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2 **Consistent Controls on Trace Metal Micronutrient Speciation in**  
3 **Wetland Soils and Stream Sediments**  
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45 **ABSTRACT**

46 Trace metal are essential for microbially-mediated biogeochemical processes occurring in  
47 anoxic wetland soils and stream bed sediments, but low availability of these elements may inhibit  
48 anaerobic element cycling and transformations. Solid-phase speciation is likely a critical control  
49 on trace metal availability but has seen limited study in anoxic systems having concentrations  
50 similar to geological background levels, where metal limitations may be most prevalent. We have  
51 investigated trace metal concentrations and solid-phase speciation in three freshwater subsurface  
52 aquatic systems: marsh wetland soils, riparian wetland soils, and the sediments of a streambed.  
53 These systems displayed low solid-phase trace metal concentrations, generally at or below  
54 geological background levels, which generally followed the trend  $Zn > Cu \approx Ni > Co$  and showed  
55 no correlation with major element compositions. All soils and sediments were dominated by quartz  
56 but varied in clay mineralogy as well as the organic matter, total sulfur, and total iron contents. X-  
57 ray absorption near-edge structure (XANES) spectroscopy shows that sulfur speciation in both  
58 wetlands is dominated by organic sulfur. Elemental sulfur and iron sulfides together made up  
59 <25% of the sulfur in the wetland soils, but the distribution between inorganic and organic forms  
60 was reversed in the stream sediments. Ferrous and ferric iron in clay minerals were common  
61 species identified by both XANES and extended X-ray absorption fine structure (EXAFS)  
62 spectroscopies at all sites. Iron(III) oxides were substantial components in all but the marsh  
63 wetland soils. Quantitative analysis of copper, nickel, and zinc XANES spectra revealed similar  
64 metal speciation across all sites. Copper speciation was dominated by sulfides, adsorbed species,  
65 and minor amounts of copper bound to organic matter; no metallic copper was detected. Nickel  
66 speciation also varied little and was dominated by nickel in clay mineral octahedral sheets and  
67 nickel sulfide, with adsorbed species also present. Zinc speciation was slightly more varied, with

68 the marsh wetland soils and stream bed sediments containing adsorbed species, zinc associated  
69 with clay mineral structures, and zinc bound to reduced sulfur groups on organic matter, whereas  
70 the riparian wetland soils lacked clay-associated zinc but contained zinc sulfide. Trace metals  
71 bound to reduced sulfur occurred at every site, with a greater sulfur-bound fraction for copper. The  
72 fractional abundance of sulfur-bound species showed no relationship with soil or sediment total  
73 sulfur content, which varied by two orders of magnitude. More broadly, the observations in this  
74 study suggest that trace metal speciation in freshwater wetland soils and stream sediments is  
75 consistently dominated by a small set of recurring components which are distinct for each metal.  
76 This may represent a general geochemical phenomenon in anoxic soils and sediments containing  
77 trace metals at background concentrations (as low as  $3 \mu\text{g g}^{-1}$ ) that was not predicted from systems  
78 that are contaminated with or naturally-enriched in copper, nickel, or zinc.

79

## 80 **1. INTRODUCTION**

81           Subsurface aquatic systems in freshwater environments, such as wetland soils and stream  
82 sediments, are foci of diverse anaerobic biogeochemical processes (Bowden, 1987; McClain et al.,  
83 2003; Kocar and Fendorf, 2009; Zarnetske et al., 2011; Arora et al., 2016; Neumann et al., 2016).  
84 Denitrification and methanogenesis in freshwater aquatic systems serve as important sources to  
85 the atmosphere of the greenhouses gases N<sub>2</sub>O and CH<sub>4</sub>, respectively (Groffman et al., 1998;  
86 Glatzel et al., 2008; Bouwman et al., 2013; Kirschke et al., 2013; Tian et al., 2015; Zhang et al.,  
87 2017). Substantial mercury methylation and subsequent emission also occur in these settings (St  
88 Louis et al., 1994; Skjellberg, 2008; Schaefer et al., 2014; Riscassi et al., 2016; Singer et al., 2016;  
89 Yang et al., 2016). Many of these processes in subsurface aquatic systems also couple to the  
90 biogeochemical cycling of other elements, such as iron and sulfur (Koretsky et al., 2003;  
91 Hlaváčová et al., 2005; Hansel et al., 2015; Segarra et al., 2015; Glodowska et al., 2020).

92           While the biogeochemistry of aquatic systems has been widely studied from the  
93 perspective of redox conditions, substrate availability, and thermodynamic controls on metabolic  
94 processes (Falkowski et al., 1998; LaRowe and Van Cappellen, 2011; Flynn et al., 2014; Arora et  
95 al., 2016; Danczak et al., 2016; Janot et al., 2016), there is a growing recognition of the importance  
96 of trace metal availability in affecting biogeochemical processes (Basiliko and Yavitt, 2001; Glass  
97 and Orphan, 2012; Jacquot et al., 2014). Elements that include cobalt, nickel, copper, and zinc  
98 serve as key reaction centers in metalloenzymes (Gärtner et al., 1993; Ermler et al., 1997; Thauer,  
99 1998; Brown et al., 2000; Parks et al., 2013; Zheng et al., 2016) and their low availability has been  
100 documented in laboratory studies to inhibit methanogenesis, nitrous oxide reduction to nitrogen,  
101 and mercury methylation (Schönheit et al., 1979; Granger and Ward, 2003; Ekstrom and Morel,  
102 2008; Glass and Orphan, 2012; Lu et al., 2018). The geochemistry of trace metals in aquatic

103 systems may thus have a direct impact on biogeochemical cycling of carbon, nutrients, and  
104 contaminants.

105         The availability of trace metals in wetland soils and stream sediments is expected to be  
106 controlled by their chemical speciation rather than bulk concentration (Worms et al., 2006;  
107 Harmsen, 2007; Zhao et al., 2016). Past studies have identified an array of chemical forms of trace  
108 metals in similar types of anoxic settings, although these have often focused on locations  
109 containing elevated metal concentrations because of environmental contamination or geogenic  
110 enrichments. Of the trace metals, copper is the most widely studied in anoxic subsurface aquatic  
111 systems. Copper sulfides and metallic copper are generated in contaminated soils, often in  
112 nanoparticulate form, upon flooding (Weber et al., 2009a; Weber et al., 2009b; Fulda et al., 2013a;  
113 Hofacker et al., 2013b; Xia et al., 2018; Cervi et al., 2021), with organic matter then potentially  
114 stabilizing these phases under oxic conditions (Fulda et al., 2013b; Mantha et al., 2019). Copper  
115 sulfides were also observed in contaminated paddy soils (Yang et al., 2015; Sun et al., 2016) and  
116 a copper-rich bog (Lett and Fletcher, 1980). Binding to organic matter, including as Cu(I) species,  
117 may be important in contaminated, flooded soils with limited sulfate (Fulda et al., 2013a; Fulda et  
118 al., 2013b). A recent study examined uncontaminated transitional and anoxic soils and found  
119 copper binds extensively to organic matter but also forms copper sulfide and minor metallic copper  
120 in the anoxic zone (Mehlhorn et al., 2018).

121         Fewer studies have examined the speciation of other essential trace metals in subsurface  
122 aquatic systems. In diverse coastal sediments that drain ultramafic laterite deposits highly elevated  
123 in nickel, this metal occurs coprecipitated with iron sulfides and in the octahedral sheets of clay  
124 minerals (Noël et al., 2015; Noël et al., 2017; Merrot et al., 2019). While nickel speciation in other  
125 anoxic systems has seen limited study, contaminated or enriched oxic soils contain nickel in

126 phyllosilicate and iron oxide structures, associated with gibbsite phases in clay interlayers, and  
127 bound to organic matter (Manceau et al., 2005; McNear et al., 2007; Dublet et al., 2012; Siebecker  
128 et al., 2017; Siebecker et al., 2018). Zinc sulfide forms under anoxic conditions in metalliferous  
129 peatlands (Yoon et al., 2012), contaminated lake sediments (Webb and Gaillard, 2015), and in a  
130 contaminated wetland, along with zinc carbonate, adsorbed zinc, and possibly zinc oxide (Bostick  
131 et al., 2001). Similar to nickel, exploration of zinc in other anoxic soil systems is generally lacking,  
132 but in contaminated oxic soils zinc sulfide, zinc in phyllosilicate octahedral sheets, and adsorbed  
133 species are common (Manceau et al., 2004; Voegelin et al., 2005; Jacquat et al., 2009; Voegelin et  
134 al., 2011; Williams et al., 2011). Less is known regarding cobalt speciation because of the difficulty  
135 probing this element using available methods in the presence of iron. Binding studies suggest that  
136 adsorbed species and complexes with organic matter are important in aerobic soils (Woodward et  
137 al., 2018) and that cobalt readily incorporates into pyrite under anoxic conditions (Swanner et al.,  
138 2019).

139         Prior studies provide important insight into trace metal species in anoxic subsurface  
140 systems that are contaminated with or naturally elevated in metals. All metals show occurrence as  
141 sulfide phases, although some results, such as nickel associating with pyrite in coastal sediments,  
142 may not be transferable to freshwater systems that are generally low in sulfur (Brown, 1985;  
143 Wieder et al., 1985; Spratt and Morgan, 1990; Prietzel et al., 2009). Similarly, nickel and zinc  
144 often occur in clay structures, likely produced through a neoformation process (Manceau et al.,  
145 1999; Ford and Sparks, 2000; Schlegel et al., 2001; Dähn et al., 2002), but it is unclear whether  
146 such species form when metals are not elevated in concentration from contamination or natural  
147 enrichment. The solid-phase trace metal concentrations in most prior studies of metal speciation  
148 in subsurface aquatic systems (Lett and Fletcher, 1980; Bostick et al., 2001; Weber et al., 2006;

149 Weber et al., 2009b; Weber et al., 2009a; Yoon et al., 2012; Fulda et al., 2013b; Fulda et al., 2013a;  
150 Hofacker et al., 2015; Noël et al., 2015; Yang et al., 2015; Sun et al., 2016; Noël et al., 2017; Xia  
151 et al., 2018; Mantha et al., 2019; Merrot et al., 2019; Cervi et al., 2021) far exceed geological  
152 background levels (Rudnick and Gao, 2003): 280 to 23000  $\text{ug g}^{-1}$  Cu versus  $28\pm4 \text{ ug g}^{-1}$   
153 background, 255 to 6044  $\text{ug g}^{-1}$  Ni versus  $47\pm11 \text{ ug g}^{-1}$  background, 1900 to 71000  $\text{ug g}^{-1}$  Zn versus  
154  $67\pm6 \text{ ug g}^{-1}$  background. Investigations of speciation in wetland soils and stream sediments with  
155 metal concentrations similar to geological background levels are rare (Webb and Gaillard, 2015;  
156 Mehlhorn et al., 2018). However, such metal concentrations are widespread in nature (Caritat et  
157 al., 2018) and likely representative of the large majority of freshwater wetlands and streambeds  
158 where biogeochemical carbon, nitrogen, and mercury cycling occur in terrestrial ecosystems.  
159 Anoxic subsurface systems displaying background metal levels should more frequently exhibit  
160 metal limitations on biogeochemical processes than contaminated or naturally-enriched systems  
161 However, the dominant controls on metal speciation in environments with low metal contents are  
162 unclear. A central challenge to investigating metal speciation in relevant anoxic subsurface  
163 systems is the difficulty in applying X-ray spectroscopic techniques to soils and sediments having  
164 low ( $<100 \text{ ug g}^{-1}$ ) element concentrations.

165 In this work, we assess the variability in trace metal speciation in the subsurface of diverse  
166 freshwater aquatic systems. Three field sites are investigated that are distributed over a  $\sim 1000 \text{ km}$   
167 transect, each occurring in geologically-distinct regions representing different types of subsurface  
168 aquatic system: marsh wetland soils, riparian wetland soils, and stream bed sediments. Duplicate  
169 soil or sediment cores and overlying surface waters were collected in two locations at each of the  
170 three study areas. Major element and trace metal concentrations, dissolved and extractable  
171 nutrients, and soil and sediment mineralogy were evaluated at these sites. X-ray absorption

172 spectroscopy evaluated bulk sulfur and iron speciation and quantified the solid-phase speciation  
173 of nickel, copper, and zinc. The latter measurements overcame the low concentrations of trace  
174 metals by utilizing a synchrotron beamline configuration optimized for high-sensitivity  
175 measurements. Metal speciation was compared across sites to identify the dominant forms of trace  
176 metals occurring in subsurface aquatic systems.

177

## 178 **2. MATERIALS AND METHODS**

### 179 **2.1 Study Sites Descriptions**

180 Three field sites (Fig. S1) were investigated to explore diverse wetland and stream settings,  
181 each of which has been previously studied in other contexts (Van Lonkhuyzen and LaGory, 1994;  
182 Van Lonkhuyzen et al., 2004; Southworth et al., 2013; Donovan et al., 2014; Riscassi et al., 2016;  
183 Flynn et al., 2017; Schwartz et al., 2019; Kaplan et al., 2020). At each site, two locations were  
184 studied to examine the variability in metal speciation in similar settings. Marsh wetlands at  
185 Argonne National Laboratory (ANL) in Lemont, Illinois, U.S.A. were sampled in October 2018.  
186 Sampling location “Marsh 1” (41°42’19.69”N 87°59’55.04”W WGS 84) is an ~1800 m<sup>2</sup> marsh  
187 surrounded by deciduous forest. It contained shallow surface water at the time of sampling but  
188 becomes unsaturated during prolonged dry periods. Location “Marsh 2” (41°42’6.78”N  
189 87°59’39.66”W WGS 84) consists of a ~3700 m<sup>2</sup> marsh containing open water in the center  
190 surrounded by an ~15 m vegetated border; this site maintains water saturation throughout most  
191 years. Sampling locations Marsh 1 and 2 have internal designations as wetlands 401 and 405 at  
192 ANL (Van Lonkhuyzen and LaGory, 1994).

193 Sediments and surface waters were sampled at East Fork Poplar Creek (EFPC) in Oak  
194 Ridge, Tennessee, U.S.A. in October 2018. Sampling locations “Stream 1” (35°57’57.89”N



195 84°21'32.81"W WGS 84) and "Stream 2" (36°00'06.13"N 84°14'55.10"W WGS 84) are located  
196 approximately 15 km apart. Location Stream 1 is 5.4 km upstream of the mouth of the creek, and  
197 Stream 2 is in a broader stretch 22 to 23 km upstream from the mouth. Sediments were collected  
198 from the northwest or north stream bank for locations Stream 1 and Stream 2, respectively.  
199 Sampling occurred 25 days after the last significant rain event (rain >12 mm). EFPC is impacted  
200 by point source contamination with mercury, but not other metals, originating at its headwaters  
201 (Brooks and Southworth, 2011) and receives discharge from a wastewater treatment plant about  
202 midway between the two sampling locations.

203 The third study site is a riparian wetland in the Tims Branch (TB) watershed at the  
204 Savannah River Site in Aiken County, South Carolina, U.S.A. Samples were collected in February  
205 2019 at two locations, "Riparian 1" (33°20'15.36"N 81°43'5.88"W WGS 84) and "Riparian 2"  
206 (33°20'9.24"N 81°43'8.04"W WGS 84). These differed in their apparent local hydrology, with  
207 Riparian 2 showing evidence of shallow groundwater inputs at the time of sampling, which was  
208 not observed at the other location. This forested site varies in its water inundation throughout the  
209 year and in summer may lack surface water for short periods of time. Note that a section of the  
210 Tims Branch watershed 1 to 2 km downstream of the sampling locations received historical  
211 discharge of drainage waters containing metals, including uranium (Kaplan et al., 2020). This  
212 discharge was introduced through a tributary and did not impact the study areas.

213

## 214 **2.2. Soil and Water Sampling**

215 At each sampling site, surface water samples were collected prior to soil or sediment  
216 coring. Subsamples of the surface waters were filtered using 0.22 µm mixed cellulose ester (MCE)  
217 syringe filters. A portion of the filtered water was acidified to 2% nitric acid (HNO<sub>3</sub>) in the field

218 using concentrated trace metal grade HNO<sub>3</sub> for dissolved major elements and trace metal analysis.  
219 The rest of the filtered water was stored on ice and immediately divided and frozen or stored at 4°  
220 C upon return to the laboratory for nutrient and anion analysis, respectively. Field process blanks  
221 were prepared using ultrapure water (>18.2 MΩ cm) following the same procedures as the surface  
222 water samples to monitor possible metal contamination during the sampling. The pH, temperature,  
223 and conductivity of the surface waters were also measured during sampling at the marsh wetlands  
224 and stream sites using a Hanna Instruments HI98194 multiparameter meter. A meter malfunction  
225 prevented data collection at the riparian wetlands; temperature and pH data for these sampling  
226 locations are taken from measurements made in January 2019, a few weeks prior to soil and water  
227 sample collection. Measurements of pH made in August 2020 at the same sampling locations in  
228 the riparian wetlands were within 0.03 pH units of those measured in January 2019, suggesting  
229 that the pH values are temporally stable at these sites.

230 Duplicate soil and sediment samples from each of the two studied locations at the marsh  
231 wetland and stream sites were collected by hand-coring using 7.6 cm diameter acid-cleaned  
232 polycarbonate tubes, which were then capped and sealed in the field. Cores from the marsh site  
233 were placed in a cooler on ice and transported to Washington University, where they were extruded  
234 within six hours of sampling and then transferred to an anaerobic chamber (Coy Laboratory  
235 Products, 3% H<sub>2</sub>/97% N<sub>2</sub> with Pd catalyst). Cores from the stream site were extruded within one  
236 hour of sampling in a laboratory at Oak Ridge National Laboratory, transferred to an anaerobic  
237 chamber, and then impulse sealed in polyethylene pouches. An inner pouch contained each  
238 extruded sample, and this was then sealed in an outer pouch. The gas headspace in both pouches  
239 were that of the anaerobic chamber and an oxygen scavenging sachet (BD GasPak™ EZ Anaerobic  
240 Container System Sachets with Indicator) was placed in the outer pouch to minimize the risk of

241 leakage causing oxidation. These doubly-contained samples were removed from the anaerobic  
242 chamber after sealing, stored at 4°C, and then transported on ice to Washington University, where  
243 they were immediately transferred to an anaerobic chamber. Duplicate soil samples from each of  
244 the two study locations at the riparian wetland site were collected using 2.5 cm diameter  
245 polycarbonate tubes. These were sealed in the field, stored overnight at 4°C, and then shipped  
246 overnight to Washington University packaged with reusable ice packs, where they were then  
247 extruded and transferred to an anaerobic chamber. All cores were divided during extrusion based  
248 on visual changes in properties (e.g., soil horizons) when apparent or to divide the core length into  
249 two equal sections, with the resulting surface layer comprising the upper 4.5 to 9 cm (Table 1). All  
250 cores were segmented into two sections except for the second core from location Stream 2, which  
251 was longer and thus divided into three segments. For naming purposes, the core segments of each  
252 type were labeled using the convention CX-Y, with X being 1 or 2 to distinguish the duplicate  
253 cores and Y being 1, 2, or 3 (only for core 2 from location Stream 2) to indicate the segment  
254 starting with a value of 1 for the surface layer.

255

### 256 **2.3 Surface Water Characterization**

257 The field acidified surface water samples were used to determine dissolved major elements  
258 (sodium, magnesium, aluminum, silicon, potassium, calcium) using a Thermo Scientific iCap 7400  
259 Duo inductively-coupled plasma optical emission spectrometer (ICP-OES). Dissolved trace metal  
260 (cobalt, nickel, copper, and zinc), iron, and manganese concentrations were determined by  
261 inductively-coupled plasma mass spectrometry (ICP-MS) using a PerkinElmer Elan DRC II  
262 instrument. Dissolved anions ( $\text{Br}^-$ ,  $\text{Cl}^-$ ,  $\text{F}^-$ , and  $\text{SO}_4^{2-}$ ) in the unacidified, filtered water samples  
263 were measured using a Thermo Scientific Dionex Integrion High Pressure Ion Chromatograph (IC)

264 using a conductivity detector. The anion concentrations were determined using a Dionex IonPac™  
265 AS19 analytical column (4×250 mm) and Dionex IonPac™ AG19 guard column (4×50 mm) set  
266 to 30°C with the eluent generated from a Dionex eluent generator cartridge (EGC 500 KOH). The  
267 eluent concentration was set to 10 mM KOH for 10 minutes then gradually increased to 45 mM  
268 KOH over 15 minutes before holding steady at 45 mM KOH for 5 minutes. The flow rate remained  
269 constant at 1 mL min<sup>-1</sup>. Dissolved nitrate, ammonium, and orthophosphate were measured  
270 spectrophotometrically on a Seal Analytical AQ300 Discrete Multi-Chemistry Analyzer using  
271 frozen, field-filtered surface water samples that were thawed overnight at 4°C before analysis.  
272 Reagents and calibration standards were prepared using reagent grade chemicals. Ammonia was  
273 measured by reaction of a sample with hypochlorite in an alkaline solution, which is subsequently  
274 reacted with phenol in the presence of nitroferricyanide, with absorbance of the resulting blue  
275 indophenol dye measured at 660 nm. Nitrate was analyzed by reduction to nitrite by cadmium  
276 which then reacts with sulfanilamide in dilute phosphoric acid and binds to N-(1-naphthyl)-  
277 ethylenediamine dihydrochloride, with the resulting azo dye detected at 520 nm. Phosphate  
278 analysis involves the reaction of samples with ammonium molybdate and potassium antimonyl  
279 tartrate in in dilute sulfuric acid, forming a complex that is then reduced by ascorbic acid, with the  
280 resulting colored complex quantified at 660 nm. Charge balance calculations and estimation of  
281 dissolved CO<sub>2</sub> concentrations were performed in the React module of The Geochemist's  
282 Workbench® (Bethke, 2007) version 12.0.6 using the Lawrence Livermore National Laboratory  
283 thermochemical database V8 R6 (Delaney and Lundeen, 1990). Although not measured during  
284 initial analyses because of an oversight, dissolved inorganic carbon (DIC) and dissolved organic  
285 carbon (DOC) were measured in May 2021 using a Shimadzu TOC-L total organic carbon analyzer  
286 for archived filtered (0.45 µm MCE) water samples that had been stored at 4°C.

287

## 288 **2.4 Solid Phase Characterization**

289           Soil and sediment core segments were mechanically mixed in the anaerobic chamber to  
290 homogenize the materials prior to analysis. Approximately 20 g of wet sediment from each core  
291 segment were dried in a convection oven at 40 °C for determining the moisture content of the soils  
292 and sediments and for further bulk analyses. Sediments were then ground with a pestle and mortar  
293 and stored in glass vials. 1 g of each sediment sample was micronized in isopropanol using a  
294 McCrone Micronizing Mill to obtain a fine powder. The fine powders were then analyzed by  
295 powder X-ray diffraction (XRD) on a Bruker d8 Advance diffractometer using Cu K $\alpha$  radiation  
296 (40 kV, 40 mA) and a LynxEye XE energy-dispersive silicon-strip detector. XRD patterns of all  
297 sediment segments were performed from 3° to 80° 2 $\theta$  with a 0.02° 2 $\theta$  step-size, a collection time  
298 of 1.0 s per step, and no sample rotation to enable measurement to low angle. The detector consists  
299 of 192 independent strips, each of which measured each 2 $\theta$  position for the 1.0 s collection time,  
300 yielding a net integration time of 192 s per data point. The Bruker Diffrac.Eva application was  
301 used for phase identification using patterns generated from crystal structures contained in the  
302 Crystallography Open Database (Graulis et al., 2009).

