1	Methane Fluxes of Vegetated Areas in Natural Freshwater
2	Ecosystems:
3	Assessments and Global Significance
4	
5	Pascal Bodmer <sup>1,2,3,*</sup> , Renske Vroom <sup>1</sup> , Tatiana Stepina <sup>1,4</sup> , Paul del Giorgio <sup>3</sup> , and Sarian Kosten <sup>1</sup>
6	
7 8	<sup>1</sup> Department of Aquatic Ecology and Environmental Biology, Institute for Water and Wetland Research, Radboud University Nijmegen, Nijmegen, The Netherlands
9	<sup>2</sup> Institute for Environmental Sciences, University of Koblenz-Landau, Landau, Germany
10	<sup>3</sup> Université du Québec à Montréal, Montréal, Québec, Canada
11	<sup>4</sup> Siberian Federal University, 79 Svobodny prospect, Krasnoyarsk, 660041, Russia
12	
13	* Corresponding author: bodmerpascal@gmail.com (Twitter: @BodmerPascal)
14	
15	

## 16 Abstract

- 17
- 18 Freshwater ecosystems, including wetlands, lakes, and running waters, are estimated to contribute
- 19 roughly 40% to global emissions of methane (CH<sub>4</sub>), a highly potent greenhouse gas. The
- 20 emission of CH<sub>4</sub> to the atmosphere entails the diffusive, ebullitive, and plant-mediated pathway.
- 21 The latter, in particular, has been largely understudied and is neither well understood nor
- 22 quantified. We have conducted a semi-quantitative literature review to (i) provide a synthesis of
- the different ways vegetated habitats can influence CH<sub>4</sub> dynamics (i.e., production, consumption,
- and transport) in freshwater ecosystems, (ii) provide an overview of methods applied to study the
- 25 fluxes from vegetated habitats, and (iii) summarize the existing data on CH<sub>4</sub> fluxes associated to
- 26 different types of vegetated habitats and their range of variation. Finally, we discuss the
- 27 implications of CH<sub>4</sub> fluxes associated with aquatic vegetated habitats for current estimates of
- aquatic CH<sub>4</sub> emissions at the global scale. We identified 13 different aspects in which plants
- 29 impact CH<sub>4</sub> dynamics (three related to gaseous CH<sub>4</sub> flux pathways) and ten approaches used to
- 30 study and quantify fluxes from vegetated habitats. The variability of the fluxes from vegetated
- areas was very high, varying from -454.4 mg CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup> (uptake) to 2882.4 mg CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup>
- 32 (emission). This synthesis highlights the need to incorporate vegetated habitats into CH<sub>4</sub> emission
- budgets from natural freshwater ecosystems and further identifies understudied research aspects
- 34 and relevant future research directions.

## 36 1 Introduction

37

38 Freshwater ecosystems, including wetlands, lakes, reservoirs, rivers, and streams, play an important role in global carbon cycling and are estimated to contribute roughly 40% to global 39 methane (CH<sub>4</sub>) emissions: 149 TgCH<sub>4</sub> yr<sup>-1</sup> for wetlands and 159 TgCH<sub>4</sub> y<sup>-1</sup> for lakes (1, 2). 40 Estimates of CH<sub>4</sub> emissions are, however, highly variable, and freshwater systems present an 41 important source of uncertainty in the global CH<sub>4</sub> budget (1). Of the identified natural sources, 42 the contribution of vegetation to the global CH<sub>4</sub> budget is least well understood (3). This is an 43 important caveat as studies focusing on single systems indicate that emissions from vegetated 44 areas can contribute up to 46% of the CH<sub>4</sub> efflux of the entire system (4) and sometimes even 45 surpasses 90% (5). 46

Clearly, the contribution of vegetated areas to the total system emission depends on the 47 vegetation coverage on the one hand and the impact of plants on CH<sub>4</sub> emissions on the other. CH<sub>4</sub> 48 emissions from vegetated sites can differ considerably – being higher or lower - from those from 49 unvegetated sites (see, e.g., an overview in 6). These contrasting findings have been attributed to 50 differences in the impact on CH<sub>4</sub> production, CH<sub>4</sub> oxidation, and CH<sub>4</sub> transport (7–10). The 51 impact of vegetation on these processes varies among species (5, 11, 12) and depends on a 52 variety of variables, including plant biomass (particularly below-ground biomass; e.g., 13), plant 53 54 growth dynamics (14), plant tissue composition (8), sediment temperature (15), type of sediment (16), water depth (17) and herbivory (18) which all vary seasonally leading to seasonal 55 differences in CH<sub>4</sub> emissions. This long list highlights that mechanistic understanding of the 56 impact of plants on CH<sub>4</sub> fluxes is needed to accurately extrapolate single CH<sub>4</sub> emissions to annual 57 emissions and upscale to system-wide or even global CH<sub>4</sub> emissions from vegetated areas. 58

Insight into the quantitative effect of vegetation on CH<sub>4</sub> fluxes is in part hampered by the 59 fact that, at least up to recently, most studies have been conducted in high-latitude areas. 60 Moreover, a wide range of different methods has been used to assess plant effects on CH<sub>4</sub> 61 emissions. These methods vary in the type of fluxes they include. Some studies, for instance, 62 63 explicitly exclude ebullition, while others have found that the effect of plants on ebullition can be substantial (19). These discrepancies complicate comparison among studies. The inconsistent use 64 of the term "plant-mediated" flux further hinders comparisons. While some authors define it as 65 the flow of CH<sub>4</sub> through the plant tissue (20), others use the term to indicate the overall effect of 66 67 plants on CH<sub>4</sub> emission, i.e., a combination of different flux pathways (21, 22). While both definitions are linguistically correct, the corresponding flux intensities can vary substantially. 68 Therefore, in this review, we refrain from using the term "plant-mediated transport" and instead 69 refer to CH<sub>4</sub> fluxes of vegetated areas, acknowledging the complexity of affected aspects of 70 vegetation in relation to CH<sub>4</sub> dynamics. 71

In this review, we aim to advance the understanding of the effect of vegetation on aquatic
 CH<sub>4</sub> fluxes, to advance the quantification of CH<sub>4</sub> emissions from vegetated areas, and to identify
 knowledge gaps. We conducted a semi-quantitative literature review to (i) provide an overview
 of the different ways aquatic vegetation can influence CH<sub>4</sub> production, consumption, and

- transport, (ii) provide an overview of methods applied to study the effect of aquatic vegetation on
- 77 CH<sub>4</sub> processes, and (iii) obtain insight in CH<sub>4</sub> fluxes of vegetated areas and its variation. Finally,
- 78 we discuss the implications of plant-habitat-mediated  $CH_4$  fluxes for current global  $CH_4$
- 79 estimates.

# 81 2 Material and Methods

82

# 83 2.1 Literature search

To obtain an overview of studies on CH<sub>4</sub> fluxes of vegetated areas, we used two approaches. 84 Firstly, we selected four key papers (cited at least 200 times in GoogleScholar; 26<sup>th</sup> of November 85 2020) dealing with the topic: Laanbroek (23), Brix et al. (24), Sebacher et al. (25), and Ström et al. 86 (26). We then exported the references citing these key papers. Secondly, a literature search was 87 done using Web of Science on the 26<sup>th</sup> of November 2020. Search inquiry for Web of Science 88 consisted of the following keywords: *plant*\* (asterisk was used to consider singular and plural), 89 mediated, and methane. The combination of keywords resulted in the following formula: TS= 90 (plant\* AND mediated AND methane), TS meant that the search was done in the title, keywords, 91 92 and the abstract of papers. The search terms were used with no restriction on publication year.

The data from the search inquiry, together with studies that had cited above mentioned key 93 papers, were integrated into a database, focusing on flux data without any experimental alterations 94 (e.g., nutrient additions). Data were taken from the main text and of supplementary material. If the 95 relevant data was not reported in the text, we used the WebPlotDigitizer tool 96 97 (https://automeris.io/WebPlotDigitizer/) to read values from graphs. One study can have multiple entries, while one entry is an average of multiple observations. Non-English and non-peer-reviewed 98 99 papers were excluded, the rest of the papers were screened according to the title (if the title was not clear, the abstract was screened), and some were removed due to the irrelevance of study, 100 101 meaning that they were not focusing on CH<sub>4</sub> fluxes. Additionally, we constrained this review by focusing on natural freshwater ecosystems or mesocosms mimicking natural systems. Hence papers 102 103 on rice, constructed wetlands receiving wastewater, and water treatment facilities were not considered. 104

Although we excluded papers dealing with CH<sub>4</sub> fluxes from rice fields from our vegetation flux database, we did review the methods used in rice field research and discuss their applicability
 in freshwater ecosystems.

Based on the data from relevant articles (n = 159), we compiled a database with specific information, such as the genus of the dominant plant, as well as mean and range of CH<sub>4</sub> fluxes. To explore the geographical distribution of performed studies, climate zones were assigned according to the latitude of reported coordinates:  $60 - 90^{\circ}$  arctic;  $60 - 45^{\circ}$  boreal;  $45 - 20^{\circ}$  temperate;  $20 - 0^{\circ}$ tropic. For experimental studies (e.g., based on mesocosms), we considered the location of the experiment, which does not necessarily represent the geographical location of the mimicked ecosystem.

115

# 116 2.2 Data analysis

117 2.2.1 Differences between types of plants and ecosystems

118 After compiling the data obtained from the literature, we conducted the following steps: 119 Firstly, for 37% of the 372 entries, we calculated the average since it was not presented in the 120 respective publications. This was done by using the minimum and maximum value (62%), a time range presented (36%), or from replicates (2%). Secondly, all fluxes were converted to mg CH<sub>4</sub> m<sup>-</sup>  $^2 d^{-1}$ . Thirdly, we classified studies into three main groups: running waters (stream + river), lakes (ponds + lakes), and wetlands (bog + marsh + fen + mire + swamp).

