter et al., 2012), the effects of the  
11 impacts of LULC change in these systems are not well studied as they are for other  
12 regions of the world (Luke et al., 2017).

13 The initial effects of LULC change on the hydrochemistry of rivers have often been  
14 observed in low-order streams (Hope et al., 2004; Neill et al., 2001; Richey et al., 1997),  
15 which connect the terrestrial environment to large rivers and integrate environmental  
16 processes, especially landscapes undergoing change (Alexander et al., 2000; Moreira-  
17 Turcq et al., 2003). These characteristics qualify small streams as sensitive indicators of  
18 changes in ecosystems due to LULC change and allow their use as important references  
19 in carbon exportation studies and as early warning systems for ecological change  
20 (Christophersen et al., 1994). Although many studies have evaluated the dynamics of  
21 carbon and nutrients in streams in several regions of the world (e.g. Southeastern USA  
22 (Marchman et al., 2015), subtropical China (Yan et al., 2015), Germany (Strohmeier et  
23 al., 2013) and Canada (Jollymore et al., 2012)), studies of carbon export dynamics in low-  
24 order tropical catchments are still scarce (de Paula et al., 2016). There is increasing  
25 research interest in high-temporal-resolution data collection in low-order fluvial systems  
26 that should also be taken into account in hydrochemistry studies (Hughes et al., 2005;  
27 Richey et al., 2011; Wohl et al., 2012) due to their importance to the global carbon  
28 dynamics (Bass et al., 2014).

29 The dynamics of stream hydrochemistry that have remained largely invisible due to the  
30 monitoring schemes that only consider weekly or monthly sampling (Kirchner and Neal,  
31 2013), have been gradually unveiled due to approaches that use subdaily sampling

1 intervals (Tang et al., 2008). However, the high-frequency water sampling approach that  
2 has been shown to be useful for these studies in temperate regions (Clark et al., 2007)  
3 has been discredited in tropical regions (Chaussê et al., 2016). Moreover, findings in  
4 Amazonian headwater streams that have used subhourly sampling routines have found  
5 that the conversion of forests to fertilized agricultural lands changed neither the stream  
6 water chemistry nor nutrient output per unit of catchment area (Neill et al., 2017; Riskin  
7 et al., 2017).

8 Our study aims to identify the differences in stream carbon and nutrient (CAN)  
9 concentrations and output fluxes during prevalent baseflow and stormflow conditions in  
10 headwater catchments under contrasting LULC (native vegetation vs. pasture), thereby  
11 contributing to the understanding of CAN drivers in low-order streams on the AAF. Our  
12 hypothesis is that LULC change is impacting stream hydrochemistry in active  
13 deforestation zones of the Amazon and Cerrado biomes, with the stormflow, which is  
14 often neglected in studies in these regions, as a substantial contributor to the total CAN  
15 fluxes.

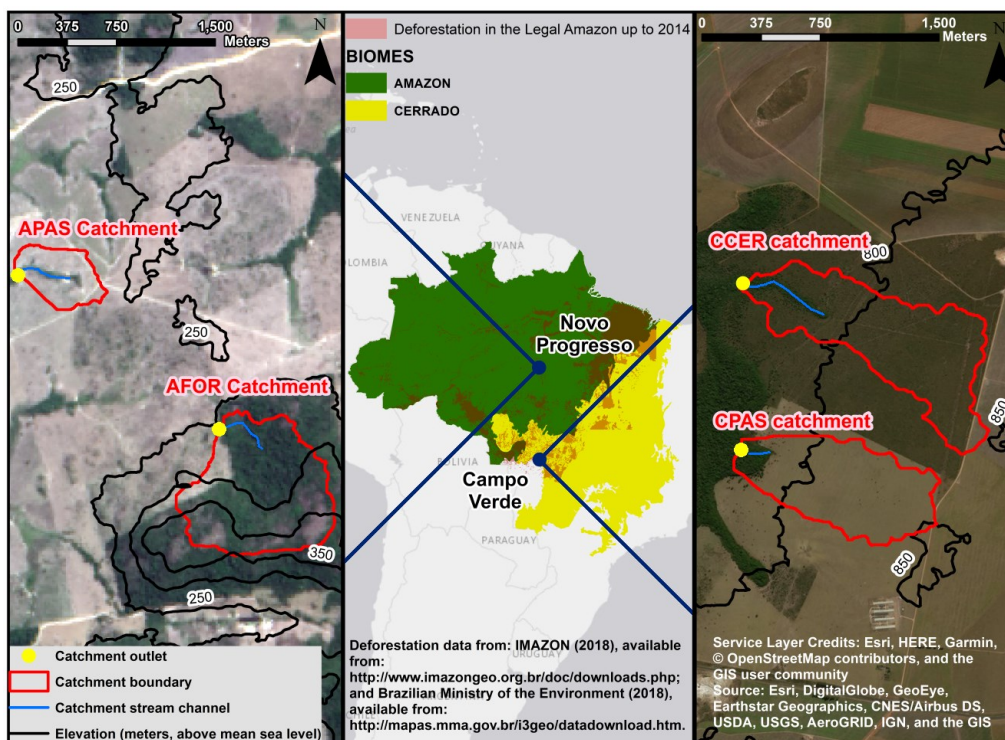
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## 17 2. Study area

18 Our study follows the space-for-time substitution approach to compare adjacent  
19 headwater catchments with different LULC but with similar characteristics, i.e. slope,  
20 geology, soils, aspect and climate (Troch et al., 2015). Studies have often used this  
21 approach to understand the effects of vegetation and land use on hydrological responses  
22 in small catchments (Brown et al., 2005; de Moraes et al., 2006; Germer et al., 2010;  
23 Muñoz-Villers and McDonnell, 2013; Ogden et al., 2013; Roa-García et al., 2011). It has  
24 also been applied to compare the impacts of LULC change on stream hydrochemistry of  
25 contrasting catchments (Sun et al., 2013; Zhao et al., 2010).

26 We used two pairs of microcatchments on the AAF (Fig. 1) with contrasting LULC. Each  
27 pair of catchments consists of a catchment with predominantly native vegetation land  
28 cover and a catchment with predominantly pasture land cover used for extensive cattle  
29 ranching. One pair of catchments is in the municipality of Novo Progresso (Brazilian state  
30 of Pará), which is a hotspot of deforestation in the Amazon biome (Pinheiro et al., 2016;

1 Rufin et al., 2015), and the other pair is in the municipality of Campo Verde (Brazilian  
 2 state of Mato Grosso), which is a region that has been massively deforested since the  
 3 1970s and is now a well-established agro-industrial area in the Cerrado biome. The  
 4 catchments in Novo Progresso, hereafter referred to as the Amazonian catchments, are  
 5 in the *Jamanxim* River watershed, which is one of the major southern subtributaries of  
 6 the Amazon River. The catchments in Campo Verde, hereafter referred to as the Cerrado  
 7 catchments, are in the *das Mortes* River watershed, the principal tributary of the *Araguaia*  
 8 River.



10 Figure 1. Study areas in the Amazon and Cerrado biomes.

11 The Amazonian catchments consist of one catchment covered with evergreen rainforest,  
 12 with signs of logging and tree regrowth (AFOR), and another catchment covered by  
 13 degraded pasture grassland (APAS). The AFOR catchment is the only catchment that is  
 14 drained by a non-perennial stream; it typically flows from November to July. The Cerrado  
 15 catchments are approximately 200 m apart, consisting of one catchment covered with  
 16 cerrado sensu stricto vegetation (CCER) and another catchment covered by pasture  
 17 grassland with signs of degradation (CPAS). The cerrado sensu stricto is characterized  
 18 as dense orchard-like vegetation consisting of many species of grasses and sedges, and

1 mixed with a great diversity of forbs and trees with an average height of 6 m (Canadell et  
2 al., 1996; Furley, 1999; Goodland, 1971; Goodland and Pollard, 1973; Ratter et al., 1997).  
3 The APAS catchment was established in 1984, and the CPAS catchment was established  
4 in 1994. Both pasture catchments are mostly covered by grasses (*Brachiaria* grass  
5 species) that exhibit low productivity rates. Lime (calcium carbonate,  $\text{CaCO}_3$ ) was applied  
6 in the pasture catchments several years before the study period. The climate in the  
7 Amazonian catchments is humid tropical, with a mean precipitation of ca. 1,900 mm yr<sup>-1</sup>,  
8 and a tropical wet and dry climate in the Cerrado catchments, with a mean precipitation  
9 of ca. 1,700 mm yr<sup>-1</sup>. More details regarding the climate, soils, morphology and hydrology  
10 of this region can be found in Lamparter et al. (2018), and Guzha et al. (2015) and in  
11 Nóbrega et al. (2017) for the Amazonian and Cerrado catchments, respectively. For  
12 clarity and to simultaneously compare the contrasting catchments within their respective  
13 biomes, we use the term native vegetation catchments to refer to the AFOR and CCER  
14 catchments, and the term pasture catchments to refer to the APAS and CPAS  
15 catchments, whose main characteristics are shown in Table 1. We instrumented these  
16 catchments during the dry season of 2012 and continuously monitored them from October  
17 of 2012 until the September of 2014.

18

### 19 3. Methods

#### 20 3.1 Soil physical and chemical properties

21 To support our findings related to CAN stream dynamics, we used evidence from soil  
22 chemical and textural analyses. We collected disturbed soil samples from the topsoil (0–  
23 10 cm soil depth), from 6 to 8 approximately equally spaced points along a topographic  
24 sequence of landscape positions from a gently sloping upper plateau, to a middle slope  
25 and a low-gradient valley bottom on the basis of digital elevation models (DEMs) derived  
26 from a topographic survey in each catchment. The topsoil of these catchments was  
27 chosen because it has a strong synergy with the surface waters and it is the soil layer  
28 under most direct influence of the LULC change (Lamparter et al., 2018). The topographic  
29 survey conducted in the Cerrado catchments is described in detail in Nóbrega et al.  
30 (2017); the described procedure was also used for the Amazonian catchments. We

1 analyzed these soil samples to determine pH, total carbon (TC), total nitrogen (TN),  
2 aluminum (Al), calcium (Ca), iron (Fe), potassium (K), magnesium (Mg), sodium (Na),  
3 phosphorus (P), sulfur (S) and particle size distribution. The particle size distribution was  
4 measured using the Köhn pipette method (DIN ISO 11277:2002-08, 2002). pH was  
5 measured using the potentiometric method (inoLAB<sup>®</sup> pH Level 2, Wissenschaftlich-  
6 Technische Werkstätten GmbH). TC and TN were quantified using an elemental analysis  
7 method (TruSpec<sup>®</sup> CHN, LECO Instrumente GmbH). For chemical analysis, a total  
8 digestion of 100–150 mg of soil was created with HClO<sub>4</sub>, HF and HNO<sub>3</sub> in 30-mL  
9 polytetrafluoroethylene (PTFE) vessels (Pressure Digestion System DAS 30, PicoTrace  
10 GmbH), and chemical concentrations were determined using inductively coupled plasma  
11 atomic emission spectroscopy (ICP-OES, Optima 4300<sup>™</sup> DV for the Cerrado catchments  
12 and ICP-OES Optima 5300<sup>™</sup> for the Amazonian catchments, PerkinElmer, Germany).  
13 Chemical analyses of soils from the Amazonian catchments were conducted at the  
14 Laboratory of the Department of Plant Ecology and Ecosystems Research and those of  
15 the Cerrado catchments were conducted at the Laboratory of the Department of  
16 Landscape Ecology, both at the University of Goettingen, Germany.