303           Total nitrogen, carbon, and sulfur contents were determined using these dried soil and  
304 sediment samples by combustion elemental analysis using an Elementar vario MACRO cube  
305 CHNS analyzer. Approximately 0.1 to 0.2 g of the dried, ground soils or sediments from all core  
306 segments were measured in triplicate. The total metal concentrations in the soils and sediments  
307 were estimated using a microwave digestion analysis method (Hassan et al., 2007). Specifically, 3  
308 mL of concentrated trace metal grade HNO<sub>3</sub> and 0.3 mL of 30% hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) were  
309 added to 0.030 g of dried soil or sediment in a PTFE microwave digester vessel. Samples were

310 left to react at room temperature in the fume hood for 30 minutes and then digested in a CEM  
311 MARS6 microwave digestion system. The temperature was ramped from ambient to 175 °C over  
312 10 min and then maintained at 175°C for 15 min to promote complete digestion. After the vessels  
313 cooled, samples were filtered using 0.22 µm MCE filters followed by dilution to 2% HNO<sub>3</sub>.  
314 Triplicate digestions were prepared for each soil or sediments. Major elements (Al, Ca, Fe, K, Mg,  
315 Na, P, and S) in the resulting fluid were analyzed using ICP-OES and trace metals (Cu, Ni, Co,  
316 and Zn) plus Mn using ICP-MS.

317 Extractable nitrate, ammonium, and phosphate in the sediments were measured by an  
318 equilibrium extraction method adapted from previous studies (Sparks et al., 1996; Pansu and  
319 Gautheyrou, 2006). The extractions were performed in the anaerobic chamber with 1 g of moist  
320 soil or sediments suspended in 10 mL of 2 M potassium chloride (KCl). Samples were rotated end-  
321 over-end for 1 h, allowed to settle for 30 min, and then filtered using 0.22 µm MCE syringe filters.  
322 The filtrate was stored at 4 °C prior to analysis. Nitrate, ammonium, and phosphate in the  
323 extraction fluid were analyzed using a Seal Analytical AQ300. Samples from both cores at each  
324 site were treated as duplicate measurements to assess variability. The measured moisture content  
325 was used to calculate the concentration of the nutrients per gram of soil or sediment.

326

## 327 **2.5 X-ray Absorption Spectroscopy Data Collection and Analysis**

328 To determine the speciation of sulfur, iron, and trace metals present in the soils and  
329 sediments, X-ray absorption spectra were measured for nickel, copper, and zinc in the C1-1  
330 segments of the cores collected at each sampling location. C2-1 from location Stream 1 was also  
331 studied because XRD identified pyrite in C1-1 but not C2-1 (see Section 3.2). Cobalt was not  
332 investigated because its signal is masked by fluorescence from iron. Samples were prepared in an

333 anaerobic chamber as wet pastes sealed in polycarbonate sample holders using two layers of  
334 Kapton tape backed with acrylic glue. The beam-facing side of samples prepared for sulfur analysis  
335 was sealed with Saint-Gobain CHR sulfur-free polyester tape with an acrylic backing. To preserve  
336 hydration and anoxic conditions, the sealed samples were then impulse sealed in polyethylene  
337 pouches, which also contained a cellulose fiber wiper moistened with deoxygenated ultrapure  
338 water to inhibit desiccation. Reference standards (Fig. S3-8) that required stable hydration, such  
339 as adsorbed trace metal standards, were prepared similarly. Standards that are stable as dry solids  
340 were prepared by grinding using a mortar and pestle and spreading the resulting fine powder on  
341 cellulose acetate tape. Lists of reference standards are available in Tables S7-11, including their  
342 source, synthesis procedure, or, if data were taken from the literature, the relevant citation.

343         Primary data collection occurred at the Stanford Synchrotron Radiation Lightsource.  
344 Nickel, copper, and zinc K-edge spectra were collected at beamline 11-2, which employs a Si (220)  
345 double crystal, variable-exit monochromator ( $\phi = 90^\circ$  orientation) to scan the incident beam  
346 energy. A Rh-coated Si collimating mirror and detuning the monochromator to ~80% of peak  
347 intensity were used to minimize the harmonic content of the X-ray beam. Nickel, copper, and zinc  
348 metal foils were used for energy calibration, with their K-edges set to 8333, 8979, and 9659 eV,  
349 respectively. Data were collected in fluorescence-yield using a 100-pixel energy-dispersive  
350 monolithic Ge detector. This large array detector captured a substantial solid-angle of the  
351 associated X-ray fluorescence emitted by each sample, maximizing the count rate originating from  
352 trace metals in the sample. Data collection focused on the X-ray absorption near-edge structure  
353 (XANES) region because of the low signal level associated with the concentrations of the trace  
354 metals. The detector was operated in a high-resolution mode to separate the fluorescence line of  
355 interest from background, which dominated the total count rate and was thus the dominant

356 contributor to noise in the data. Detector saturation was avoided because of the relatively low Fe  
357 content in the samples and the relative attenuation of Fe fluorescence by Al foil placed between  
358 the sample and detector. The high flux of the beamline enabled this substantial attenuation while  
359 still utilizing the full linear-range of the detector. Multiple scans were collected for each sample  
360 and then averaged. No detectable changes in the spectra were observed, such as those caused by  
361 beam-induced damage, and no drift in energy calibration was detected.

362         Iron and sulfur K-edge spectra were collected at beamlines 4-3 configured with a Si (111)  
363 double-crystal monochromator ( $\phi = 90^\circ$  orientation) and a Si mirror for harmonic rejection.  
364 Sulfur XANES spectra were collected in fluorescence-yield using a Vortex ME7 silicon drift  
365 detector. All samples for sulfur measurements were removed from their polyethylene pouches and  
366 immediately chilled to  $\sim 20$  K using a liquid He cryostream to prevent beam-induced redox changes  
367 while minimizing beam attenuation. Iron spectra were collected through the extended X-ray  
368 absorption fine structure (EXAFS) region in transmission at ambient temperatures. Repeated scans  
369 of sulfur and iron in a single position on each sample showed negligible changes over time.  
370 However, attempts to evaluate other positions on the samples were not successful because of  
371 diffraction into the fluorescence detector and spatially heterogeneous absorption, both of which  
372 are attributed to the substantial quartz contents of the samples. Data for some standards were  
373 collected at beamlines 5-BM-D and 12-BM-B at the Advanced Photon Source at Argonne National  
374 Laboratory. These beamlines employ Si (111) fixed-offset monochromators with a flat Rh-coated  
375 Si mirror for harmonic rejection; a toroidal Rh-coated Si mirror at 12-BM-B also provides beam  
376 focusing. Data for pure compounds were collected in transmission while those with lower target  
377 element concentrations, such as elements substituting in a mineral structure, were collected in  
378 fluorescence yield using a 12-element energy-dispersive solid-state Ge detector.



379 XANES spectra were averaged in the Sixpack interface (Webb, 2005) to IFEFFIT  
380 (Newville, 2001) and then background subtracted and normalized in Athena (Ravel and Newville,  
381 2005). For the trace metal spectra,  $E_0$  was set to 8342 eV for nickel, 8988 eV for copper, and 9659  
382 eV for zinc. These values reflect the typical edge position of the collection of samples and are thus  
383 shifted relative to the energies expected for these elements in their metallic forms. A linear pre-  
384 edge background was fitted to the pre-edge region (150 to 30 eV below  $E_0$  for copper and nickel,  
385 and 60 to 30 eV below  $E_0$  for zinc) and removed. The data were then normalized by fitting a linear  
386 function to the post-edge region (75 to 225 eV above  $E_0$ ). For iron,  $E_0$  was set to 7125 eV and a  
387 linear background was fitted 27 to 20 eV below  $E_0$ . This background range was narrow because  
388 some spectra had a glitch at lower energy and a consistent background removal was used for all  
389 samples. Separate post-edge normalizations were used for the iron XANES and EXAFS spectra.  
390 For the XANES, normalization was again performed using a linear function fitted to 75 to 225 eV  
391 above  $E_0$ . To account for slight curvature in the data over the larger energy range covering the  
392 EXAFS region, the data were normalized using a quadratic function fitted between 150 and 540  
393 eV above  $E_0$ . The EXAFS were extracted after fitting a spline using the Autobk algorithm in  
394 Athena ( $R_{\text{bkg}} = 0.7$ ;  $k$ -weight = 3;  $k$ -range 3 – 11  $\text{\AA}^{-1}$ ). The  $E_0$  for the sulfur XANES data was set  
395 to 2472 eV, with a linear background fitted between 20 and 7 eV below  $E_0$  and a linear post-edge  
396 normalization performed between 35 to 56 eV above  $E_0$ .

397 For the spectra of the trace metals, Athena was used for principal component analysis  
398 (PCA), target transformation (TT) of the spectra of standards, and linear combination fitting (LCF)  
399 of the spectra (Manceau et al., 2002). Statistical constraints on the amount of variability in a set of  
400 spectra are provided by PCA but this cannot uniquely identify the exact number of fitting  
401 components required because of limited data set size and possible co-occurrence of real species in

402 multiple spectra (Manceau et al., 2002). Similarly, TT provides statistical support for identifying  
403 the standards most likely to occur in a set of samples, but this is also ambiguous and needs to be  
404 guided by chemical and mineralogical constraints. Further, similar species (e.g., trace metals in  
405 clay mineral structures) may display subtle variations in their spectra that would require an  
406 impractical number of standards to fully capture. The spectrum of the actual species present in a  
407 sample may thus not be perfectly matched by that of an individual standard, but a linear mixture  
408 of two or more chemically similar standards may reproduce this well. Given these limitations and  
409 the inability to independently identify the individual trace metal species present, a combinatorial  
410 approach was used where all combinations of the spectra of relevant standards, identified by TT,  
411 were fit to the sample spectra, with the combination that minimized the reduced chi-squared value,  
412  $\chi^2$  (Kelly et al., 2008) used as the final result. Fitted components were then grouped in species  
413 categories (see Section 3.4). Linear-combination fitting of sulfur XANES spectra also employed a  
414 combinatorial approach using Athena, but standards used as fitting components were selected to  
415 reproduce the features present. The spectra clearly indicated the co-occurrence of more sulfur  
416 species than there were samples, precluding use of PCA. Similarly, fitting of iron XANES and  
417 EXAFS spectra in Athena used standards based on common iron phases and mineralogy  
418 determined by XRD. We note that the insight gained from all linear combination fitting is limited  
419 by the standard spectra available. Inclusion of additional standard spectra, such as through future  
420 reuse of the data presented, may enable greater insight into the species present.

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423

424

## 425 3. RESULTS

### 426 3.1. Soil and Sediment Mineralogy and Composition

427 The mineralogy at all sites is dominated by quartz with only minor phases showing  
428 substantial variations in occurrence (Fig. 1). The investigated duplicate core samples at each  
429 location yielded similar mineralogy (data from only one core are plotted) except at location Stream  
430 1, with the surface layer of one of the two cores containing pyrite, the only detection of this phase  
431 (Fig. 1). Minor phases in the marsh wetland soils included smectite, illite, and kaolinite as well as  
432 plagioclase feldspar. The riparian wetland soils differ from the marsh wetland soils, with location  
433 Riparian 2 having minor K-feldspar, kaolinite, chlorite, and gibbsite, and trace illite. In contrast,  
434 the location Riparian 1 soil contains only trace amounts of kaolinite, chlorite, and gibbsite, with  
435 no other phases detected besides the ubiquitous quartz. Note that prior studies at the Savannah  
436 River Site, which hosts the riparian wetlands, suggest that clays identified by XRD as chlorite may  
437 instead be hydroxy-interlayered vermiculite (Kaplan et al., 1997; Zaunbrecher et al., 2015). The  
438 stream sediments at both sampling locations contain minor K-feldspar, chlorite, smectite, kaolinite,  
439 and illite. No carbonate minerals were detected in any sample.

440 The total carbon contents varied among the sites (Fig. 2), with highest concentration in the  
441 marsh soils and lesser amounts at the other two sites. Notably, the core sample containing pyrite  
442 at location Stream 1 also contained elevated organic carbon, and this was substantially lower in  
443 the pyrite-free duplicate core. It is unclear whether there is a genetic relationship between organic  
444 carbon and pyrite as the latter had sharp X-ray diffraction lines (Fig. 1), suggestive of a detrital  
445 phase. Sulfur showed a similar pattern of variation as carbon despite the substantially lower  
446 concentration. The cores at location Stream 1 showed a large difference in sulfur content, again  
447 because of the presence of pyrite in one of the two duplicates. Iron contents were similar among

448 all samples except for location Riparian 1, which contained an order of magnitude less iron,  
449 consistent with the mineralogy being dominated nearly exclusively by quartz. Manganese contents  
450 were two to three orders of magnitude lower than iron contents, with the lowest values observed  
451 in the riparian soils.

452

## 453 **3.2. Trace Metal Concentrations**

### 454 *3.2.1. Dissolved trace metals*

455 Dissolved trace metal concentrations (Fig. 3) generally followed the trend of  $Zn > Cu \approx Ni$   
456  $> Co$  at all the investigated sites. Dissolved Zn concentrations showed the largest absolute  
457 variation, ranging from 46 to 230 nM with no clear trend among the sites, and the values were in  
458 all cases above the 10 to 20 nM concentrations in the process blanks (Table S2). Similarly,  
459 dissolved Cu concentrations showed no trend among the three field sites and ranged from 20 to 80  
460 nM. Measured Cu concentrations were substantially greater than in the process blanks except for  
461 marsh wetland location Marsh 2. Ni concentrations were similar at the marsh wetlands and stream  
462 sites, occurring in the range of 30-60 nM, but location Riparian 2 contained substantially less  
463 dissolved Ni (12 nM). One process blank from the marsh wetland contained ~130 nM dissolved  
464 Ni while all others were  $<10$  nM. We attribute this to contamination, possibly from dust associated  
465 with  $>40$  km hr<sup>-1</sup> wind gusts during sampling at this location. Dissolved Co concentrations were  
466 low, often  $<10$  nM, but in most cases were above the approximately 2 to 4 nM concentrations in  
467 the process blanks.

468

469

470

### 471 **3.2.2. Solid phase trace metals**

472 Overall patterns of solid-phase trace metal concentrations (Fig. 3), as determined by  
473 microwave-assisted acid digestion, followed the same general trend as for the surface waters: Zn  
474  $> \text{Cu} \approx \text{Ni} > \text{Co}$ . Variations in the abundance of each trace metal among the field sites generally  
475 tracked one another. The marsh wetlands contained the highest solid-phase Cu, Ni, and Zn  
476 contents, with Co concentrations the largest in the stream site sediments, although this was likely  
477 affected by the presence of pyrite in one core. The riparian wetlands, especially location Riparian  
478 2, had the lowest solid-phase trace metal contents, consistent with the low abundance of non-quartz  
479 minerals and low major element (C, S, Fe, Mn) concentrations. The concentrations of the trace  
480 metals (0.3 to 18  $\mu\text{g g}^{-1}$  Co, 3 to 27  $\mu\text{g g}^{-1}$  Ni, 3 to 45  $\mu\text{g g}^{-1}$  Cu, and 11 to 144  $\mu\text{g g}^{-1}$  Zn) were  
481 similar to or well below geological background levels of  $17.3 \pm 0.6 \mu\text{g g}^{-1}$  Co,  $47 \pm 11 \mu\text{g g}^{-1}$  Ni,  
482  $28 \pm 4 \mu\text{g g}^{-1}$  Cu, and  $67 \pm 6 \mu\text{g g}^{-1}$  Zn (Rudnick and Gao, 2003).

483

### 484 **3.3. Spectroscopic Analysis of Sulfur and Iron Speciation**

485 Seven samples in total were examined from the three field sites using sulfur K-edge  
486 XANES and iron K-edge XANES and EXAFS spectroscopy. The surface layer sample from the  
487 first core at each sampling location was examined except for location Stream 1. The duplicate  
488 cores from this location differed in their mineralogy (core 1 contained pyrite) as well as their bulk  
489 carbon, iron, sulfur, and trace metal contents. Samples from both cores from location Stream 1  
490 were thus investigated.

491

492

493

### 494 3.3.1. Sulfur Speciation

495 The sulfur XANES spectra of wetland soils and stream sediments display two primary  
496 whitenline regions at 2472.6 to 2474.1 eV and 2481.1 to 2482.5 eV, with some displaying a shoulder  
497 near ~2470.5 eV (Fig. 4). This array of features indicate the presence of multiple sulfur oxidation  
498 states occurring in both inorganic and organic forms (Prietz et al., 2009; Manceau and Nagy,  
499 2012), potentially consistent with inorganic mono- and disulfide minerals, elemental sulfur,  
500 exocyclic reduced sulfur, sulfones, sulfonates, and organic or inorganic sulfates. Preliminary  
501 principal component analysis (PCA) of these spectra revealed that fine-structure features were  
502 present in all components. This indicates that the number of species in the samples was similar to  
503 or exceeded the number of samples analyzed (seven). Further evaluation using statistical  
504 techniques, including evaluation of possible standard spectra via target transform (TT), was thus  
505 not possible as the sample set is inadequate to describe the variability in the data.

506 A combinatorial fitting approach was used (Manceau and Nagy, 2012) to identify the  
507 species present because of the large number of species likely present. We modified the previously-  
508 published procedure by using the reduced chi-squared ( $\chi^2$ ) as the goodness-of-fit parameter to  
509 minimize because it accounts for the removal of degrees of freedom when adding additional fitting  
510 components. The fitting results are considered semi-quantitative because the spectra of standards  
511 were taken from prior studies (Table S7) that may have differed in their energy resolutions. This  
512 is most apparent for pyrite and elemental sulfur fitting components. Attempts to collect data on  
513 both phases after micronizing and dispersing on sulfur-free polyester tape resulted in whitenlines  
514 that were lower intensity and broader than the diagnostic features in the sample spectra. This  
515 indicates that the standards suffered from over-absorption, an effect that has been widely reported  
516 in previous studies (George et al., 2008; Almkvist et al., 2010; Manceau and Nagy, 2012; Lau et

517 al., 2017). While the systematic errors associated with the standards used could not be rectified,  
518 the fitting results still constrain which sulfur species are present and their relative abundance.

519 Fitting demonstrates that the wetland soils and stream sediments all contain elemental  
520 sulfur, exocyclic reduced sulfur, sulfonate, and sulfate. The latter species was best modeled using  
521 inorganic sulfate, with the combinatorial fitting not identifying an organosulfate in any sample.  
522 All samples except core 2 of location Stream 1 contained a small FeS component. However, the  
523 abundance of this phase may be underrepresented in some samples as fitting did not always fully  
524 reproduce the intensity of the shoulder at  $\sim 2470.5$  eV associated with the phase. Pyrite was only  
525 observed in both samples from location Stream 1. It is likely that the C1 sample measured by  
526 XANES did not contain the substantial amount of pyrite identified by XRD. Sample alignment  
527 prior to XANES data collection involves identifying an area with spatially-uniform fluorescence,  
528 and this process likely selected for areas free of coarser pyrite grains. The sharp XRD peaks  
529 indicate pyrite in the C1 samples from location Stream 1 is well crystalline, and such grains would  
530 produce locations of spatially-varying fluorescence that would be avoided during alignment. The  
531 two marsh wetland soils and the Riparian 2 sample also contained minor amounts of sulfoxides  
532 and sulfones.

533

### 534 **3.3.2. Iron Speciation**

535 The speciation of iron was probed using both XANES and EXAFS spectra for all sampling  
536 locations except Riparian 1 (Fig. 5). The high quartz content in this sample prevented identification  
537 of a spatially homogeneous location and all EXAFS spectra that were attempted contained  
538 substantial artifacts; only the XANES spectrum was analyzed for this sample. Preliminary PCA of  
539 the XANES and EXAFS spectra revealed fine-structure features present in all components

540 obtained for each spectra region. This indicates that the number of species present in the samples  
541 was equal to or greater the number of spectra measured (seven for XANES, six for EXAFS). This  
542 precluded further statistical analyses, including evaluation of potential standard spectra using TT.

543         Similar trends in overall speciation were observed among the samples through analysis of  
544 the XANES and EXAFS regions. However, quantitative agreement between the XANES and  
545 EXAFS fitting results did not occur for all samples. Such differences can result if the standards  
546 are more crystalline than the species present, as such cases would produce similar XANES spectra  
547 but distinct EXAFS spectra associated with weaker oscillations from more disordered second shell  
548 features in less crystalline materials. These discrepancies do not affect the overall results of this  
549 study as iron speciation was investigated primarily to constrain the potential host phases for trace  
550 metals. Any solid iron species detected would occur at two to four orders of magnitude greater  
551 abundance than the trace metals.

552         Iron at all sites partially occurs as Fe(II) and Fe(III) in clay mineral structures, with all but  
553 location Marsh 1 showing evidence for Fe(III) oxide minerals as well. Both core samples from  
554 location Stream 1 contained FeS, with both XANES and EXAFS showing that this phase occurs  
555 in greater abundance in core 1. No pyrite was apparent in either core from location Stream 1.  
556 Similar to the sulfur case, the need to obtain data on a portion of the sample having spatially  
557 uniform absorbance may have biased the measurement by avoiding pyrite grains. Note that the  
558 iron, sulfur, and trace metal spectra (Section 3.4.), as well as XRD patterns, were each collected  
559 on different subsamples of physically homogenized core segments because of the distinct  
560 measurement requirements. The detection of pyrite in the Stream 1 samples by sulfur XANES  
561 (Fig. 4) is thus not necessarily contradictory to the iron speciation results. We interpret these results  
562 to indicate that iron sulfide phases are present in the samples. The XANES and EXAFS spectra



563 are not consistent regarding FeS or pyrite occurrences in other samples. These phases have low  
564 abundances (<10 mol% of Fe) in the fitting results and may thus either be artifacts or reflect  
565 different sensitivities to minor phases. The weak shoulder at ~2470.5 in many sulfur XANES  
566 spectra is more diagnostic for a minor FeS component in many samples, suggesting this is present  
567 in small quantities. Note that iron concentrations are an order of magnitude greater than sulfur in  
568 most samples so an ~10% FeS component in the sulfur XANES spectra would correspond to ~1%  
569 FeS in the iron spectra. Finally, most samples required a contribution from dissolved Fe(II) to  
570 reproduce the iron XANES and EXAFS spectra. This may reflect Fe(II) adsorbed to or cation  
571 exchanged onto minerals or complexed by organic matter. While no iron-bearing primary silicates  
572 were detected in XRD, we lack available XANES and EXAFS spectra for such phases, and the  
573 dissolved Fe(II) component may at least partially account for minor occurrences of such minerals.  
574 Finally, fitting of the XANES spectra identified magnetite as a minor component of only the Marsh  
575 1 sample, and this was not observed for any EXAFS spectrum. We thus conclude that magnetite  
576 is not an observable component of the iron speciation in the sites examined.