In a first linear mixed-effect model (LME), we tested the fixed effects ecosystem type 124 (running waters, lakes, wetlands), study period (growing, non-growing), plant type (emergent, 125 submerged, floating), and climate zone (arctic, boreal, temperate, tropic) on CH<sub>4</sub> fluxes of 126 vegetated areas (response variable). Because the model with the full range of CH4 fluxes of 127 vegetated areas data (-454.4 to 2882.4 mg CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup>) was not valid (i.e., residuals were not 128 normally distributed), we built the model for the range of -96.0 to 2882.4 mg CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup>, excluding 129 the minimum value of the dataset (-454.4 mg CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup>). In a second LME, we zoomed in on the 130 emergent plants, for which we had most data. We tested the fixed effects plant genus (34 individual 131 genera), ecosystem type (lakes, wetlands; there were no studies of emergent plants in running 132 waters), study period (growing, non-growing), and climate zone (arctic, boreal, temperate, tropic) 133 on CH<sub>4</sub> fluxes of emergent plants (response variable). We used the "lmer" function of the R-134 package "lme4" (27) with Maximum Likelihood estimation. For the LMEs, we included study ID 135 136 as a random effect on the intercept to account for the fact that one study can have multiple entries. 137 Statistical significances of fixed effects were assessed with likelihood ratio tests using the function "drop1" (28). The LMEs were followed by a model validation, checking the residuals for normal 138 distribution and homogeneity of variances. For both LMEs, we transformed the response variable 139 140  $(\log_{10}(\operatorname{plant_flux}) + \min(\operatorname{plant_flux}) + 4)$  to improve the model performance and validity. Both LMEs were followed by a pairwise comparison posthoc test (Tukey adjustment for multiple 141 comparisons), comparing the individual levels of the significant fixed effects using the R-package 142 "emmeans" (29). 143

- 144
- 145 2.2.2 Global importance

To estimate the global importance of plant effects on CH<sub>4</sub> fluxes in lakes, we performed a
Monte Carlo simulation including and excluding the plant effects of CH<sub>4</sub> fluxes.

148 We used the following two equations for the Monte Carlo simulations (10'000 iterations):

149 Model A (vegetation not taken into account): Global lake  $CH_4$  emission =

- 150 global lake area \* (diffusive  $CH_4$  flux + ebbulitive  $CH_4$  flux)
- 151 Model B (vegetation taken into account):  $Global lake CH_4 emission =$
- 152  $((global lake area vegetated area) * (diffusive CH_4 flux + ebullitive CH_4 flux)) +$
- 153 (vegetated area  $*CH_4$  fluxes of vegetated areas)

We used the global distribution of lake areas provided by DelSontro et al. (30) (based on 31), which includes the global area of lakes and impoundments, separated in eight size class bins (bin 1:  $0.001 - 0.01 \text{ km}^2$ , bin 2:  $0.01 - 0.1 \text{ km}^2$ , bin 3:  $0.1 - 1 \text{ km}^2$ , bin 4:  $1 - 10 \text{ km}^2$ , bin 5:  $10 - 100 \text{ km}^2$ , bin 6:  $100 - 1000 \text{ km}^2$ , bin 7:  $1000 - 10'000 \text{ km}^2$ , bin 8:  $10'000 - 1 * 10^6 \text{ km}^2$ ). Subsequently, we subset the global dataset of diffusive and ebullitive fluxes (from 30) according to the same size class bins (both fluxes in mg C-CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup>). For the areal vegetation coverage, we used 83 South American shallow lakes from the Salga data set (32) and 109 lakes around the world from the data set of Zhang et al. (33). For the Salga data set, we summed the areal coverage of all three vegetation types (i.e., floating, emergent, and submerged); if the sum was higher than 100%, we fixed the value to 100%. For the Zhang data set, we calculated the average coverage (either floating, emergent, or submerged, or different combinations of these; in %) of all available years. For the CH<sub>4</sub> fluxes of vegetated areas, we used the lake fluxes of the compiled database of this study (in mg C-CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup>).

As for the CH<sub>4</sub> fluxes of vegetated areas from our data set, we combined all data from different study periods (i.e., growing, non-growing, and all year data), as well as from all climate zones since none of these parameters has a significant effect on the CH<sub>4</sub> fluxes of vegetated areas (see Table 2). Although the vegetation type significantly influences CH<sub>4</sub> fluxes of vegetated areas (Table 2), we combined all data of different vegetation types to simplify the analysis and increase the data power (i.e., number of observations).

Both models (A and B) were run using random sampling of the Monte Carlo simulations 173 based on the probability density functions (PDF) fit to the original data with few exceptions: We 174 fit the ebullitive and diffusive CH<sub>4</sub> fluxes of the first seven bins to a PDF (ebullitive: all log-normal 175 176 distribution except normal distribution for first bin; diffusive: all log-normal distribution). Bin 8 177 had too few ebullitive and diffusive CH<sub>4</sub> flux data to fit a proper PDF; hence the data were sampled from the original data. Lake CH<sub>4</sub> fluxes of vegetated areas and lake vegetation coverage data were 178 as well fit to a log-normal PDF. Since fitting PDF to data also introduces negative values, we 179 180 truncated the PDF at zero when negative values are physically impossible (vegetation coverage, ebullitive flux) or when no negative data were in the original data sets (diffusive fluxes as well as 181 CH<sub>4</sub> fluxes of vegetated areas). 182

For global estimates, we summed the estimates of all eight bins for each simulation and multiplied these data times 365 days and 10<sup>-15</sup>, expressing the data in Tg C-CH<sub>4</sub> yr<sup>-1</sup>. We tested if the resulting distributions of models A and B are different with a Two-sample Kolmogorov-Smirnov test using the "ks.test" function.

We consider this global upscaling exercise as a first attempt to obtain a rough idea about the potential relevance of vegetated lake areas for global CH<sub>4</sub> fluxes. The advantage of using Monte Carlo simulations is that we can run the models with and without vegetation with partially the same input data, in a consistent way, and get an idea about and highlight the uncertainty of the global estimates in the form of the interquartile range. We acknowledge that figures obtained in this way should be viewed with caution due to the limited data of CH<sub>4</sub> fluxes of vegetated areas and areal estimates of vegetation cover in lakes.

For running waters, even less data was available, which impeded us from assessing the potential impact of vegetation on global riverine CH<sub>4</sub> emissions. The incorporation of vegetation in existing wetland CH<sub>4</sub> models varies, which we discuss in a separate section (see section 3.4.1).

197 All statistical analyses were performed using the statistical programming language R (34).

C

6

h

d 02

#### **Results and discussion** 198 3

**∿**<sup>∆</sup>∕

0

Ш

#### 200 3.1 The impact of vegetation on CH4 emission

201

199





203	Figure 1. Plant-mediated process	ses that influence methar	ne (CH <sub>4</sub> ) emis	sions from	freshwat	er
		1 1 1 1 0				

systems. The depicted plant represents a hybrid of emergent, floating, and submerged vegetation 204 to illustrate the main processes of these three different functional groups. The water layer is 205

Δ

divided into an oxic (light blue) and anoxic (dark blue) zone. The processes illustrated are related 206

207 to CH<sub>4</sub> production (1 - 6, red), CH<sub>4</sub> oxidation (a - d, blue), and CH<sub>4</sub> transport (I - III, yellow).

Plant-mediated effects on CH<sub>4</sub> production include the provision of organic carbon through root 208

exudates (1) and decaying organic matter (2), sediment accumulation due to reduced water flow

- (3), dissolved carbon release followed by oxic methanogenesis in the water column (4),  $CH_4$
- production within the plant (5), and reduced sediment resuspension (6). CH<sub>4</sub> oxidation is affected
- by radial oxygen loss (ROL) in the root zone (a) potentially related to photosynthesis (b), CH<sub>4</sub>
- oxidation by epiphytic and endophytic methanotrophs (c), and oxygen  $(O_2)$  availability in the
- oxic water or sediment layer (d). Plants can alter the transport of  $CH_4$  by serving as a bypass
- between the anoxic sediment and the atmosphere (I), by impeding the movement of bubbles from
- the water to the atmosphere (II), and by reducing water-atmosphere gas exchange (III).
- 217
- 218 3.1.1 CH<sub>4</sub> production

Decaying plant matter can fuel CH<sub>4</sub> production in the anoxic sediment by providing organic matter, a precursor of the substrate for methanogens (35, 36). The intensity of methanogenesis depends on the water content and quality of this organic matter (e.g., C:N and C:P ratios) (7, 8). In a study of a boreal lake, for instance, *Lemna trisicula* produced twice as

223 much CH<sub>4</sub> per gram biomass as *Phragmites australis* (14).

224 In addition to decaying organic matter, an important pool of labile carbon substrate is 225 formed by root exudates, such as acetate (37, 38). Acetate exudation per gram root can vary 226 considerably between species. For instance, Eriophorum vaginatum may release over seven times 227 more acetate than Carex rostrata and Juncus effusus (26). Furthermore, Turner et al. (39) found a 228 suppressing effect of root exudates on CH<sub>4</sub> oxidation in a thermokarst bog, presumably resulting 229 from competition for electron acceptors. In addition to enhancing sediment methanogenesis, the 230 release of labile organic carbon could also fuel methanogenesis in the oxic water column, as has 231 been suggested for algae in rivers and lakes (40–42).

Besides producing organic matter as a precursor for methanogenesis, aquatic vegetation can reduce water flow velocity and thus increase sedimentation rates (43, 44). In particular, fine sediments rich in organic matter form an optimal environment for methanogens due to high sediment surface area and a high potential for microbial biofilm formation (8, 45, 46). Reduced flow velocity may therefore enhance methanogenesis in vegetated water bodies (16, 47).

Although oxygen  $(O_2)$  generally suppresses methanogenesis, Wilmoth et al. (48) showed that temporary exposure of Sphagnum peat to  $O_2$  can increase CH<sub>4</sub> yields up to 2000- fold during subsequent anoxic conditions compared to Sphagnum peat without  $O_2$  exposure due to functional shifts in the microbiome of the redox- oscillated peat.

