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pyisotopomer: A Python package for obtaining nitrous oxide isotopocules from isotope ratio mass spectrometry

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Abstract

RATIONALE Obtaining nitrous oxide isotopocule measurements with isotope ratio mass spectrometry (IRMS) requires measuring the m/z ratios of the nitrous oxide (N_2O) molecule as well as those of the NO^+ fragment ion. This measurement depends on correcting for a phenomenon referred to as “scrambling” in the ion source, whereby the NO^+ fragment ion contains the outer N atom from the N_2O molecule. While descriptions of the scrambling correction exist in the literature, there has yet to be published a unified software package and method for performing this correction.

METHODS We developed a user-friendly Python package (pyisotopomer), with a MATLAB alternative, to determine two coefficients that describe scrambling in the ion source of a given IRMS, and then to use this calibration to obtain N_2O isotopocule measurements.

RESULTS We assess the sensitivity of pyisotopomer to its input parameters and discuss the relevant assumptions. We show that the scrambling behavior of an IRMS can vary with time, necessitating regular calibrations. We show that to obtain a relative uncertainty in site preference of $<1\%$, the relative uncertainty in each scrambling coefficient should be $<0.2\%$. Finally, we present an intercalibration between two IRMS laboratories, using pyisotopomer to calculate scrambling and obtain N_2O isotopocule data.

CONCLUSIONS Given these considerations, we discuss how to use this software package to obtain high-quality N_2O isotopocule data from IRMS systems, including the use of appropriate reference materials and frequency of calibration.

48 1. Introduction

49 Nitrous oxide (N₂O) is a potent greenhouse gas, with a greenhouse gas potential
50 approximately 265 times that of carbon dioxide, over 100 years and on a per-molecule basis.^{1,2}
51 N₂O is also likely to be the most important ozone depletion agent of the 21st century, due to
52 production of NO radicals in the stratosphere that interact destructively with ozone.³⁻⁶
53 Historically, the bulk stable isotopes of nitrogen and oxygen in N₂O have been used to quantify
54 its microbial cycling in soils,^{7,8} its destruction by photolysis and cycling in the atmosphere,^{9,10}
55 and microbial cycling in the ocean.¹¹⁻¹⁴ This approach often does not provide a unique solution,
56 because the bulk nitrogen and oxygen isotope ratios of N₂O depend on the isotopic composition
57 of the substrate, as well as the isotope effects of production and consumption processes.¹⁴
58 Furthermore, in the context of microbial N₂O cycling in soils and the ocean, bacterial
59 nitrification and denitrification produce N₂O with similar bulk δ¹⁵N values, which prevents
60 partitioning between these processes on the basis of bulk δ¹⁵N alone.^{15,16} This leaves room for a
61 more nuanced measurement, which may distinguish between N₂O production processes in the
62 soils, atmosphere, and ocean.

63 Given the asymmetry of the N₂O molecule, its two nitrogen atoms exist in unique
64 chemical environments, making the molecule particularly rich in isotopic information. The
65 individual isotopic compositions of each nitrogen atom were first measured by Friedman and
66 Bigeleisen, who quantified the yields of isotopomers ¹⁴N¹⁵N¹⁶O and ¹⁵N¹⁴N¹⁶O from enriched
67 ammonium nitrate by measuring the ion beam signal of the fragment ion NO⁺ in an isotope ratio
68 mass spectrometer.¹⁷ The natural abundance isotopomers of N₂O were quantified 50 years later
69 with a similar technique, by measuring the molecular N₂O mass to charge (m/z) ratios 44, 45,
70 and 46 as well as the m/z ratios 30 and 31 of the NO⁺ fragment ion.^{18,19} Toyoda and Yoshida
71 (1999) defined the site-specific isotope ratios of the central (α) nitrogen atom and outer (β)
72 nitrogen atom as follows:

$$73 \quad {}^{15}R^{\alpha} = \frac{{}^{14}N{}^{15}NO}{{}^{14}N{}^{14}NO} \quad (1)$$

$$74 \quad {}^{15}R^{\beta} = \frac{{}^{15}N{}^{14}NO}{{}^{14}N{}^{14}NO} \quad (2)$$