577

#### 578 **3.4. Spectroscopic Analysis of Trace Metal Speciation**

579 XANES spectroscopy (Fig. 6-8) was employed to investigate solid-phase trace metal  
580 speciation using the same sample selection rationale as for the sulfur and iron measurements  
581 described above. EXAFS measurements were not possible because of the low total metal contents  
582 of the samples. Cobalt was not investigated because its fluorescence signal is masked by the iron  
583  $K_{\beta}$  fluorescence line. The resulting seven spectra per element were initially examined by PCA to  
584 evaluate variability in the XANES spectra for each element and to provide statistical guidance for  
585 the minimum number of components contained in each dataset. Target transform (TT) was then

586 used to assess whether the spectra of various standards for each metal (Fig. S6-8) were possibly  
587 present in each dataset. Those standard spectra which could be reconstructed by TT, minus species  
588 inconsistent with the observed mineralogy or the sulfur and iron speciation, were then used in  
589 linear-combination fitting (LCF) to quantitatively assess trace metal speciation. A combinatorial  
590 LCF approach was used, with all possible combinations of two or more standards evaluated. The  
591 fit that yielded the lowest reduced chi-squared value was then selected.

592 This approach was motivated by the lower sensitivity of XANES spectra to specific  
593 structural environments compared to EXAFS spectra, which are unmeasurable because of the low  
594 trace metal concentrations. In recognition of this limitation, the LCF results were binned by broad  
595 species type to more accurately reflect the information obtained. While XANES spectra reflect the  
596 general coordination environment of the target element, they lack the specific structural sensitivity  
597 of EXAFS spectra. The adsorbed metal groupings included XANES fitting contributions from the  
598 spectra of aqueous ions because these are spectrally similar to some adsorbed species (Fig. S6-8).  
599 In addition, their spectra are expected to be indistinguishable from the spectra of cation exchanged  
600 species. Dissolved species themselves are likely a negligible contribution to the spectra for all  
601 samples. Assuming maximum porewater trace metal concentrations of 1  $\mu\text{M}$ , well above the  
602 dissolved concentrations in overlying surface waters (Fig. 3), aqueous species would constitute  
603 <1% of the metals per unit volume in the soil and sediment samples. The spectra of metals  
604 coordinated to carboxylic acids were also similar to the spectra of aqueous and adsorbed species  
605 and were thus binned into the adsorbed metal groupings. An analogous approach was used to  
606 establish the “clay structures” grouping, consisting of metals substituted into octahedral sheets in  
607 phyllosilicates and related phases. Clay minerals vary widely in their compositions, and the range  
608 of possible second-shell cation neighbors produces subtle variations in the fine structure features

609 of trace metal XANES spectra. There is no method available to independently determine the  
610 composition of clay(s) hosting a trace metal in a sample. Sample spectra were thus fitted using the  
611 spectra of a series of clay standards to account for these subtle spectral variations. All such fitting  
612 components were binned into the “clay structures” grouping. The standards utilized in all analyses  
613 are thus interpreted as proxies for general classes of metal species distinguishable by their local  
614 coordination environments.

615

### 616 *3.4.1. Copper Speciation*

617 Five of the seven copper XANES spectra (Fig. 6), spanning samples from all three sites,  
618 displayed a clear shoulder at ~8986 eV indicative of copper bound to reduced sulfur. These include  
619 the spectra of the duplicate core samples from location Stream 1, one of which contained pyrite.  
620 Spectra from location Riparian 1 and Stream 2 displayed a weaker low-energy shoulder and  
621 stronger white line. PCA required three components to adequately reconstruct the fine structure of  
622 all of the sample spectra. TT identified covellite (CuS), chalcocite (Cu<sub>2</sub>S), aqueous Cu(II), Cu(II)  
623 adsorbed to goethite, copper(II) citrate, copper(II) hydroxide, and Cu(II) complexed to cysteine  
624 and histidine as potential standards. While observed in prior studies (Weber et al., 2009b; Weber  
625 et al., 2009a; Hofacker et al., 2013b; Hofacker et al., 2013a; Hofacker et al., 2015), metallic copper  
626 was poorly reconstructed and thus unlikely to be present in the samples. Combinatorial LCF results  
627 utilized all of the well-reconstructed standards in modelling the XANES spectra except for  
628 copper(II) citrate, copper(II) hydroxide, and Cu(II) complexed to cysteine, which were not present  
629 in any optimized fit. In the fitting results, the histidine complex is classified as representing amine-  
630 bound copper, and both aqueous Cu(II) and Cu(II) adsorbed to goethite representing adsorbed  
631 copper.

632 CuS was present in all sites studied and accounted for >70% of the copper in five of the  
633 seven samples (Fig. 6). Adsorbed forms of copper were the next most abundant species overall,  
634 present in all but location Marsh 1. This species type occurred in equal abundance as CuS at  
635 location Riparian 1 and was the dominant copper species in the sediments at location Stream 2.  
636 Amine-bound copper was a minor component in four samples and fitting only identified Cu<sub>2</sub>S  
637 (30±4%) at location Riparian 1. No significant difference in CuS abundance was observed for the  
638 duplicate core samples from location Stream 1 despite one of these (C1) containing pyrite. While  
639 fitting identified a minor amine-bound copper component in one of the two duplicate cores, its  
640 occurrence is of marginal statistical significance (9±7 wt.%) and adsorbed plus amine-bound  
641 copper are effectively identical for the two samples.

642

### 643 *3.4.2. Nickel Speciation*

644 The nickel XANES spectra (Fig. 7) show, at most, subtle variation among all samples  
645 examined. These are limited to slight variations in the slope of the absorption edge, the height and  
646 shape of the white line at ~8352 eV, and the intensity of the weak post-edge feature at ~8367 eV.  
647 Reconstruction of the spectra during PCA required three components to reproduce major fine-  
648 structure features. A fourth component contained features consistent with XANES fine-structure  
649 and further improved reconstructions of some spectra. TT identified nickel sulfide, Ni(II)  
650 substituting in magnetite and the octahedral sheets of serpentine and a trioctahedral Fe(II)-Mg  
651 smectite, dissolved Ni(II), Ni(II) adsorbed to goethite, and Ni(II) complexed to histidine, cysteine,  
652 glutathione, and cellulose as potential standards. Of these standards, none of the organic complexes  
653 occurred in the optimized fits to the data, and Ni(II) substituting in magnetite was not used because  
654 magnetite was not clearly identified by XRD or the iron speciation results. For the remaining

655 standards, spectra of the Ni(II) in the octahedral sheets of serpentine and smectite were grouped as  
656 “clay structures”, and Ni(II) adsorbed to goethite and in aqueous solution were grouped as  
657 adsorbed Ni.

658 Fitting identified Ni in clay structures, nickel sulfide, and adsorbed Ni in all samples (Fig.  
659 7). Nickel in clay mineral structures was the dominant species in all samples, ranging from 50%  
660 to 70% of the total Ni. Nickel sulfide was the next most abundant species in the marsh wetlands  
661 and location Stream 1, with adsorbed nickel occurring in higher abundance in the other stream site  
662 and the riparian wetlands. Despite the presence of pyrite in one of the duplicate cores, the sulfide  
663 component of the nickel species at stream location Stream 1 were statistically identical.

664 The quantitative nickel speciation results likely contain systematic errors associated with  
665 the presence of a species not represented in the set of standards available. The spectra of a number  
666 of samples, most notably the riparian wetland samples, contain a weaker feature at ~8367 eV and  
667 a splitting in the broad features from 8380 to 8420 eV that are not reproduced in the fits (Fig. 7).  
668 Examination of published spectra of diverse organic and mineral-associated forms of nickel  
669 indicates that such features are rare in Ni K-edge XANES spectra (Kopittke et al., 2011; Van Der  
670 Ent et al., 2018; Ratié et al., 2018; Siebecker et al., 2018). The 8380 to 8420 eV feature corresponds  
671 to the 3 to 4 Å<sup>-1</sup> region of the EXAFS spectrum, and this has been shown to produce a splitting  
672 when nickel occurs in Mg-rich serpentine, gibbsite (Siebecker et al., 2018), and aluminum  
673 hydroxide clusters in smectite interlayers or associated with smectite as small gibbsite particles  
674 (Nachtegaal et al., 2005), with the Al-modified smectites producing the deepest node in this split  
675 feature. The misfit of the XANES data in the 8380 to 8420 eV region is most substantial in samples  
676 containing detectable gibbsite in the XRD data, supporting the assessment that this originates from  
677 nickel species associated with aluminum hydroxides. In the grouping approach used in this study,

678 these would be categorized as clay-associated species and thus the misfits do not fundamentally  
679 alter the observation of this group as the predominant nickel species in the samples examined.  
680 Further, the nickel sulfide content determined by the fitting is likely unaffected by the lack of an  
681 appropriate standard for this type of clay-bound nickel. Nickel sulfide has an absorption edge  
682 shifted to lower energy from all clay species and its contribution reproduces broadening of the data  
683 in the low-energy portion of the absorption edge.

684

### 685 *3.4.3. Zinc Speciation*

686 Similar to copper and nickel, the XANES spectra of zinc show only subtle variations except  
687 for sample Riparian 2, which lacks fine structure features in the whiteness (Fig. 8). At least three  
688 components are required to reconstruct the spectra during PCA, with four clearly improving  
689 reconstruction of features in some spectra. In addition, an examination of the individual  
690 components showed that up to five contained non-noise features. TT identified a large number of  
691 possible candidate species: zinc(II) oxide, sphalerite (ZnS), zinc(II) in the octahedral sheet of a  
692 trioctahedral Fe(II)-Mg smectite, a zinc-aluminum layered double hydroxide, zinc(II) adsorbed on  
693 and incorporated into goethite, dissolved zinc(II), zinc(II) acetate, zinc(II) oxalate, zinc(II) sulfate,  
694 and zinc(II) complexed to cysteine and glutathione. Combinatorial LCF fitting did not utilize the  
695 spectra of the layered double hydroxide, zinc oxalate, and zinc incorporated into goethite in the  
696 final results. Species were grouped by category in reporting the fitting results as described for the  
697 other metals.

698 Zinc speciation (Fig. 8) was similar in the marsh wetland soils and stream site sediments,  
699 consisting of adsorbed species, zinc associated with clay mineral structures, and complexed to  
700 thiol groups. The riparian wetland soils lacked Zn in clay structures and contained sphalerite, with

701 location Riparian 2 lacking thiol-bound zinc and location Riparian 1 containing a large adsorbed  
702 pool. In addition, a substantial zinc oxide component ranging from 10% to 54% of the zinc species  
703 occurred in all samples. Prior studies of zinc-contaminated soils have indicated that zinc oxide  
704 introduced into such settings does not persist, dissolving and transforming to new phases (Voegelin  
705 et al., 2005; Voegelin et al., 2011). An inverse correlation was observed between XANES-derived  
706 zinc oxide content and solid-phase zinc concentrations ( $R = -0.81$ ,  $p = 0.03$ ). These considerations  
707 suggest that the zinc oxide component may be a contaminant or data collection artifact. Notably,  
708 the cellulose acetate tape widely used for sample mounting and other purposes at synchrotron  
709 beamlines contains zinc oxide (Supplementary material text and Fig.S9). Zinc oxide introduced  
710 during field sampling is unlikely to have persisted and process blanks during microwave digestion  
711 did not identify zinc concentrations above the detection limit. The zinc oxide components in the  
712 XANES spectra are thus likely attributable to a background signal associated with sample cells  
713 and beamline components.

714

## 715 **4. DISCUSSION**

### 716 **4.1 Lack of Elemental Correlations in Bulk Compositions**

717 A linear regression employing maximum likelihood estimation and weighted by the  
718 bivariate uncertainties (Thirumalai et al., 2011) determined correlation coefficients ( $R$ ) and  $p$ -  
719 values among pairs of the various major element and trace metal concentrations for each site. The  
720 datasets used in the analysis consisted of compositional data and associated measurement  
721 uncertainties for all sections of the duplicate cores from both sampling locations at each field site,  
722 e.g., cores 1 and 2 from the Marsh 1 and Marsh 2 locations at the marsh wetlands site. For the  
723 stream sediment site, the surface layer from core 1 at location Stream 1 contained pyrite and was

724 removed from the dataset because it serves as an outlier that then yields spuriously large R values.  
725 In many regression analyses the pyrite-bearing sample occupied one end of a linear trend far  
726 removed from all other samples, which form a separate cluster that anchors to the opposite end of  
727 the linear trend. Removal of this one outlier datum in each analysis substantially changed the  
728 regression line and decreased the R value.

729         Regression of trace metal and major element abundances in the marsh wetland soils and  
730 stream sediments generally yield low R values (Fig. 9), indicating weak correlation among these  
731 elements. No clear relationships can be identified between the trace metal and major element  
732 contents of these soils and sediments. In contrast, large R values between the trace metals and all  
733 major elements were obtained for the riparian wetland soils (Fig. 9), with all having high statistical  
734 significance ( $p \leq 0.003$ ). Further examination, however, shows that the major elements are also  
735 highly correlated with each other, and all element pairs yield large R values with small p-values.  
736 We hypothesize that the correlations among all elements (C, S, Fe, Mn, and trace metals) in the  
737 riparian wetland soils reflect variations in the minor non-quartz content in which all of these  
738 elements reside. Variations in major element abundance thus do not provide insight into processes  
739 controlling solid-phase trace metal speciation in the soils and sediments.

740

## 741 **4.2. Importance of Reduced Sulfur and Clay Minerals to Background Metal Speciation**

### 742 ***4.2.1 Reduced Sulfur***

743         In the present study, substantial fractions of copper, nickel, and zinc in all samples were  
744 bound to reduced sulfur (Fig. 10A), primarily as sulfide minerals for copper and nickel and as thiol  
745 groups on organic matter for zinc. Trace metals are widely reported to associate with reduced  
746 sulfur in anoxic soils and sediments in freshwater systems (Huerta-Diaz et al., 1998; Koretsky et



747 al., 2007), but the relationship between metal speciation and sulfur content has not been previously  
748 explored. Larger fractions of copper are associated with sulfur compared to nickel and zinc,  
749 consistent with past studies of both natural and contaminated sites (Bostick et al., 2001; Weber et  
750 al., 2009b; Fulda et al., 2013a; Noël et al., 2015; Webb and Gaillard, 2015; Siebecker et al., 2018;  
751 Mantha et al., 2019). However, no clear relationship exists between the fractions of each metal  
752 bound to reduced sulfur and the total sulfur content of the soil and sediment samples (Fig. 10B),  
753 despite nearly two orders of magnitude variation in the concentration of sulfur (Fig. 2). Prior  
754 studies suggest that metal binding to reduced sulfur in flooded soils (Weber et al., 2009b; Fulda et  
755 al., 2013a) is limited by sulfide availability. Such limitations are not apparent in the present work  
756 despite the bulk sulfur content spanning the range of concentrations in these prior investigations.  
757 This different observation may originate from the focus of prior studies on either contaminated  
758 soils with substantially higher metal concentrations or soils to which sulfate was added during  
759 anoxic incubations. At the low metal concentrations observed in the marsh and riparian wetland  
760 soils and stream sediments under examination, which are similar to crustal averages (Rudnick and  
761 Gao, 2003), sulfur contents and fractional abundances of reduced sulfur were adequate to allow  
762 similar formation of reduced sulfur-bound metals in all systems. The total solid-phase sulfur  
763 concentrations observed in the present work span the range reported in prior studies of freshwater  
764 subsurface aquatic systems (Casagrande et al., 1977; Altschuler et al., 1983; Brown, 1985; Wieder  
765 et al., 1985; Spratt and Morgan, 1990; Dail and Fitzgerald, 1999; Rozan et al., 2002; Prietzel et  
766 al., 2009; Mehlhorn et al., 2018), indicating that sulfur-bound trace metals have broad occurrences  
767 as key species in such systems regardless of the soil or sediment sulfur content.

768

769

#### 770 **4.2.2. Clay Minerals**

771 Substantial prior work has demonstrated that nickel and zinc enter into clay mineral and  
772 layered double hydroxide structures in contaminated soils (Manceau et al., 2004; Voegelin et al.,  
773 2005; McNear et al., 2007; Jacquat et al., 2009; Voegelin et al., 2011), and this has also been  
774 observed for regions hosting Ni-rich ultramafic rocks (Dublet et al., 2012; Noël et al., 2015; Noël  
775 et al., 2017; Siebecker et al., 2017; Siebecker et al., 2018). However, the role of clay mineral  
776 structures in hosting trace metals in anoxic freshwater aquatic systems containing trace metals at  
777 low, background concentrations has not been previously demonstrated. The neoformation of such  
778 solids may depend on saturation state or dissolved metal concentrations (Manceau et al., 1999;  
779 Schlegel et al., 2001) and may be inhibited by organic matter (Nachtegaal and Sparks, 2003). This  
780 study demonstrates that the low trace metal contents of wetland soils and stream sediments still  
781 enable the incorporation of both nickel and zinc in clay structures, with this being the dominant  
782 nickel species in all systems. In addition, the occurrence of nickel and zinc in clay structures is  
783 independent of sulfur content and speciation, suggesting that they may be inherited from the  
784 weathering process and are resistant to disruption in the presence of sulfide.

785

#### 786 **4.3 Consistent Trace Metal Speciation in Wetland Soils and Stream Sediments**

787 The study sites are distributed over a ~1000 km transect and represent different aquatic  
788 system types in geologically-distinct regions, with the marsh wetlands occurring in glacial  
789 sediments, the stream sediments draining carbonate and shale lithologies, and the riparian wetlands  
790 underlain by clastic coastal plain sediments. The riparian wetland soils are also heavily weathered  
791 compared to the other two sites, lacking smectite clay minerals at both study areas with no  
792 detectable feldspar at location Riparian 1, yielding more acidic and cation-poor local waters (Table

793 S1). Despite these substantial differences, the three sites investigated have overall similar trace  
794 metal speciation, with variations between the sampling locations at each site often larger than  
795 among the different field areas. These observations suggest that a small set of components  
796 consistently dominates trace metal speciation in uncontaminated freshwater subsurface aquatic  
797 systems: sulfide and adsorbed forms predominate in diverse systems for copper, with nickel and  
798 zinc also occurring in clay mineral structures. Further, similar distributions of trace metal species  
799 occur at multiple, unrelated and spatially separate field sites, which may indicate that these specific  
800 phase associations are a common feature of freshwater aquatic systems.

801         Limited prior studies of uncontaminated, anoxic soils and sediments generally support our  
802 hypothesis that trace metal speciation has consistent controls across many freshwater aquatic  
803 systems. A study of an anoxic soil and a peat lens formed at a natural CO<sub>2</sub> seep found copper  
804 bound primarily to reduced sulfur with a lesser component complexed to organic matter (Mehlhorn  
805 et al., 2018). As organic-complexed copper is classified into the adsorbed pool in the grouping  
806 scheme used in the present study, these results are consistent with our observations. Metallic  
807 copper was not observed by XANES in either our work or this prior study, suggesting it may be a  
808 substantial species only in contaminated flooded soils, where it has been frequently observed  
809 (Weber et al., 2009a; Fulda et al., 2013a; Hofacker et al., 2013b). Nickel forms sulfide and clay-  
810 associated species in anoxic coastal sediments (Noël et al., 2015; Noël et al., 2017; Merrot et al.,  
811 2019), which were also found in the present study. The greater importance of the sulfide species  
812 in these prior studies results from substantial pyrite formation caused by the reduction of the ample  
813 sulfate in seawater. These coastal systems are thus not directly comparable to freshwater aquatic  
814 systems, which typically have lower sulfate inputs (Wieder and Lang, 1988; Castro et al., 2002;  
815 Vile et al., 2003; Knorr and Blodau, 2009). However, locations within terrestrial-aquatic interfaces

816 where sulfur-rich groundwater mixes with fresh surface waters represent potential hot spots of  
817 reactivity which may require additional study. Further, these prior studies investigated systems  
818 with naturally elevated nickel concentrations (up to 0.5 wt.%) associated with an ultramafic  
819 laterite, potentially leading to distinct speciation. Despite these differences, the nickel sulfide pool  
820 was ubiquitous in the aquatic systems investigated here. This is likely supported by high rates of  
821 sulfate reduction in freshwater aquatic systems (Segarra et al., 2015) despite the lower total sulfate  
822 levels observed in the absence of seawater inputs. We are unaware of studies of zinc speciation in  
823 anoxic sites that are uncontaminated or lack geogenic zinc accumulations.

824 This study also highlights that the observations of trace metal behavior in contaminated  
825 systems often are not predictive of the behavior of these elements when they occur at natural,  
826 background levels. As noted above, metallic copper was not observed at any site in this study  
827 despite frequently being reported for contaminated soils (Weber et al., 2009a; Weber et al., 2009b;  
828 Fulda et al., 2013a; Hofacker et al., 2015). Similarly, our studies find that zinc sulfide is limited in  
829 occurrence, with zinc more commonly binding to reduced organic sulfur species. The greater  
830 abundance of zinc sulfide in contaminated settings (Bostick et al., 2001; Webb and Gaillard, 2015)  
831 may thus be more prevalent in systems receiving substantial zinc inputs.

832 Finally, this study suggests that greater insight into the speciation of trace metals occurring  
833 at geological background levels may be gained using modern analytical tools. The data obtained  
834 in this study for trace metals occurring at or below geological background levels was enabled by  
835 recent developments in detectors and enhancements in beam flux at synchrotron lightsources. This  
836 demonstrates that spectroscopic investigation of trace metal in other uncontaminated terrestrial  
837 aquatic systems is viable, allowing further evaluation of whether the consistent speciation observed  
838 in the present work is a general geochemical phenomenon.

839 **5. CONCLUSIONS**

840 Trace metals in marsh wetland soils, riparian wetland soils, and stream bed sediments at  
841 three field sites display a series of similar features. Relative solid-phase concentrations of trace  
842 metals followed the trend  $Zn > Cu \approx Ni > Co$ , but their absolute abundance showed no predictive  
843 correlations with major element contents. Nickel, copper, and zinc all partially occurred as sulfur-  
844 bound species, yet the fractional abundance of these phases were insensitive to the bulk sulfur  
845 content, which varied by two orders of magnitude. Freshwater subsurface aquatic systems appear  
846 to contain adequate sulfur to stabilize sulfide minerals and metals complexed to reduced organic  
847 sulfur species despite their generally low total sulfur contents. Zinc and nickel also widely occurred  
848 in clay mineral structures, and all trace metals also occurred complexed to mineral surfaces and  
849 organic matter.

850 The lack of a relationship between sulfur-bound metal contents and total sulfur  
851 concentration may reflect the stability of other metal species or heterogeneous microenvironments  
852 in wetlands and stream beds. While reaction with iron might limit the availability of reduce sulfur  
853 to bind trace metals, it is difficult to envision how this would consistently generate similar fractions  
854 of metal-sulfur species across an array of soils and sediments. More broadly, similar species of  
855 each trace metal occurred in the soils and sediments at all field sites, often in similar proportions.  
856 The consistent trace metal speciation among distant and unrelated sites suggests that a small  
857 number of species may dominate freshwater wetland soils and stream sediments. Future  
858 investigation of the temporal dynamics of trace metal species in such settings may provide  
859 additional insight into the processes that yield consistent trace metal speciation across diverse  
860 freshwater aquatic systems.