An additional CH<sub>4</sub> production pathway could be oxic, non-enzymatic production of CH<sub>4</sub> within plants (49, 50). This process has been observed in living and litter material of several terrestrial plant species and is likely related to the oxic chemical breakdown of pectine when a plant is exposed to UV radiation or other stressors (51, 52).

245

246 3.1.2 CH<sub>4</sub> oxidation

To overcome anoxia-related problems in waterlogged soils, much vascular wetland plants
transport oxygen from their shoots to below-ground tissues. O<sub>2</sub> is transported within gas-filled
aerenchyma via diffusion and/or pressurized flow-through (53, 54). This process supplies O<sub>2</sub> to

- roots and rhizomes and can also result in radial O<sub>2</sub> loss (ROL) to the rhizosphere. ROL enables
- the detoxification of harmful substances (e.g., sulphide), enhances nutrient uptake, and alters the
- microbial community structure (55, 56). The increased  $O_2$  availability in the rhizosphere
- enhances aerobic CH<sub>4</sub> oxidation. The importance of ROL in regulating CH<sub>4</sub> emissions is strongly
- dependent on plant species and developmental stage (57). For instance, Ström et al. (26) showed
- in peat monoliths that Carex-dominated vegetation emitted more than twice as much  $CH_4$  as
- Eriophorum-dominated vegetation due to a lower degree of rhizospheric CH<sub>4</sub> oxidation in Carex.
  In a Patagonian bog colonized by cushion plants (*Astelia pumila* and *Donatia fascicularis*), CH<sub>4</sub>
- 258 oxidation as a result of ROL even eliminated CH<sub>4</sub> emissions completely (9).

Besides enhancing CH<sub>4</sub> oxidation, O<sub>2</sub> release in the rhizosphere can also suppress 259 260 methanogenesis, as O<sub>2</sub> availability results in the oxidation of the alternative electron acceptors 261 nitrate, manganese(IV), iron(III), and sulphate. The use of these alternative electron acceptors in organic matter decomposition is energetically favorable compared to methanogenesis. O<sub>2</sub> release 262 in the rhizosphere can, therefore, indirectly lead to repression of methanogenesis, depending on 263 264 the pool of alternative electron acceptors and the magnitude of ROL (23). In addition to altering 265 the availability of substrate and electron acceptors, plants can influence methanogenesis and methanotrophy by altering nutrient availability, affecting the microbial community, including 266 methanogens and methanotrophs (58, 59). 267

268 The presence of vegetation can alter  $O_2$  concentrations not only in the sediment but also in 269 the water column. Floating vegetation forms a barrier at the water-air interface, limiting gas 270 exchange. This reduces O<sub>2</sub> diffusion into the water column and dissolved CH<sub>4</sub> release to the 271 atmosphere (6, 60). Additionally, vegetation can limit light intrusion into the water and thus reduce primary production and CH<sub>4</sub> oxidation (61). The physical barrier formed by floating 272 273 plants not only impedes diffusion but also captures bubbles, enhancing their residence time and 274 potential oxidation (6). ROL can also occur in floating plants and adventitious roots, reducing 275 dissolved CH<sub>4</sub> concentrations in the water column (6, 62, 63). O<sub>2</sub> availability and, therefore, 276 potential CH<sub>4</sub> oxidation will thus strongly depend on the plant species and water column depth.

277 In addition to impacting microbial processes through alteration of their physical 278 environments, certain plant species also harbor methanotrophs directly on or within their tissues. 279 Epiphytic methanotrophs have been found on the shoots of a range of emergent and submersed plant species (12, 64-66). Endophytic methanotrophs have been found in Sphagnum mosses and 280 281 several vascular peatland plants (67, 68). Beyond harboring, Iguchi et al. (69) showed that duckweeds living in freshwater lakes are not only inhabited by methanotrophs, but the duckweed 282 plant actually has an enhancing effect on methane oxidation. Duckweeds may stimulate 283 methanotrophic growth, presumably by contributing certain metabolites (69). 284

285

**286** 3.1.3 CH<sub>4</sub> transport

Besides affecting CH<sub>4</sub> production and consumption processes, aquatic vegetation plays a vital role in CH<sub>4</sub> transport. Most notably, aerenchymous plants form a direct conduit for CH<sub>4</sub> from the sediment to the atmosphere, causing CH<sub>4</sub> to bypass oxidation in oxic sediment and water layers (70). This has also been observed for floating plants rooting in the sediments of

- 291 shallow waters (22). Depending on plant species characteristics, CH<sub>4</sub> transport through the plant 292 can take place by diffusion (driven by differences in concentration) or by convective flow (driven 293 by differences in pressure) (71). These processes result in the abovementioned transport of  $O_2$  to 294 the root zone and the opposite flow of CH<sub>4</sub> from roots to shoots. Plant-mediated CH<sub>4</sub> transport is 295 affected by, among other factors, temperature, light intensity, plant porosity, rates of 296 photosynthesis, and stomatal conductance, and may therefore vary diurnally and seasonally (e.g., 297 72–74). This "chimney effect" is estimated to contribute 55 to 85% to total CH<sub>4</sub> fluxes in 298 peatlands (10). Damage or herbivory may result in enhanced CH<sub>4</sub> transport: CH<sub>4</sub> emissions of 299 damaged (clipped) plants increased to 160% of control values for Carex aquatilis (75), and similar results were observed for herbivore-induced damage in a wetland plant community (18). 300 301 Plant-mediated transport may reduce sediment CH<sub>4</sub> concentrations to such an extent that
- it reduces ebullition. In an experimental set-up with *Phragmites australis*, for instance, plant
- 303 clipping resulted in a 5 to 10-fold increase in ebullition, whereas the total emission, including
- 304 plant-mediated transport, was about 1.8 times higher in intact plants (76).

#### 3.2 Methods to assess CH<sub>4</sub> fluxes of vegetated areas and geographical distribution of 306 307 studies

308

309 A wide variety of methods is used to assess CH<sub>4</sub> fluxes in vegetated areas, all with different strengths and drawbacks. The principal difference between the approaches is the scale of the CH<sub>4</sub> 310 311 fluxes they incorporate, thereby incorporating single or multiple emission pathways in the 312 vegetated areas. In addition, there is a strong bias towards growing season-only data and focus on 313 a small subset of genera and plant types. These biases all contribute to the large uncertainty of 314 freshwater systems in global CH<sub>4</sub> budgets (77).

In the screened literature (159 studies, 372 entries; some studies applied multiple methods), in 315 316 total ten methods were applied to measure CH<sub>4</sub> fluxes of vegetated areas (Fig. 2; Table 1): (i) 317 chamber (141 studies; 343 entries; Fig. 2A), (ii) porewater sampling (14 studies; 23 entries: Fig. 2B), (iii) eddy covariance flux tower (10 studies; 19 entries; Fig. 2C), (iv) inverted funnel (6 318 studies; 10 entries; Fig. 2D), (v) concentration measurement and k value (6 studies; 9 entries; Fig. 319 2E), (vi) bottle incubation (2 studies; 7 entries; Fig. 2F), (vii) gas sampling from plant tissue (2 320 studies; 4 entries; Fig. 2G), (viii) submerged chamber (2 studies; 3 entries; Fig. 2H), (ix) leaf 321

- chamber (1 studies; 3 entries; Fig. 2I), and (x) inverted water-filled vial (1 study; 1 entry; Fig. 322 2J).
- 323

324



- Figure 2. Methods to investigate CH<sub>4</sub> fluxes of vegetated areas (field and laboratory): chamber 326
- (A), porewater sampling (B), eddy covariance flux tower (C), inverted funnel (D), concentration 327
- measurement and k value (E), bottle incubation (F), gas sampling from plant tissue (G), 328
- submerged chamber (H), leaf chamber (I), inverted water-filled vial (J), stacked chamber (K), 329
- diffusion potential of specific plant parts (L). 330

# **Table 1.** Summary of methods used to determine CH<sub>4</sub> fluxes of vegetated areas.

Method	Description	Exemplary references
Natural freshwater systems		
Chamber	Transparent plexiglass chamber	
(Fig. 2A)	with a fan placed over the	
	vegetation (floating or on a	(78–80)
	frame/base; measurement of	
	headspace CH <sub>4</sub> increase over	
	time)	
Porewater sampling	Sampling of CH <sub>4</sub> concentration	(80, 81)
(Fig. 2B)	in porewater near and away from	
	roots (with vials connected to	
	rhizons)	
Eddy covariance (EC) flux	Placed in such a way that the EC	(76, 82, 83)
tower	tootprint represents vegetated	
(Fig. 2C)	and/or unvegetated areas	
Inverted funnel	Using an inverted funnel to trap	
(F1g. 2D)	and measure volume and	(20, 84, 85)
	concentration of $CH_4$ bubbles	(20, 84, 85)
	(e.g., placed at the surface and a	
	deeper depui)	
Concentration measurement and	Developing a model based on	(86)
k value	laboratory and/or field results to	
(Fig. 2E)	estimate the gas exchange	
	coefficient based on temperature	
	and wind speed	
Pottle insubstion	Massuring plant associated CH	(97)
(Fig. 2E)	production and/or oxidation by	(87)
(119.21)	incubating in the dark e.g.	
	roots rhizomes or tillers in	
	glass bottles measuring	
	headspace CH <sub>4</sub> partial pressure	
	over time	
Gas sampling from plant tissue	Measurement of CH <sub>4</sub>	(88, 89)
(Fig. 2G)	concentration within plant stem	
	(e.g., 1 cm below the waterline	
	via a syringe)	
Submerged chamber	Capturing CH <sub>4</sub> release of	
(Fig. 2H)	submerged plants by submerged	(4, 64)
	bags (e.g., made of a multi-laver	

	clear film of saran and PVC with low CH <sub>4</sub> permeability)	
Leaf chamber (Fig. 2I)	Transparent plastic storage boxes sealed with rimmed	
	weather stripping and large binder clips (measurement of headspace CH4 increase over	(11)
	time)	
Inverted water-filled vial	Clipping of main stem and	
(Fig. 2J)	placing an inverted water-filled	(20)
	vial over it (leaving vial for a	
	defined amount of time)	
Rice literature		
Stacked chamber	Installing two chambers with	
(Fig. 2K)	different diameters: larger one	
	covering water surface with lid	
	sealed around the plant, smaller	(90, 91)
	one only enclosing plant	
Diffusion potential of specific	A single plant part (e.g., leaf,	(92)
plant parts	panicle, node, or internode) is	
(Fig. 2L)	inserted in the hole of a rubber	
	stopper fitted onto a flask filled	
	with $CH_4$ enriched water; the	
	flask stands in a water-filled	
	container to provide a water lock	
	for the chamber covering the	
	plant part. $CH_4$ emission rates	
	are determined by temporal	
	increase in concentration in the	
	cnamber.	