75 The N₂O isotopomer measurement was initially performed with two sequential
76 measurements of the same sample on an isotope ratio mass spectrometer, one for m/z ratios 44,
77 45, and 46, and one for m/z ratios 30 and 31.¹⁸ Subsequent advances led to the measurement of
78 all five m/z ratios simultaneously with the correct configuration of cups.²⁰

79 The slight difference in zero-point energies between the isotopomers of N₂O result in
80 different isotopic fractionations during photolysis in the stratosphere,²¹ making the isotopomers
81 of N₂O a powerful tool for understanding its atmospheric cycling.²²⁻²⁶ Likewise, N₂O site
82 preference, defined as δ¹⁵N^α - δ¹⁵N^β, was shown in microbial culture experiments to be largely a
83 function of reaction mechanism, independent of source composition.²⁷⁻³² This allowed for the
84 differentiation between N₂O deriving from bacterial nitrification and denitrification, although
85 some debate exists about whether the site preference of N₂O produced by denitrifying bacteria is
86 closer to 0‰ or 25‰,^{31,33} the latter possibility being largely ignored in subsequent literature.
87 During N₂O consumption, δ¹⁵N^α and δ¹⁸O_{N₂O} were shown in microbial culture³⁴ and soil
88 mesocosm³⁵ experiments to exhibit a characteristic relationship, allowing subsequent studies to

89 use this expected relationship to distinguish between oxidative and reductive regimes of N₂O
90 cycling.^{36,37}

91 The site-specific isotopomers of N₂O provide a far more nuanced constraint on the
92 biogeochemical cycling of N₂O than its bulk composition alone. N₂O isotopomers have been
93 used extensively to quantify its biogeochemical cycling in soils^{35,38–40} the atmosphere,^{22,29} and
94 the ocean.^{27,36,37,41–48} Nonetheless, there is as yet no unified method for calibrating isotope ratio
95 mass spectrometry systems for the N₂O isotopomer (and isotopocule) measurement. The need for
96 such a calibration is largely due to a phenomenon called “scrambling,” whereby the NO⁺
97 fragment ion contains the outer, β nitrogen atom, rather than the expected α nitrogen. A number
98 of approaches have been taken to calibrate an IRMS system for this effect: the use of a single
99 “rearrangement factor” to describe scrambling,^{18,49} the use of nine coefficients to describe the
100 different fragmentation behaviors of the different isotopocules of N₂O,⁵⁰ and finally the use of
101 two coefficients to describe scrambling in the ion source.³² While descriptions exist of each of
102 these approaches, and international intercalibration efforts have been made,^{51,52} there has yet to
103 be published a package of code for implementing one of the above isotopomer calibrations.

104 We developed a Python software package that implements the two-coefficient approach
105 described by Frame and Casciotti (2010) to calibrate an IRMS system for scrambling and use
106 that calibration to obtain high-quality N₂O isotopocule data. To quantify the performance of the
107 software, we tested its sensitivity to solver parameters and the assumptions inherent to the
108 scrambling equations. Next, we quantified the uncertainty associated with different pairings of
109 reference materials used to calculate scrambling and compared the outputs from the MATLAB
110 and Python versions of the software. To quantify the variability of the fragmentation behavior of
111 an instrument over time, we examined the scrambling coefficients for one isotope ratio mass
112 spectrometer over the course of four years of measurements. To assess the effect of uncertainty
113 in each scrambling coefficient on calculated isotopocule values, we used a Monte Carlo
114 simulation approach to quantify the effect of uncertainty in the scrambling coefficients on the
115 final, output N₂O isotopocule values. Finally, we performed an intercalibration using this
116 software across two labs at Stanford and the University of Basel.