17

### 18 3.2 Water-sampling design and analysis

19 An automatic water sampler (BL2000<sup>®</sup>, Hach-Lange GmbH) was installed at the outlet of  
20 each catchment to collect stream water ca. 20 cm below the water surface and 2–4 m  
21 upstream from the catchment weir. The sampling procedure was simultaneously based  
22 on both time intervals and water-level variations to characterize the streamflow  
23 hydrochemistry during baseflow- and stormflow-prevailing conditions, respectively. The  
24 time sampling routine was based on filling a 1-L sample bottle over 1–3 days using an  
25 extraction of 200 mL from the stream at equal intervals. The stormflow sampling was  
26 determined using a subhourly routine activated by water-level increase and detected by  
27 a pressure bell switch (FD-01, Profimess GmbH). The pressure bell switches and the  
28 automatic samplers were calibrated throughout the year according to the water-level  
29 variation to maximize the coverage of the catchment stormflows, which considered the  
30 time of every sampling procedure and its respective hydrograph.



1 The samples from the Cerrado catchments were transported to the *Ecofisiologia Vegetal*  
2 Laboratory (EVL) at the Federal University of Mato Grosso (UFMT) in Cuiabá, Mato  
3 Grosso. The samples from the Amazonian catchments were also brought to this  
4 laboratory with prior preparation at a field facility ca. 5 km from the catchments and stored  
5 in light-free freezers until their transportation to the EVL. Transport of all water samples  
6 to the EVL was made using light-free coolers packed with ice. After transportation, the  
7 water in each bottle was used to fill two 50-mL aliquots in high-density polyethylene  
8 bottles prewashed with deionized water. One aliquot was used for the analysis of TC,  
9 total organic carbon (TOC), total inorganic carbon (TIC) and TN, and the other was filtered  
10 with pre-ashed glass fiber filters (0.7- $\mu\text{m}$  nominal pore size, Whatman GF/F) prewashed  
11 with 20 mL of water sample for the remaining analyses. The samples were then frozen  
12 and shipped in Styrofoam coolers for analysis at the Laboratory of the Department of  
13 Landscape Ecology, University of Goettingen, Germany (total travel time of ca. 22 h).

14 TC, TIC, TOC, total dissolved carbon (DC), dissolved inorganic carbon (DIC) and DOC  
15 contents were determined using high-temperature catalytic oxidation (TC-Analyzer,  
16 DIMATOC 100 (R), Dimatec GmbH). TN and DN were quantified using the  
17 chemiluminescence detection method (DIMA\_N module (CLD), Dimatec GmbH). Fluorine  
18 (F), chlorine (Cl), nitrate ( $\text{NO}_3$ ) and sulfate ( $\text{SO}_4$ ) concentrations were determined using  
19 ion chromatography (761 Compact IC, Metrohm, Switzerland). Dissolved Ca, Fe, K, Mg,  
20 Na, P and S concentrations were quantified using atomic spectroscopy (ICP-OES,  
21 Optima 4300™ DV, PerkinElmer). Prior to the analyses of the dissolved solutes, the water  
22 samples were filtered through membrane filters (0.45- $\mu\text{m}$  nominal pore size, cellulose  
23 acetate, Sartorius Stedim Biotech GmbH). These filters were prewashed with ultrapure  
24 water and transferred to high density polyethylene (HDPE) bottles that were prewashed  
25 with nitric acid solution (2.6%  $\text{HNO}_3$ ) and rinsed with ultrapure water.

26 For quality control, during the entire study period, approximately 20% of the water  
27 samples were analyzed for DOC within 12 hours after collection using a UV-Vis  
28 spectrometric device (spectro::lyser™ UV-Vis, scan Messtechnik GmbH) to cross-check  
29 with the final DOC results. This comparison indicated a linear correlation ( $r = .96$ ,  $n = 200$ ,  
30  $p < .001$ , Pearson's correlation), which is considered adequate because of the  
31 insignificant differences in DOC estimation by the spectrometric device calibration

1 (Avagyan et al., 2014; Bass et al., 2011). Additionally, a 1-L water sample was manually  
 2 collected in an automatic sampler bottle and kept in a separate automatic water sampler  
 3 unit at the EVL to check DOC fluctuations resulting from the storage of the samples in  
 4 this instrument. This water sample was analyzed using the spectrometric device up to 8  
 5 days after sampling, which was the average time interval of the field trips for sample  
 6 collection. This procedure was conducted during the first wet season (January–May of  
 7 2013) and did not indicate any significant changes in the DOC concentrations.

8

### 9 3.3. Streamflow and CAN output fluxes

10 At the outlet of each catchment, an adjustable weir was installed. During the rainy season,  
 11 the weirs were rectangular, whereas a v-notch contraction section was inserted during  
 12 the dry season. A multiparameter probe (DS 5X, OTT) was installed 2–4 m upstream of  
 13 each catchment's weir to obtain data on water level at 10 or 15-min intervals. To quantify  
 14 catchment discharge (flow rate), we used the standard flow equation (Eq. (1)) based on  
 15 the Bernoulli equation for the rectangular weir, and the Kindsvater–Shen equation (Eq.  
 16 (2)) together with calibration adjustment functions (Eqs. (3) and (4)) for the v-notch weir  
 17 (Shen, 1981), as follows:

$$18 \quad Q = \frac{2}{3} C_{dR} b \sqrt{2g} h^{\frac{3}{2}}, \quad (1)$$

$$19 \quad Q = \frac{8}{15} C_e \sqrt{2g} \tan\left(\frac{\theta}{2}\right) h_e^{\frac{5}{2}}, \quad (2)$$

$$20 \quad K_h = 0.001[\theta(1.395\theta - 4.296) + 4.135], \quad (3)$$

$$21 \quad C_e = \theta(0.02286\theta - 0.05734) + 0.6115, \quad (4)$$

22 where Q is the discharge over the weir ( $\text{m}^3 \text{s}^{-1}$ );  $C_{dR}$  and  $C_e$  are the effective  
 23 dimensionless discharge coefficients for the rectangular and v-notch weirs, respectively;  
 24 b is the weir length (m);  $\theta$  is the angle of the v-notch (radians); h is the upstream head  
 25 above the crest of the weir (m);  $h_e$  is the effective head ( $h + K_h$ ); and  $K_h$  is the head-  
 26 adjustment factor. For the Amazonian catchments, we adopted a  $C_{dR}$  of 0.62 based on  
 27 the geometric characteristics of the weirs (Kindsvater and Carter, 1957). For the Cerrado  
 28 catchments, we conducted discharge calibration measurements using an acoustic digital

1 current meter (ADC, OTT) and estimated  $C_{dR}$  values of 0.74 for the CCER catchment and  
2 0.65 for the APAS catchment.

3 We classified the streamflow as base streamflow ( $S_b$ ) and storm streamflow ( $S_s$ ), which  
4 represent the total stream discharge during baseflow- and stormflow-prevailing  
5 conditions, respectively.  $S_s$  was computed as the flow change in response to event  
6 precipitation and ending at the point separating the stormflow components, i.e. the  
7 surface and subsurface stormflow, from the baseflow recession. These flows were  
8 determined using a recursive digital filter (Eckhardt, 2005) implemented in the Web GIS-  
9 based Hydrograph Analysis Tool (WHAT) for baseflow separation (Lim et al., 2010, 2005).  
10 Using this information, we calculated the ratio of  $S_s$  to total streamflow ( $S_t$ ) discharge.

11 The annual CAN stream output fluxes for each catchment were calculated multiplying the  
12 annual mean CAN concentration by the respective annual  $S_b$  and  $S_s$  volumes (Eqs. 5 and  
13 6) as follows:

$$14 \quad F_{TS_b} = \frac{C_{S_b} \times V_{S_b}}{A \times 10^6}, \quad (5)$$

$$15 \quad F_{TS_s} = \frac{C_{S_s} \times V_{S_s}}{A \times 10^6}, \quad (6)$$

16 where  $F_{TS_b}$  and  $F_{TS_s}$  are, respectively, the annual CAN output fluxes of  $S_b$  and  $S_s$  ( $\text{kg ha}^{-1}$   
17  $\text{yr}^{-1}$ );  $C_{S_b}$  is the mean CAN concentration in  $S_b$  ( $\text{mg L}^{-1}$ );  $C_{S_s}$  is the volume-weighted  
18 mean CAN concentration obtained using Eq. 7 ( $\text{mg L}^{-1}$ );  $V_{S_b}$  and  $V_{S_s}$  are the mean annual  
19  $S_b$  and  $S_s$  discharges ( $\text{L yr}^{-1}$ ), respectively; and  $A$  is the catchment area (ha).

$$20 \quad C_{S_s} = \frac{\sum_{j=1}^m \left( \sum_{i=1}^n \frac{C_{S_s(i)}}{n} \right) \times V_j}{\sum_{j=1}^m V_j}, \quad (7)$$

21 where  $C_{S_s(i)}$  is the CAN concentration per  $S_s$  event interval  $i$  for the number of event  
22 intervals  $n$  ( $\text{mg L}^{-1}$ ) and  $V_j$  is the volume per event  $j$  for the number of  $S_s$  events  $m$  (L).

23

### 24 *3.4. Statistical analysis*

25 We used principal component analysis (PCA) to identify the most representative  
26 hydrochemical parameters causing most of the total variance in  $S_b$  and  $S_s$ . PCA is

1 commonly used to identify the variables that contain the most information and to provide  
2 future data collection criteria in ecological studies (King and Jackson, 1999; Zhang et al.,  
3 2009). It is useful for the identification of important surface water-quality parameters  
4 (Ouyang, 2005; Zeinalzadeh and Rezaei, 2017).