The majority of the reviewed studies applied floating chambers. In a practical sense, this 333 334 method seems straightforward, but it comes along with certain issues. Firstly, it is difficult to place the floating chamber on top of emerging and floating vegetation without disturbing the 335 plant. Disturbance of vegetation may liberate gas bubbles inducing a peak emission; in addition, 336 the chamber alters moisture, light, and carbon dioxide (CO<sub>2</sub>) concentrations which impacts 337 338 stomata and may consequently influence CH<sub>4</sub> fluxes. Secondly, often the chamber does not cover 339 the entire plant; when part of the root system exchanges gas with the overlying water outside the chamber area, these fluxes may be missed (e.g., 93). Thirdly, fluxes measured in floating 340 chambers integrate plant-mediated and diffusive fluxes, complicating the disentangling of 341 342 different flux pathways.

The stacked chamber approach, used in rice studies (90, 91), could be used to better differentiate between actual plant-mediated CH<sub>4</sub> fluxes, diffusion, and/or ebullition, avoiding the potentially misleading integration discussed above. The diffusion potential method (92) can be used to scan for possible within-plant hotspots of CH<sub>4</sub> flux.

Most of all reviewed studies (159 studies, 372 entries; one study performed both approaches) were performed in the field (134 studies; 297 entries) compared to mesocosm experiments (26 studies; 75 entries). Studies on all scales are valuable to both obtain mechanistic understanding and enable upscaling efforts. Furthermore, mesocosm experiments can be used to simulate running waters/flow conditions, which would help to constrain and to understand CH<sub>4</sub> fluxes of vegetated areas in running waters.

From the 159 compiled studies (of which one study was performed on two continents), 70 were performed in North America (154 entries), followed by 62 in Europe (161 entries), and 21 (48 entries), 5 (7 entries), 1 (1 entry), and 1 (1 entry) in Asia, South America, Oceania, and Australia, respectively (Fig. 3). Besides missing data in, e.g., Africa, there are only a handful of studies in South America (e.g., 94, 95), where some of the largest inland aquatic systems are located (e.g., Amazon and Pantanal). Consequently, our current understanding of the magnitude of CH<sub>4</sub> fluxes of vegetated areas is strongly biased.



Figure 3. Geographical distribution of studies related to CH<sub>4</sub> fluxes of vegetated areas (A) with a
zoom-in of Europe (B). The number of studies is indicated on the respective countries
accompanied by different color intensities indicating the higher frequency (higher color intensity)
and lower frequency (lower color intensity) of performed studies. Grey areas indicate no
available studies according to our literature search.

367

# 368 3.3 CH4 flux intensity and variability from vegetated areas

369

Although our systematic literature review points out that there are spatial and temporaldata gaps and that different methods cannot readily be compared, we provide a first overview.

- From an ecosystem perspective, of total 159 studies (372 entries), 136 were done in wetlands (318 entries), whereas only 22 (53 entries) and 1 (1 entry) in lakes and running waters, respectively. This implies that the available data are strongly imbalanced, urging for more studies in standing and running waters.
- 376 CH<sub>4</sub> fluxes of vegetated areas ranged from -454.4 (uptake) to 2882.4 mg CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup> 377 (emission) (62.1 [8.8, 183.8]; median [interquartile range (IQR)]) in wetlands, from 0.2 to 1960.0 378 mg CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup> (76.0 [34.6, 230.0]) in lakes, and on average 5.4 mg CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup> in running

- waters (only one entry available) (Fig. 4). There was neither a significant effect of ecosystem
- type nor climate zone on  $CH_4$  fluxes of vegetated areas (Table 2A).

- **Table 2.** Results of the linear mixed models testing effects of ecosystem type (running waters,
- lakes, wetlands), study period (growing, non-growing), plant type (emergent, floating,
- submerged), climate zone (arctic, boreal, temperate, tropic) on CH<sub>4</sub> fluxes of vegetated areas (A),
- and testing the effects of plant genus (34 individual genera), ecosystem type (lakes, wetlands),
- study period (growing, non-growing), and climate zone (arctic, boreal, temperate, tropic), on CH<sub>4</sub>
- 387 fluxes of vegetated areas of emergent plants (B). Study ID was included as a random effect on the
- intercept. Significances of fixed effects were assessed with likelihood ratio tests with degrees of
- 389 freedom = 1. Significant p values <0.05 are in bold.

Response	Fixed effect	$\chi^{2}(1)$	р
variable			
A) Testing cor	nbined CH4 fluxes of vegetated	areas	
CH <sub>4</sub> fluxes of	Ecosystem type	1.068	0.586
vegetated areas*	Study period	2.698	0.100
	Plant type	9.268	0.010
	Climate zone	4.919	0.178
<b>B</b> ) <b>Testing CH</b>	I4 fluxes of vegetated areas of e	nergent pla	ants
	Plant genus	64.601	< 0.001
CH <sub>4</sub> fluxes of	Ecosystem type	0.112	0.738
vegetated areas*	Study period	2.146	0.143
	Climate zone	3.140	0.371

\* Transformation: log10 (CH<sub>4</sub> fluxes of vegetated areas + abs (min (CH<sub>4</sub> fluxes of vegetated

**391** areas)) +4)

A) Marginal  $R^2 = 0.065$ , conditional  $R^2 = 0.537$ , sample size = 301.

B) Marginal  $R^2 = 0.260$ , conditional  $R^2 = 0.614$ , sample size = 192.





Figure 4. CH<sub>4</sub> fluxes of vegetated areas of different ecosystem types (wetlands, lakes, and
running waters). Boxplots represent median (black line), first and third quartiles (hinges), range
(whiskers), and outliers (black dots).

395

From all the 159 studies (372 entries), only 11 studies (20 entries) are from the non-400 growing season, compared to 117 studies (282 entries) performed in the growing season, while 401 29 studies (61 entries) include measurements throughout the year. The very similar median fluxes 402 (Fig. 5) in the growing and non-growing season calls for more attention to CH<sub>4</sub> fluxes of 403 vegetated areas outside the growing season. For example, the genus Phragmites does not grow in 404 winter but may facilitate CH<sub>4</sub> emissions by creating a direct pathway or "chimney" from the 405 406 sediments to the atmosphere while the water column is potentially sealed off from the atmosphere 407 by ice. Fluxes outside the growing period may be considerable and need further attention.

Study period did not significantly influence CH<sub>4</sub> fluxes of vegetated areas (p = 0.100; Table 2A) with fluxes ranging from -454.4 to 2882.4 mg CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup> (55.7 [8.5, 200.3]) in the growing season, and from 0.1 to 553.2 mg CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup> (74.3 [7.7, 131.8]) in the non-growing season (Fig. 5). One explanation of the absent significant effect might be that organic matter supply rate (96), e.g., due to dying vegetation, could actually be more relevant in driving CH<sub>4</sub> production than the division of growing and non-growing seasons.



Figure 5. CH<sub>4</sub> fluxes of vegetated areas from different plant types combined, measured in 3
different seasons (growing and non-growing season, and year-round studies). Boxplots represent

418 median (black line), first and third quartiles (hinges), range (whiskers), and outliers (black dots).

419

415

There was a strong imbalance in the focus on different plant growth forms/types, with the majority of studies focusing on emergent plants and only a few studies focusing on floating and particularly submerged plants (Fig. 6).

The fluxes for emergent plants ranged from -454.4 to 2882.4 mg CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup> (62.4 [12.0, 423 184.4]), for floating plants from 3.9 to 1860.8 mg CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup> (77.0 [43.5, 391.8]), and for 424 submerged plants from 5.4 to 380.9 mg CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup> (50.6 [15.4, 192.7]). Fluxes from mixed 425 stands with a mix of different plant types ranged from 4.2 to 9.9 mg CH<sub>4</sub> m<sup>-2</sup> d<sup>-1</sup> (7.1 [5.6, 8.5]) 426 (Fig. 6). These differences led to a significant effect of plant type on CH<sub>4</sub> fluxes of vegetated 427 areas (p = 0.010; Table 2A), with significantly higher fluxes from floating plants compared to 428 emergent plants (post hoc pairwise comparison; p = 0.011). Different processes may underlie the 429 high emissions from areas with floating plants. Particularly the low oxygen conditions below the 430 431 floating plants related to the inhibition of photosynthesis and the hampered oxygen intrusion into the water distinguish the floating plants from other plants (see also section 3.1). The low oxygen 432 433 availability in the water column and sediment creates favorable circumstances for methanogenesis. When the floating plants are rooted in the sediment – thereby connecting the 434 high CH<sub>4</sub> concentrations in the sediment directly to the atmosphere through plant-mediated 435 transport, emissions may be particularly high (22, 94). However, only a small fraction of studies 436 indicated whether the studied floating plants were free-floating or rooted in the sediment. This 437 438 information should be better considered and reported in future studies.



Figure 6. CH<sub>4</sub> fluxes of vegetated areas from different plant types. Emergent: 147 studies, 337
entries; floating: 15 studies, 24 entries; submerged: 2 studies, 9 entries; mixed: 1 study, 2 entries.
Boxplots represent median (black line), first and third quartiles (hinges), range (whiskers), and

444 outliers (black dots).