861

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882

883 **Appendix A. Supplementary Material**

884 The Supplementary Materials include information on the synthesis and sources of standards for  
885 XANES and EXAFS analyses, a summary of surface water compositions and extractable nutrients,

886 a map of the field sites, plots of the standards used for XANES and EXAFS analyses, tabulations  
887 of the chemical analyses and spectroscopic fitting results, and evidence for a zinc oxide  
888 contaminant source.

889

890 **Research Data**

891 Research Data associated with this article can be accessed at <https://doi.org/10.15485/1773008>.

892

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1260 **TABLES**

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1262 **Table 1.** Sample depth (cm) for soil and sediment core segments.

| <b>Sample Location</b> | <b>C1-1</b> | <b>C1-2</b> | <b>C2-1</b> | <b>C2-2</b> |
|------------------------|-------------|-------------|-------------|-------------|
| Marsh 1                | 0-7.5       | 7.5-15.3    | 0-9         | 9-18.4      |
| Marsh 2                | 0-6.5       | 6.5-11      | 0-9         | 9-12        |
| Riparian 1             | 0-4.5       | 4.5-9.5     | 0-4.5       | 4.5-10.5    |
| Riparian 2             | 0-4.5       | 4.5-9.5     | 0-4         | 4-10        |
| Stream 1               | 0-8         | 8-21.5      | 0-3         | 3-19.5      |
| Stream 2*              | 0-8         | 8-12        | 0-8         | 8-18        |

1263 \*A third segment was obtained for C2 from this location over the depth range of 18 to 28.5 cm.

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1266 **FIGURE CAPTIONS**

1267 **Figure 1.** Powder X-ray diffraction (XRD) patterns of the surface soil or sediment layer over the  
1268 full range (top) or the low angle region more sensitive to clay minerals. Data for duplicate cores  
1269 are shown for location Stream 1 because of their distinct mineralogy.

1270 **Figure 2.** Total carbon, sulfur, iron, and manganese contents in the surface layer of the soils and  
1271 sediments at the study sites. The error bars reflect the standard deviations of triplicate analyses of  
1272 samples of individual cores added in quadrature to the standard deviation of the mean values for  
1273 the two duplicate cores collected at each site (Data tabulated in Table S6).

1274 **Figure 3.** Dissolved (top) and total solid phase (bottom) trace metal concentrations for the study  
1275 sites. The error bars for the dissolved concentrations are the uncertainties of individual analyses  
1276 obtained from linear regression of the calibration curves. Error bars from the solid phase  
1277 concentration reflect the calibration-derived uncertainties for samples from individual cores  
1278 added in quadrature to the standard deviation of the mean values for the two duplicate cores  
1279 collected at each site (Data tabulated in Tables S3 and S4).

1280 **Figure 4.** Sulfur K edge XANES spectra (solid) and fits (dashed) and the associated distribution  
1281 of species for the surface layer soils and sediments at the study locations (Results tabulated in  
1282 Table S12).

1283 **Figure 5.** Iron K-edge XANES (top) and EXAFS (bottom) spectra (solid) and fits (dashed) and  
1284 the associated distribution of species for the surface layer soils and sediments at the study  
1285 locations. (Results tabulated in Tables S13 and S14)

1286 **Figure 6.** Cu-K edge XANES spectra (solid) and fits (dashed) and the associated distribution of  
1287 species for the surface layer soils and sediments at the study locations (Results tabulated in Table  
1288 S15).

1289 **Figure 7.** Ni-K edge XANES spectra (solid) and fits (dashed) and the associated distribution of  
1290 species for the surface layer soils and sediments at the study locations (Results tabulated in Table  
1291 S16).

1292 **Figure 8.** Zn-K edge XANES spectra (solid) and fits (dashed) and the associated distribution of  
1293 species for the surface layer soils and sediments at the study locations (Results tabulated in Table  
1294 S17).

1295 **Figure 9.** Correlation coefficients (R) and associated p-values derived from weighted linear  
1296 regression of the solid-phase concentrations of each element pair. Data for the stream bed site  
1297 excluded the single sample containing pyrite.

1298 **Figure 10.** (A) Average and standard deviation of the fraction of trace metals bound to reduced  
1299 sulfur as determined by XANES spectroscopy. (B) Variation in the fraction of each trace metal  
1300 bound to reduced sulfur compared to the total sulfur content of the samples compared to trend  
1301 lines for each metal (Cu:  $R = -0.06$ ,  $p = 0.03$ ; Ni:  $R = 0.37$ ,  $p = 0.001$ ; Zn:  $R = 0.18$ ,  $p = 0.1$ )  
1302 determined through linear regression weighted by the bivariate uncertainties.

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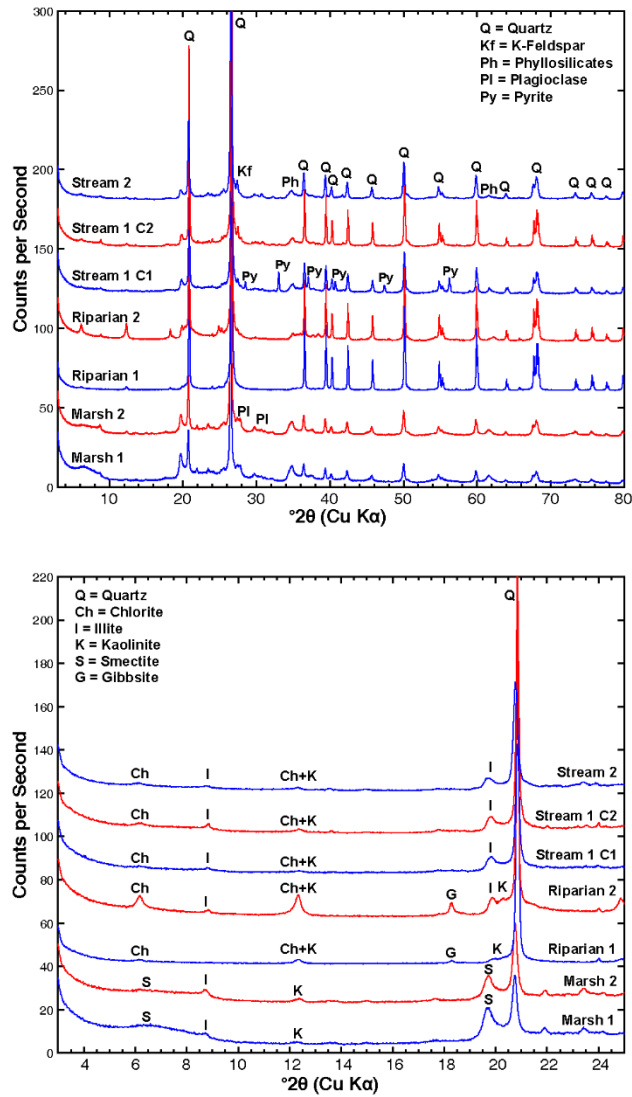
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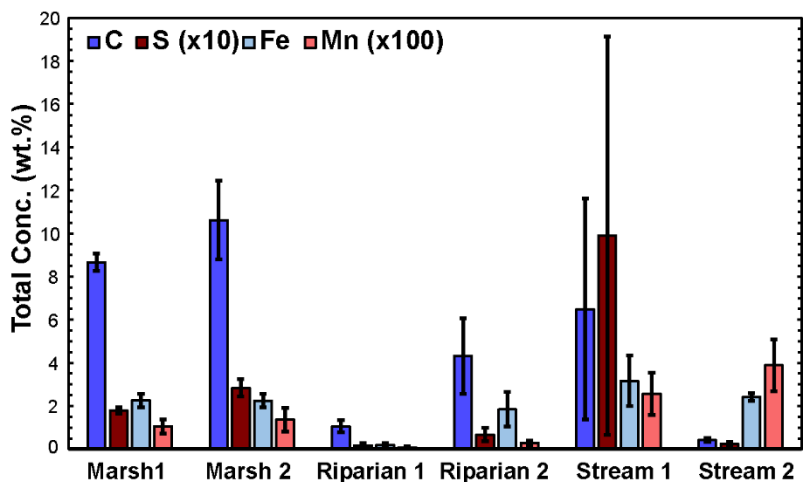
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1313 **Figure 1.** Powder X-ray diffraction (XRD) patterns of the surface soil or sediment layer over the

1314 full range (top) or the low angle region more sensitive to clay minerals. Data for duplicate cores

1315 are shown for location Stream 1 because of their distinct mineralogy.

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**Figure 2.** Total carbon, sulfur, iron, and manganese contents in the surface layer of the soils and

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sediments at the study sites. The error bars reflect the standard deviations of triplicate analyses of

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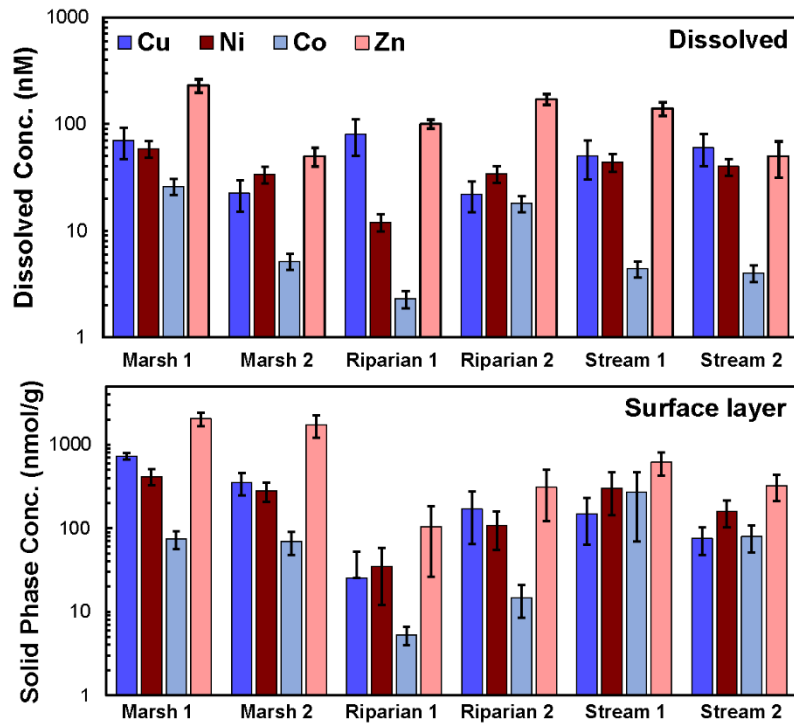
samples of individual cores added in quadrature to the standard deviation of the mean values for

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the two duplicate cores collected at each site (Data tabulated in Table S6).

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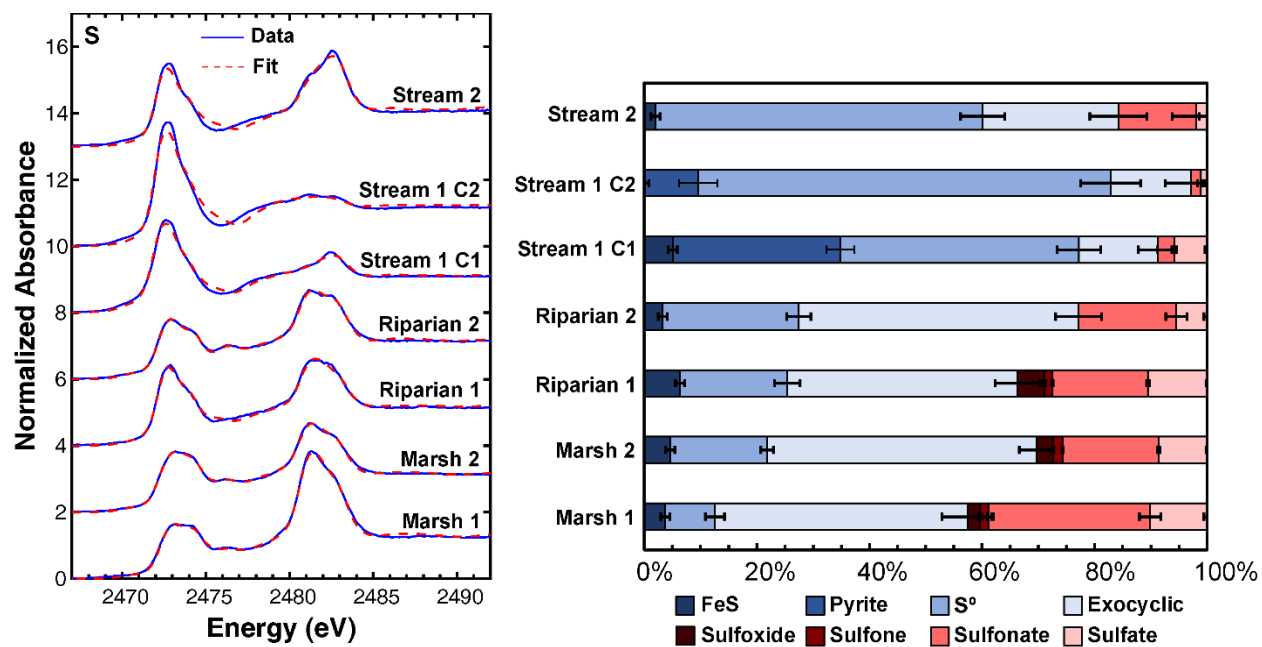
1325 **Figure 3.** Dissolved (top) and total solid phase (bottom) trace metal concentrations for the study  
1326 sites. The error bars for the dissolved concentrations are the uncertainties of individual analyses  
1327 obtained from linear regression of the calibration curves. Error bars from the solid phase  
1328 concentration reflect the calibration-derived uncertainties for samples from individual cores  
1329 added in quadrature to the standard deviation of the mean values for the two duplicate cores  
1330 collected at each site (Data tabulated in Tables S3 and S4).

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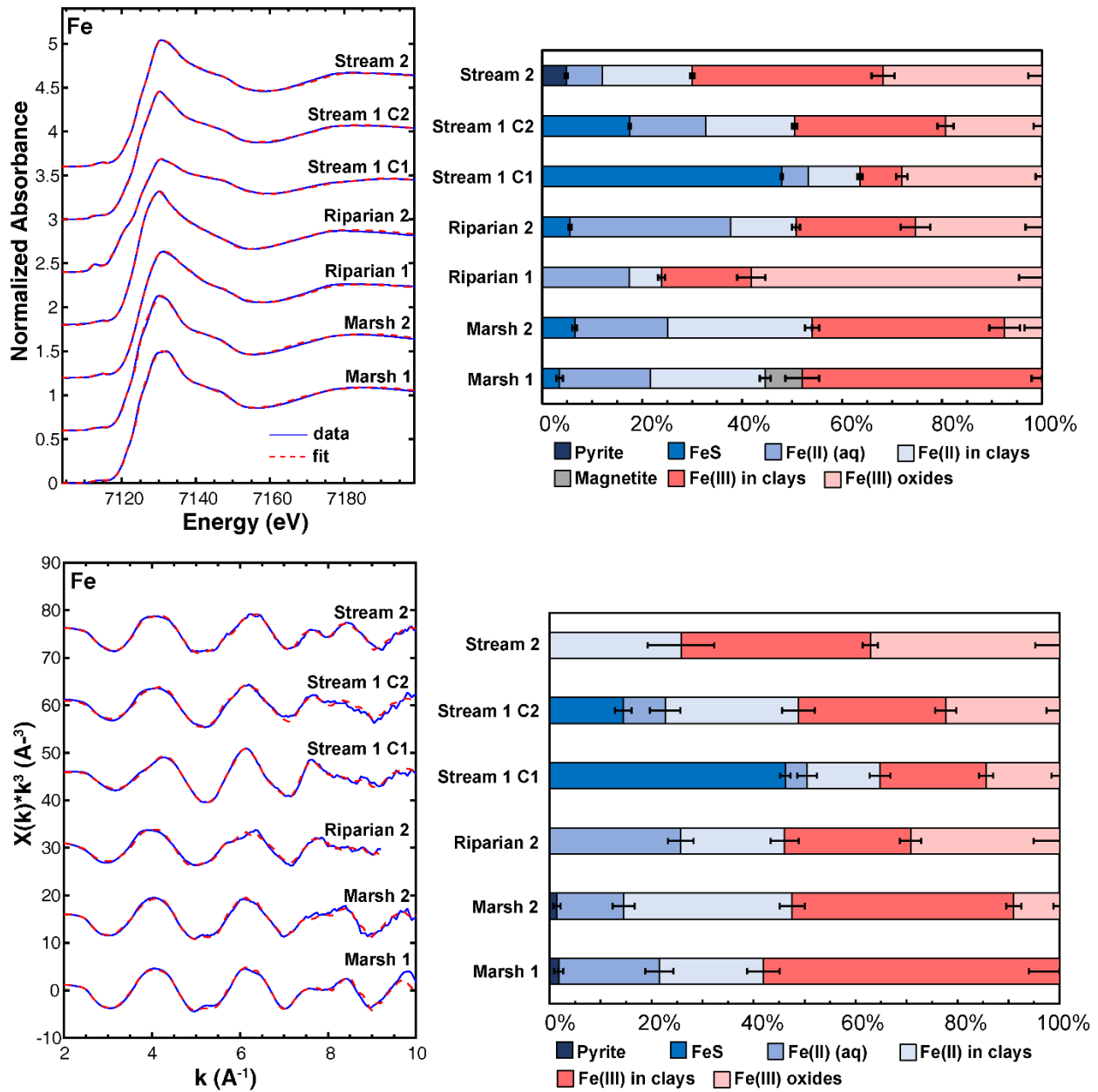
1335

1336 **Figure 4.** Sulfur K edge XANES spectra (solid) and fits (dashed) and the associated distribution

1337 of species for the surface layer soils and sediments at the study locations (Results tabulated in

1338 Table S12).

1339



1341

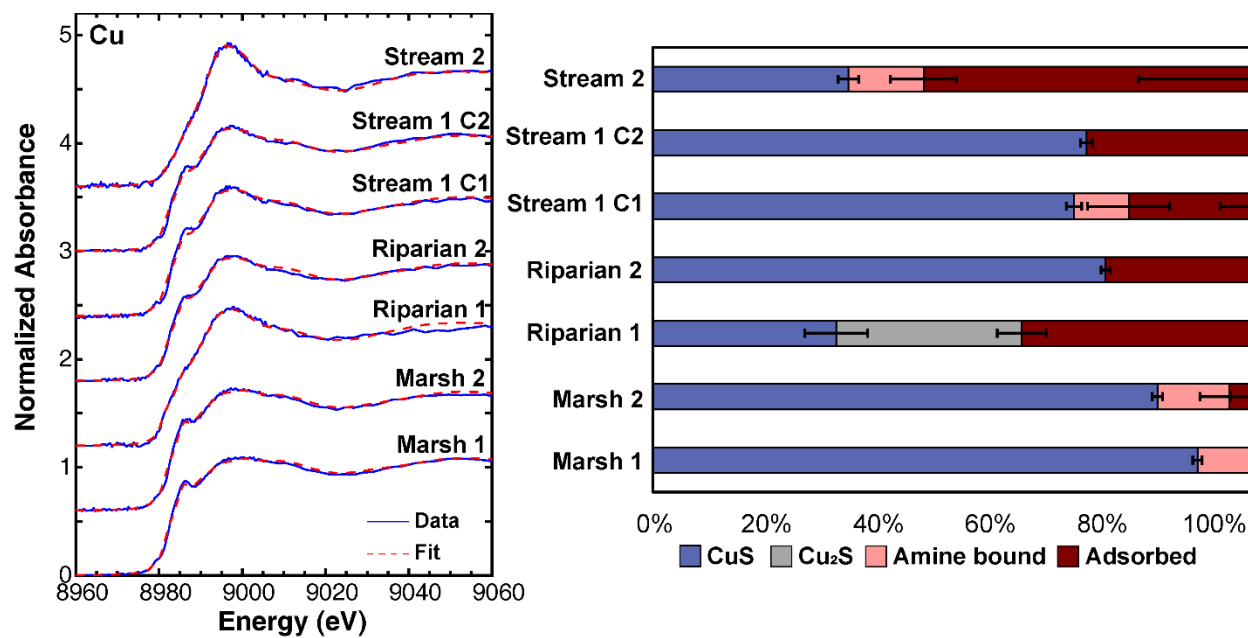
1342 **Figure 5.** Iron K-edge XANES (top) and EXAFS (bottom) spectra (solid) and fits (dashed) and

1343 the associated distribution of species for the surface layer soils and sediments at the study

1344 locations. (Results tabulated in Tables S13 and S14)

1345

1346



1347

1348 **Figure 6.** Cu-K edge XANES spectra (solid) and fits (dashed) and the associated distribution of

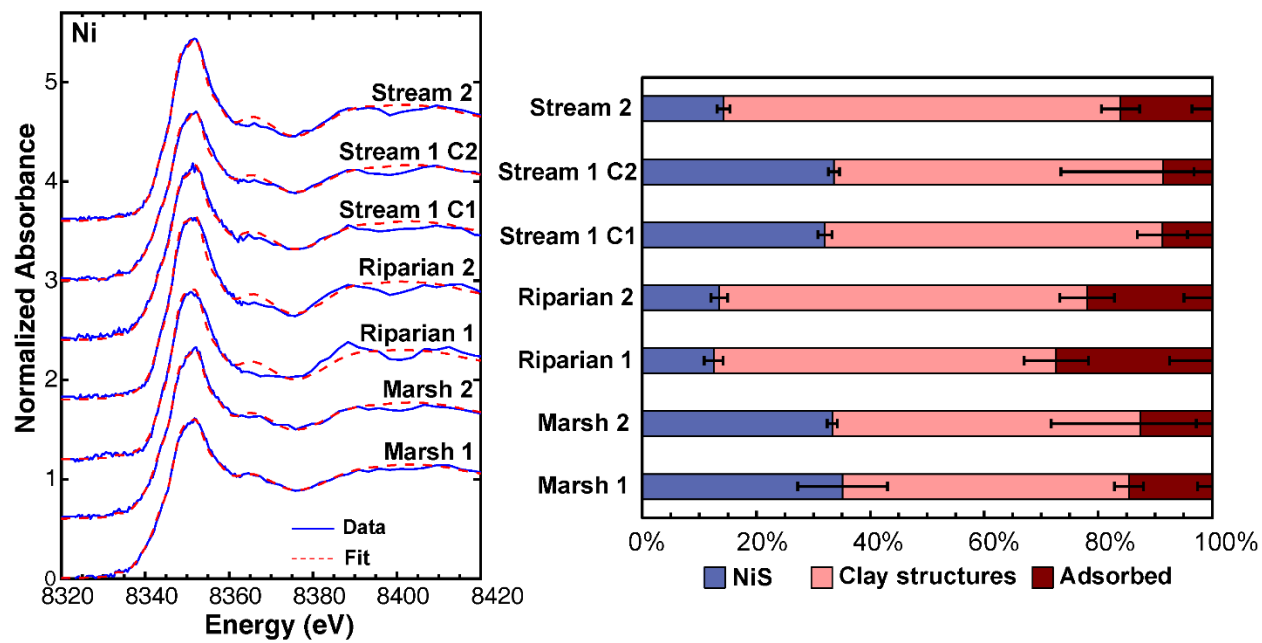
1349 species for the surface layer soils and sediments at the study locations (Results tabulated in Table

1350 S15).

1351

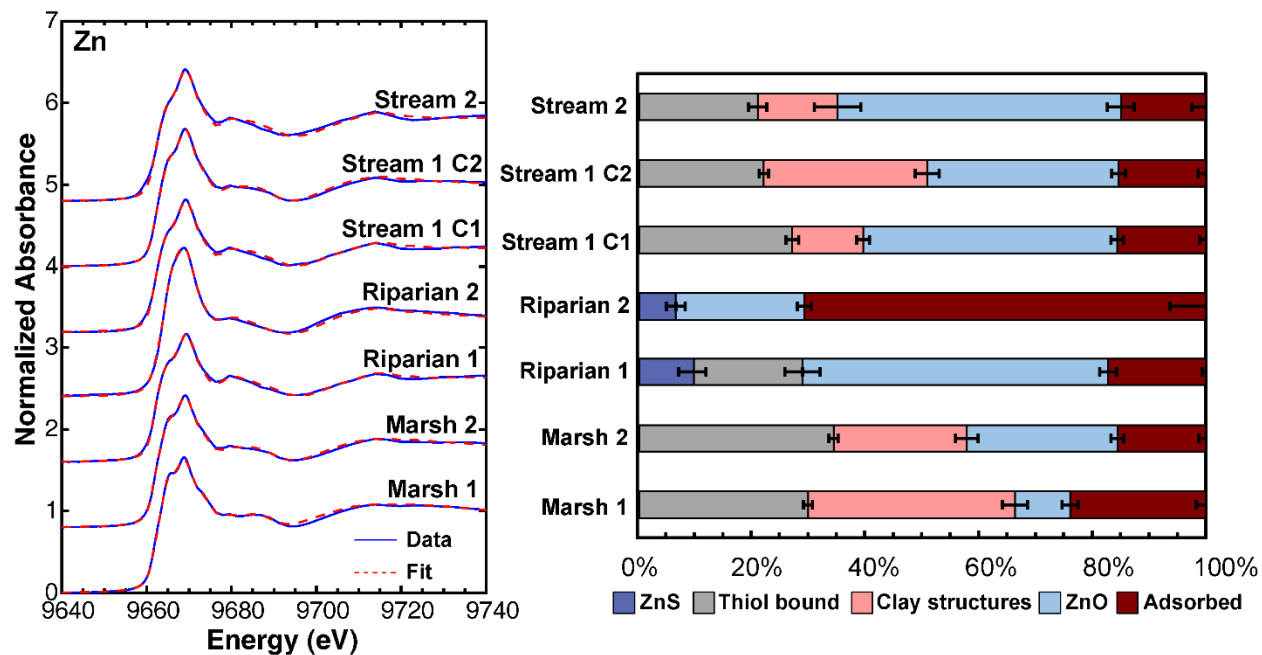
1352





1353  
 1354 **Figure 7.** Ni-K edge XANES spectra (solid) and fits (dashed) and the associated distribution of  
 1355 species for the surface layer soils and sediments at the study locations (Results tabulated in Table  
 1356 S16).