5 We conducted PCAs separately for each biome (Amazon and Cerrado) and flow condition  
6 ( $S_b$  and  $S_s$ ) in order to avoid the dominance of the PCA by the data variance of only one  
7 specific region or streamflow condition. We used the Kaiser–Meyer–Olkin (KMO) test  
8 (Kaiser, 1974) as a measure of quality control in the PCAs. The KMO test measures the  
9 sampling adequacy of each variable for the complete analysis. We only considered CAN  
10 parameters with individual KMO values greater than the bare minimum of .5; therefore  
11 we repeated the PCAs, excluding the unacceptable CAN parameters from the analyses,  
12 until we obtained acceptable individual KMO results. We applied the orthogonal rotation  
13 varimax with Kaiser normalization to the PCAs to maximize the dispersion of loadings  
14 within the factors and considered the results with the most significant components  
15 (eigenvalues > 1).

16 We used the Kolmogorov-Smirnov test of normality for each dataset to determine the  
17 adequate statistical test, i.e., parametric or nonparametric, for comparison of catchments  
18 within the same biome. We used the two-sample t-test to compare the soil chemistry and  
19 the Mann–Whitney (MW) U-test to compare the CAN concentrations by means of sample  
20 ranks to determine whether  $S_b$  and  $S_s$  were significantly different between the native  
21 vegetation and pasture catchments. Additionally to the MW test, we used Mood’s median  
22 test, given its robustness for outliers to detect differences in the median. We used the  
23 language and environment R (R Core Team, 2017) and the significance threshold at .05  
24 for all statistical analyses.

25

## 26 4. Results

### 27 4.1. Soil physical and chemical properties

28 The soils exhibited textural similarities within each pair of catchments, with mostly sandy  
29 clay loams in the Amazonian and loamy sand textures in the Cerrado catchments (Table  
30 2). The soil pH was between 10 to 25% higher in the pasture catchments, being

1 significantly different ( $p < .01$ ) between the CCER and CPAS catchments. The soils from  
2 all catchments have a high content of Al and Fe and low nutrient contents (Table 2). K,  
3 Mg and Mn contents exhibited significant differences ( $p < .05$ ) between the Amazonian  
4 catchments, with higher Mn content in the AFOR than that of the APAS catchment. In the  
5 Cerrado catchments, Ca was the only element to exhibit significant differences ( $p < .01$ )  
6 between the CCER ( $0.03 \text{ g kg}^{-1}$ ) and CPAS catchments ( $0.18 \text{ g kg}^{-1}$ ).

#### 7 4.2. Hydrochemistry results

8 TOC, DOC, K and  $\text{NO}_3$  exhibited the highest mean concentrations ( $> 1 \text{ mg L}^{-1}$ ) in the  
9 Amazonian catchments under both flow conditions. For these catchments, our results  
10 indicate low mean streamflow concentrations for Cl,  $\text{SO}_4$ , Na, Ca and Mg ( $< 0.4 \text{ mg L}^{-1}$ ).  
11 In the Cerrado catchments, TOC, DOC,  $\text{NO}_3$  and Ca showed the highest mean  
12 concentrations. Other elements, such as Mg and Na, exhibited relatively low  
13 concentrations in the CCER catchment. Fe, F, P, S and  $\text{SO}_4$  had the lowest  
14 concentrations in all catchments, with most values less than the limit of detection (Tables  
15 A.1 and A.2).

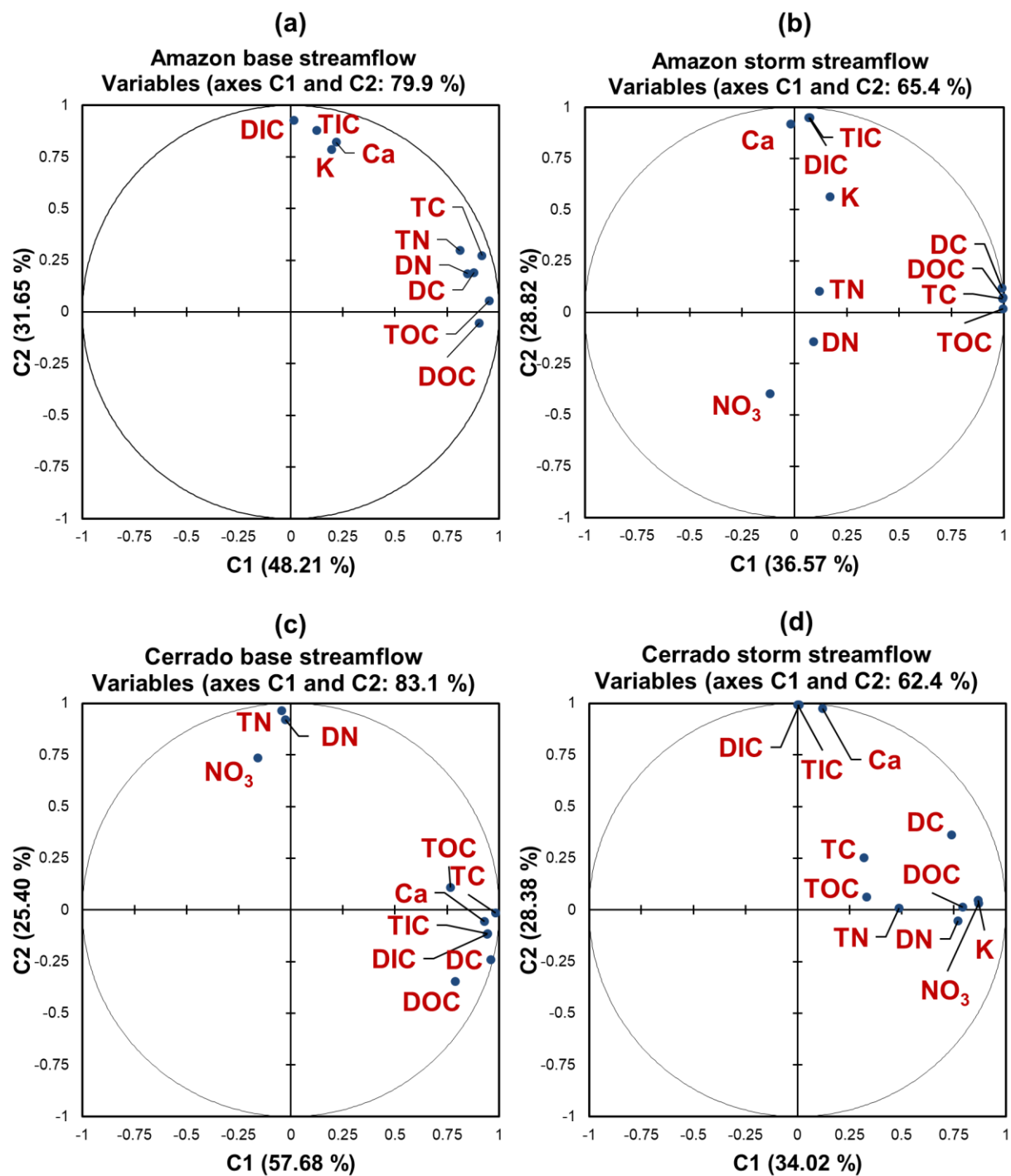
16 The varimax rotation applied to the PCA on the water quality parameters exhibited  
17 individual KMO values greater than .5 (Table 3). The overall KMO was .70 for  $S_b$  and .63  
18 for the  $S_s$  PCAs in the Amazonian catchments, and .68 for both the  $S_b$  and  $S_s$  PCAs in  
19 the Cerrado catchments, which are acceptable values of sampling adequacy for PCA  
20 (Kaiser, 1974). Bartlett's test of sphericity for the parameters indicated that correlations  
21 between items were sufficiently great for PCA ( $p < .001$ ). Kaiser's criterion of eigenvalues  
22 greater than 1 was met by two components in the  $S_b$  PCAs and by three components in  
23 the stormflow PCAs for the Amazonian and Cerrado catchments. In combination, these  
24 components explained 80% and 86% of the variance in the  $S_b$  and  $S_s$  values in the  
25 Amazonian catchments, and 83% and 88% of the variance in the  $S_b$  and  $S_s$  values in the  
26 Cerrado catchments, respectively. Some parameters, such as TC, TOC, DC and DOC,  
27 cluster in the same components in all PCAs with high factor loadings.

28 In all of the PCAs, the first two components account for more than 60% of the total  
29 variance (Fig. 2). For the Amazonian catchments, the first component of the  $S_b$  PCA (Fig  
30 2a) was mostly correlated with nitrogen and organic carbon, which showed the highest