445

440

Within the emergent plants (337 entries), most data are from mixed plants (28.2%), 446 followed by Carex (16.6%), Eriophorum (9.8%), Phragmites (9.2%), and Typha (4.5%). Not 447 surprisingly, seen in the distribution of the studies (Fig 3), emphasis has been on plants widely 448 occurring in the arctic, subarctic, and temperate regions. The remaining 46 genera/combination of 449 genera represents less than 4.0% of the entries (Table S1). Within floating plants (24 entries), 450 451 33.3% of the data are from the genus Nuphar, followed by Eichhornia (25.0%) and Nelumbo and 452 Nymphaea with 12.5% each (Table S1). Finally, most data for submerged plants (9 entries) are from Myriophyllum (33.3%) and Ranunculus (22.2%) (Table S1). 453

454 CH<sub>4</sub> fluxes from different genera within the different plant types are highly variable (Fig. 7 & S1). Being the strongest predictor in the LME run on fluxes of emergent plants, the genus 455 456 had a significant effect (p < 0.001; Table 2B). Particularly Glyceria stands out with significantly higher CH<sub>4</sub> fluxes than Carex and Spartina (pairwise comparison posthoc; p = 0.032 and p =457 458 0.043). As mentioned above, there is a strong imbalance related to the number of entries of individual genera. A higher number of entries for a specific genus (e.g., for Phragmites) likely 459 leads to higher variability since it has been studied in very different systems, which creates a 460 certain bias and an imbalanced model. We are therefore currently unable to predict CH<sub>4</sub> fluxes 461 based on occurring genera on a large scale. Given the large variability in CH<sub>4</sub> fluxes within 462 463 genera, it is also questionable if reasonable genera inferred predictions of CH<sub>4</sub> fluxes can be developed. Predictions made based on vegetation types - aggregating several genera at a specific 464 site, thereby "controlling" for climate and soil type - are likely more promising (see e.g., 56, 97). 465



467

Figure 7. CH<sub>4</sub> fluxes of vegetated areas from individual genera of investigated emergent plants.
Boxplots represent median (black line), first and third quartiles (hinges), range (whiskers), and
outliers (black dots).

471

# 472 3.4 Plant effects on CH<sub>4</sub> fluxes: Implications for current global CH<sub>4</sub> estimates 473

## 474 3.4.1 Wetlands

While vegetation presence and vegetation composition are commonly used to model CH<sub>4</sub> 475 emissions at local to regional scales (e.g., 98–100), the effect of vegetation on CH<sub>4</sub> emissions is 476 often not included in larger-scale models. Global models of wetland-CH<sub>4</sub> feedbacks indicate that 477 wetland CH<sub>4</sub> emissions could drive 21<sup>st</sup>-century climate change, with global wetland emissions 478 matching or exceeding anthropogenic emissions by 2100 (101). However, modeled CH<sub>4</sub> 479 480 emissions vary strongly (102, 103), partly due to a poor understanding of the role of vegetation in 481 CH<sub>4</sub> production, oxidation, and transport (104, 105). Many wetland models (e.g., 106–109) incorporate the primary production of vegetation as it fuels carbon stocks that are subsequently 482 available for decomposition. Only a few take the transport through plants into account (e.g., 483 Kleinen et al. (110) using the CH<sub>4</sub> transport model by Riley et al. (105) and Ito & Inatomi (111)). 484 Potentially influential factors such as the impact of vegetation community composition on 485 486 transport through plants, as well as plant effects on CH<sub>4</sub> oxidation, are generally not included in larger-scale models (but see 101, who included CH<sub>4</sub> oxidation during plant-mediated transport in 487 their global wetland model). Upscaling of the many ways plants can impact CH<sub>4</sub> –as summarized 488 in Fig. 1- is no sinecure but the lack of mechanistic processes (see examples in section 3.1). 489 490 Describing CH<sub>4</sub> emissions from wetlands is an important caveat as it is estimated that herbaceous plants represent ca. 28 to 90% of the total ecosystem-level CH<sub>4</sub> flux from wetlands (3) which is a 491

492 globally significant amount considering the total annual wetland  $CH_4$  emission of 149 TgCH<sub>4</sub> yr<sup>-1</sup> 493 (1).

494

495 3.4.2 Inland waters

The global annual CH<sub>4</sub> emission for rivers is estimated to be 27 TgCH<sub>4</sub> yr<sup>-1</sup> (112). This estimate is based on a simple bootstrapping approach using published diffusive riverine CH<sub>4</sub> fluxes. These fluxes exclude emissions from emergent vegetation, and it is unclear to which extent they include submerged vegetation. Our literature search, unfortunately, yielded only one study (20) where both CH<sub>4</sub> emissions from vegetated river sections and unvegetated sections were compared. We could therefore not estimate the relevance of vegetation for riverine CH<sub>4</sub> fluxes.

In a current global CH<sub>4</sub> emission estimate of lakes of 104 TgCH<sub>4</sub> yr<sup>-1</sup> (30), vegetated 503 areas are not explicitly considered. In a recent overview study on the global regulation of CH<sub>4</sub> 504 emissions from lakes, the emission via vegetation is even explicitly excluded, and only open 505 water fluxes are considered (113). Our estimate of the global importance of CH<sub>4</sub> fluxes of 506 vegetation in lakes, based on a comparison between a simulated global CH<sub>4</sub> flux with and without 507 considering the effect of vegetation on CH<sub>4</sub> fluxes, indicates that the median global CH<sub>4</sub> emission 508 of lakes neglecting vegetation is 123.9 Tg C-CH<sub>4</sub> yr<sup>-1</sup> (IQR: 80.9, 202.3). The simulated CH<sub>4</sub> 509 emission from lakes taking the vegetation effect into account is 180.8 Tg C-CH<sub>4</sub> yr<sup>-1</sup> (IQR: 122.8, 510 511 275.5), which has a significantly different distribution compared to without vegetation (Two-512 sample Kolmogorov-Smirnov test; p < 0.001). These estimations imply that we potentially underestimate global CH<sub>4</sub> emissions from lakes by 46% (IQR: 36; 52). These 46% 513 514 underestimations are in line with a study of Jeffrey et al. (114) which showed that plant-mediated CH<sub>4</sub> fluxes (seasons aggregated and weighting for changes in lily coverage) accounted for ~59% 515 of the annual CH<sub>4</sub> emissions. In contrast, ebullition and diffusion each accounted for ~20%. As 516 517 pointed out before in this review, it is important to consider that the flux data used for this upscaling exercise do not reflect the entirety of the habitat, and it is unclear which aspects and 518 519 how much of the vegetated habitat is covered (see section 3.2). Hence, it is paramount to consider CH<sub>4</sub> fluxes from a vegetated habitat perspective and standardize related measurement approaches 520 521 and terminologies, to improve these estimates.

- 522
- 523

# 524 **4 Conclusions and outlook**

525

526 CH<sub>4</sub> fluxes from vegetated areas are largely understudied and neither well understood nor
527 quantified. The here presented study is, according to our knowledge, the first attempt to
528 synthesize the many ways in which vegetation impacts CH<sub>4</sub> fluxes across different freshwater

529 ecosystems, combined with a compilation of methodological approaches used to assess

vegetation effects on CH<sub>4</sub> related processes. Moreover, we provide an overview of flux intensities

across a geographical, seasonal, and vegetation type related spectrum and provide a first-order

estimate of the significance of CH<sub>4</sub> fluxes from vegetated areas for global lake CH<sub>4</sub> emissions.

533 From a methodological perspective, we urge for a more standardized way of measuring 534 CH<sub>4</sub> fluxes from vegetated areas. We consider not only measuring the growing season but also the non-growing season as pertinent to reduce biased estimates on an annual basis. To measure 535 536 specifically plant-mediated CH<sub>4</sub> fluxes, we consider the stacked chamber method (Table 1) as the most appropriate one since it allows to separate the plant-mediated flux from the diffusive flux. 537 538 Alternatively, we suggest trying to capture CH<sub>4</sub> fluxes from the vegetated areas as a whole by measuring diffusion, ebullition, and plant-mediated fluxes combined. Further, we suggest 539 assessing if plants are rooted or non-rooted when measuring fluxes from floating plants since this 540 information is relevant for future upscaling or generalization efforts. 541

542 By showing a comprehensive overview of how vegetated areas can affect CH<sub>4</sub> dynamics 543 in numerous aspects, we urge further studies to consider a vegetated habitat view instead of 544 focusing solely on plant-mediated CH<sub>4</sub> fluxes. The comprehensive overview given in this study 545 may pave the way for future studies tackling vegetation-related aspects of CH<sub>4</sub> production, 546 oxidation, and/or transport in the light of global and land-use change. We believe that linking the 547 elaborated processes to quantifiable species traits is an important next step.

548 Despite being beyond the scope of this study, we recommend considering CO<sub>2</sub> and nitrous 549 oxide fluxes beside CH<sub>4</sub> fluxes to fully capture the role of vegetated areas in the carbon and 550 greenhouse gas cycle. If CH<sub>4</sub> fluxes are described on their own, it is important to consider that 551 those vegetated areas also store carbon; otherwise, we might exaggerate their potential radiative 552 effect. Although not specifically accounting for plant-mediated CH<sub>4</sub> fluxes, it has been shown 553 that CH<sub>4</sub> emissions partially offset "blue carbon" burial in mangroves (115). Similar assessments 554 in freshwater systems are scarce (but see 116, 117).

Although general global trends in plant coverage in the light of global change are difficult to predict (33), they seem to differ among vegetation types (i.e., emergent, floating, submerged). Specifically, global change trends seem to favor floating over submerged vegetation (118, 119). However, the data basis for this trend is still weak. It was furthermore shown that invasive vegetation has the potential to increase CH<sub>4</sub> emissions from a subtropical tidal estuarine wetland (120). Hence, the understudied and highly likely underestimated CH<sub>4</sub> fluxes from vegetated areas might get even more relevant due to global warming and land-use change.