1357  
 1358  
 1359



1360

1361 **Figure 8.** Zn-K edge XANES spectra (solid) and fits (dashed) and the associated distribution of  
 1362 species for the surface layer soils and sediments at the study locations (Results tabulated in Table  
 1363 S17).

1364

**Marsh Wetlands**

|    |                       |                        |                        |                     |                      |                      |    |
|----|-----------------------|------------------------|------------------------|---------------------|----------------------|----------------------|----|
| Fe | <b>-0.91</b><br>0.001 |                        |                        |                     |                      |                      |    |
| S  | <b>0.92</b><br><0.001 | <b>-0.78</b><br><0.001 |                        |                     |                      |                      |    |
| Mn | <b>0.45</b><br>0.2    | <b>0.68</b><br>0.05    | <b>-0.23</b><br>0.2    |                     |                      |                      |    |
| Co | <b>-0.47</b><br>0.6   | <b>0.59</b><br>0.1     | <b>-0.46</b><br>0.2    | <b>0.80</b><br>0.1  |                      |                      |    |
| Ni | <b>-0.14</b><br>0.03  | <b>0.13</b><br>0.9     | <b>-0.42</b><br>0.001  | <b>-0.24</b><br>0.8 | <b>0.42</b><br>0.04  |                      |    |
| Cu | <b>0.40</b><br>0.01   | <b>-0.39</b><br>0.003  | <b>-0.14</b><br><0.001 | <b>-0.66</b><br>0.3 | <b>-0.12</b><br>0.05 | <b>0.79</b><br>0.001 |    |
| Zn | <b>0.69</b><br>0.06   | <b>-0.68</b><br>0.01   | <b>0.49</b><br>0.6     | <b>-0.38</b><br>0.1 | <b>0.02</b><br>0.05  | <b>0.64</b><br>0.004 |    |
|    | <b>0.74</b><br>0.002  |                        |                        |                     |                      |                      |    |
|    | C                     | Fe                     | S                      | Mn                  | Co                   | Ni                   | Cu |

**Riparian Wetlands**

|    |                       |                       |                       |                       |                       |                       |    |
|----|-----------------------|-----------------------|-----------------------|-----------------------|-----------------------|-----------------------|----|
| Fe | <b>0.95</b><br><0.001 |                       |                       |                       |                       |                       |    |
| S  | <b>0.98</b><br><0.001 | <b>0.94</b><br>0.004  |                       |                       |                       |                       |    |
| Mn | <b>0.98</b><br><0.001 | <b>0.97</b><br><0.001 | <b>0.98</b><br>0.001  |                       |                       |                       |    |
| Co | <b>0.95</b><br><0.001 | <b>0.96</b><br><0.001 | <b>0.96</b><br>0.003  | <b>0.98</b><br><0.001 |                       |                       |    |
| Ni | <b>0.97</b><br><0.001 | <b>0.92</b><br><0.001 | <b>0.98</b><br>0.003  | <b>0.97</b><br><0.001 | <b>0.96</b><br><0.001 |                       |    |
| Cu | <b>0.99</b><br><0.001 | <b>0.93</b><br><0.001 | <b>0.98</b><br>0.003  | <b>0.98</b><br><0.001 | <b>0.97</b><br><0.001 | <b>1.00</b><br><0.001 |    |
| Zn | <b>0.93</b><br><0.001 | <b>0.88</b><br><0.001 | <b>0.93</b><br><0.001 | <b>0.95</b><br><0.001 | <b>0.95</b><br><0.001 | <b>0.98</b><br><0.001 |    |
|    |                       |                       |                       |                       |                       | <b>0.98</b><br><0.001 |    |
|    | C                     | Fe                    | S                     | Mn                    | Co                    | Ni                    | Cu |

**Stream Bed**

|    |                        |                       |                       |                      |                       |                      |    |
|----|------------------------|-----------------------|-----------------------|----------------------|-----------------------|----------------------|----|
| Fe | <b>-0.78</b><br><0.001 |                       |                       |                      |                       |                      |    |
| S  | <b>0.91</b><br><0.001  | <b>-0.57</b><br>0.001 |                       |                      |                       |                      |    |
| Mn | <b>-0.58</b><br>0.001  | <b>0.85</b><br>0.001  | <b>-0.09</b><br>0.002 |                      |                       |                      |    |
| Co | <b>0.32</b><br>1.0     | <b>0.74</b><br><0.001 | <b>-0.20</b><br>0.001 | <b>0.64</b><br>0.001 |                       |                      |    |
| Ni | <b>0.52</b><br>0.9     | <b>0.60</b><br><0.001 | <b>0.07</b><br>0.002  | <b>0.40</b><br>0.001 | <b>0.97</b><br><0.001 |                      |    |
| Cu | <b>0.48</b><br>0.2     | <b>-0.16</b><br>0.03  | <b>0.55</b><br>0.3    | <b>-0.32</b><br>0.09 | <b>0.14</b><br>0.02   | <b>0.48</b><br>0.004 |    |
| Zn | <b>0.61</b><br>0.2     | <b>0.22</b><br>0.001  | <b>0.66</b><br>0.002  | <b>0.41</b><br>0.002 | <b>0.73</b><br>0.001  | <b>0.90</b><br>0.001 |    |
|    |                        |                       |                       |                      |                       | <b>0.90</b><br>0.003 |    |
|    | C                      | Fe                    | S                     | Mn                   | Co                    | Ni                   | Cu |

1365

1366

**Figure 9.** Correlation coefficients (R) and associated p-values derived from weighted linear

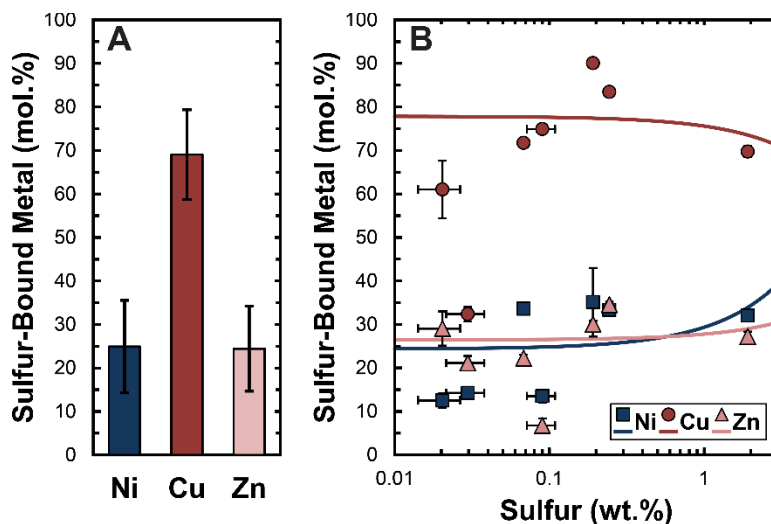
1367

regression of the solid-phase concentrations of each element pair. Data for the stream bed site

1368

excluded the single sample containing pyrite.

1369



1370  
 1371 **Figure 10.** (A) Average and standard deviation of the fraction of trace metals bound to reduced  
 1372 sulfur as determined by XANES spectroscopy. (B) Variation in the fraction of each trace metal  
 1373 bound to reduced sulfur compared to the total sulfur content of the samples compared to trend  
 1374 lines for each metal (Cu:  $R = -0.06$ ,  $p = 0.03$ ; Ni:  $R = 0.37$ ,  $p = 0.001$ ; Zn:  $R = 0.18$ ,  $p = 0.1$ )  
 1375 determined through linear regression weighted by the bivariate uncertainties.

1376

1 SUPPLEMENTARY MATERIAL FOR

2  
3 **Consistent Controls on Trace Metal Micronutrient Speciation in**  
4 **Wetland Soils and Stream Sediments**

5  
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20

## 21 **Synthesis of Reference Standards**

22 Copper citrate was made using equal volumes (100 mL) of 0.15 M copper sulfate ( $\text{CuSO}_4$ )  
23 and 0.1 M sodium citrate. The solutions were heated to  $100^\circ\text{C}$  on a heated magnetic stir plate. The  
24  $\text{CuSO}_4$  solution was added to the citrate solution while continuously stirring in the fume hood. The  
25 mixture was stirred for 2 hours at  $100^\circ\text{C}$  then centrifuged at 4500 rpm for 5 minutes (Beckman  
26 Coulter SX4250 rotor) and the supernatant discarded. Copper citrate was washed three times with  
27 50 mL of ultrapure water, centrifuged, and the supernatant discarded. It was then dried in a vacuum  
28 desiccator and ground with a mortar and pestle.

29 Zinc oxalate was prepared by mixing an equimolar ratio of zinc nitrate ( $\text{Zn}(\text{NO}_3)_2$ ) and  
30 oxalic acid solutions as described by (Raj et al., 2011). While stirring, 10 mL of 1 M  $\text{Zn}(\text{NO}_3)_2$   
31 was added to 20 mL of 0.5 M oxalic acid. A milky white precipitate formed immediately. The  
32 solution was centrifuged at 4500 rpm for 10 minutes and the supernatant discarded. The precipitant  
33 was then rinsed three times with 50 mL of ultrapure water, centrifuged, and the supernatant  
34 discarded. Zinc oxalate was dried in a vacuum desiccator and then ground with a mortar and pestle.

35 To synthesize metal-cysteine, metal-histidine, and metal-glutathione complexes, solutions  
36 with a 1:10 ratio of metal to ligand were reacted based on published methods (Kelly et al., 2002;  
37 Beauchemin et al., 2004; Montargès-Pelletier et al., 2008; Dokken et al., 2009; Manceau et al.,  
38 2013; Van Der Ent et al., 2017). All solutions were prepared in the anaerobic chamber and using  
39 deoxygenated ultrapure water. Solutions containing 7 mM of copper chloride ( $\text{CuCl}_2$ ), nickel  
40 chloride ( $\text{NiCl}_2$ ), or zinc nitrate ( $\text{Zn}(\text{NO}_3)_2$ ) and 70 mM of L-cysteine hydrochloride, L-histidine  
41 hydrochloride, or glutathione (reduced) were adjusted to  $\text{pH } 7 \pm 0.2$  using NaOH. The solutions  
42 containing the metal-ligand complexes were then removed from the anaerobic chamber and frozen.  
43 The samples were freeze-dried before being ground using a mortar and pestle.

44 Standards of Ni, Cu, and Zn adsorbed to goethite were also prepared. Goethite was  
45 synthesized using established methods (Schwertmann and Cornell, 2000). Briefly, 50 mL of 1 M  
46 iron nitrate ( $\text{Fe}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$ ) was added to 90 mL of 5 M potassium hydroxide (KOH) over 1.5  
47 min while continuously stirring. The solution was immediately diluted to 1 L using ultrapure water  
48 and heated at 70°C for 60 h. After cooling, the supernatant was discarded, and the goethite was  
49 rinsed three times with approximately 150 mL of ultrapure water through a vacuum filter. The  
50 goethite was stored as a suspension in ultrapure water and powder X-ray diffraction (XRD) was  
51 used to confirm that no other crystalline minerals precipitated. A solution of 0.01 M sodium  
52 chloride (NaCl), 0.001 M MOPS buffer, and 0.004 M  $\text{CuCl}_2$ ,  $\text{NiCl}_2$ , or  $\text{Zn}(\text{NO}_3)_2$  was reacted with  
53 8 g/L goethite at pH 7.5. Samples were wrapped in Al foil to prevent photochemical reactions and  
54 rotated end-over-end for 24 h. They were then centrifuged at 4500 rpm and the supernatant  
55 decanted, filtered, acidified to 2% nitric acid, and analyzed via ICP-MS to confirm substantial  
56 adsorption had occurred.

57 Ni- and Zn-substituted trioctahedral Fe(II)-Mg smectite standards were synthesized by  
58 modifying the recipe for “Clay D” in Chemtob et al., (2015) to include 0.5 wt.% Ni or Zn,  
59 respectively, replacing an equivalent amount of Mg in the structure. Ni- and Fe(II)-substituted  
60 serpentines were prepared using a similar sol-gel procedure as used for the smectite standards. 18  
61 mL of 0.5 M sodium metasilicate was added to 13 mL of a solution containing 1 M magnesium  
62 chloride and 0.003 M nickel chloride or iron(II) chloride. The resulting gels were aged overnight  
63 and then washed via centrifugation and decanting of the fluid, which was replaced with ultrapure  
64 water and centrifuged and decanted again. This process was repeated two additional times. The  
65 gels were then resuspended in ultrapure water and heated in a PTFE-lined hydrothermal vessel at  
66 200°C for 14 days. After cooling the resulting solids were separated by vacuum filtration and dried

67 in a vacuum desiccator. XRD confirmed that poorly crystalline turbostratic lizardite was produced.  
68 Ni-substituted brucite was also prepared by mixing 20 mL of 1 M sodium hydroxide with 15 mL  
69 of 1 M magnesium chloride and 0.003 M nickel chloride. A white precipitate formed quickly and  
70 was aged overnight before washing via centrifugation and then drying in a vacuum desiccator. The  
71 formation of brucite was confirmed by XRD.

72 Ni- and Zn-substituted magnetite, as well as metal-free magnetite, were prepared by mixing  
73 stoichiometric amounts of iron(II) chloride and iron(III) chloride solutions. Nickel chloride and  
74 zinc chloride were added into the iron(III) chloride solution before mixing to yield an ~0.3 mol%  
75 substitution level for each metal; this step was not used in the metal-free magnetite synthesis.  
76 Sodium hydroxide was used to adjust the pH to  $7.0\pm 0.1$  after fluid mixing, yielding a black  
77 precipitate. This suspension was continuously mixed overnight before being separated by vacuum  
78 filtration and then dried in a vacuum desiccator. All steps were performed in an anaerobic chamber.  
79 Magnetite was the sole product identified by XRD.

80 Goethite, hematite, lepidocrocite, and 2-line ferrihydrite were prepared following  
81 established procedures (Schwertmann and Cornell, 2000). The lepidocrocite was provided by  
82 Walter Schenkeveld. Mackinawite was prepared through direct precipitation in an anaerobic  
83 chamber following previously-described procedures (Swanner et al., 2019). Smectites from the  
84 Source Clay Repository were size fractionated and Na-saturated as previously described (Catalano  
85 and Brown, 2005). Preparation of all standards for data collection are described in the full  
86 manuscript.

87

88

89



90 **Surface Water Composition**

91 Major element concentrations and pH were similar among the two sampling locations at  
92 each of the field sites investigated (Table S1). While differences were observed between the three  
93 field sites, the overall water compositions were more distinct at the riparian wetlands compared to  
94 the marsh wetlands and the stream sediment sites. Surface waters in the riparian wetlands are dilute  
95 with sodium as the dominant cation (~280  $\mu\text{M}$ ) and pH values of 5.5 to 5.6. The marsh wetlands  
96 and stream system were more alkaline, having pH values of 7.5 to 8.1, with substantially higher  
97 calcium, magnesium, and sulfate concentrations. Sodium and chloride concentrations are higher  
98 in the stream system compared to the marsh wetlands, and the stream sites have the highest ionic  
99 strengths of the sites examined. Dissolved silica concentrations indicate that all fluids were  
100 supersaturated with respect to quartz but undersaturated with respect to amorphous silica.

101 Dissolved nutrient concentrations (Fig. S2; Table S5) varied substantially among the sites.  
102 Nitrate was greater than ammonium at all sampling locations except for location Riparian 2, where  
103 both were below detection limit. Notably, the stream sites had substantially elevated nitrate  
104 concentrations (160 to 210  $\mu\text{M}$ ) compared to the marsh and riparian wetland sites (<15  $\mu\text{M}$ ),  
105 because of contamination at the headwaters and wastewater treatment plant discharge. Dissolved  
106 phosphate concentrations were similar at the marsh and riparian wetlands, but the stream sites  
107 displayed slightly higher concentrations.

108 Alkalinity and dissolved organic carbon (DOC) were not measured at the time of sampling  
109 because of an oversight. Measurements of DOC on archived samples provide an upper bound on  
110 values in the original waters. Bicarbonate concentrations and  $\text{pCO}_2$  were estimated using two  
111 approaches: (1) from dissolved inorganic carbon (DIC) measurements on archived samples and  
112 (2) via charge balance constraints. For the DIC-based estimation approach, the measured water

113 composition as well as the DIC and pH from the time of DIC measurement were input into a fully-  
114 speciated geochemical model. The pH of most samples were greater than the field-measured values  
115 because of CO<sub>2</sub> degassing. However, alkalinity is conserved during degassing. Field values of  
116 bicarbonate and pCO<sub>2</sub> were thus determined by increasing the pCO<sub>2</sub> in the geochemical model  
117 until the original, field-measured pH was obtained. For the charge-balance approach, measured  
118 anion concentrations were inadequate to balance the cation charge, indicating a missing  
119 bicarbonate component. This bicarbonate was added to the system in a geochemical model to reach  
120 charge balance while maintaining the field-measured pH and other aspects of the fluid  
121 composition. Both estimation approaches indicate that bicarbonate is the dominant anion in all  
122 systems and yield similar pCO<sub>2</sub> values (Table S1).

123         The marsh wetlands host Ca-Mg-HCO<sub>3</sub> fluids, the stream sites Ca-HCO<sub>3</sub> fluids, and the  
124 riparian wetlands Na-HCO<sub>3</sub> fluids. pCO<sub>2</sub> values were lowest in the marsh wetlands and highest in  
125 the riparian wetlands, with all but one location sampled being elevated above equilibrium with the  
126 atmospheric concentration. Filtered but non-acidified water samples returned to the laboratory  
127 from the riparian wetland field sites had pH values of 7.2 to 7.3 immediately upon arrival and 7.7  
128 after longer storage. This is consistent with a large pH increase associated with degassing of  
129 fluids containing highly elevated pCO<sub>2</sub> values and suggesting that the estimated bicarbonate  
130 concentrations are relatively accurate.