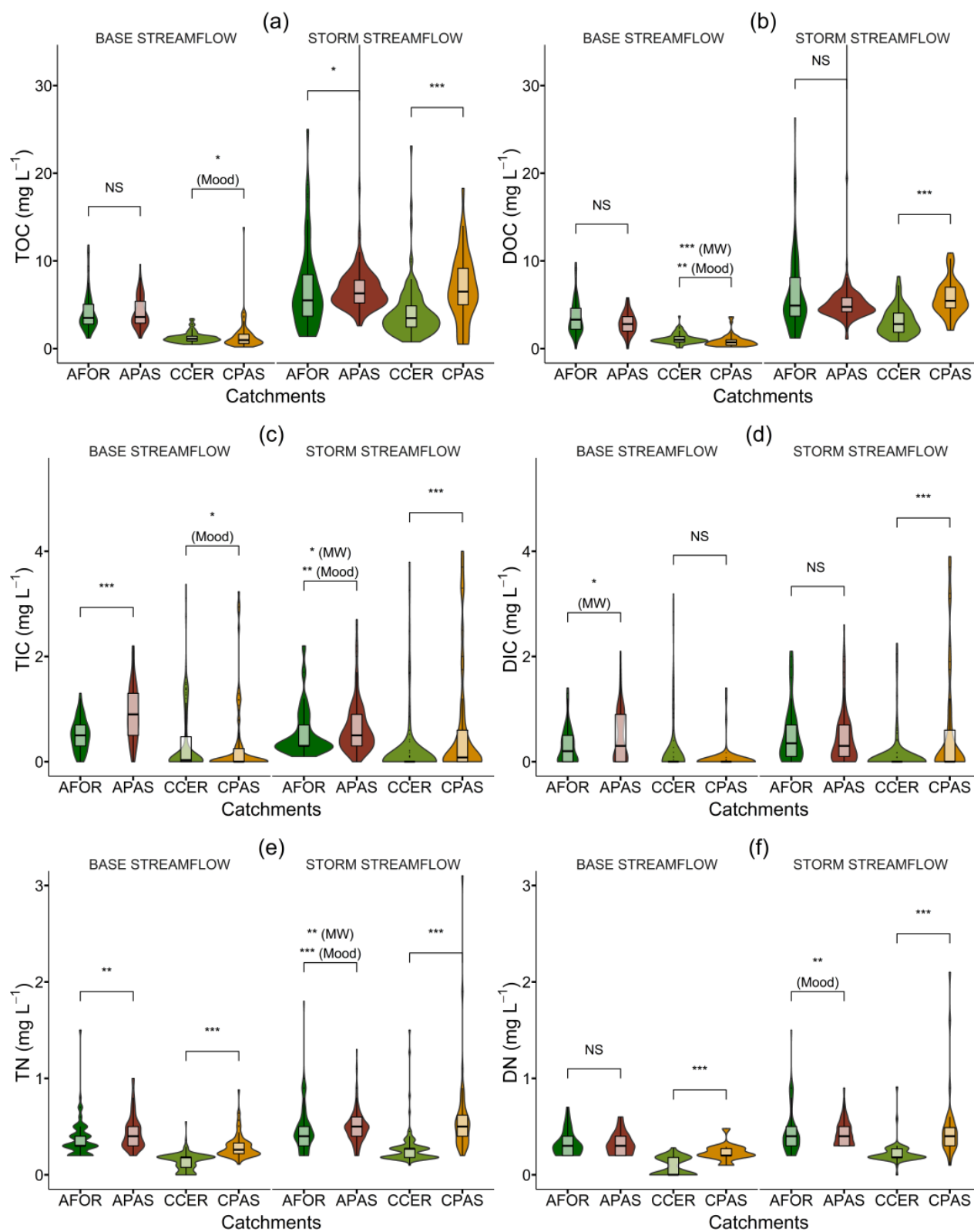
1 standard deviations. The items that cluster in the second component represent the  
2 inorganic carbon and cations (Ca and K). The main difference between the  $S_b$  and  $S_s$   
3 PCAs (Fig. 2b) is the clustering of  $NO_3$ , TN and DN in the third component of the  $S_s$  PCA,  
4 suggesting that during stormflow events, nitrogen fluxes have a distinct dynamic from that  
5 of the other nutrients. For the Cerrado catchments, the first component of the  $S_b$  PCA  
6 (Fig. 2c) groups carbon and Ca, and the second component groups TN, DN and  $NO_3$ .  
7 This is the only PCA where the organic and inorganic carbon compounds cluster in the  
8 same component. The  $S_s$  PCA (Fig. 2d) shows that the first component groups DOC with  
9 DN,  $NO_3$  and K, and the second component shows a high factor loading grouping of TIC,  
10 DIC and Ca. The third component of this PCA groups TC, TOC and TN. This is the only  
11 PCA where TOC does not group together with DOC, which indicates the importance of  
12 particulate organic carbon (POC) in these catchments. We did not directly measure POC  
13 in our study, but the differences between TOC and DOC, which could be interpreted as  
14 POC (Zhou et al., 2013), were the highest in the Cerrado catchments, representing an  
15 average of 19% of the TOC.

16 Based on the results of the PCAs, we compared TOC, DOC, TIC, DIC, TN and DN (Fig.  
17 3), and  $NO_3$ , Ca and K (Fig. 4). With the exception of higher TOC in the APAS catchment,  
18  $S_s$  carbon concentrations between the Amazonian catchments did not exhibit significant  
19 differences. In the Cerrado catchments, the highest differences were found in  $S_s$ , with  
20 higher TOC and DOC concentrations in the CPAS catchment compared to those of the  
21 CCER (Fig. 3a–b). For DIC, the differences in concentration between the Amazonian  
22 catchments in  $S_b$  and between the Cerrado catchments in  $S_s$  (Fig. 3c–d) were significant.

23 Except for DN in  $S_b$  of the Amazonian catchments, the pasture catchments exhibited  
24 higher TN and DN concentrations than those of the native vegetation catchments. The  
25 differences in  $NO_3$  were significant between the Cerrado catchments, with higher  
26 concentrations in the CPAS catchment, whereas there was no significant difference in the  
27 Amazonian catchments (Fig. 4a). Differences in Ca concentrations (Fig. 4b) were  
28 significant in the catchments of both biomes, but not for the same flow conditions. While  
29 the difference in Ca was significant only in  $S_b$  of the Amazonian catchments, this was only  
30 observed in  $S_s$  of the Cerrado catchments. There were significantly higher K  
31 concentrations in both  $S_b$  and  $S_s$  for the pasture catchments (Fig. 4c).



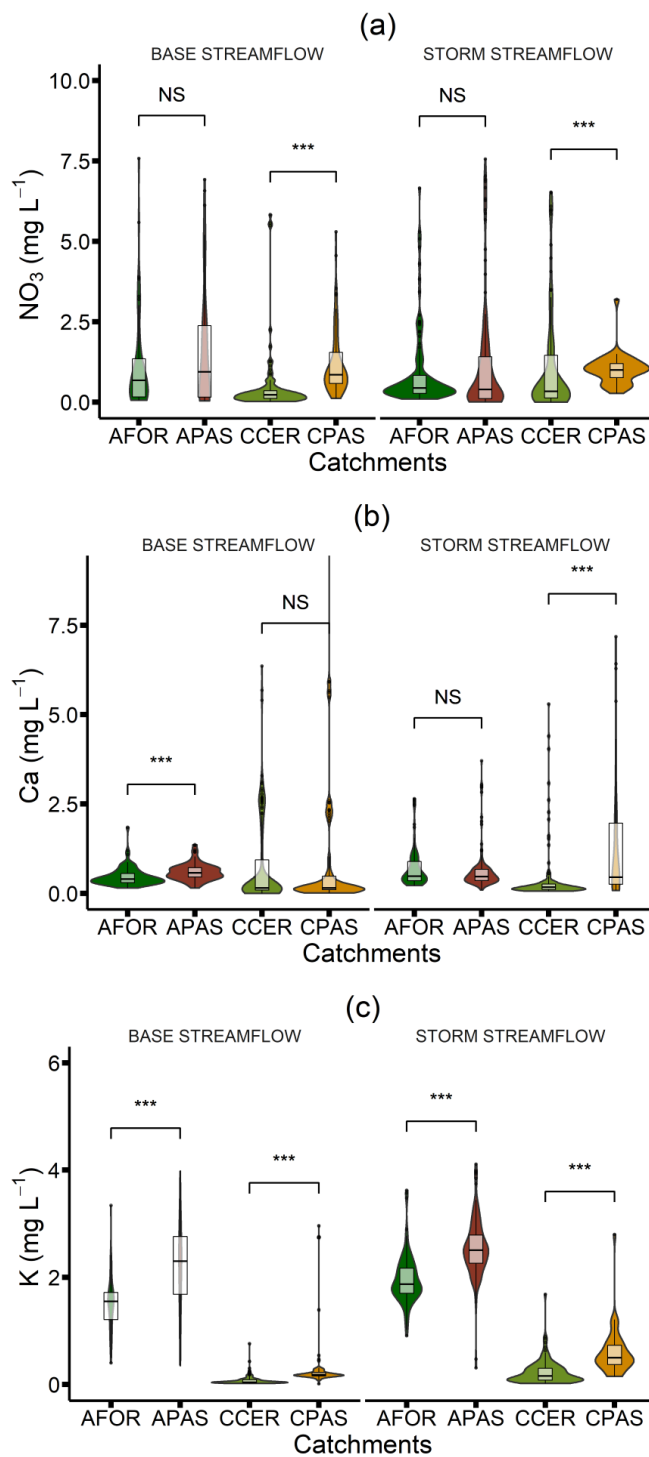
1  
 2 Figure 2. Biplots of the PCAs after varimax rotation for the first (C1) and second (C2) components of the: a) Amazon  
 3 catchments base streamflow ( $S_b$ ); b) Amazon catchments storm streamflow ( $S_s$ ); c) Cerrado catchments base  
 4 streamflow ( $S_b$ ); and d) Cerrado storm streamflow ( $S_s$ ).



1  
 2 Figure 3. Boxplot and violin plots of non-flow weighted carbon and nitrogen concentrations in base streamflow and  
 3 storm streamflow. The violin plots indicate the density of the sample distribution across the y-values. The y-axis was  
 4 limited to exclude some outliers (only graphically) for better visualization of the results. NS stands for not significant



1 and \*, \*\* and \*\*\* indicate statistical significance at the .05, .01 and .001 probability levels, respectively. The significance  
 2 of the results was based on the MW and Mood tests. When the test type is not indicated, the result is valid for both  
 3 tests.



4  
 5 Figure 4. Boxplot and violin plots of  $\text{NO}_3$ , Ca and K non-flow weighted concentrations in base streamflow and storm  
 6 streamflow. The violin plots indicate the density of the sample distribution across the y-values. The y-axis was limited

1 to exclude some outliers (only graphically) for better visualization of the results. NS stands for not significant and \*, \*\*  
2 and \*\*\* indicate the statistical significance at the .05, .01 and .001 probability levels, respectively. The significance  
3 results were based on the MW and Mood tests. When the test type is not indicated, the result is valid for both tests.

4

#### 5 4.3. Hydrological and CAN output fluxes

6 The Amazonian catchments exhibited the greater annual average stream discharge with  
7  $23.2 \text{ L s}^{-1}$  for the AFOR catchment and  $18.3 \text{ L s}^{-1}$  for the APAS catchment, whereas the  
8 stream discharge for the Cerrado catchments were  $11.6 \text{ L s}^{-1}$  for the CCER catchment  
9 and  $13.4 \text{ L s}^{-1}$  for the CPAS catchment. The average stream discharge during stormflow  
10 events were  $94.2 \text{ L s}^{-1}$  for the AFOR catchment,  $89.5$  for the APAS catchment,  $11.6 \text{ L s}^{-1}$   
11 for the CCER catchment and  $30.9 \text{ L s}^{-1}$  for the CPAS catchment.