117

118 2. Theory

119 Toyoda and Yoshida (1999) introduced the isotopomer notation in equations 1 and 2,
120 designating the inner and outer nitrogen atoms as the α and β nitrogen atoms, respectively. The
121 expressions for molecular m/z ratios 45/44 (⁴⁵R) and 46/44 (⁴⁶R), used for two different
122 calibration approaches,^{18,49} follow from this notation:

123

$$^{45}R = ^{15}R^{\alpha} + ^{15}R^{\beta} + ^{17}R \quad (3)$$

$$^{46}R = (^{15}R^{\alpha} + ^{15}R^{\beta})^{17}R + ^{18}R + (^{15}R^{\alpha})(^{15}R^{\beta}) \quad (4)$$

124 where ¹⁵R^α denotes the isotopocule ratio ¹⁴N¹⁵N¹⁶O/¹⁴N¹⁴N¹⁶O, ¹⁵R^β denotes the isotopocule ratio
125 ¹⁵N¹⁴N¹⁶O/¹⁴N¹⁴N¹⁶O, ¹⁷R denotes the isotopocule ratio ¹⁴N¹⁴N¹⁷O/¹⁴N¹⁴N¹⁶O, and ¹⁸R denotes
126 the isotopocule ratio ¹⁴N¹⁴N¹⁸O/¹⁴N¹⁴N¹⁶O.

127 While the oxygen triple isotopes of N₂O (Δ¹⁷O) provide additional information about the
128 sources and sinks of N₂O,^{53,54} they will not be discussed further here. Thus, we will assume that
129 ¹⁷R covaries with ¹⁸R according to the oxygen isotope content of Vienna Standard Mean Ocean
130 Water (VSMOW)^{55,56} and a mass-dependent relationship between ¹⁷R and ¹⁸R:⁵³

131

$$^{17}R/0.0003799 = (^{18}R/0.0020052)^{0.516} \quad (5)$$

132 The simplest formulation for the NO⁺ fragment ion ratio 31/30 (³¹R) comes from Toyoda
133 and Yoshida (1999):

$$^{31}R = ^{15}R^\alpha + ^{17}R \quad (6)$$

134 This equation would represent the ³¹R measured by IRMS if no scrambling occurred.
135 Indeed, given a known ¹⁵R^α and ¹⁷R for a reference gas, this equation defines the theoretical ³¹R
136 of that reference gas. Toyoda and Yoshida (1999) account for scrambling by defining the
137 rearrangement factor γ (which was later redefined as γ) as “the fraction of NO⁺ bearing the β
138 nitrogen of the initial N₂O to the total NO⁺ formed,” to yield:
139

$$^{31}R = (1 - \gamma)^{15}R^\alpha + \gamma^{15}R^\beta + ^{17}R \quad (7)$$

140 Kaiser et al. (2004) introduce a more complete representation of ³¹R, adding terms for
141 ¹⁵N¹⁵N¹⁶O, ¹⁴N¹⁵N¹⁷O, and ¹⁵N¹⁴N¹⁷O to m/z=31, and terms for ¹⁵N¹⁴N¹⁶O and ¹⁴N¹⁵N¹⁶O to
142 m/z=30:
143

$$^{31}R = \frac{(1 - \gamma)^{15}R^\alpha + \gamma^{15}R^\beta + (^{15}R^\alpha)(^{15}R^\beta) + ^{17}R(1 + \gamma^{15}R^\alpha + (1 - \gamma)^{15}R^\beta)}{1 + \gamma^{15}R^\alpha + (1 - \gamma)^{15}R^\beta} \quad (8)$$