At present, upscaling CH<sub>4</sub> flux from vegetated areas to the landscape and global scale is challenging. To narrow down estimates of the global contribution of this source depends on (i) more standardized methods with clear accounting for and disentangling of different CH<sub>4</sub> flux pathways or making sure that the vegetated habitat is captured as a whole, (ii) flux measurements taken across the complete geographic extent and vegetation type (i.e., emergent, floating, and submerged), and (iii) a better and more complete assessment of vegetation cover using, e.g.,

- remote sensing techniques (e.g., 121). Despite the uncertainties, it is clear that CH<sub>4</sub> emissions
- from vegetated areas are an important source of global  $CH_4$  emissions from freshwater systems.
- 570 Especially studies of lakes and running waters may be missing an important pathway of the total
- 571 CH<sub>4</sub> flux. This omission introduces a major degree of uncertainty in the global estimation of CH<sub>4</sub>
- 572 emissions from freshwater systems.
- 573
- 574

# 575 Acknowledgments

- 576 We thank Vincent Fugère for his help in setting up the linear mixed models and Joan Pere Casas-
- 577 Ruiz for his support with the Monte-Carlo approach. We also thank Karelle Desrosiers for fruitful
- 578 discussions and input into the wetland section. Pascal Bodmer was partially financed by an
- individual research grant from the German Research Foundation (BO 5050/1-1).

#### 580 **References**

- M. Saunois, *et al.*, The global methane budget 2000–2017. *Earth Syst. Sci. Data* 12, 1561–
   1623 (2020).
- J. A. Rosentreter, *et al.*, Half of global methane emissions come from highly variable
  aquatic ecosystem sources. *Nat. Geosci.* (2021) https://doi.org/10.1038/s41561-021-007152.
- M. J. Carmichael, E. S. Bernhardt, S. L. Bräuer, W. K. Smith, The role of vegetation in
  methane flux to the atmosphere: Should vegetation be included as a distinct category in the
  global methane budget? *Biogeochemistry* 119, 1–24 (2014).
- J. W. H. Dacey, M. J. Klug, Methane efflux from lake sediments through water lilies. *Science (80-. ).* 203, 1253–1255 (1979).
- 5. A. Holzapfel-Pschorn, R. Conrad, W. Seiler, Effects of vegetation on the emission of
  methane from submerged paddy soil. *Plant Soil* 92, 223–233 (1986).
- 593 6. S. Kosten, *et al.*, Fate of methane in aquatic systems dominated by free-floating plants.
  594 *Water Res.* 104, 200–207 (2016).
- 595 7. C. Grasset, *et al.*, Large but variable methane production in anoxic freshwater sediment
  596 upon addition of allochthonous and autochthonous organic matter. *Limnol. Oceanogr.* 63,
  597 1488–1501 (2018).
- C. Grasset, G. Abril, R. Mendonça, F. Roland, S. Sobek, The transformation of macrophyte-derived organic matter to methane relates to plant water and nutrient contents. *Limnol. Oceanogr.* 64, 1737–1749 (2019).
- 601 9. C. Fritz, *et al.*, Zero methane emission bogs: extreme rhizosphere oxygenation by cushion
  602 plants in Patagonia. *New Phytol.* 190, 398–408 (2011).
- J. F. Dean, *et al.*, Methane Feedbacks to the Global Climate System in a Warmer World. *Rev. Geophys.* 56, 207–250 (2018).
- J. A. Villa, *et al.*, Plant-mediated methane transport in emergent and floating-leaved
  species of a temperate freshwater mineral-soil wetland. *Limnol. Oceanogr.* 65, 1635–1650
  (2020).
- N. Yoshida, H. Iguchi, H. Yurimoto, A. Murakami, Y. Sakai, Aquatic plant surface as a niche for methanotrophs. *Front. Microbiol.* 5, 30 (2014).
- S. Liu, Y. Chen, J. Liu, Methane emissions from the littoral zone of Poyang lake during
  drawdown periods. *J. Freshw. Ecol.* 34, 37–48 (2019).
- P. Kankaala, T. Käki, A. Ojala, Quality of detritus impacts on spatial variation of methane
  emissions from littoral sediment of a boreal lake. *Arch. Fur Hydrobiol.* 157, 47–66 (2003).
- P. Kankaala, A. Ojala, T. Käki, Temporal and spatial variation in methane emissions from
  a flooded transgression shore of a boreal lake. *Biogeochemistry* 68, 297–311 (2004).
- 616 16. P. Kankaala, *et al.*, Methane efflux in relation to plant biomass and sediment
  617 characteristics in stands of three common emergent macrophytes in boreal mesoeutrophic
  618 lakes. *Glob. Chang. Biol.* 11, 145–153 (2005).

619 620	17.	W. Ding, Z. Cai, H. Tsuruta, X. Li, Effect of standing water depth on methane emissions from freshwater marshes in northeast China. <i>Atmos. Environ.</i> <b>36</b> , 5149–5157 (2002).
621 622	18.	B. J. J. Dingemans, E. S. Bakker, P. L. E. Bodelier, Aquatic herbivores facilitate the emission of methane from wetlands. <i>Ecology</i> <b>92</b> , 1166–1173 (2011).
623 624	19.	T. A. Davidson, <i>et al.</i> , Synergy between nutrients and warming enhances methane ebullition from experimental lakes. <i>Nat. Clim. Chang.</i> <b>8</b> , 156–160 (2018).
625 626	20.	I. A. Sanders, <i>et al.</i> , Emission of methane from chalk streams has potential implications for agricultural practices. <i>Freshw. Biol.</i> <b>52</b> , 1176–1186 (2007).
627 628 629	21.	P. Milberg, L. Törnqvist, L. M. Westerberg, D. Bastviken, Temporal variations in methane emissions from emergent aquatic macrophytes in two boreonemoral lakes. <i>AoB Plants</i> <b>9</b> (2017).
630 631 632	22.	E. S. Oliveira-Junior, <i>et al.</i> , The impact of water hyacinth (Eichhornia crassipes) on greenhouse gas emission and nutrient mobilization depends on rooting and plant coverage. <i>Aquat. Bot.</i> <b>145</b> , 1–9 (2018).
633 634	23.	H. J. Laanbroek, Methane emission from natural wetlands: interplay between emergent macrophytes and soil microbial processes. A mini-review. <i>Ann. Bot.</i> <b>105</b> , 141–153 (2010).
635 636	24.	H. Brix, B. K. Sorrell, HH. Schierup, Gas fluxes achieved by in situ convective flow in Phragmites australis. <i>Aquat. Bot.</i> <b>54</b> , 151–164 (1996).
637 638	25.	D. I. Sebacher, R. C. Harriss, K. B. Bartlett, Methane Emissions to the Atmosphere Through Aquatic Plants 1. <i>J. Environ. Qual.</i> <b>14</b> , 40–46 (1985).
639 640	26.	L. Ström, M. Mastepanov, T. R. Christensen, Species-specific effects of vascular plants on carbon turnover and methane emissions from wetlands. <i>Biogeochemistry</i> <b>75</b> , 65–82 (2005).
641 642	27.	D. Bates, M. Mächler, B. Bolker, S. Walker, Fitting Linear Mixed-Effects Models Using lme4. J. Stat. Software; Vol 1, Issue 1 (2015) https://doi.org/10.18637/jss.v067.i01.
643 644	28.	A. Zuur, E. N. Ieno, N. Walker, A. A. Saveliev, G. M. Smith, <i>Mixed effects models and extensions in ecology with R</i> (Springer Science & Business Media, 2009).
645 646	29.	R. Lenth, Estimated marginal means, aka least-squares means. R package version 1.3. 5.1 (2019).
647 648 649	30.	T. DelSontro, J. J. Beaulieu, J. A. Downing, Greenhouse gas emissions from lakes and impoundments: Upscaling in the face of global change. <i>Limnol. Oceanogr. Lett.</i> <b>3</b> , 64–75 (2018).
650 651	31.	C. Verpoorter, T. Kutser, D. A. Seekell, L. J. Tranvik, A global inventory of lakes based on high-resolution satellite imagery. <i>Geophys. Res. Lett.</i> <b>41</b> , 6396–6402 (2014).
652 653	32.	S. Kosten, <i>et al.</i> , Ambiguous climate impacts on competition between submerged macrophytes and phytoplankton in shallow lakes. <i>Freshw. Biol.</i> <b>56</b> , 1540–1553 (2011).
654 655	33.	Y. Zhang, <i>et al.</i> , Global loss of aquatic vegetation in lakes. <i>Earth-Science Rev.</i> <b>173</b> , 259–265 (2017).
656 657	34.	R Core Team, R: A Language and Environment for Statistical Computing. <i>R Found. Stat. Comput.</i> (2021).