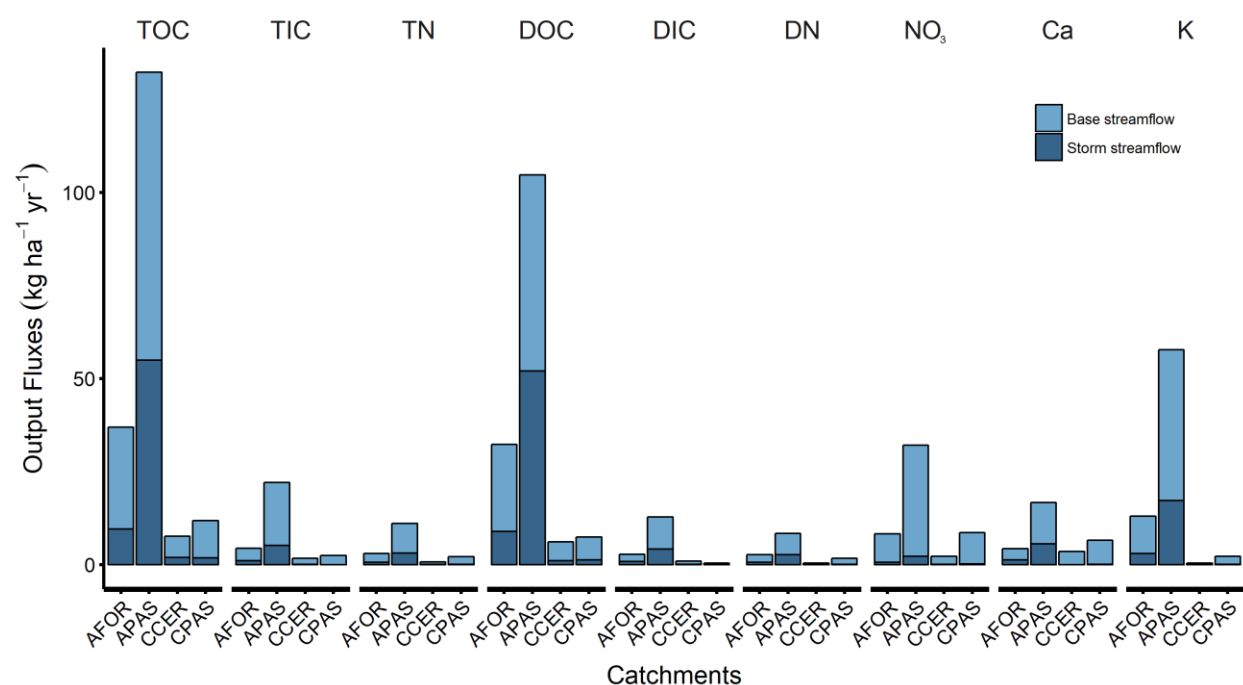
12 In the Amazonian catchments, TOC output fluxes were between  $35$  and  $135 \text{ kg ha}^{-1} \text{ yr}^{-1}$ ,  
13 and K and  $\text{NO}_3$  values ranged from  $8$  to  $60 \text{ kg ha}^{-1} \text{ yr}^{-1}$  (Fig. 5). In the Cerrado catchments,  
14 TOC, Ca and  $\text{NO}_3$  had total output fluxes between  $2$  and  $12 \text{ kg ha}^{-1} \text{ yr}^{-1}$ , and DIC and DN  
15 had output fluxes less than  $2 \text{ kg ha}^{-1} \text{ yr}^{-1}$ . Although the two biomes show different  
16 magnitudes of CAN fluxes with higher fluxes in the Amazonian catchments, the  $S_b$  CAN  
17 fluxes were higher than those of the  $S_s$  in all catchments. Furthermore, the fluxes in the  
18 pasture catchments were generally higher compared to those of the native vegetation  
19 catchments.

## 20 5. Discussion

### 21 5.1. Stream hydrochemistry

22 Our results showed significantly higher CAN concentrations in the pasture catchments  
23 compared to those of the native vegetation catchments, especially for TIC, TN and K.  
24 Some other macronutrients (Mg, P and S) and micronutrients (F, Cl, Fe and Na) exhibited  
25 concentrations of  $< 1 \text{ mg L}^{-1}$  in all of the studied catchments. Our DOC results for the  
26 Amazonian streams are in accordance with other studies of  $S_b$  of major tributaries of the  
27 Amazon River (Moreira-Turcq et al., 2003; Tardy et al., 2005) and in  $S_s$  of small  
28 Amazonian streams (Johnson et al., 2006). Although stream hydrochemistry data are  
29 scarce in these regions, studies have reported low stream concentrations for nutrients in  
30 a forested catchment in the central Amazon (Zanchi et al., 2015) as well in natural and

1 disturbed catchments in the central and southwestern Cerrado (Silva et al., 2012, 2011).  
 2 For some nutrients, i.e. F and Fe, we attributed this to the absence of fertilizer application  
 3 in the pasture catchments during our study period and the poor soil nutrient conditions in  
 4 both regions, which is typical of Lixisols (Driessen and Deckers, 2001) and Arenosols  
 5 (Markewitz et al., 2006) because of their strongly weathered substrate. Additionally, the  
 6 highly weathered soils fix available nutrients, especially P, in the form of Fe and Al  
 7 sesquioxides (Uehara and Gillman, 1981). Indeed, the soils from all catchments exhibited  
 8 a high content of Al and Fe and, a characteristic often found in Amazon (dos Santos and  
 9 Alleoni, 2013; Quesada et al., 2011) and Cerrado soils (Buol, 2009).



10

11 Figure 5. Annual carbon and nutrient output fluxes of base streamflow (S<sub>b</sub>) and storm streamflow (S<sub>s</sub>).

12 Soil pH in the pasture catchments was higher than that in the native vegetation  
 13 catchments, which has also been reported in other studies in other regions of the Amazon  
 14 (Mazzetto et al., 2016) and Cerrado (Carvalho et al., 2007; Hunke et al., 2015b; Neufeldt  
 15 et al., 2002). This is owing to liming practices in the pasture catchments. Lime (CaCO<sub>3</sub>)  
 16 is often applied to acidic soils in these regions to increase soil pH (Couto et al., 1997;  
 17 Jepson et al., 2010; Moreira and Fageria, 2010). Therefore, Ca content was higher in the  
 18 soils of the pasture catchments than in the soils of the native vegetation catchments. The  
 19 pasture catchments exhibited significantly higher stream Ca concentrations, which

1 reported in in other studies in the Amazon (Biggs et al., 2002; Figueiredo et al., 2010) and  
2 Cerrado (Markewitz et al., 2011; Silva et al., 2011).

3 The significantly higher  $S_s$  Ca concentrations exhibited in the CPAS catchment compared  
4 to those of the CCER catchment indicates that liming practices are increasing Ca content  
5 in the topsoil of the CPAS catchment and facilitating the leaching of this element to the  
6 stream during stormflow events. Other studies have already reported that the high rainfall  
7 rates in the Cerrado are sufficient to solubilize and leach fertilizers such as Ca (Hunke et  
8 al., 2015a; Villela and Haridasan, 1994). Conversely, between the Amazonian  
9 catchments, the Ca concentrations in stream water were significantly higher in the APAS,  
10 but only in  $S_b$ . Such an enrichment of Ca in the  $S_b$  has been observed in other studies in  
11 Brazil (Da Silva et al., 1998; Gonzatto, 2014), and we attribute this to the slow percolation  
12 of the residual lime through the soil profile (Rowe, 1982). Because Lixisols are in an  
13 advanced weathering stage (Quesada et al., 2011) and characterized by a low cation  
14 exchange capacity (Driessen and Deckers, 2001), the percolating soil water carries the  
15 residual Ca, thereby increasing its concentration in the  $S_b$ . In contrast, during storm  
16 events, the surface runoff dilutes the Ca concentration in the  $S_b$ , resulting in similar  
17 concentrations between the Amazonian catchments. Biggs et al. (2002) found strong  
18 correlations between the soil exchangeable cation content and the concentration of  
19 stream solutes and suggested that pasture age may help explain the substantial variation  
20 in solute concentration responses to deforestation, especially for Ca. DIC presented  
21 dynamics similar to Ca; its differences within the Amazonian and Cerrado catchments  
22 occur in the same flow types, and they are grouped in the same components in all PCAs.  
23 We ascribe this to be a consequence of liming practices. As lime is applied, the  $\text{CaCO}_3$   
24 reacts with water, increasing the soil pH and producing  $\text{HCO}_3^-$ , which is one of the main  
25 DIC components and has been identified as a main driver of DIC fluxes in small streams  
26 in the Amazon (Cak et al., 2015; Johnson et al., 2006).

27 We found  $\text{NO}_3^-$  concentrations to be significantly different only between the Cerrado  
28 catchments, with higher values in the CPAS catchment. The increase in  $\text{NO}_3^-$   
29 concentrations due to deforestation in Amazonian streams are not as clear (Figueiredo  
30 et al., 2010; Silva et al., 2007; Williams and Melack, 1997) as they are in the Cerrado  
31 (Silva et al., 2011). It has been reported that the high percentage of mineralized N nitrified

1 in forests is the cause of a high potential for  $\text{NO}_3$  loss in soil solution and streamwater  
2 when these forests are cleared and burned (Neill et al., 2006; Vourlitis and Hentz, 2016),  
3 which has occurred in small catchments under recent or ongoing deforestation (Williams  
4 and Melack, 1997). The fact that we could not find this same relationship between the  
5  $\text{NO}_3$  concentrations of the Amazonian catchments is consistent with patterns of N cycling  
6 and N availability, which shows high soil solution  $\text{NO}_3$  concentrations in Amazonian  
7 forests (Neill et al., 2001). The Amazonian forest behaves rather similar to old and  
8 temperate forests, which present high nitrification rates and  $\text{NO}_3$  pool losses that occur  
9 under normal conditions (Aber et al., 1989; Neill et al., 2001; Stevens et al., 1994). These  
10 forests may become net sources of nitrogen, thereby causing  $\text{NO}_3$  leaching to streams  
11 (Aber et al., 1995).

## 12 5.2. Stream CAN output fluxes

13 Except for DIC in the Cerrado catchments, the CAN fluxes were greater in the pasture  
14 catchments (Table 4). The Amazonian catchments exhibited the greatest differences in  
15 CAN fluxes. In these catchments,  $S_s$  showed a greater difference between the APAS and  
16 AFOR catchments, with an average APAS:AFOR ratio 37% higher than that in  $S_b$ .  
17 Conversely, for the Cerrado catchments, the CPAS:CCER CAN ratios were, on average,  
18 56% less in  $S_s$  than in  $S_b$ . This is consistent with that fact that nutrients, especially K and  
19 Ca, have been shown to have higher stream fluxes in pastures than in forests in the  
20 Amazon (Germer et al., 2009; Williams and Melack, 1997) and Cerrado (Figueiredo et  
21 al., 2010; Silva et al., 2011).