131

## 132 **Extractable Nutrients in the Soils and Sediments**

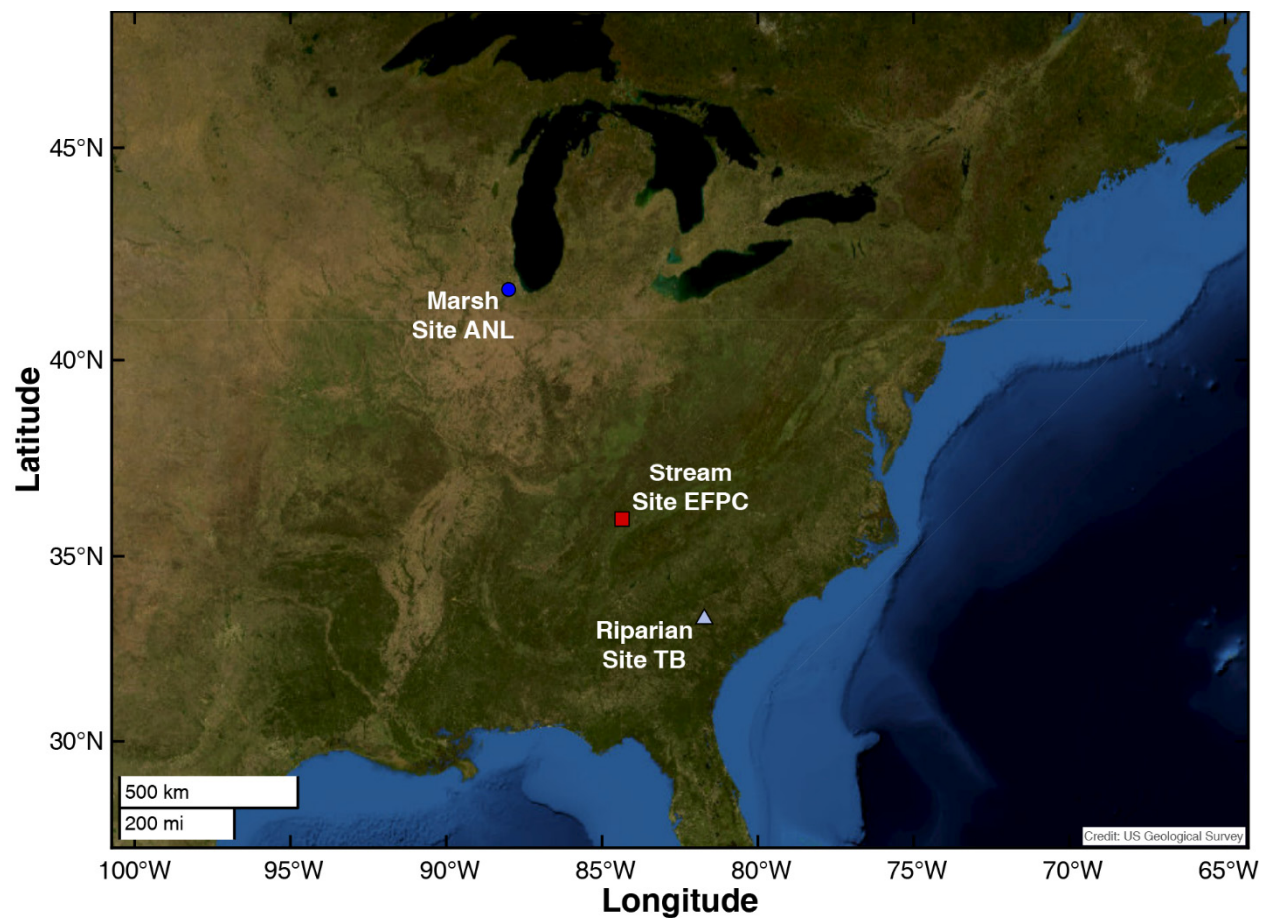
133         Extractable ammonium (Fig. S2) was substantially greater than extractable nitrate at all  
134 sites, often by an order of magnitude, showing an opposite pattern of the surface waters. This is  
135 consistent with ammonium retention via cation exchange but may also reflect denitrification. The

136 marsh soils contained the highest extractable ammonium concentrations and were the only soils  
137 to show clear evidence for smectites in XRD (Fig. 1), suggesting that cation exchange was at  
138 least partially responsible for the higher ammonium concentrations. Extractable phosphate was  
139 highest in the marsh soils (Fig. S2), with concentrations obtained from location Stream 2 and  
140 both riparian wetland sites an order of magnitude lower.

141

#### 142 **Zinc Speciation in Cellulose Acetate Tape**

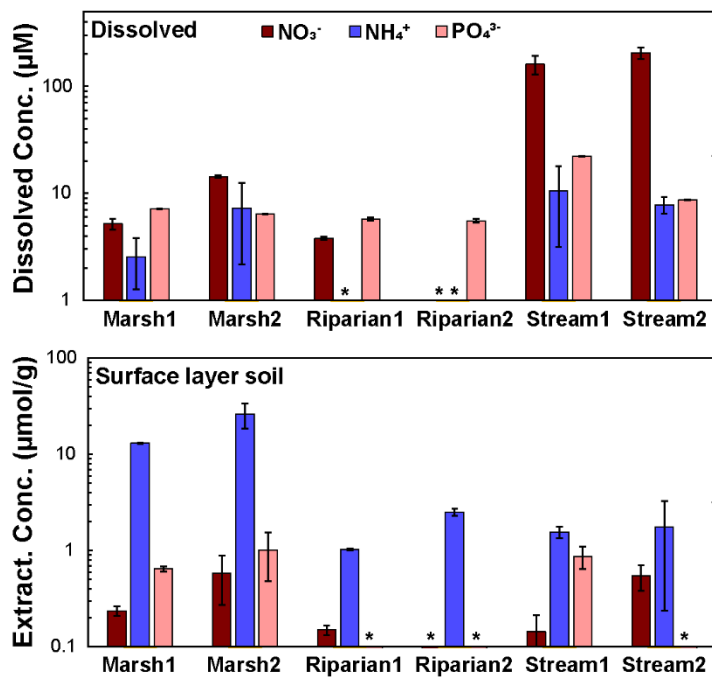
143 Cellulose acetate tape (e.g., Scotch<sup>TM</sup> tape in the United States) is widely used for sample  
144 mounting at synchrotron lightsources. It was noted during measurements at Advanced Photon  
145 Source beamline 12-BM-B that the tape contained zinc based on its X-ray fluorescence spectrum.  
146 A Zn K-edge XANES spectrum was collected in fluorescence yield from a stack of 10 layers of  
147 tape. This is spectrally similar to the spectrum of zinc oxide (Fig. S9), indicating that this is the  
148 dominant form of zinc in the tape.



149

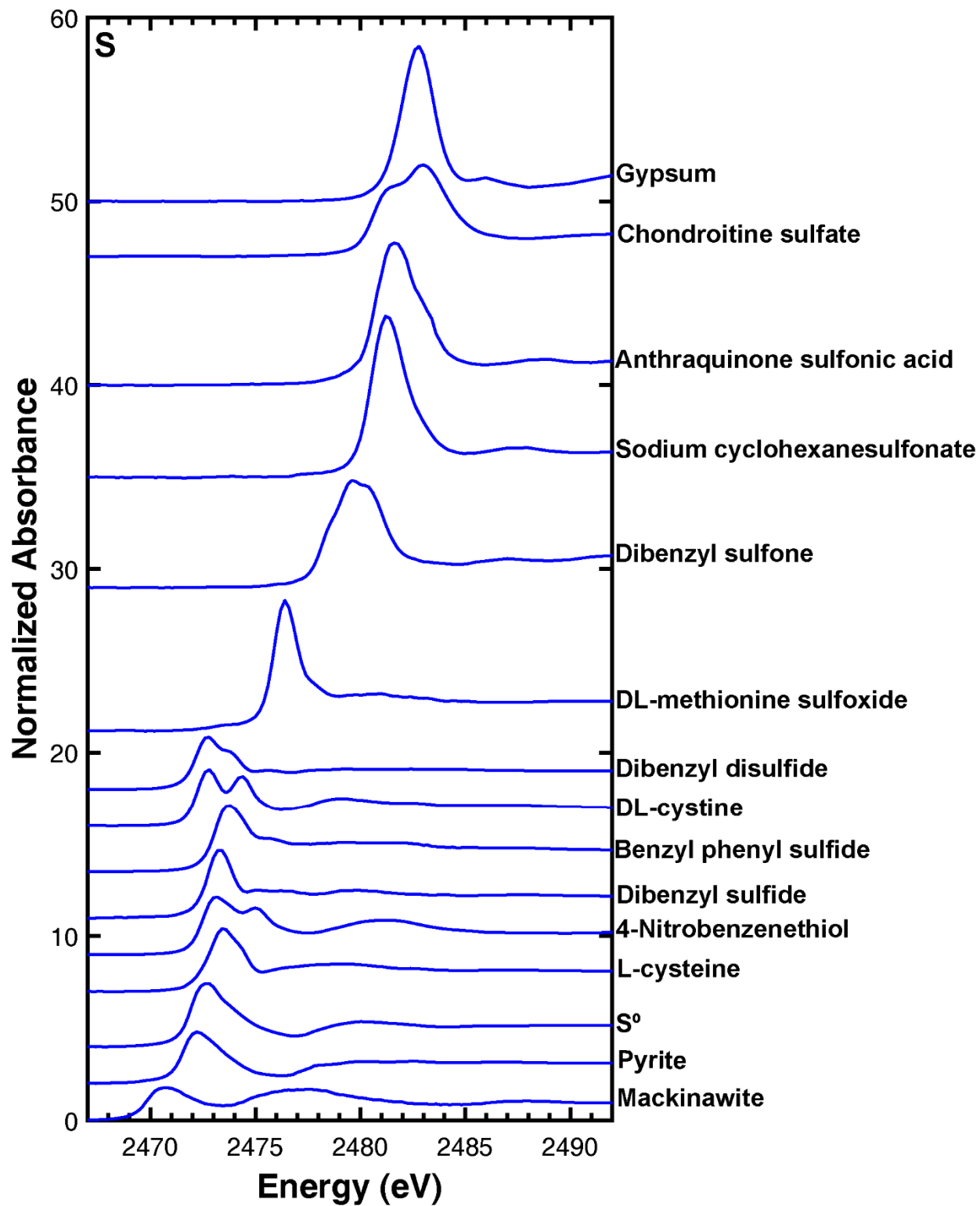
150 **Figure S1.** Location of the three field sites: Marsh wetlands at Argonne National Laboratory  
151 (Site ANL), stream bed sediments in the East Fork Poplar Creek (Site EFPC), and riparian  
152 wetlands in the Tims Branch watershed (Site TB). Details site descriptions and coordinates for  
153 these locations are provided in the Materials and Methods.

154



156

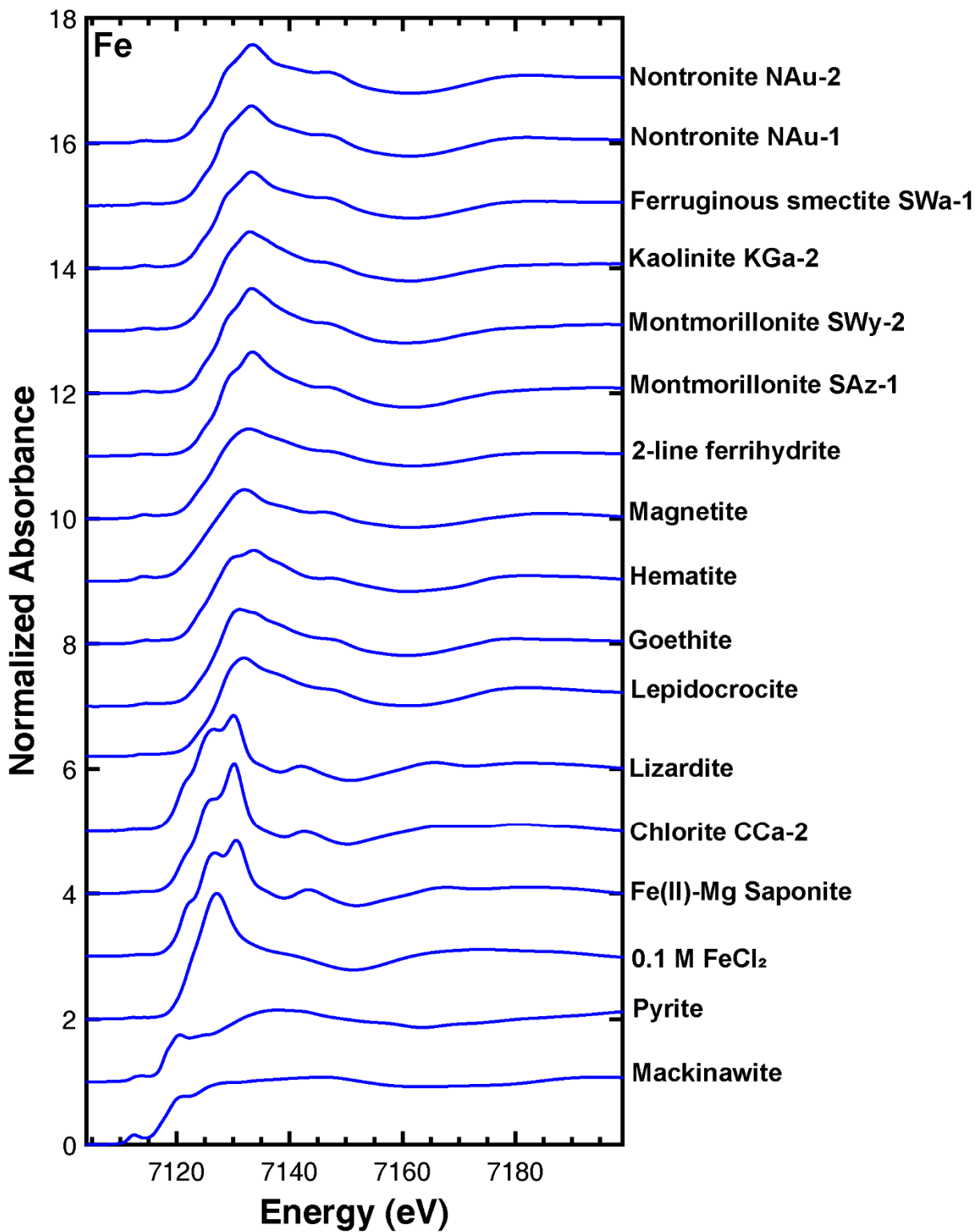
157 **Figure S2.** Dissolved (top) and solid-phase extractable (bottom) nitrate, ammonium, and phosphate  
 158 concentrations for the study sites. Sample with an asterisk were below the  $\sim 2 \mu\text{M}$  or  $\sim 0.04 \mu\text{mol/g}$   
 159 detection limit.



160

161 **Figure S3.** XANES spectra of sulfur reference standards. See Table S7 for more information.

162

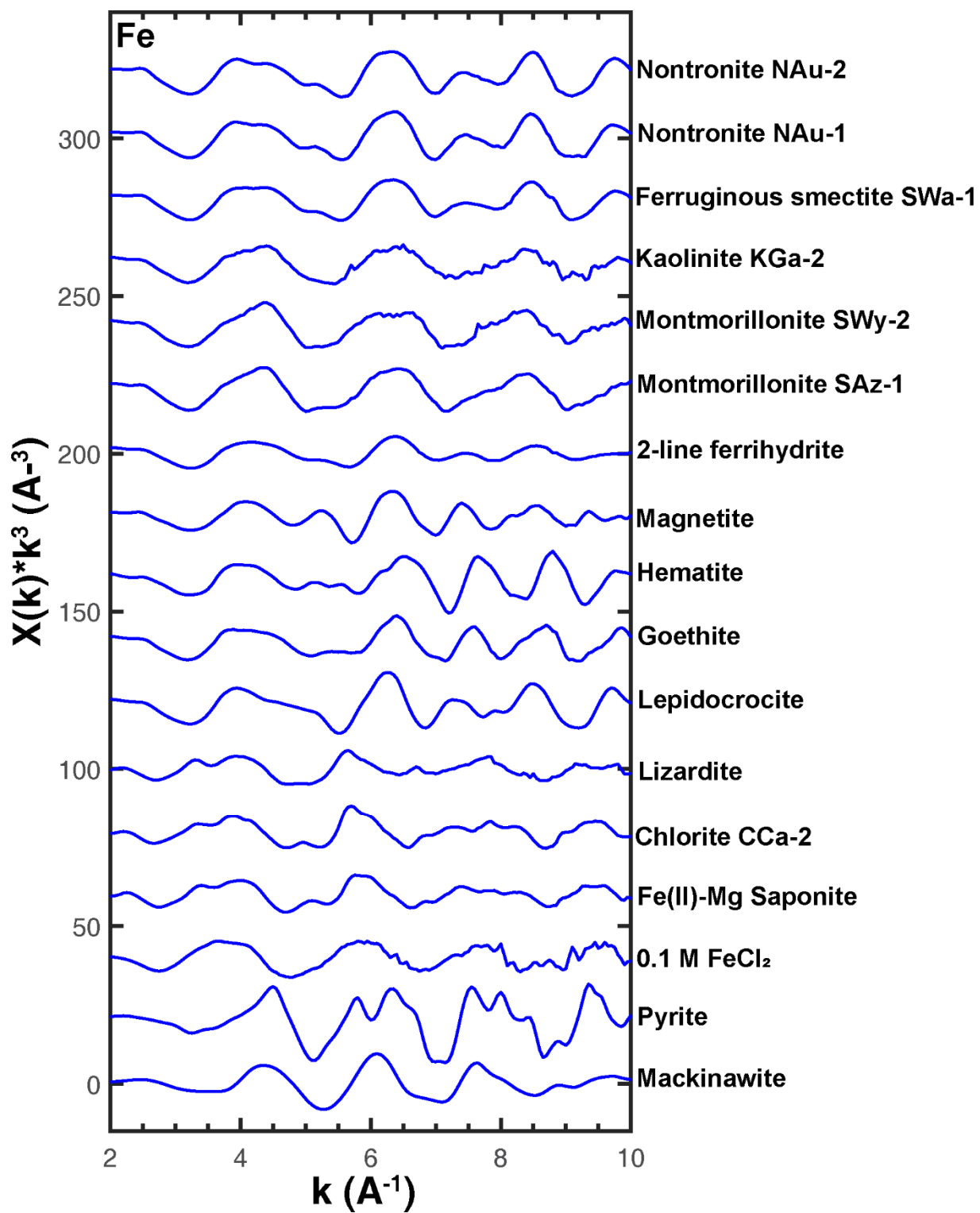


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164

165

**Figure S4.** XANES spectra of iron reference standards. See Table S8 for more information.



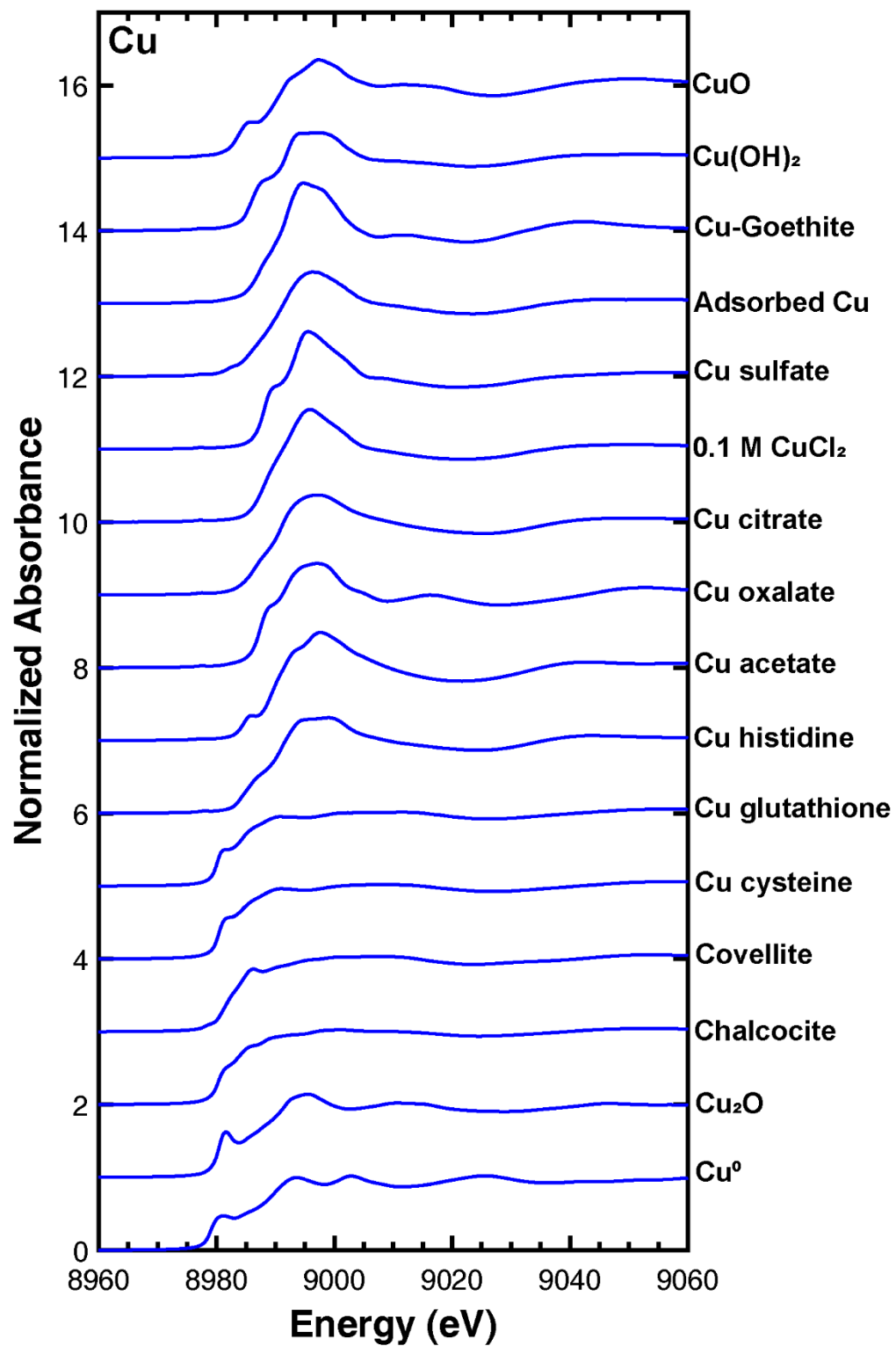
166

167

168

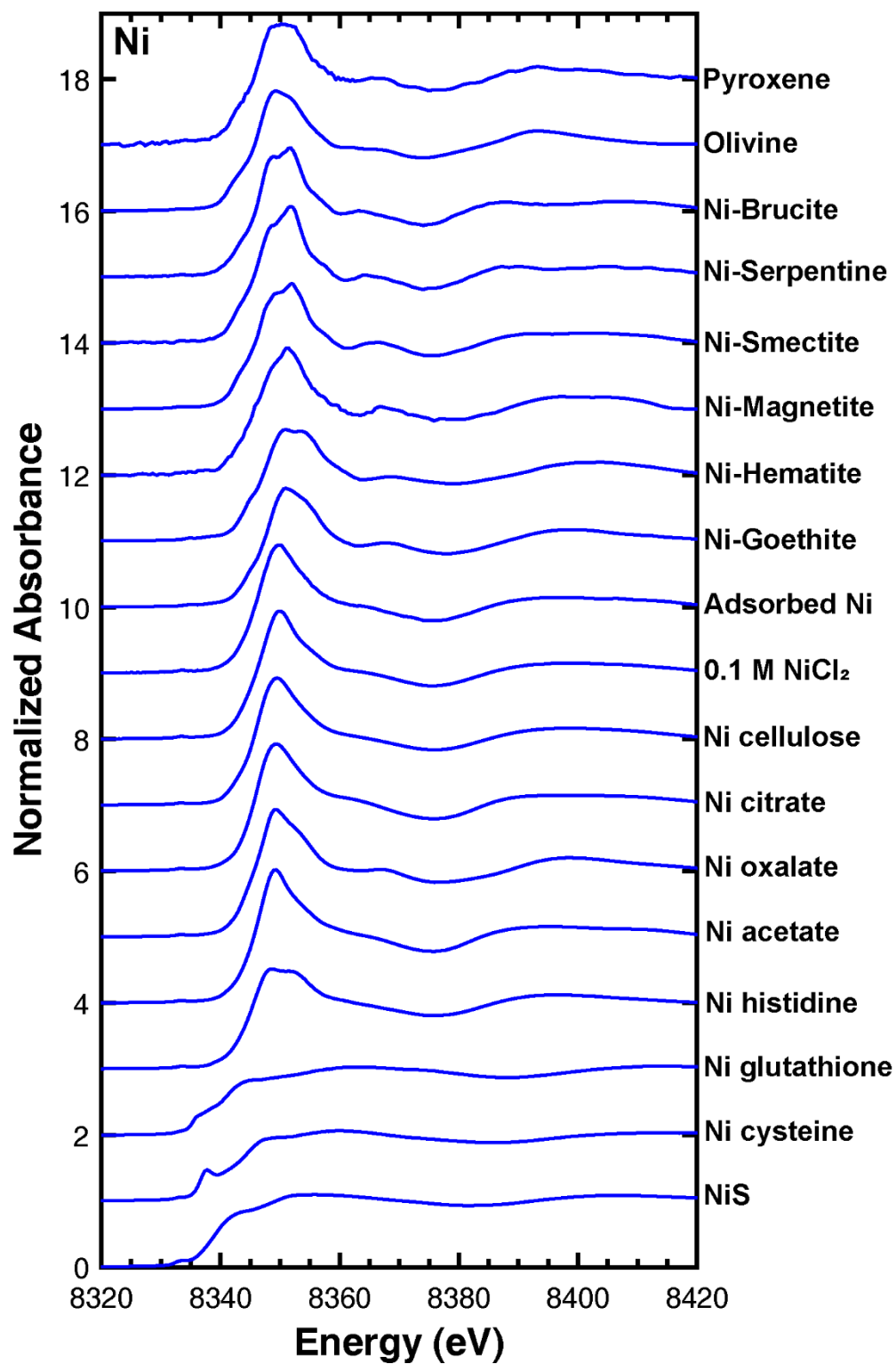
**Figure S5.** EXAFS spectra of iron reference standards. See Table S8 for more information.