144 Note that Kaiser et al. (2004), in their Eqn. (3), use the term “s” in place of γ, ¹⁵R_{1,ref} in
145 place of ¹⁵R^β, and ¹⁵R_{2,ref} in place of ¹⁵R^α. To account for different fragmentation rates from
146 different isotopic species of N₂O, Westley et al. (2007) split the fragmentation factor γ into nine
147 coefficients:
148

$$^{31}R = \frac{a_{31}^{15}R^\alpha + b_{31}^{15}R^\beta + c_{31}(^{15}R^\alpha)(^{15}R^\beta) + ^{17}R(d_{31} + e_{31}^{15}R^\alpha + f_{31}^{15}R^\beta)}{1 + a_{30}^{15}R^\alpha + b_{30}^{15}R^\beta + c_{30}(^{15}R^\alpha)(^{15}R^\beta)} \quad (9)$$

149 While this approach considers the possibility of different rates of fragmentation for every
150 isotopic species of N₂O (i.e., isotopic fractionation) measurable with this IRMS configuration, it
151 also requires solving for six to nine coefficients, depending on whether the m/z=30 coefficients
152 are considered separate from the m/z=31 coefficients. Frame and Casciotti (2010) simplify this
153 equation by reducing the number of fragmentation factors to two coefficients, γ and κ, which
154 represent the yield of ³⁰NO⁺ from ¹⁴N¹⁵NO (for both ¹⁷O and ¹⁶O) and the yield of ³¹NO⁺ from
155 ¹⁵N¹⁴NO, respectively. This yields the equation:
156

$$^{31}R = \frac{(1 - \gamma)^{15}R^\alpha + \kappa^{15}R^\beta + (^{15}R^\alpha)(^{15}R^\beta) + ^{17}R(1 + \gamma^{15}R^\alpha + (1 - \kappa)^{15}R^\beta)}{1 + \gamma^{15}R^\alpha + (1 - \kappa)^{15}R^\beta} \quad (10)$$

157 The important pieces of information contained within the two scrambling factors are the
158 unequal rates of fragmentation for the isotopomers ¹⁴N¹⁵NO and ¹⁵N¹⁴NO, which the equation
159 from Kaiser et al. (2004) does not account for. It is assumed that the isotopic substitution of ¹⁷O
160 in the oxygen atom has a negligible effect on the scrambling coefficients, or, in terms of the

161 equation from Westley et al. (2007), that $e_{31}=(1-a_{31})$ and $f_{31}=(1-b_{31})$. It is also assumed that the
 162 coefficient c_{31} from Westley et al. (2007) is equal to 1, or that the yield of $^{31}\text{NO}^+$ from $^{15}\text{N}^{15}\text{N}^{16}\text{O}$
 163 is equal to the yield of $^{30}\text{NO}^+$ from $^{14}\text{N}^{14}\text{N}^{16}\text{O}$. Given that naturally occurring N_2O contains very
 164 little of the $^{15}\text{N}^{15}\text{N}^{16}\text{O}$ isotopocule, a few per mil difference in the fractionation during
 165 fragmentation of $^{15}\text{N}^{15}\text{N}^{16}\text{O}$ and $^{14}\text{N}^{14}\text{N}^{16}\text{O}$ would not drastically alter the measured ^{31}R .⁵⁷
 166 Finally, the coefficient d_{31} is assumed to be equal to 1, or that the yield of $^{31}\text{NO}^+$ from $^{14}\text{N}^{14}\text{N}^{17}\text{O}$
 167 is equal to the yield of $^{30}\text{NO}^+$ from $^{14}\text{N}^{14}\text{N}^{16}\text{O}$, again, an assumption yielding little potential error
 168 in ^{31}R , given the low natural abundance of ^{17}O in N_2O .⁵³