22 The total and dissolved carbon stream outputs were higher from the pasture catchments.  
23 Strey et al. (2016) found that degraded pasture areas exhibit lower organic carbon (OC)  
24 content than that of areas with native vegetation in the Cerrado and Amazon biomes,  
25 which is likely connected to larger losses of forest-derived OC after deforestation. In these  
26 biomes, the reduced organic carbon due to native vegetation clearing for pasture has  
27 been shown to be associated with reduced aggregate stability (Longo et al., 1999), which,  
28 in turn, has resulted in degraded pasture soils storing less carbon than soils covered with  
29 natural vegetation (Fonte et al., 2014). This facilitates carbon leaching and, consequently,  
30 increases the TOC and DOC fluxes. Kindler et al. (2011) affirmed that the quantification

1 of DOC leaching from soil is crucial for the carbon balance. These authors found that  
2 losses of biogenic carbon from grasslands account for ca. 22% of the net ecosystem  
3 exchange, whereas leaching from forest sites hardly affects net ecosystem carbon  
4 balances. In the Amazon, the decreased soil carbon storage as a consequence of forest  
5 conversion to pastures has been reported to be directly correlated with pasture age  
6 (Asner et al., 2004). In the Cerrado, while well-managed pastures may sustain soil carbon  
7 content, most pastures in this biome are in advanced stages of degradation (Davidson et  
8 al., 2012). In this region, the sandy soils, such as the Arenosols, are commonly found and  
9 the decrease of their organic matter content owing to their increasingly use for agricultural  
10 practices (Speratti et al., 2017) is likely to increase the leaching of nutrients (Hunke et al.,  
11 2015a).

12 The results of C content and C:N ratios for the Amazonian catchments are in accordance  
13 with studies on primary forests and old pastures in the Amazon (McGrath et al., 2001).  
14 For the Cerrado catchments, the C:N ratios are also similar to other results for topsoil in  
15 areas with cerrado vegetation and pasture in this biome (Figueiredo et al., 2010; Neufeldt  
16 et al., 2002). Similar to C, N output fluxes were higher in the pasture catchments. In  
17 comparison to the Cerrado catchments, the Amazonian catchments exhibited a lower C:N  
18 ratio, which is typical for Oxisols in the uppermost horizon (Tardy et al., 2005), and has  
19 been identified as an important controlling factor of total ecosystem N retention. High C:N  
20 promotes N immobilization, reduces net nitrification and consequently contributes to  
21 greater N retention (Templer et al., 2012). This has direct implications for the net N fluxes  
22 in this region, as the atmospheric deposition of N ( $3.5\text{--}10\text{ kg N ha}^{-1}\text{ year}^{-1}$ ) (Bobbink et  
23 al., 2010; Salemi et al., 2015)) is exceeded by N output via streamflow in the APAS  
24 catchment. This indicates that the pastures in this region might be a sink for N, as has  
25 been found in other studies in the Amazon (e.g., Germer et al., 2009 and Salemi et al.,  
26 2015).

27 Our results show the importance of  $S_s$  as a significant contributor to  $S_t$  CAN fluxes in  
28 catchments of the Amazon and Cerrado biomes. To illustrate this, we provide the ratios  
29 between the short-lived events ( $S_s$ ) to the  $S_t$  duration, volume and CAN fluxes in Table 5.  
30 The  $S_s:S_t$  duration ratios were only 4.9–5.3% in the Amazonian catchments and 1.7–2.1%  
31 in the Cerrado catchments. Nevertheless, the relatively small durations of the  $S_s$  events

1 caused an increase of 15.9–26.5% and 2.8–5.5% in the  $S_t$  volume in the Amazonian and  
2 Cerrado catchments, respectively. Moreover, in nearly all cases the  $S_s$  contribution to the  
3  $S_t$  CAN output fluxes was greater than its contribution to the  $S_t$  volume. In the APAS  
4 catchment, 50% of the  $S_t$  DOC output fluxes were caused by  $S_s$ . In the Cerrado  
5 catchments,  $S_s$  fluxes accounted for 16–26% of the TOC total streamflow output fluxes,  
6 despite the  $S_s$  contribution to  $S_t$  volume of only approximately 2–5%. This shows that  $S_s$   
7 is especially important as a rapid hydrological pathway for CAN losses in areas on the  
8 AAF where deforestation reduces the infiltration capacity rates, which are in turn  
9 exceeded by the rainfall intensities, causing greater stormflow contributions  
10 (Zimmermann et al., 2006). The substantial contribution exhibited by  $S_s$  to  $S_t$  CAN fluxes  
11 is mainly owing to their higher CAN concentrations compared to those of  $S_b$ . These  
12 concentrations may be higher in  $S_s$  because of the rapid subsurface response in streams  
13 dominated by pre-event water, where a rapid mobilization of old water occurs (Kirchner,  
14 2003), and to surface flow paths that contribute to higher CAN concentrations (Johnson  
15 et al., 2006).

16 DIC also exhibits a rapid response during stormflows in wet tropical catchments under  
17 pristine rainforest and agriculture LULC (Bass et al., 2014). In the Amazonian catchments,  
18 we found that  $S_s$  represented slightly more than 30% of  $S_t$  DIC fluxes, with similar  $S_s:S_t$   
19 DIC fluxes between these catchments. In contrast,  $S_s$  DIC fluxes represented only 6% of  
20 the total output fluxes in the CCER catchment and 10% in the CPAS catchment.

21 While many recent studies showed insights of high-temporal monitoring schemes in areas  
22 with fairly easy access (e.g., close to urban centers accessed via paved roads) in Europe  
23 (e.g., Blaen et al., 2016; Cuomo and Guida, 2016) and North America (e.g., Jollymore et  
24 al., 2012; Sherson et al., 2015) as a valid and new approach to ensure appropriate  
25 management of the natural resources (Skeffington et al., 2015), our study uses this  
26 method to assess the impacts of LULC change in catchments located in data-scarce  
27 active zones of deforestation of the two largest biomes of South America.

28 Despite the contribution of our study contributes to the understanding of the  
29 hydrochemical fluxes on the AFF, the magnitude and duration of these impacts depend  
30 on several catchments characteristics (e.g., soils, morphology and geology) that should

1 also be addressed in further studies (Birkinshaw et al., 2010). Long-term measurements  
2 (over 10 years) of stormflow events including quantifying changes in groundwater quality  
3 are required to analyze trends in water quality. Biggs et al. (2006) found evidence of long-  
4 term increases in solute fluxes following the conversion of forest to pasture in the Amazon.  
5 Hence, empirical studies that contemplate the comparison of pastures with different ages  
6 are fundamental to quantify the effect pasture age in CAN fluxes.

7 The degree to which the chemical changes of the streamwater in the Amazon and  
8 Cerrado biomes are affecting the CAN delivery to the ocean is poorly understood and  
9 difficult to assess (Bouchez et al., 2014). Notwithstanding, the changes in stream  
10 hydrochemistry are likely to unfold greater impacts due to several large dams under  
11 construction in this region (Pavanato et al., 2016; Tollefson, 2015), which will receive and  
12 store the increased loads of CAN and negatively affect their suitability as aquatic habitats.  
13 To that end, we recommend studies that take into account the long-term effects of LULC  
14 change on stream hydrochemistry in nested scales and their impacts in large watershed  
15 systems in this region.

## 16 6. Conclusions

17 Our research demonstrates how the conversion of natural vegetated landscapes (forest  
18 and cerrado) to pasture changes stream hydrochemistry, which can disturb the natural  
19 carbon and nutrient balance in the Amazon and Cerrado biomes. Stream carbon and  
20 nutrient concentrations were significantly higher in catchments where the native  
21 vegetation was replaced by pastures. These higher concentrations underlie further  
22 implications for carbon and nutrient fluxes as streamflow increase occurs, which is widely  
23 reported in this region as a consequence of the conversion of native vegetation into  
24 agricultural lands.

25 We found that most of the carbon and nutrient flux contributions of stormflow to total  
26 streamflow is proportionately greater than its respective volumetric contribution to stream  
27 discharge. This shows that stormflow is a substantial hydrological pathway for carbon and  
28 nutrient losses, including areas with small stormflow contribution, as shown in the Cerrado  
29 catchments. This indicates that the unaccounted stream carbon and nutrient fluxes  
30 derived from sampling approaches on a daily or weekly basis are substantially great. Our



1 study confirms the need for detailed temporal data on stream hydrochemistry that include  
2 the sampling of short-lived stormflow events to not only to understand natural tropical  
3 ecosystems, but also to unveil impacts of anthropogenic changes in these environments.

4 Although the acquisition of high-temporal resolution data in tropical forests is often limited  
5 by logistical restraints, we recommend that further studies use novel monitoring  
6 techniques such as automatic overland flow sampling and real-time water-quality sensors  
7 to improve the understanding of hydrochemical pathways and fluxes in forest ecosystems  
8 under anthropogenic changes such as the Amazonian agricultural frontier.

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Table 1. Main characteristics of the catchments.

	Amazonian catchments		Cerrado catchments	
	AFOR	APAS	CCER	CPAS
<b>Biome</b>	Amazon		Cerrado	
<b>Area (ha)</b>	93.4	23.1	77.8	58.4
<b>Mean precipitation (mm yr<sup>-1</sup>)</b>	1,900		1,700	
<b>Wet season</b>	Nov–May		Oct–Apr	
<b>Farm property</b>	<i>Paraiso farm</i>		<i>Rancho do Sol farm</i>	<i>Gianetta farm</i>
<b>Coordinates</b>	7.032° S, 55.363° W	7.023° S, 55.375° W	15.797° S, 55.332° W	15.805° S, 55.336° W
<b>Soil classification (IUSS Working Group WRB, 2015, and Soil Survey Staff, 2014)</b>	Lixisols, Oxisols		Arenosols, Entisols Quartzipsamments	
<b>Predominant land cover</b>	Rainforest	Pasture	Cerrado sensu stricto	Pasture
<b>Aspect</b>			E-W	
<b>Average slope (%)</b>	23.6	7.5	8.4	7.7
<b>Average elevation (m, above mean sea level)</b>	292.4	223.0	811.1	817.8

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1 Table 2. Mean, one standard deviation and sample size (*n*) of soil physical and  
 2 chemical properties.