169  
 170  
 171  
 172

**Figure S6.** XANES spectra of copper reference standards. See Table S9 for more information.

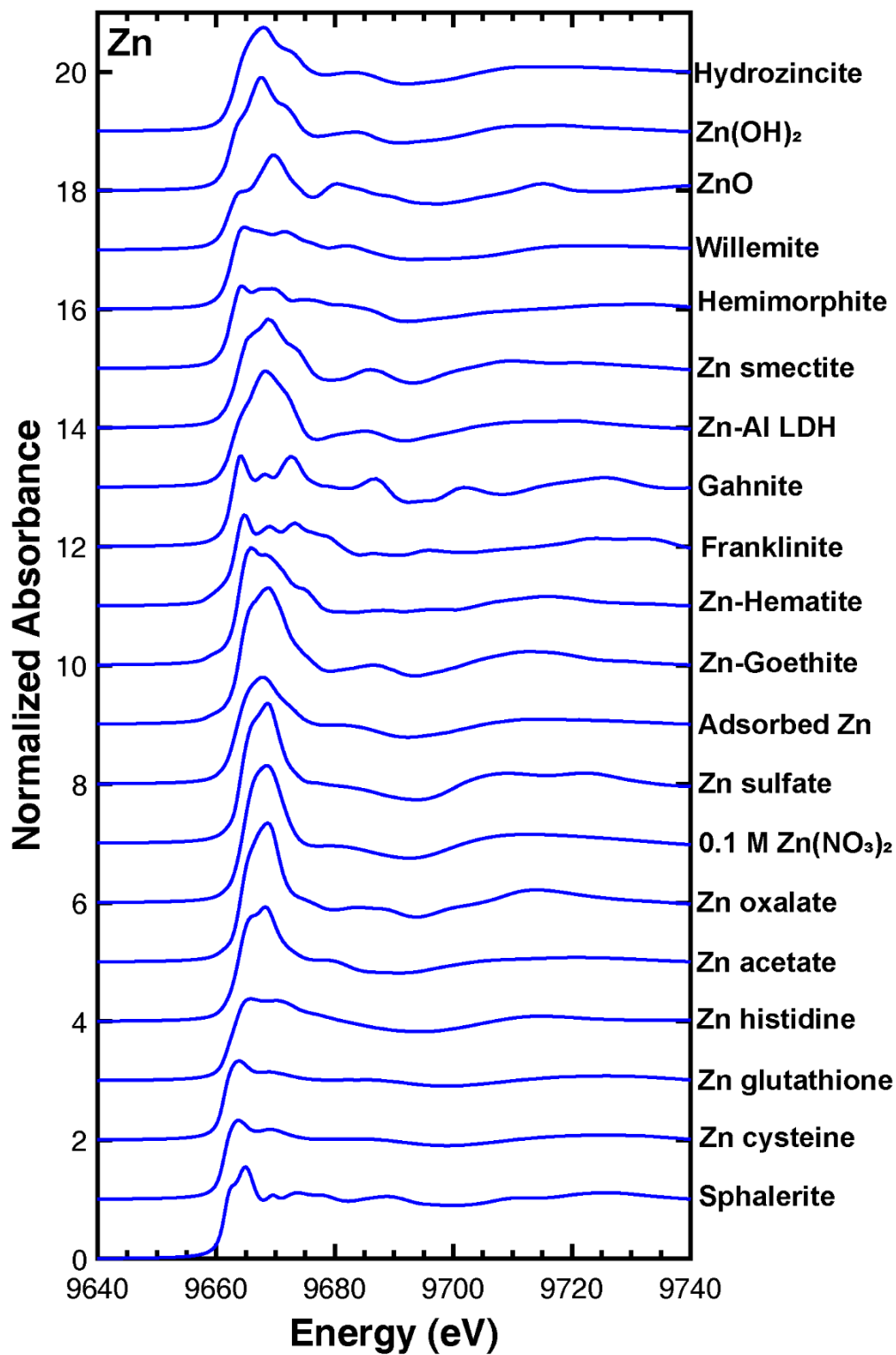


173

174 **Figure S7.** XANES spectra of nickel reference standards. See Table S10 for more information.

175

176



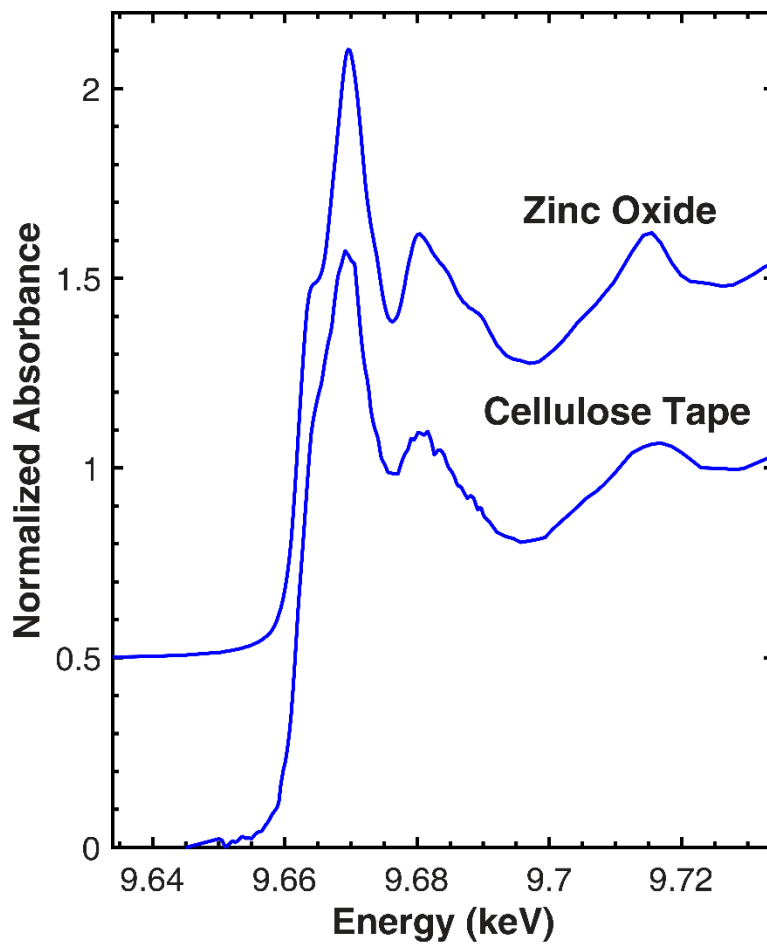
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180

**Figure S8.** XANES spectra of zinc reference standards. See Table S11 for more information.



181

182 **Figure S9.** XANES spectrum of zinc in cellulose acetate tape compared to zinc oxide.

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184

185

186

**Table S1.** Major elements and nutrients concentrations in surface waters.

| Parameter                                       | ANL 401        | ANL 405   | TBL               | TBG       | EFPC 5.4   | EFPC 22-23 |
|---|----------------|-----------|-------------------|-----------|------------|------------|
| pH  | 8.10           | 7.62      | 5.58              | 5.49      | 7.51       | 7.60       |
| T (°C)  | 4.2            | 4.7       | 13.8              | 14.2      | 12.3       | 14.2       |
| Conductivity (µS)                               | 220            | 371       | n.a. <sup>a</sup> | n.a.      | 293        | 348        |
| Ca (µM)   | 430±10         | 590±10    | 18±1              | 47±1      | 1280±30    | 1360±30    |
| Mg (µM)   | 369±5          | 557±4     | 6.3±0.3           | 9.1±0.3   | 456±5      | 633±5      |
| Na (µM)   | 70±10          | 220±10    | 296±2             | 258±3     | 670±20     | 800±10     |
| K (µM)  | 13±2           | 22±2      | 0.8±0.1           | 0.8±0.2   | 12±1       | 9±3        |
| Si (µM)   | 206±4          | 187±3     | 146.5±0.5         | 140.2±0.6 | 225±8      | 250±10     |
| Al (µM)   | - <sup>b</sup> | -         | 6.5±0.9           | 5.6±0.5   | -          | -          |
| Fe (µM)   | 2.6±0.7        | 5±1       | 2.3±0.6           | 70±6      | 0.6±0.2    | 1.0±0.3    |
| Mn (µM)   | 0.07±0.01      | 0.8±0.3   | 0.17±0.03         | 0.43±0.09 | 1.9±0.6    | 0.01±0.06  |
| PO <sub>4</sub> <sup>3-</sup> (µM)              | 7.18±0.02      | 6.44±0.02 | 5.7±0.2           | 5.5±0.2   | 22.17±0.07 | 8.65±0.06  |
| NO <sub>3</sub> <sup>-</sup> (µM)               | 5.2±0.6        | 14.4±0.4  | 3.8±0.1           | -         | 160±30     | 210±30     |
| NH <sub>4</sub> <sup>+</sup> (µM)               | 3±1            | 7±5       | -                 | -         | 11±7       | 8±1        |
| SO <sub>4</sub> <sup>2-</sup> (µM)              | 410±30         | 332.5±0.8 | 10±1              | 13.6±0.04 | 217±3      | 396±4      |
| F <sup>-</sup> (µM)                             | 2.26±0.06      | 3.7±0.4   | 1.9±0.1           | 1.28±0.07 | 14.6±0.3   | 22.4±0.6   |
| Cl <sup>-</sup> (µM)                            | 68±4           | 221.8±0.4 | 61±2              | 77±1      | 465±5      | 578±7      |
| Br <sup>-</sup> (µM)                            | 0.66±0.06      | 0.6±0.2   | 5±2               | 5±1       | 0.9±0.1    | 6.2±0.5    |
| HCO <sub>3</sub> <sup>-</sup> (µM) <sup>c</sup> | 776            | 1620      | 256               | 265       | 3100       | 3190       |
| DIC <sub>calc</sub> (µM) <sup>c</sup>           | 801            | 1740      | 2050              | 2530      | 3350       | 3390       |
| log pCO <sub>2</sub> <sup>c</sup>               | -3.56          | -2.76     | -1.44             | -1.34     | -2.33      | -2.40      |
| I (mM) <sup>c</sup>                             | 2.8            | 3.9       | 0.38              | 0.44      | 6.0        | 6.9        |
| DOC (mg C L <sup>-1</sup> ) <sup>d</sup>        | 39.4           | 47.1      | 8.5               | 12.5      | 27.9       | 27.6       |
| DIC (µM) <sup>d</sup>                           | 790            | 2270      | 300               | 330       | 3920       | 4090       |
| HCO <sub>3</sub> <sup>-</sup> (µM) <sup>e</sup> | 764            | 2206      | 295               | 326       | 3877       | 4057       |
| DIC <sub>calc</sub> (µM) <sup>e</sup>           | 783            | 2369      | 2354              | 3110      | 4180       | 4304       |
| log pCO <sub>2</sub> <sup>e</sup>               | -3.56          | -2.62     | -1.38             | -1.25     | -2.23      | -2.29      |
| I (mM) <sup>e</sup>                             | 2.8            | 3.7       | 0.39              | 0.45      | 6.8        | 7.4        |
| SI <sub>Quartz</sub>                            | 0.81           | 0.75      | 0.43              | 0.40      | 0.65       | 0.65       |
| SI <sub>Am. Silica</sub>                        | -0.66          | -0.71     | -0.95             | -0.98     | -0.75      | -0.73      |

188 <sup>a</sup> Not measured. <sup>b</sup> Value below detection limit, 1.1 µM for Al, 2.1 µM for NO<sub>3</sub><sup>-</sup>, 1.1 µM for  
189 NH<sub>4</sub><sup>+</sup>. <sup>c</sup> Estimated values from charge balance; HCO<sub>3</sub><sup>-</sup> concentration is sum of free and  
190 complexed species, with 0.1% to 1.6% occurring in complexes at all sites. <sup>d</sup> Measured on  
191 archived samples in May 2021. <sup>e</sup> Estimated values from DIC measurement after pCO<sub>2</sub>  
192 adjustment to match field pH; HCO<sub>3</sub><sup>-</sup> concentration is sum of free and complexed species, with  
193 0.1% to 1.6% occurring in complexes at all sites.

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**Table S2.** Field blank major element and trace metal concentrations.

| <b>Conc.</b>         | <b>ANL 401</b> | <b>ANL 405</b> | <b>EFPC 5.4</b>   | <b>EFPC 22-23</b> |
|----------------------|----------------|----------------|-------------------|-------------------|
| Na ( $\mu\text{M}$ ) | 11.5 $\pm$ 0.8 | 13 $\pm$ 0.7   | 39 $\pm$ 1        | 45 $\pm$ 1        |
| Al ( $\mu\text{M}$ ) | - <sup>a</sup> | -              | -                 | -                 |
| Si ( $\mu\text{M}$ ) | -              | -              | -                 | -                 |
| Mg ( $\mu\text{M}$ ) | -              | -              | -                 | -                 |
| K ( $\mu\text{M}$ )  | -              | -              | -                 | -                 |
| Ca ( $\mu\text{M}$ ) | -              | -              | -                 | -                 |
| Fe ( $\mu\text{M}$ ) | 0.3 $\pm$ 0.1  | 0.3 $\pm$ 0.1  | n.a. <sup>b</sup> | n.a.              |
| Mn (nM)              | 8 $\pm$ 1      | 41 $\pm$ 7     | 1.5 $\pm$ 0.2     | 1.9 $\pm$ 0.3     |
| Co (nM)              | 1.7 $\pm$ 0.3  | 3.7 $\pm$ 0.7  | 1.5 $\pm$ 0.3     | 1.6 $\pm$ 0.3     |
| Ni (nM)              | 7 $\pm$ 1      | 134 $\pm$ 24   | 2.4 $\pm$ 0.5     | 2.7 $\pm$ 0.5     |
| Cu (nM)              | 15 $\pm$ 5     | 22 $\pm$ 7     | 7 $\pm$ 2         | 8 $\pm$ 3         |
| Zn (nM)              | 19 $\pm$ 3     | 16 $\pm$ 2     | 7 $\pm$ 1         | 11 $\pm$ 1        |

198 <sup>a</sup> Below the limit of detection: Al =1.6  $\mu\text{M}$ , Si=1.0  $\mu\text{M}$ , Mg=0.51  $\mu\text{M}$ , K=0.83  $\mu\text{M}$ , Ca=4.7  $\mu\text{M}$ .

199 <sup>b</sup> Not measured.

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**Table S3.** Trace metal concentrations in surface water samples.

| <b>Sample</b> | <b>Cu (nM)</b> | <b>Ni (nM)</b> | <b>Co (nM)</b> | <b>Zn (nM)</b> |
|---------------|----------------|----------------|----------------|----------------|
| ANL 401       | 70±20          | 60±10          | 26±5           | 230±30         |
| ANL 405       | 20±7           | 34±6           | 5±1            | 50±10          |
| TBL           | 80±30          | 12±2           | 2.3±0.4        | 100±10         |
| TBG           | 22±7           | 34±6           | 18±3           | 170±20         |
| EFPC 5.4      | 50±20          | 44±8           | 4.4±0.8        | 140±20         |
| EFPC 22-23    | 60±20          | 40±7           | 4.0±0.7        | 50±20          |

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**Table S4.** Trace metal concentrations in soils and sediments.

| <b>Sample</b>        | <b>Cu (nmol/g)</b> | <b>Ni (nmol/g)</b> | <b>Co (nmol/g)</b> | <b>Zn (nmol/g)</b> |
|----------------------|--------------------|--------------------|--------------------|--------------------|
| <i>Surface layer</i> |                    |                    |                    |                    |
| ANL 401              | 730±70             | 420±90             | 70±20              | 2100±400           |
| ANL 405              | 400±100            | 280±70             | 70±20              | 1700±500           |
| TBL                  | 30±30              | 40±20              | 5±1                | 100±80             |
| TBG                  | 200±100            | 110±50             | 20±6               | 300±200            |
| EFPC 5.4             | 150±80             | 300±200            | 300±200            | 600±200            |
| EFPC 22-23           | 80±30              | 160±60             | 80±30              | 300±100            |
| <i>Second layer</i>  |                    |                    |                    |                    |
| ANL 401              | 764±200            | 462±70             | 78±10              | 1833±300           |
| ANL 405              | 366±100            | 377±100            | 102±40             | 1547±400           |
| TBL                  | 10±10              | 33±10              | 6±3                | 84±60              |
| TBG                  | 96±80              | 73±40              | 13±5               | 180±100            |
| EFPC 5.4             | 84±60              | 184±90             | 95±40              | 483±300            |
| EFPC 22-23           | 41±20              | 180±90             | 115±60             | 324±200            |

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**Table S5.** KCl-extractable nutrients concentrations in the soils and sediments.

| Sample               | NO <sub>3</sub> <sup>-</sup> (μmol/g soil) | NH <sub>4</sub> <sup>+</sup> (μmol/g soil) | PO <sub>4</sub> <sup>3-</sup> (μmol/g soil) |
|----------------------|--|--|---|
| <i>Surface layer</i> |  |  |   |
| ANL 401              | 0.24±0.03                                  | 13.0±0.2                                   | 0.64±0.04                                   |
| ANL 405              | 0.6±0.3                                    | 26±8                                       | 1.0±0.5                                     |
| TBL                  | 0.15±0.02                                  | 1.03±0.02                                  | 0.057±0.001                                 |
| TBG                  | 0.01±0.01                                  | 2.5±0.2                                    | 0.06±0.04                                   |
| EFPC 5.4             | 0.14±0.07                                  | 1.6±0.2                                    | 0.9±0.2                                     |
| EFPC 22-23           | 0.5±0.2                                    | 2±2  | 0.036±0.006                                 |
| <i>Second layer</i>  |  |  |   |
| ANL 401              | 0.245±0.009                                | 4.9±0.3                                    | 0.35±0.02                                   |
| ANL 405              | 0.3±0.1                                    | 5±2  | 0.45±0.05                                   |
| TBL                  | 0.10±0.03                                  | 0.21±0.02                                  | 0.01±0.03                                   |
| TBG                  | 0.02±0.03                                  | 1±1  | 0.022±0.008                                 |
| EFPC 5.4             | 0.17±0.09                                  | 0.9±0.2                                    | 0.16±0.08                                   |
| EFPC 22-23           | 0.6±0.5                                    | 2±1  | 0.030±0.007                                 |

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**Table S6.** Major element content of the soils and sediment samples.

| Sample               | C (wt.%)    | N (wt.%)    | S (wt.%)    | Fe (wt.%) | Mn (wt.%)      |
|----------------------|-------------|-------------|-------------|-----------|----------------|
| <i>Surface layer</i> |             |             |             |           |                |
| ANL 401              | 8.7±0.4     | 0.79±0.02   | 0.18±0.01   | 2.3±0.3   | 0.011±0.003    |
| ANL 405              | 11±2        | 1.0±0.2     | 0.28±0.04   | 2.2±0.3   | 0.014±0.005    |
| TBL                  | 1.1±0.3     | 0.3±0.1     | 0.018±0.009 | 0.20±0.08 | 0.0008±0.0005  |
| TBG                  | 4±2         | 0.05±0.02   | 0.07±0.03   | 1.8±0.8   | 0.003±0.001    |
| EFPC 5.4             | 6±5         | 0.2±0.4     | 1.0±0.9     | 3±1       | 0.026±0.009    |
| EFPC 22-23           | 0.42±0.09   | 0.05±0.07   | 0.02±0.01   | 2.4±0.2   | 0.04±0.01      |
| <i>Second layer</i>  |             |             |             |           |                |
| ANL 401              | 6.45±0.02   | 0.65±0.02   | 0.100±0.002 | 2.5±0.3   | 0.009±0.001    |
| ANL 405              | 3.24±0.05   | 0.337±0.009 | 0.068±0.003 | 3.0±0.1   | 0.020±0.006    |
| TBL                  | 0.19±0.03   | 0.017±0.002 | 0.014±0.008 | 0.19±0.08 | - <sup>a</sup> |
| TBG                  | 2.8±0.2     | 0.168±0.010 | 0.045±0.006 | 2.0±0.5   | 0.0023±0.0004  |
| EFPC 5.4             | 2.14±0.03   | 0.160±0.004 | 0.065±0.005 | 1.4±0.2   | 0.016±0.006    |
| EFPC 22-23           | 0.482±0.006 | 0.06±0.01   | 0.019±0.003 | 2.60±0.04 | 0.09±0.01      |

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<sup>a</sup> Below detection limit.

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**Table S7.** Sulfur K-edge XANES standards and sources.

| <b>Standard</b>             | <b>Source/Reference</b> |
|-----------------------------|-------------------------|
| Mackinawite                 | Swanner et al. (2019)   |
| Pyrite                      | Manceau and Nagy (2012) |
| Elemental S                 | Manceau and Nagy (2012) |
| L-cysteine                  | Manceau and Nagy (2012) |
| 4-Nitrobenzenethiol         | Manceau and Nagy (2012) |
| Dibenzyl sulfide            | Manceau and Nagy (2012) |
| Benzyl phenyl sulfide       | Manceau and Nagy (2012) |
| DL-cystine                  | Manceau and Nagy (2012) |
| Dibenzyl disulfide          | Manceau and Nagy (2012) |
| DL-methionine sulfoxide     | Manceau and Nagy (2012) |
| Dibenzyl sulfone            | Manceau and Nagy (2012) |
| Sodium cyclohexanesulfonate | Manceau and Nagy (2012) |
| Anthraquinone sulfonic acid | Bohic et al. (2012)     |
| Chondroitine sulfate        | Cuif et al. (2003)      |
| Gypsum                      | Manceau and Nagy (2012) |

\*Data were taken from the literature when a reference is given.