- R. Segers, Methane production and methane consumption: a review of processes
  underlying wetland methane fluxes. *Biogeochemistry* 41, 23–51 (1998).
- 36. J. R. Webster, E. F. Benfield, Vascular plant breakdown in freshwater ecosystems. *Annu. Rev. Ecol. Syst.* 17, 567–594 (1986).
- A. Joabsson, T. R. Christensen, B. Wallén, Vascular plant controls on methane emissions
  from northern peatforming wetlands. *Trends Ecol. Evol.* 14, 385–388 (1999).
- 38. L. Ström, A. Ekberg, M. Mastepanov, T. Røjle Christensen, The effect of vascular plants
  on carbon turnover and methane emissions from a tundra wetland. *Glob. Chang. Biol.* 9, 1185–1192 (2003).
- 39. J. C. Turner, et al., Getting to the Root of Plant-Mediated Methane Emissions and
  Oxidation in a Thermokarst Bog (2020).
- 40. M. J. Bogard, *et al.*, Oxic water column methanogenesis as a major component of aquatic
  670 CH 4 fluxes. *Nat. Commun.* 5, 1–9 (2014).
- 41. D. Donis, *et al.*, Full-scale evaluation of methane production under oxic conditions in a mesotrophic lake. *Nat. Commun.* 8, 1–12 (2017).
- 42. D. Mei, *et al.*, Filamentous green algae Spirogyra regulates methane emissions from eutrophic rivers. *Environ. Sci. Pollut. Res.* 28, 3660–3671 (2020).
- 43. S. J. Clarke, Vegetation growth in rivers: influences upon sediment and nutrient dynamics. *Prog. Phys. Geogr.* 26, 159–172 (2002).
- K. Sand-Jensen, O. Pedersen, Velocity gradients and turbulence around macrophyte stands
  in streams. *Freshw. Biol.* 42, 315–328 (1999).
- 45. J. M. Sanchez, S. Arijo, M. A. Munoz, M. A. Morinigo, J. J. Borrego, Microbial
  colonization of different support materials used to enhance the methanogenic process. *Appl. Microbiol. Biotechnol.* 41, 480–486 (1994).
- 46. P. Bodmer, J. Wilkinson, A. Lorke, Sediment Properties Drive Spatial Variability of
  Potential Methane Production and Oxidation in Small Streams Journal of Geophysical
  Research : Biogeosciences. 1–15 (2020).
- 47. J. T. Crawford, E. H. Stanley, Controls on methane concentrations and fluxes in streams draining human-dominated landscapes. *Ecol. Appl.* 26, 1581–1591 (2016).
- 48. J. L. Wilmoth, *et al.*, The role of oxygen in stimulating methane production in wetlands. *Glob. Chang. Biol.* n/a (2021).
- 49. D. Bruhn, I. M. Møller, T. N. Mikkelsen, P. Ambus, Terrestrial plant methane production and emission. *Physiol. Plant.* 144, 201–209 (2012).
- 50. F. Keppler, J. T. G. Hamilton, M. Braß, T. Röckmann, Methane emissions from terrestrial plants under aerobic conditions. *Nature* 439, 187–191 (2006).
- 51. F. Keppler, *et al.*, Methoxyl groups of plant pectin as a precursor of atmospheric methane:
  evidence from deuterium labelling studies. *New Phytol.* 178, 808–814 (2008).
- 52. Z. Wang, F. Keppler, M. Greule, J. T. G. Hamilton, Non-microbial methane emissions
  from fresh leaves: effects of physical wounding and anoxia. *Atmos. Environ.* 45, 4915–

- **697 4921 (2011)**.
- W. Armstrong, "Aeration in higher plants" in *Advances in Botanical Research*, (Elsevier, 1980), pp. 225–332.
- T. D. Colmer, Long-distance transport of gases in plants: a perspective on internal aeration and radial oxygen loss from roots. *Plant. Cell Environ.* 26, 17–36 (2003).
- 702 55. P. E. Galand, H. Fritze, R. Conrad, K. Yrjälä, Pathways for methanogenesis and diversity
  703 of methanogenic archaea in three boreal peatland ecosystems. *Appl. Environ. Microbiol.*704 71, 2195–2198 (2005).
- 56. B. J. M. Robroek, *et al.*, Peatland vascular plant functional types affect methane dynamics
  by altering microbial community structure. *J. Ecol.* 103, 925–934 (2015).
- F.-J. W. A. van der Nat, J. J. Middelburg, Seasonal variation in methane oxidation by the
  rhizosphere of Phragmites australis and Scirpus lacustris. *Aquat. Bot.* 61, 95–110 (1998).
- 709 58. P. L. E. Bodelier, H. J. Laanbroek, Nitrogen as a regulatory factor of methane oxidation in soils and sediments. *FEMS Microbiol. Ecol.* 47, 265–277 (2004).
- A. J. Veraart, A. K. Steenbergh, A. Ho, S. Y. Kim, P. L. E. Bodelier, Beyond nitrogen: the importance of phosphorus for CH4 oxidation in soils and sediments. *Geoderma* 259, 337–346 (2015).
- K. Attermeyer, *et al.*, Invasive floating macrophytes reduce greenhouse gas emissions
  from a small tropical lake. *Sci. Rep.* 6, 1–10 (2016).
- 61. G. M. King, Regulation by light of methane emissions from a wetland. *Nature* 345, 513–
  515 (1990).
- A. L. dos S. Fonseca, C. C. Marinho, F. de A. Esteves, Floating Aquatic Macrophytes
  Decrease the Methane Concentration in the Water Column of a Tropical Coastal Lagoon:
  Implications for Methane Oxidation and Emission. *Brazilian Arch. Biol. Technol.* 60
  (2017).
- 63. E. J. W. Visser, T. D. Colmer, C. Blom, L. Voesenek, Changes in growth, porosity, and
  radial oxygen loss from adventitious roots of selected mono-and dicotyledonous wetland
  species with contrasting types of aerenchyma. *Plant. Cell Environ.* 23, 1237–1245 (2000).
- M. A. Heilman, R. G. Carlton, Methane oxidation associated with submersed vascular
  macrophytes and its impact on plant diffusive methane flux. *Biogeochemistry* 52, 207–224
  (2001).
- B. K. Sorrell, M. T. Downes, C. L. Stanger, Methanotrophic bacteria and their activity on submerged aquatic macrophytes. *Aquat. Bot.* 72, 107–119 (2002).
- M. P. Ávila, *et al.*, The water hyacinth microbiome: link between carbon turnover and nutrient cycling. *Microb. Ecol.* 78, 575–588 (2019).
- A. A. Raghoebarsing, *et al.*, Methanotrophic symbionts provide carbon for photosynthesis
  in peat bogs. *Nature* 436, 1153–1156 (2005).
- 734 68. Z. Stępniewska, *et al.*, Methane oxidation by endophytic bacteria inhabiting Sphagnum sp.
  735 and some vascular plants. *Wetlands* 38, 411–422 (2018).

- 739 70. J. P. Chanton, J. W. H. Dacey, *Effects of vegetation on methane flux, reservoirs, and carbon isotopic composition*, H. Sharkey, T., Holland, E., Mooney, Ed., Trace Gas (Academic Press, 1991).
- 742 71. H. Brix, Macrophyte-mediated oxygen transfer in wetlands: transport mechanisms and rates. *Constr. Wetl. water Qual. Improv.*, 391–398 (1993).
- 744 72. J. Kim, S. B. Verma, D. P. Billesbach, Seasonal variation in methane emission from a
  745 temperate Phragmites-dominated marsh: Effect of growth stage and plant-mediated
  746 transport. *Glob. Chang. Biol.* 5, 433–440 (1999).
- 747 73. G. J. Whiting, J. P. Chanton, Control of the diurnal pattern of methane emission from
  748 emergent aquatic macrophytes by gas transport mechanisms. *Aquat. Bot.* 54, 237–253
  749 (1996).
- 750 74. J. B. Yavitt, A. K. Knapp, Methane emission to the atmosphere through emergent cattail
  (Typha latifolia L.) plants. *Tellus B Chem. Phys. Meteorol.* 47, 521–534 (1995).
- 752 75. J. P. Schimel, Plant transport and methane production as controls on methane flux from arctic wet meadow tundra. *Biogeochemistry* 28, 183–200 (1995).
- 754 76. M. van den Berg, *et al.*, Contribution of plant-induced pressurized flow to CH4 emission
  755 from a Phragmites fen. *Sci. Rep.* 10, 1–10 (2020).
- 756 77. M. Saunois, *et al.*, The global methane budget 2000–2012. *Earth Syst. Sci. Data* 8, 697–
  757 751 (2016).
- 758 78. A. Joabsson, T. R. Christensen, Methane emissions from wetlands and their relationship
  759 with vascular plants: an Arctic example. *Glob. Chang. Biol.* 7, 919–932 (2001).
- 760 79. M. G. Öquist, B. H. Svensson, Vascular plants as regulators of methane emissions from a subarctic mire ecosystem. *J. Geophys. Res. Atmos.* 107 (2002).
- 80. S. J. Davidson, *et al.*, Vegetation type dominates the spatial variability in CH4 emissions across multiple Arctic tundra landscapes. *Ecosystems* 19, 1116–1132 (2016).
- F.-J. Van Der Nat, J. J. Middelburg, Methane emission from tidal freshwater marshes. *Biogeochemistry* 49, 103–121 (2000).
- H. Chu, *et al.*, Net ecosystem methane and carbon dioxide exchanges in a Lake Erie
  coastal marsh and a nearby cropland. *J. Geophys. Res. Biogeosciences* 119, 722–740
  (2014).
- J. Kim, S. B. Verma, D. P. Billesbach, R. J. Clement, Diel variation in methane emission
  from a midlatitude prairie wetland: significance of convective throughflow in Phragmites
  australis. *J. Geophys. Res. Atmos.* 103, 28029–28039 (1998).
- A. Sugimoto, N. Fujita, Characteristics of methane emission from different vegetations on a wetland. *Tellus B Chem. Phys. Meteorol.* 49, 382–392 (1997).
- 85. S. Flury, D. F. McGinnis, M. O. Gessner, Methane emissions from a freshwater marsh in
  response to experimentally simulated global warming and nitrogen enrichment. *J.*