169 Measurement of two reference materials against a common third reference gas, each with
 170 known $^{15}\text{R}^\alpha$ and $^{15}\text{R}^\beta$ values, produces two equations to solve for two unknowns, γ and κ ,

$$171 \frac{{}^{31}\text{R}_{\text{sample1}}}{{}^{31}\text{R}_{\text{reference}}} - \frac{[r\text{R}31/30_{\text{sample1}}]}{[r\text{R}31/30_{\text{reference}}]} = 0 \quad (11)$$

$$172 \frac{{}^{31}\text{R}_{\text{sample2}}}{{}^{31}\text{R}_{\text{reference}}} - \frac{[r\text{R}31/30_{\text{sample2}}]}{[r\text{R}31/30_{\text{reference}}]} = 0 \quad (12)$$

173 Where ${}^{31}\text{R}_{\text{sample1}}$ is the theoretical ${}^{31}\text{R}$ of the first reference material, calculated from
 174 equation 10, ${}^{31}\text{R}_{\text{reference}}$ is the theoretical ${}^{31}\text{R}$ of the common reference gas against which sample
 175 peaks are normalized, and ${}^{31}\text{R}_{\text{sample2}}$ is the theoretical ${}^{31}\text{R}$ of the second reference material. These
 176 “ratios of ratios” should be equivalent to the measured ${}^{31}\text{R}$ of each reference material normalized
 177 to the common reference injection, leaving a set of cost functions to be minimized. To aid in
 178 finding a solution, equations 11 and 12 may be simplified by assuming that the ${}^{31}\text{R}$ of the direct
 179 reference injection is constant (“C”, below), and thus that the flat-topped reference peak has a
 180 defined scrambling behavior that could differ from that of a sample peak. Calculating ‘C’ for the
 181 common reference injection from equation 10, using assumed γ and κ values, produces the
 182 following two equations:

$$183 \frac{{}^{31}\text{R}_{\text{sample1}}}{C} - \frac{[r\text{R}31/30_{\text{sample1}}]}{[r\text{R}31/30_{\text{reference}}]} = 0 \quad (13)$$

$$184 \frac{{}^{31}\text{R}_{\text{sample2}}}{C} - \frac{[r\text{R}31/30_{\text{sample2}}]}{[r\text{R}31/30_{\text{reference}}]} = 0 \quad (14)$$

185 Given a pair of reference materials with known $^{15}\text{R}^\alpha$ and $^{15}\text{R}^\beta$, pyisotopomer⁵⁸ solves
 186 equations 13 and 14 for scrambling coefficients γ and κ . Pyisotopomer uses the “least_squares”
 187 function from the scipy optimization library to vary γ and κ until the calculated values of
 188 ${}^{31}\text{R}_{\text{sample}}/{}^{31}\text{R}_{\text{reference}}$ are as close to $[r\text{R}31\text{NO}/30\text{NO}_{\text{sample}}/r\text{R}31\text{NO}/30\text{NO}_{\text{reference}}]$, as described in
 189 Frame and Casciotti (2010). In MATLAB, the solver lsqnonlin is used to perform the
 190 optimization.

191 After the scrambling coefficients are calculated from measured reference materials,
 192 pyisotopomer can be used to calculate the $^{15}\text{R}^\alpha$ and $^{15}\text{R}^\beta$ of unknown samples using equations 3,
 193 4, and 10. These equations are solved for the $^{15}\text{R}^\alpha$ and $^{15}\text{R}^\beta$, from measured isotope ratios ${}^{31}\text{R}$,
 194 ^{45}R , and ^{46}R of the unknown. As in the scrambling function, pyisotopomer uses least_squares
 195 from scipy.optimize to solve this set of equations, producing the isotope ratios $^{15}\text{R}^\alpha$, $^{15}\text{R}^\beta$, ^{17}R ,

197 and ^{18}R , and from these, the delta values $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, site preference, $\delta^{15}\text{N}^{\text{bulk}}$, $\delta^{17}\text{O}_{\text{N}_2\text{O}}$, and
198 $\delta^{18}\text{O}_{\text{N}_2\text{O}}$ are calculated relative to primary reference scales (^{15}R from atmospheric N_2 and ^{17}R and
199 ^{18}R from VSMOW).