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**Table S8.** Iron K-edge XANES standards and sources.

| <b>Standard</b>            | <b>Source</b>                       |
|----------------------------|-------------------------------------|
| Mackinawite                | Synthesized                         |
| Pyrite                     | Ward's Science                      |
| 0.1 M FeCl <sub>2</sub>    | Salt from Sigma-Aldrich             |
| Fe(II)-Mg Saponite         | Chemtob et al. (2015)               |
| Chlorite CCa-2             | Source Clay Repository, as received |
| Lizardite                  | Synthesized                         |
| Lepidocrocite              | Synthesized                         |
| Goethite                   | Synthesized                         |
| Hematite                   | Synthesized                         |
| Magnetite                  | Synthesized                         |
| 2-line Ferrihydrite        | Synthesized                         |
| Montmorillonite SAz-1      | Source Clay Repository, <2 μm       |
| Montmorillonite SWy-2      | Source Clay Repository, <2 μm       |
| Kaolinite KGa-2            | Source Clay Repository, as received |
| Ferruginous Smectite SWa-1 | Source Clay Repository, <2 μm       |
| Nontronite NAu-1           | Source Clay Repository, <0.5 μm     |
| Nontronite NAu-2           | Source Clay Repository, <0.5 μm     |

217 \*Synthesis methods of the reference standards are described in the section "Synthesis of  
218 Reference Standards". Data were taken from the literature when a reference is given.  
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**Table S9** Cu reference standards for EXAFS measurement and their sources.

| <b>Standard</b>                | <b>Source/Reference</b>  |
|--------------------------------|--|
| Copper(II) Acetate             | Acros Organics   |
| Copper(II) Oxalate             | Alfa Aesar   |
| Copper(II) Citrate             | Synthesized  |
| Copper(II) Sulfate             | Fisher Scientific  |
| Chalcocite (Cu <sub>2</sub> S) | Alfa Aesar   |
| Cu(II)-Cysteine Complex        | Synthesized  |
| Cu(II)-Histidine Complex       | Synthesized  |
| Cu(II)-Glutathione Complex     | Synthesized  |
| Covellite (CuS)                | Butte, Montana, USA  |
| Cu(II) adsorbed to goethite    | Synthesized  |
| Cu metal                       | EXAFS Materials  |
| 0.1 M CuCl <sub>2</sub>        | Salt from Sigma-Aldrich  |
| Cu-substituted Goethite        | Friedrich and Catalano (2012)  |
| Copper(II) oxide               | Friedrich and Catalano (2012)  |
| Copper(I) oxide                | Friedrich and Catalano (2012)  |
| Copper(II) hydroxide           | Alfa Aesar; See Friedrich and Catalano (2012) for data collection information. |

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\*Synthesis methods of the reference standards are described in the section "Synthesis of Reference Standards". Data were taken from the literature when a reference is given.

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**Table S10.** Ni reference standards for EXAFS measurement and their sources.

| <b>Standard</b>                                 | <b>Source/Reference</b>  |
|---|--------------------------|
| Nickel(II) Acetate                              | Acros Organics           |
| Nickel(II) Oxalate                              | Alfa Aesar               |
| Nickel(II) Citrate                              | Alfa Aesar               |
| Nickel(II) Sulfide                              | Alfa Aesar               |
| Ni(II)-Cysteine Complex                         | Synthesized              |
| Ni(II)-Histidine Complex                        | Synthesized              |
| Ni(II)-Glutathione Complex                      | Synthesized              |
| Ni(II)-Cellulose Complex                        | Synthesized              |
| Ni-substituted Hematite                         | Friedrich et al. (2011)  |
| Ni-substituted Goethite                         | Friedrich et al. (2011)  |
| Ni-substituted Magnetite                        | Synthesized              |
| Pyroxene (Augite)                               | Ontario, Canada          |
| Olivine   | San Carlos, Arizona, USA |
| Ni-substituted Brucite                          | Synthesized              |
| Ni-substituted Serpentine                       | Synthesized              |
| Ni-substituted Fe(II)-Mg Trioctahedral Smectite | Synthesized              |
| Ni(II) adsorbed to goethite                     | Synthesized              |
| 0.1 M NiCl <sub>2</sub>                         | Salt from Sigma-Aldrich  |

225 \*Synthesis methods of the reference standards are described in the section “Synthesis of  
 226 Reference Standards”. Data were taken from the literature when a reference is given.  
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**Table S11.** Zn reference standards for EXAFS measurement and their sources.

| <b>Standard</b>                         | <b>Source/Reference</b>        |
|---|--------------------------------|
| Zinc(II) Acetate                        | Fisher Scientific              |
| Zinc(II) Oxalate                        | Synthesized                    |
| 0.1 M Zn(NO <sub>3</sub> ) <sub>2</sub> | Salt from J.T. Baker           |
| Zinc(II) Sulfate                        | Fisher Scientific              |
| Zn(II)-Cysteine Complex                 | Synthesized                    |
| Zn(II)-Histidine Complex                | Synthesized                    |
| Zn(II)-Glutathione Complex              | Synthesized                    |
| Franklinite                             | Luo et al. (2011)              |
| Gahnite                                 | Luo et al. (2011)              |
| Willemite                               | Luo et al. (2011)              |
| Hemimorphite                            | Luo et al. (2011)              |
| Zinc(II) Hydroxide                      | Luo et al. (2011)              |
| Zn-Al layered double hydroxide          | Luo et al. (2011)              |
| Zn smectite                             | Synthesized                    |
| Zinc(II) Oxide                          | Luo et al. (2011)              |
| Hydrozincite                            | Luo et al. (2011)              |
| Sphalerite                              | Pettus County, Missouri, USA   |
| Zn-substituted Hematite                 | Friedrich and Catalano (2012a) |
| Zn-substituted Goethite                 | Friedrich and Catalano (2012a) |
| Zn adsorbed to goethite                 | Synthesized                    |

229 \*Synthesis methods of the reference standards are described in the section “Synthesis of  
 230 Reference Standards”. Data were taken from the literature when a reference is given.

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**Table S12.** Sulfur XANES linear combination fitting results.

| <b>Standard</b>             | <b>Marsh 1</b> | <b>Marsh 2</b> | <b>Riparian 1</b> | <b>Riparian 2</b> | <b>Stream 1 C1</b> | <b>Stream 1 C2</b> | <b>Stream 2</b> |
|-----------------------------|----------------|----------------|-------------------|-------------------|--------------------|--------------------|-----------------|
| <i>Fitting Components</i>   |                |                |                   |                   |                    |                    |                 |
| Gypsum                      | 10.1±0.6%      | 8.6±0.2        | 5.5±0.6%          | 10.5±0.2%         | 5.8±0.4%           | 1.1±0.7%           | 16±1%           |
| Anthraquinone sulfonic acid | 6±2%           | - <sup>a</sup> | 9±2%              | -                 | 3.0±0.4%           | 1.7±0.6%           | 7±3%            |
| Sodium cyclohexanesulfonate | 23±1%          | 17.1±0.2       | 9±1%              | 17.0±0.2          | -                  | -                  | 5±2%            |
| Dibenzyl sulfone            | 1.6±0.4%       | 1.6±0.2%       | -                 | 1.5±0.2           | -                  | -                  | -               |
| DL-methionine sulfoxide     | 2.2±0.4%       | 3.0±0.2%       | -                 | 4.8±0.4%          | -                  | -                  | -               |
| Dibenzyl disulfide          | -              | -              | 35±4%             | 5±4%              | 14±3%              | 14±5%              | 21±4%           |
| Cystine                     | 17±2%          | 16±1%          | 6±2%              | 18±1%             | -                  | -                  | -               |
| Benzyl phenyl sulfide       | 10±3%          | 13±2%          | -                 | -                 | -                  | -                  | -               |
| 4-Nitrobenzenethiol         | 10±2%          | 7±1%           | -                 | -                 | -                  | -                  | -               |
| Cysteine                    | 8±3%           | 13±2%          | 8±1%              | 18±1%             | -                  | -                  | -               |
| Elemental S                 | 9±2%           | 17±1%          | 24±2%             | 19±2%             | 42±4%              | 73±5%              | 50±3%           |
| Pyrite                      | -              | -              | -                 | -                 | 30±3%              | 10±3%              | -               |
| Mackinawite                 | 3.7±0.8%       | 4.6±0.5%       | 3.3±0.7%          | 6.4±0.6%          | 5±1%               | -                  | 2±1%            |
| Component sum               | 1.075          | 0.977          | 1.063             | 1.006             | 1.009              | 1.093              | 1.006           |
| R factor                    | 0.003          | 0.002          | 0.005             | 0.003             | 0.01               | 0.02               | 0.02            |
| $\chi^2$                    | 0.002          | 0.0009         | 0.003             | 0.001             | 0.004              | 0.01               | 0.008           |
| <i>Groupings</i>            |                |                |                   |                   |                    |                    |                 |
| FeS                         | 3.7±0.8%       | 4.6±0.5%       | 3.3±0.7%          | 6.4±0.6%          | 5±1%               | -                  | 2±1%            |
| Pyrite                      | -              | -              | -                 | -                 | 30±3%              | 10±3%              | -               |
| S <sup>0</sup>              | 9±2%           | 17±1%          | 24±2%             | 19±2%             | 42±4%              | 73±5%              | 50±3%           |
| Exocyclic                   | 45±5%          | 48±3%          | 50±4%             | 41±4%             | 14±3%              | 14±5%              | 21±4%           |
| Sulfoxide                   | 2.2±0.4%       | 3.0±0.2%       | -                 | 4.8±0.4%          | -                  | -                  | -               |
| Sulfone                     | 1.6±0.4%       | 1.6±0.2%       | -                 | 1.5±0.2%          | -                  | -                  | -               |
| Sulfonate                   | 29±2%          | 17.1±0.2%      | 17±2%             | 17.0±0.2%         | 3.0±0.4%           | 2±1%               | 12±4%           |
| Sulfate (inorganic)         | 10.1±0.6       | 8.6±0.2%       | 5.5±0.6%          | 10.5±0.2%         | 5.8±0.4%           | 1±1%               | 16±1%           |

234 <sup>a</sup> Not present in final fit.

**Table S13.** Iron XANES linear combination fitting results.

| <b>Standard</b>           | <b>Marsh 1</b> | <b>Marsh 2</b> | <b>Riparian 1</b> | <b>Riparian 2</b> | <b>Stream 1 C1</b> | <b>Stream 1 C2</b> | <b>Stream 2</b> |
|---------------------------|----------------|----------------|-------------------|-------------------|--------------------|--------------------|-----------------|
| <i>Fitting Components</i> |                |                |                   |                   |                    |                    |                 |
| Pyrite                    | -              | -              | -                 | -                 | -                  | -                  | 4.8±0.3%        |
| FeS                       | 3.5±0.7%       | 6.5±0.5%       | -                 | 5.6±0.3%          | 48.0±0.2%          | 17.5±0.2%          | -               |
| Fe(II) (aq)               | 18.2±0.7%      | 19±1%          | 17.5±0.4%         | 32.1±0.4%         | 5.3±0.3%           | 15.2±0.3%          | 7.2±0.3%        |
| Fe(II)-Mg Smectite        | 23±1%          | 29±1%          | -                 | -                 | 10.3±0.5%          | -                  | -               |
| Chlorite                  | -              | -              | 6.4±0.7%          | 13.1±0.8%         | -                  | 17.7±0.5%          | 18.0±0.4%       |
| Magnetite                 | 7%±3%          | -              | -                 | -                 | -                  | -                  | -               |
| Montmorillonite           | 48±2%          | 39±3%          | -                 | -                 | 8±1%               | -                  | -               |
| Ferruginous smectite      | -              | -              | -                 | -                 | -                  | 30±2%              | 38±2%           |
| Kaolinite                 | -              | -              | 18±3%             | 24±3%             | -                  | -                  | -               |
| Lepidocrocite             | -              | 7±4%           | -                 | -                 | -                  | -                  | -               |
| Goethite                  | -              | -              | 11±3%             | 25±3%             | 28±1%              | 19±2%              | -               |
| 2-line ferrihydrite       | -              | -              | 48±2%             | -                 | -                  | -                  | 32±3%           |
| Component sum             | 1.01           | 1.015          | 0.996             | 1.006             | 0.999              | 1.004              | 0.996           |
| R factor                  | 0.0004         | 0.0006         | 0.0001            | 0.0002            | 0.0001             | 0.00008            | 0.0001          |
| $\chi^2$                  | 0.0001         | 0.0002         | 0.00004           | 0.00005           | 0.00002            | 0.00002            | 0.00004         |
| <i>Groupings</i>          |                |                |                   |                   |                    |                    |                 |
| Pyrite                    | -              | -              | -                 | -                 | -                  | -                  | 4.8±0.3%        |
| FeS                       | 3.5±0.7%       | 6.5±0.5%       | -                 | 5.6±0.3%          | 48.0±0.2%          | 17.5±0.2%          | -               |
| Fe(II) (aq)               | 18.2±0.7%      | 19±1%          | 17.5±0.4%         | 32.1±0.4%         | 5.3±0.3%           | 15.2±0.3%          | 7.2±0.3%        |
| Fe(II) in clays           | 23±1%          | 29±1%          | 6.4±0.7%          | 13.1±0.8%         | 10.3±0.5%          | 17.7±0.5%          | 18.0±0.4%       |
| Magnetite                 | 7±3%           | -              | -                 | -                 | -                  | -                  | -               |
| Fe(III) in clays          | 48±2%          | 39±3%          | 18±3%             | 24±3%             | 8±1%               | 30±2%              | 38±2%           |
| Fe(III) oxides            | -              | 7±4%           | 58±5%             | 25±3%             | 28±1%              | 19±2%              | 32±3%           |

237 <sup>a</sup> Not present in final fit.

238

239

**Table S14.** Iron EXAFS linear combination fitting results.

| <b>Standard</b>           | <b>Marsh 1</b> | <b>Marsh 2</b> | <b>Riparian 2</b> | <b>Stream 1 C1</b> | <b>Stream 1 C2</b> | <b>Stream 2</b> |
|---------------------------|----------------|----------------|-------------------|--------------------|--------------------|-----------------|
| <i>Fitting Components</i> |                |                |                   |                    |                    |                 |
| Pyrite                    | 1.8±0.9%       | 1.4±0.8%       | -                 | -                  | -                  | -               |
| FeS                       | -              | -              | -                 | 46±1%              | 14±2%              | -               |
| Fe(II) (aq)               | 20±3%          | 13±2%          | 26±3%             | 4±2%               | 8±3%               | -               |
| Fe(II)-Mg Smectite        | 20±3%          | 33±2%          | -                 | -                  | -                  | 6±3%            |
| Chlorite                  | -              | -              | 20±3%             | 14±2%              | 26±3%              | 19±3%           |
| Montmorillonite           | 24±3%          | 43±2%          | 25±2%             | 21±1%              | 29±2%              | 37±2%           |
| Ferruginous smectite      | 34±3%          | -              | -                 | -                  | -                  | -               |
| Lepidocrocite             | -              | 9±1%           | -                 | -                  | -                  | -               |
| Goethite                  | -              | -              | -                 | 14±2%              | 22±3%              | 27±2%           |
| 2-line ferrihydrite       | -              | -              | 13±4%             | -                  | -                  | 10±3%           |
| Hematite                  | -              | -              | 16±1%             | -                  | -                  | -               |
| Component sum             | 1.008          | 0.932          | 0.904             | 0.889              | 0.886              | 0.861           |
| R factor                  | 0.04           | 0.03           | 0.04              | 0.02               | 0.05               | 0.02            |
| $\chi^2$                  | 0.3            | 0.2            | 0.2               | 0.1                | 0.3                | 0.1             |
| <i>Groupings</i>          |                |                |                   |                    |                    |                 |
| Pyrite                    | 1.8±0.9%       | 1.4±0.8%       | -                 | -                  | -                  | -               |
| FeS                       | -              | -              | -                 | 46±1%              | 14±2%              | -               |
| Fe(II) (aq)               | 20±3%          | 13±2%          | 26±3%             | 4±2%               | 8±3%               | -               |
| Fe(II) in clays           | 20±3%          | 33±2%          | 20±3%             | 14±2%              | 26±3%              | 26±7%           |
| Fe(III) in clays          | 58±6%          | 43±2%          | 25±2%             | 21±1%              | 29±2%              | 37±2%           |
| Fe(III) oxides            | -              | 9±1%           | 29±5%             | 14±2%              | 22±3%              | 37±5%           |

241 <sup>a</sup> Not present in final fit.

242



243

**Table S15.** Copper XANES linear combination fitting results.

| <b>Standard</b>           | <b>Marsh 1</b> | <b>Marsh 2</b> | <b>Riparian 1</b> | <b>Riparian 2</b> | <b>Stream 1 C1</b> | <b>Stream 1 C2</b> | <b>Stream 2</b> |
|---------------------------|----------------|----------------|-------------------|-------------------|--------------------|--------------------|-----------------|
| <i>Fitting Components</i> |                |                |                   |                   |                    |                    |                 |
| Covellite                 | 90±1%          | 83±1%          | 40±4%             | 75±1%             | 70±1%              | 72±1%              | 32±2%           |
| Chalcocite                | - <sup>a</sup> | -              | 30±4%             | -                 | -                  | -                  | -               |
| Cu(II)-Histidine          | 10±1%          | 12±5%          | -                 | -                 | 9±7%               | -                  | 13±5%           |
| 0.1 M CuCl <sub>2</sub>   | -              | -              | -                 | -                 | -                  | -                  | 21±8%           |
| Ads. Cu                   | -              | 5±4%           | 40±1%             | 25±1%             | 21±6%              | 28±1%              | 34±8%           |
| Component sum             | 1.028          | 1.04           | 1.081             | 1.028             | 1.041              | 1.012              | 1.004           |
| R factor                  | 0.002          | 0.002          | 0.003             | 0.002             | 0.003              | 0.002              | 0.001           |
| $\chi^2$                  | 0.0003         | 0.0004         | 0.0007            | 0.0004            | 0.0007             | 0.0005             | 0.0003          |
| <i>Groupings</i>          |                |                |                   |                   |                    |                    |                 |
| CuS                       | 90±1%          | 83±1%          | 40±4%             | 75±1%             | 70±1%              | 72±1%              | 32±2%           |
| Cu <sub>2</sub> S         | -              | -              | 30±4%             | -                 | -                  | -                  | -               |
| Amine bound               | 10±1%          | 12±5%          | -                 | -                 | 9±7%               | -                  | 13±5%           |
| Adsorbed                  | -              | 5±4%           | 40±1%             | 25±1%             | 21±6%              | 28±1%              | 55±20%          |

244 <sup>a</sup> Not present in final fit.

245

246

247

**Table S16.** Nickel XANES linear combination fitting results.

| <b>Standard</b>           | <b>Marsh 1</b> | <b>Marsh 2</b> | <b>Riparian 1</b> | <b>Riparian 2</b> | <b>Stream 1 C1</b> | <b>Stream 1 C2</b> | <b>Stream 2</b> |
|---------------------------|----------------|----------------|-------------------|-------------------|--------------------|--------------------|-----------------|
| <i>Fitting Components</i> |                |                |                   |                   |                    |                    |                 |
| NiS                       | 35±8%          | 33±1%          | 13±2%             | 14±1%             | 32±1%              | 34±1%              | 14±1%           |
| Ni-Smectite               | 50±3%          | 45±8%          | 60±6%             | 65±5%             | 59±4%              | 45±9%              | 70±3%           |
| Serpentine                | - <sup>a</sup> | 9±8%           | -                 | -                 | -                  | 13±9%              | -               |
| 0.1 M NiCl <sub>2</sub>   | 15±3%          | 13±3%          | 27±7%             | 22±5%             | -                  | -                  | 16±4%           |
| Ads. Ni                   | -              | -              | -                 | -                 | 9±4%               | 9±3%               | -               |
| Component sum             | 1.014          | 1.03           | 0.974             | 1.028             | 1.074              | 1.024              | 1.011           |
| R factor                  | 0.002          | 0.002          | 0.007             | 0.005             | 0.005              | 0.003              | 0.003           |
| $\chi^2$                  | 0.0005         | 0.0007         | 0.002             | 0.002             | 0.001              | 0.0009             | 0.001           |
| <i>Groupings</i>          |                |                |                   |                   |                    |                    |                 |
| NiS                       | 35±8%          | 33±8%          | 13±2%             | 13±1%             | 32±1%              | 34±1%              | 14±1%           |
| Clay structures           | 50±3%          | 54±3%          | 60±6%             | 65±5%             | 59±4%              | 58±18%             | 70±3%           |
| Adsorbed                  | 15±3%          | 12±3%          | 27±7%             | 22±5%             | 9±4%               | 9±3%               | 16±4%           |

248 <sup>a</sup> Not present in final fit.

249

250

**Table S17.** Zinc XANES linear combination fitting results.

| <b>Standard</b>           | <b>Marsh 1</b> | <b>Marsh 2</b> | <b>Riparian 1</b> | <b>Riparian 2</b> | <b>Stream 1 C1</b> | <b>Stream 1 C2</b> | <b>Stream 2</b> |
|---------------------------|----------------|----------------|-------------------|-------------------|--------------------|--------------------|-----------------|
| <i>Fitting Components</i> |                |                |                   |                   |                    |                    |                 |
| Zn-Smectite               | 36±2%          | 23±2%          | - <sup>a</sup>    | -                 | 12±1%              | 29±2%              | 14±4%           |
| ZnO                       | 10±1%          | 27±1%          | 54±2%             | 23±1%             | 45±1%              | 34±1%              | 50±2%           |
| Zn cysteine               | 30±1%          | -              | -                 | -                 | 27±1%              | 22±1%              | -               |
| Zn glutathione            | -              | 34±1%          | 19±3%             | -                 | -                  | -                  | 21±2%           |
| Sphalerite                | -              | -              | 10±2%             | 7±2%              | -                  | -                  | -               |
| Zn Acetate                | 24±2%          | -              | -                 | -                 | -                  | -                  | -               |
| ZnSO <sub>4</sub>         | -              | 16±1%          | 17±1%             | -                 | 16±1%              | 15±1%              | 15±2%           |
| Zn Nitrate                | -              | -              | -                 | 41±2%             | -                  | -                  | -               |
| Ads. Zn                   | -              | -              | -                 | 29±4%             | -                  | -                  | -               |
| Component sum             | 1.006          | 0.996          | 0.995             | 0.977             | 0.996              | 0.999              | 0.984           |
| R factor                  | 0.0004         | 0.0003         | 0.0004            | 0.0005            | 0.0007             | 0.0004             | 0.001           |
| $\chi^2$                  | 0.02           | 0.0001         | 0.0002            | 0.0002            | 0.01               | 0.0001             | 0.05            |
| <i>Groupings</i>          |                |                |                   |                   |                    |                    |                 |
| ZnS                       | -              | -              | 10±2%             | 7±2%              | -                  | -                  | -               |
| Thiol bound               | 30±1%          | 34±1%          | 19±3%             | -                 | 27±1%              | 22±1%              | 21±2%           |
| Clay structures           | 36±2%          | 23±2%          | - <sup>a</sup>    | -                 | 12±1%              | 29±2%              | 14±4%           |
| ZnO                       | 10±1%          | 27±1%          | 54±2%             | 23±1%             | 45±1%              | 34±1%              | 50±2%           |
| Adsorbed                  | 24±2%          | 16±1%          | 17±1%             | 70±6%             | 16±1%              | 15±1%              | 15±2%           |

252 <sup>a</sup> Not present in final fit.

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