Soil properties	Amazonian catchments		Cerrado catchments	
	AFOR	APAS	CCER	CPAS
<b>Sand (%)</b>	67.2 ± 6.0 (8)	57.6 ± 6.4 (8)	81.1 ± 20.5 (6)	93.3 ± 1.0 (8)
<b>Silt (%)</b>	9.1 ± 3.9 (8)	22.8 ± 6.0 (8)	6.1 ± 7.3 (6)	1.5 ± 0.4 (8)
<b>Clay (%)</b>	23.7 ± 6.1 (8)	19.6 ± 5.5 (8)	14.0 ± 13.4 (6)	5.2 ± 0.7 (8)
<b>pH</b>	5.7 ± 0.3 (3) <sup>a</sup>	6.4 ± 0.7 (3) <sup>a</sup>	3.6 ± 0.3 (6) <sup>c</sup>	4.4 ± 0.5 (8) <sup>d</sup>
<b>C (%)</b>	3.19 ± 2.54 (5) <sup>a</sup>	1.47 ± 0.45 (6) <sup>a</sup>	3.41 ± 3.88 (6) <sup>c</sup>	1.33 ± 1.01 (8) <sup>c</sup>
<b>N (%)</b>	0.27 ± 0.22 (5) <sup>a</sup>	0.12 ± 0.04 (6) <sup>a</sup>	0.18 ± 0.20 (6) <sup>c</sup>	0.07 ± 0.05 (8) <sup>c</sup>
<b>C:N ratio</b>	11.9 ± 1.8	11.8 ± 0.5	17.9 ± 2.4	18.3 ± 3.3
<b>Al (g kg<sup>-1</sup>)</b>	57.8 ± 16.3 (8) <sup>a</sup>	43.1 ± 19.2 (8) <sup>a</sup>	26.5 ± 23.4 (6) <sup>c</sup>	16.1 ± 3.4 (8) <sup>c</sup>
<b>Ca (g kg<sup>-1</sup>)</b>	1.0 ± 0.6 (8) <sup>a</sup>	0.5 ± 0.2 (8) <sup>a</sup>	<0.1 ± <0.1 (6) <sup>c</sup>	0.2 ± 0.1 (8) <sup>d</sup>
<b>Fe (g kg<sup>-1</sup>)</b>	15.5 ± 6.1 (8) <sup>a</sup>	11.5 ± 6.8 (8) <sup>a</sup>	10.8 ± 4.6 (6) <sup>c</sup>	13.2 ± 6.8 (8) <sup>c</sup>
<b>K (g kg<sup>-1</sup>)</b>	3.0 ± 2.2 (8) <sup>a</sup>	5.6 ± 3.4 (8) <sup>b</sup>	1.0 ± 1.4 (6) <sup>c</sup>	0.1 ± <0.1 (8) <sup>c</sup>
<b>Mg (g kg<sup>-1</sup>)</b>	0.4 ± 0.2 (8) <sup>a</sup>	0.8 ± 0.5 (8) <sup>b</sup>	0.1 ± 0.2 (6) <sup>c</sup>	0.1 ± 0.1 (8) <sup>c</sup>
<b>Mn (g kg<sup>-1</sup>)</b>	0.8 ± 1.0 (8) <sup>a</sup>	0.2 ± 0.2 (8) <sup>b</sup>	<0.1 ± <0.1 (6) <sup>c</sup>	<0.1 ± <0.1 (8) <sup>c</sup>
<b>P (g kg<sup>-1</sup>)</b>	0.2 ± 0.1 (8) <sup>a</sup>	0.2 ± 0.1 (8) <sup>a</sup>	0.2 ± 0.2 (6) <sup>c</sup>	0.1 ± <0.1 (8) <sup>c</sup>
<b>S (g kg<sup>-1</sup>)</b>	0.2 ± 0.1 (8) <sup>a</sup>	0.2 ± 0.1 (8) <sup>a</sup>	0.2 ± 0.2 (6) <sup>c</sup>	0.1 ± <0.1 (8) <sup>c</sup>

3 Significant differences ( $p < .05$ ) are indicated by different letters. Comparisons were  
 4 performed between catchments within the same biome.

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2 Table 3. Correlations between variables and components after varimax rotation.

	Amazonian catchments					Cerrado catchments				
	S <sub>b</sub>		S <sub>s</sub>			S <sub>b</sub>		S <sub>s</sub>		
	C1	C2	C1	C2	C3	C1	C2	C1	C2	C3
TC	<b>.92</b>	.27	<b>.99</b>	.07	.07	<b>.98</b>	-.02	.32	.25	<b>.90</b>
TIC	.12	<b>.88</b>	.07	<b>.95</b>	-.17	<b>.94</b>	-.12	.00	<b>.99</b>	.05
TOC	<b>.95</b>	.05	<b>.99</b>	.02	.08	<b>.77</b>	.11	.33	.06	<b>.92</b>
TN	<b>.81</b>	.30	.12	.10	<b>.92</b>	-.04	<b>.96</b>	.49	.01	<b>.75</b>
DC	<b>.88</b>	.19	<b>.99</b>	.12	.01	<b>.96</b>	-.24	<b>.74</b>	.36	.41
DIC	.01	<b>.93</b>	.07	<b>.95</b>	-.25	<b>.94</b>	-.12	.01	<b>.99</b>	.07
DOC	<b>.91</b>	-.05	<b>1.00</b>	.07	.03	<b>.79</b>	-.35	<b>.79</b>	.01	.41
DN	<b>.85</b>	.19	.09	-.14	<b>.95</b>	-.03	<b>.92</b>	<b>.77</b>	-.05	.33
NO <sub>3</sub>	-	-	-.12	-.40	<b>.56</b>	-.16	<b>.74</b>	<b>.87</b>	.03	.12
Ca	.22	<b>.82</b>	-.02	<b>.92</b>	-.01	<b>.93</b>	-.06	.12	<b>.97</b>	.13
K	.20	<b>.79</b>	.17	<b>.56</b>	.37	-	-	<b>.87</b>	.05	.29
Eigenvalue	5.5	2.5	4.3	3.2	2.0	6.0	2.3	5.8	2.9	1.0
Variability (%)	48.2	31.7	36.6	28.8	20.9	57.7	25.4	34.0	28.4	25.4

3 Correlations between variables and components greater than .5 are bolded.

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1 Table 4. Base streamflow, storm streamflow and total streamflow ratios of stream output  
 2 fluxes for each pair of catchments.

Ratio	Flow type	TOC	TIC	TN	DOC	DIC	DN	NO <sub>3</sub>	Ca	K
<b>APAS:AFOR</b>	Base streamflow	2.8	5.0	3.4	2.3	4.5	2.8	3.9	3.6	4.1
<b>APAS:AFOR</b>	Storm streamflow	5.8	5.0	4.7	5.8	4.8	4.4	3.8	4.6	5.7
<b>APAS:AFOR</b>	Total streamflow	3.6	5.0	3.7	3.2	4.6	3.2	3.9	3.8	4.4
<b>CPAS:CCER</b>	Base streamflow	1.8	1.5	3.3	1.2	0.4	4.0	3.8	1.8	6.8
<b>CPAS:CCER</b>	Storm streamflow	1.0	0.7	1.2	1.1	0.6	1.7	2.7	2.8	1.4
<b>CPAS:CCER</b>	Total streamflow	1.6	1.4	3.0	1.2	0.4	3.7	3.7	1.8	5.5

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1 Table 5. Percentage ratio of the storm streamflow duration, volume and fluxes to the  
 2 total streamflow.

Catchment	S <sub>s</sub> :S <sub>t</sub> (CAN fluxes)										
	S <sub>s</sub> :S <sub>t</sub> (duration)	S <sub>s</sub> :S <sub>t</sub> (volume)	TOC	TIC	TN	DOC	DIC	DN	NO <sub>3</sub>	Ca	K
<b>AFOR</b>	4.9%	15.9%	26%	24%	23%	28%	31%	23%	7%	29%	23%
<b>APAS</b>	5.3%	26.5%	42%	23%	28%	50%	33%	32%	7%	34%	30%
<b>CCER</b>	2.0%	5.2%	26%	3%	14%	18%	6%	12%	4%	2%	24%
<b>CPAS</b>	1.6%	2.8%	16%	2%	6%	17%	10%	6%	3%	2%	6%

3

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Table A.1. Descriptive statistics of the base streamflow hydrochemistry<sup>a</sup>.