776 Geophys. Res. Biogeosciences 115 (2010). T. R. Barber, R. A. Burke, W. M. Sackett, Diffusive flux of methane from warm wetlands. 777 86. 778 Global Biogeochem. Cycles 2, 411–425 (1988). P. Frenzel, J. Rudolph, Methane emission from a wetland plant: the role of CH 4 oxidation 779 87. in Eriophorum. Plant Soil 202, 27-32 (1998). 780 88. J. P. Chanton, G. J. Whiting, J. D. Happell, G. Gerard, Contrasting rates and diurnal 781 patterns of methane emission from emergent aquatic macrophytes. Aquat. Bot. 46, 111-782 128 (1993). 783 J. P. Chanton, G. J. Whiting, W. J. Showers, P. M. Crill, Methane flux from Peltandra 784 89. 785 virginica: Stable isotope tracing and chamber effects. Global Biogeochem. Cycles 6, 15-31 (1992). 786 90. P. M. van Bodegom, T. Groot, B. den Hout, P. A. Leffelaar, J. Goudriaan, Diffusive gas 787 transport through flooded rice systems. J. Geophys. Res. Atmos. 106, 20861–20873 (2001). 788 K. Butterbach-Bahl, H. Papen, H. Rennenberg, Impact of gas transport through rice 789 91. cultivars on methane emission from rice paddy fields. Plant. Cell Environ. 20, 1175–1183 790 (1997). 791 92. 792 B. Wang, H. U. Neue, H. P. Samonte, Role of rice in mediating methane emission. Plant Soil 189, 107–115 (1997). 793 794 93. S. Bansal, O. F. Johnson, J. Meier, X. Zhu, Vegetation Affects Timing and Location of 795 Wetland Methane Emissions. J. Geophys. Res. Biogeosciences 125 (2020). E. S. Oliveira Junior, et al., Water Hyacinth's Effect on Greenhouse Gas Fluxes: A Field 796 94. Study in a Wide Variety of Tropical Water Bodies. *Ecosystems* (2020) 797 798 https:/doi.org/10.1007/s10021-020-00564-x. 95. J. P. Silva, T. R. Canchala, H. J. Lubberding, E. J. Peña, H. J. Gijzen, Greenhouse Gas 799 Emissions from a Tropical Eutrophic Freshwater Wetland. World Acad. Sci. Eng. Technol. 800 Int. J. Environ. Chem. Ecol. Geol. Geophys. Eng. 10, 541–547 (2016). 801 802 96. C. Grasset, et al., An empirical model to predict methane production in inland water sediment from particular organic matter supply and reactivity. *Limnol. Oceanogr.* 66, 803 3643-3655 (2021). 804 97. 805 S. Juutinen, et al., Major implication of the littoral zone for methane release from boreal lakes. Global Biogeochem. Cycles 17 (2003). 806 98. J. L. Bubier, T. R. Moore, L. Bellisario, N. T. Comer, P. M. Crill, Ecological controls on 807 methane emissions from a northern peatland complex in the zone of discontinuous 808 permafrost, Manitoba, Canada. Global Biogeochem. Cycles 9, 455–470 (1995). 809 810 99. A. T. C. Dias, B. Hoorens, R. S. P. Van Logtestijn, J. E. Vermaat, R. Aerts, Plant species composition can be used as a proxy to predict methane emissions in peatland ecosystems 811 after land-use changes. Ecosystems 13, 526-538 (2010). 812 100. B. J. M. Robroek, V. E. J. Jassey, B. Beltman, M. M. Hefting, Diverse fen plant 813 communities enhance carbon-related multifunctionality, but do not mitigate negative 814 effects of drought. R. Soc. open Sci. 4, 170449 (2017). 815

- 816 101. B. Zhang, *et al.*, Methane emissions from global wetlands: An assessment of the
  817 uncertainty associated with various wetland extent data sets. *Atmos. Environ.* 165, 310–
  818 321 (2017).
- P. Ciais, *et al.*, "Carbon and other biogeochemical cycles" in *Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*, (Cambridge University Press, 2014), pp.
  465–570.
- J. R. Melton, *et al.*, Present state of global wetland extent and wetland methane modelling:
   conclusions from a model intercomparison project (WETCHIMP). *Biogeosciences* 10,
   753–788 (2013).
- 104. C. Berrittella, J. Van Huissteden, Uncertainties in modelling CH4 emissions from northern
  wetlands in glacial climates: The role of vegetation parameters. *Clim. Past* 7, 1075–1087
  (2011).
- W. J. Riley, *et al.*, Barriers to predicting changes in global terrestrial methane fluxes:
  analyses using CLM4Me, a methane biogeochemistry model integrated in CESM. *Biogeosciences* 8, 1925–1953 (2011).
- M. Cao, S. Marshall, K. Gregson, Global carbon exchange and methane emissions from natural wetlands: Application of a process-based model. *J. Geophys. Res. Atmos.* 101, 14399–14414 (1996).
- 835 107. E. L. Hodson, B. Poulter, N. E. Zimmermann, C. Prigent, J. O. Kaplan, The El Niño–
  836 Southern Oscillation and wetland methane interannual variability. *Geophys. Res. Lett.* 38
  837 (2011).
- 838 108. V. K. Arora, J. R. Melton, D. Plummer, An assessment of natural methane fluxes
  839 simulated by the CLASS-CTEM model. *Biogeosciences* 15, 4683–4709 (2018).
- R. J. Parker, *et al.*, Evaluating year-to-year anomalies in tropical wetland methane
  emissions using satellite CH4 observations. *Remote Sens. Environ.* 211, 261–275 (2018).
- T. Kleinen, U. Mikolajewicz, V. Brovkin, Terrestrial methane emissions from the Last
  Glacial Maximum to the preindustrial period. *Clim. Past* 16, 575–595 (2020).
- A. Ito, M. Inatomi, Use of a process-based model for assessing the methane budgets of
  global terrestrial ecosystems and evaluation of uncertainty. *Biogeosciences* 9, 759–773
  (2012).
- 847 112. E. H. Stanley, *et al.*, The ecology of methane in streams and rivers: Patterns, controls, and
  848 global significance. *Ecol. Monogr.* 86, 146–171 (2016).
- L. F. Sanches, B. Guenet, C. C. Marinho, N. Barros, F. de Assis Esteves, Global regulation of methane emission from natural lakes. *Sci. Rep.* 9, 1–10 (2019).
- L. C. Jeffrey, *et al.*, Wetland methane emissions dominated by plant-mediated fluxes:
  Contrasting emissions pathways and seasons within a shallow freshwater subtropical
  wetland. *Limnol. Oceanogr.* 64, 1895–1912 (2019).
- J. A. Rosentreter, D. T. Maher, D. V. Erler, R. H. Murray, B. D. Eyre, Methane emissions
  partially offset "blue carbon" burial in mangroves. *Sci. Adv.* 4 (2018).

856 857	116.	D. Bastviken, L. J. Tranvik, J. A. Downing, P. M. Crill, A. Enrich-Prast, Freshwater Methane Emissions Offset the Continental Carbon Sink. <i>Science (80 ).</i> <b>331</b> , 50 (2011).
858 859	117.	T. J. H. M. van Bergen, <i>et al.</i> , Seasonal and diel variation in greenhouse gas emissions from an urban pond and its major drivers. <i>Limnol. Oceanogr.</i> <b>64</b> , 2129–2139 (2019).
860 861	118.	E. T. H. M. Peeters, <i>et al.</i> , Changing weather conditions and floating plants in temperate drainage ditches. <i>J. Appl. Ecol.</i> <b>50</b> , 585–593 (2013).
862 863 864	119.	J. J. C. Netten, J. van Zuidam, S. Kosten, E. T. H. M. Peeters, Differential response to climatic variation of free-floating and submerged macrophytes in ditches. <i>Freshw. Biol.</i> <b>56</b> , 1761–1768 (2011).
865 866	120.	C. Tong, <i>et al.</i> , Invasive alien plants increase CH 4 emissions from a subtropical tidal estuarine wetland. <i>Biogeochemistry</i> <b>111</b> , 677–693 (2012).
867 868	121.	T. S. F. Silva, M. P. F. Costa, J. M. Melack, E. M. L. M. Novo, Remote sensing of aquatic vegetation: Theory and applications. <i>Environ. Monit. Assess.</i> <b>140</b> , 131–145 (2008).

## 870 **Supplementary material**

871

872	Table S1.	Ind	ividual	ger	nera	ι of	inv	vest	igated	emergent	plants	(total n	= 337)	), 1	loati	ng	plan	its (	(tota	al
	• •						/		<b>a</b> >				0			× •				

873 n = 24), and submerged plants (total n = 9); percentage of the number of entries (n) for individual

874	genera in relation to the tota	l number of entries was calculated.
-----	--------------------------------	-------------------------------------

Emergent plants					
Plant genus	n	%	Plant genus	n	%
mixed	95	28.2	Scirpus	2	0.6
Carex	56	16.6	Sparganium	2	0.6
Eriophorum	33	9.8	Suaeda	2	0.6
Phragmites	31	9.2	Alunus	1	0.3
Typha	15	4.5	Anthoxanthum	1	0.3
Sphagnum	13	3.9	Arctagrostis+Carex+Dupontia+EriophorumandEquise tum	ຳ 1	0.3
Cladium	8	2.4	Betula	1	0.3
Juncus	8	2.4	Capparis	1	0.3
Typha+Schoenoplectus	7	2.1	Carex+Scirpus	1	0.3
Menyanthes	5	1.5	Cassiope	1	0.3
Schoenoplectus	4	1.2	Chamaedaphne	1	0.3
Eleocharis	3	0.9	Cyperus	1	0.3
Equisetum	3	0.9	Eleocharis+Juncus	1	0.3
Glyceria	3	0.9	Eriophorum+Carex	1	0.3
Phalaris	3	0.9	Eriophorum+Sphagnum	1	0.3
Potentilla	3	0.9	Eriophorumvaginatum	1	0.3
Spartina	3	0.9	Juncus+moss	1	0.3
Arctophila	2	0.6	Kobresia	1	0.3
Carex+moss	2	0.6	Maianthemum+Ledum	1	0.3
Deyeuxia	2	0.6	Mimulus	1	0.3
Lemna	2	0.6	Molinia	1	0.3
Peltandra	2	0.6	Onoclea+Osmunda	1	0.3
Sagittaria	2	0.6	Raphia	1	0.3
Scheuchzeria	2	0.6	Salix	1	0.3

Emergent plants (continuation)		
Plant genus	n	%
Salix+Carex	1	0.3
Tamarix	1	0.3
Verbena	1	0.3
Floating plants		
Plant genus	n	%
Nuphar	8	33.3
Eichhornia	6	25.0
Nelumbo	3	12.5
Nymphaea	3	12.5
mixed	1	4.2
Nuphar+Potamogeton+Mentha+Hippuris+Berula	1	4.2
Potamogeton	1	4.2
Тгара	1	4.2
Submerged plants		
Plant genus	n	%
Myriophyllum	3	33.3
Ranunculus	2	22.2
Cabomba	1	11.1
Ceratophyllum	1	11.1
Hottonia	1	11.1
Lagarosiphon	1	11.1

Non-peer reviewed preprint submitted to EarthArXiv



Figure S1. CH<sub>4</sub> fluxes of vegetated areas from individual genera of investigated floating plants
(upper panel) and submerged plants (lower panel). Boxplots represent median (black line), first

and third quartiles (hinges), range (whiskers), and outliers (black dots).