Parameter	Amazonian catchments														Cerrado catchments													
	AFOR							APAS							CCER							CPAS						
(mg L <sup>-1</sup> )	N	min	max	median	mean	sd	vc	n	min	max	median	mean	sd	vc	n	min	max	median	mean	sd	vc	n	min	max	median	mean	sd	vc
TC	75	1.18	12.62	4.04	4.67	2.29	0.49	96	1.17	10.27	4.67	5.12	1.90	0.37	126	0.48	5.46	1.19	1.65	1.17	0.70	86	0.19	13.81	1.04	1.78	1.89	1.06
TIC	75	< LOD <sup>b</sup>	1.33	0.50	0.51	0.30	0.59	96	< LOD <sup>b</sup>	2.21	0.86	0.92	0.51	0.56	126	< LOD <sup>b</sup>	3.37	0.03	0.38	0.66	1.75	86	< LOD <sup>b</sup>	3.23	< LOD <sup>b</sup>	0.35	0.74	2.11
TOC	75	1.18	11.78	3.50	4.16	2.18	0.52	96	1.17	9.63	3.63	4.20	1.74	0.41	126	0.48	3.42	1.10	1.28	0.62	0.48	86	0.19	13.81	0.97	1.43	1.66	1.15
TN	75	0.18	1.55	0.27	0.35	0.21	0.58	96	0.18	1.00	0.36	0.43	0.19	0.45	126	< LOD <sup>b</sup>	0.55	0.18	0.14	0.09	0.62	86	0.11	0.88	0.26	0.29	0.12	0.42
DC	73	0.48	9.76	3.54	3.83	1.99	0.51	95	0.70	6.51	3.12	3.33	1.34	0.40	82	0.01	5.58	1.00	1.37	1.13	0.82	53	0.20	4.23	0.71	0.97	0.88	0.89
DIC	73	< LOD <sup>b</sup>	1.44	0.23	0.29	0.34	1.16	95	< LOD <sup>b</sup>	2.08	0.25	0.47	0.49	1.06	101	< LOD <sup>b</sup>	3.19	0.00	0.20	0.59	2.93	73	< LOD <sup>b</sup>	1.40	< LOD <sup>b</sup>	0.05	0.23	4.53
DOC	73	< LOD <sup>b</sup>	9.76	3.29	3.54	1.95	0.55	95	< LOD <sup>b</sup>	5.76	2.84	2.86	1.21	0.42	82	0.10	3.70	1.00	1.14	0.59	0.52	53	0.20	3.62	0.71	0.89	0.73	0.81
DN	41	0.18	0.73	0.27	0.31	0.14	0.43	37	0.18	0.65	0.27	0.31	0.11	0.37	62	< LOD <sup>b</sup>	0.28	< LOD <sup>b</sup>	0.09	0.09	1.08	16	0.10	0.48	0.20	0.23	0.09	0.37
F	75	0.01	0.09	0.02	0.02	0.01	0.43	95	0.01	0.20	0.04	0.04	0.02	0.53	114	< LOD <sup>b</sup>	0.64	0.01	0.05	0.11	2.03	88	< LOD <sup>b</sup>	1.18	0.03	0.12	0.21	1.82
Cl	75	0.17	0.79	0.43	0.45	0.15	0.32	95	0.10	2.03	0.44	0.55	0.32	0.57	119	0.04	2.81	0.19	0.39	0.48	1.22	88	0.10	5.18	0.27	0.62	0.81	1.30
NO <sub>3</sub>	51	0.06	7.58	0.68	1.16	1.52	1.29	66	0.04	6.92	0.94	1.62	1.84	1.13	90	0.02	5.83	0.23	0.50	1.03	2.03	77	0.12	5.30	0.85	1.20	1.01	0.84
SO <sub>4</sub>	70	< LOD <sup>b</sup>	0.63	0.04	0.08	0.10	1.29	87	< LOD <sup>b</sup>	0.34	0.04	0.06	0.05	0.93	119	< LOD <sup>b</sup>	0.50	0.06	0.08	0.08	0.95	88	< LOD <sup>b</sup>	0.74	0.06	0.11	0.13	1.18
Ca	75	0.15	1.85	0.40	0.47	0.26	0.56	95	0.15	1.36	0.57	0.60	0.24	0.40	126	< LOD <sup>b</sup>	6.36	0.15	0.79	1.26	1.58	87	0.01	15.54	0.15	0.92	2.13	2.29
Fe	75	< LOD <sup>b</sup>	0.11	< 0.01	0.01	0.02	1.54	95	< LOD <sup>b</sup>	0.06	< 0.01	0.01	0.01	1.73	126	< LOD <sup>b</sup>	0.05	< 0.01	< 0.01	0.01	3.18	87	< LOD <sup>b</sup>	0.09	< 0.01	< 0.01	0.01	4.78
K	75	0.40	3.34	1.55	1.51	0.50	0.33	95	0.35	3.98	2.30	2.20	0.81	0.36	126	0.02	0.76	0.04	0.07	0.09	1.16	87	0.01	2.96	0.18	0.30	0.50	1.64
Mg	75	0.03	0.40	0.10	0.12	0.06	0.50	95	0.03	0.42	0.15	0.16	0.07	0.42	126	0.01	0.56	0.05	0.07	0.07	0.98	87	0.01	0.35	0.06	0.07	0.06	0.81
Na	75	0.24	1.36	0.90	0.89	0.25	0.28	95	0.21	1.65	0.93	0.90	0.31	0.34	125	< LOD <sup>b</sup>	0.73	0.10	0.16	0.13	0.86	87	< LOD <sup>b</sup>	1.40	0.23	0.27	0.16	0.59
P	75	< LOD <sup>b</sup>	0.11	0.04	0.04	0.03	0.78	95	< LOD <sup>b</sup>	0.15	0.03	0.03	0.04	1.03	126	< LOD <sup>b</sup>	0.09	< 0.01	0.01	0.02	1.92	87	< LOD <sup>b</sup>	0.20	< 0.01	0.02	0.04	1.92
S	75	< LOD <sup>b</sup>	0.27	0.03	0.05	0.05	1.07	95	< LOD <sup>b</sup>	0.19	0.04	0.05	0.03	0.66	126	< LOD <sup>b</sup>	0.06	< 0.01	0.01	0.01	1.63	87	< LOD <sup>b</sup>	0.21	< 0.01	0.01	0.04	2.51

<sup>a</sup> The results of the base streamflow chemistry are related to sampling routines performed from 04/2013 to 07/2014 in the Amazonian catchments and from 12/2012 to 07/2014 in the Cerrado catchments.

<sup>b</sup> LOD stands for limit of detection.



Table A.2. Descriptive statistics of the storm streamflow hydrochemistry<sup>a</sup>.

Parameter (mg L <sup>-1</sup> )	Amazonian catchments														Cerrado catchments													
	AFOR							APAS							CCER					CPAS								
	n	min	max	median	mean	sd	vc	n	min	max	median	mean	sd	vc	n	min	max	median	mean	sd	vc	n	min	max	median	mean	sd	vc
<b>TC</b>	108	1.56	25.80	6.08	7.39	4.91	0.66	160	2.63	96.80	7.04	8.59	9.71	1.13	119	0.77	24.90	3.57	4.27	3.16	0.74	43	0.50	20.02	7.00	7.47	3.98	0.53
<b>TIC</b>	108	0.08	2.20	0.35	0.53	0.47	0.87	160	< LOD <sup>b</sup>	2.70	0.52	0.64	0.49	0.76	119	< LOD <sup>b</sup>	3.79	< LOD <sup>b</sup>	0.17	0.58	3.44	43	< LOD <sup>b</sup>	4.00	0.08	0.64	1.11	1.73
<b>TOC</b>	108	1.38	25.01	5.50	6.86	4.81	0.70	160	2.63	95.50	6.29	7.95	9.66	1.21	119	0.77	23.10	3.47	4.10	3.00	0.73	43	0.50	18.27	6.50	6.84	3.88	0.56
<b>TN</b>	108	0.18	1.82	0.40	0.46	0.24	0.53	160	0.22	1.30	0.50	0.49	0.17	0.35	119	0.10	1.50	0.27	0.27	0.18	0.65	43	0.20	3.10	0.50	0.61	0.48	0.79
<b>DC</b>	93	1.94	27.30	5.35	6.73	4.41	0.65	148	1.12	98.60	5.18	6.94	10.58	1.52	119	0.80	10.20	2.90	3.26	1.73	0.53	38	3.30	11.40	6.21	6.50	1.96	0.30
<b>DIC</b>	46	< LOD <sup>b</sup>	2.10	0.34	0.52	0.56	1.06	125	< LOD <sup>b</sup>	2.60	0.30	0.45	0.51	1.14	115	< LOD <sup>b</sup>	2.25	< LOD <sup>b</sup>	0.12	0.40	3.43	41	< LOD <sup>b</sup>	3.90	< LOD <sup>b</sup>	0.62	1.10	1.75
<b>DOC</b>	93	1.21	26.30	4.87	6.13	4.33	0.70	148	1.12	97.60	4.73	6.47	10.49	1.61	119	0.80	8.22	2.80	3.13	1.62	0.51	38	2.10	10.90	5.45	5.81	2.03	0.34
<b>DN</b>	91	0.18	1.46	0.36	0.42	0.23	0.55	117	0.27	0.90	0.40	0.42	0.15	0.34	65	< LOD <sup>b</sup>	0.91	0.18	0.22	0.11	0.49	35	0.10	2.10	0.40	0.49	0.37	0.75
<b>F</b>	109	0.01	3.62	0.02	0.07	0.35	5.03	159	0.01	0.10	0.03	0.03	0.01	0.42	119	< LOD <sup>b</sup>	0.33	0.01	0.01	0.03	2.93	36	< LOD <sup>b</sup>	1.23	0.04	0.19	0.30	1.51
<b>Cl</b>	109	0.35	16.05	0.53	0.81	1.53	1.88	159	0.08	4.95	0.60	0.63	0.40	0.64	119	0.06	4.20	0.17	0.28	0.42	1.50	36	0.20	3.65	0.59	0.93	0.90	0.96
<b>NO<sub>3</sub></b>	107	0.10	6.66	0.44	0.93	1.21	1.29	142	0.01	7.56	0.40	1.18	1.74	1.48	109	< LOD <sup>b</sup>	6.53	0.34	1.09	1.62	1.48	35	0.27	3.20	1.00	1.02	0.50	0.48
<b>SO<sub>4</sub></b>	107	0.01	1.03	0.07	0.12	0.16	1.26	159	0.01	0.55	0.07	0.09	0.07	0.82	117	0.02	0.62	0.05	0.07	0.07	0.97	36	0.04	0.38	0.11	0.14	0.09	0.67
<b>Ca</b>	109	0.22	2.65	0.48	0.70	0.53	0.77	160	0.09	3.71	0.47	0.61	0.54	0.88	118	0.06	5.30	0.17	0.41	0.84	2.02	42	0.08	7.18	0.45	1.43	1.88	1.30
<b>Fe</b>	109	< LOD <sup>b</sup>	0.06	0.01	0.01	0.02	1.04	160	< LOD <sup>b</sup>	0.23	0.03	0.03	0.03	1.02	119	< LOD <sup>b</sup>	0.11	0.01	0.02	0.02	1.09	42	< LOD <sup>b</sup>	0.05	< 0.01	0.01	0.02	1.75
<b>K</b>	109	0.91	3.62	1.87	1.96	0.46	0.23	160	0.31	4.11	2.51	2.54	0.53	0.21	118	0.02	1.68	0.16	0.23	0.23	0.98	42	0.15	2.80	0.50	0.60	0.45	0.73
<b>Mg</b>	109	0.04	0.30	0.12	0.14	0.06	0.40	160	0.02	0.26	0.12	0.14	0.05	0.35	118	0.03	2.36	0.08	0.12	0.22	1.81	42	0.04	0.42	0.08	0.11	0.07	0.65
<b>Na</b>	109	0.56	1.95	0.92	0.96	0.22	0.23	160	0.14	1.18	0.76	0.72	0.23	0.33	118	0.05	1.57	0.11	0.22	0.22	1.01	42	0.15	1.62	0.27	0.41	0.30	0.72
<b>P</b>	109	< LOD <sup>b</sup>	0.11	< LOD <sup>b</sup>	0.02	0.03	1.45	160	< LOD <sup>b</sup>	0.14	0.01	0.04	0.04	1.13	119	< LOD <sup>b</sup>	0.11	< 0.01	0.02	0.03	1.39	42	< LOD <sup>b</sup>	0.09	< 0.01	0.02	0.03	1.82
<b>S</b>	109	< LOD <sup>b</sup>	0.52	0.05	0.07	0.08	1.18	160	< LOD <sup>b</sup>	0.21	0.07	0.07	0.05	0.78	119	< LOD <sup>b</sup>	0.26	0.02	0.03	0.03	1.18	42	< LOD <sup>b</sup>	0.09	< 0.01	0.01	0.03	1.76

<sup>a</sup> The results of the storm streamflow chemistry are related to sampling obtained from 02/2013 to 02/2014 in the Amazon and Cerrado catchments.

<sup>b</sup> LOD stands for limit of detection.