200

201 3. Experimental

202 3.1 Preparation and analysis of N_2O reference materials

203 To validate the usage of pyisotopomer to calculate scrambling coefficients, and from
204 these coefficients obtain precise N_2O isotopocule data, a series of N_2O reference materials (Table
205 1) were prepared and analyzed in sample format at Stanford University (“Lab 1”) and the
206 University of Basel (“Lab 2”). Reference materials were prepared by filling 160-mL glass serum
207 bottles (Wheaton) with de-ionized water and removing a 4-mL headspace (Lab 1) or 10-20-mL
208 headspace (Lab 2), then were capped with a gray butyl septum (National Scientific) and sealed
209 with an aluminum crimp seal. These bottles were purged with helium for 90 minutes to remove
210 all background N_2O . The purged bottles were then injected with reference gas in a range of nmol
211 amounts from 2-20 nmols N_2O (S2, EMPA1, EMPA2, EMPA3 reference gases) or with 10
212 nmols N_2O (B6 reference gas) using a gas-tight syringe. Reference materials prepared in Lab 1
213 were preserved with 100 μL saturated mercuric chloride solution; those prepared in Lab 2
214 contained no added preservative. For Lab 1, atmosphere-equilibrated seawater was prepared by
215 filtering surface seawater (collected in Half Moon Bay, CA) through a 0.22 mm Sterivex filter,
216 allowing it to undergo static equilibration with outdoor air for three days, then re-filtering into
217 160-mL serum bottles, removing a ~ 1 -mL headspace, and preserving with 100 μL saturated
218 mercuric chloride solution. For Lab 2, atmosphere-equilibrated reference materials were
219 prepared by purging either de-ionized water or a sodium chloride solution with helium, allowing
220 it to undergo static equilibration with outdoor air for three days, filling into 160-mL serum
221 bottles, and removing a 10-mL headspace. Reference materials were run in the same format as
222 samples to account for any potential fractionation associated with the extraction and purification
223 of N_2O associated with the purge-and-trap system. The magnitude of such fractionation was
224 quantified for Lab 1 by running the pure N_2O reference tank against itself, and yielded offsets of
225 $0.22 \pm 0.52\text{‰}$ for $\delta^{15}\text{N}^{\text{bulk}}$ and $0.16 \pm 0.62\text{‰}$ for $\delta^{18}\text{O}_{\text{N}_2\text{O}}$.

226 The isotopocule values for each reference gas were calibrated independently by J. Mohn
227 (EMPA; mini-QCLAS aerodyne) or S. Toyoda (Tokyo Tech; IRMS), except for one internal
228 standard used by Lab 1 (Table 1). The reported ^{31}R , ^{45}R , and ^{46}R for the pure N_2O reference
229 tanks represent the theoretical values of these ratios, assuming some amount of scrambling for
230 the reference gas: $\gamma=0.17$ and $\kappa=0.08$ (Stanford University/Lab 1), and $\gamma=0.156$ and $\kappa=0.155$
231 (University of Basel/Lab 2). The values for $\delta^{17}\text{O}_{\text{N}_2\text{O}}$ for each gas were calculated assuming a
232 mass-dependent relationship between ^{17}R and ^{18}R .⁵³

233 These reference gases and samples were measured on Thermo Finnigan DELTA V^{PLUS}
234 isotope ratio mass spectrometers (IRMS; Thermo Fisher Scientific, Waltham, MA) in Lab 1 and
235 Lab 2. Each IRMS had Faraday cups configured to simultaneously measure m/z 30, 31, 44, 45,
236 and 46. Reference materials and samples were analyzed on a custom purge-and-trap system
237 coupled to each IRMS, which was run in continuous flow mode⁵⁹ (Table 1). The two systems
238 had slight differences in the purge-and-trap system: in Lab 1, liquid from each sample bottle was
239 transferred to a sparging column to extract the dissolved gases; in Lab 2, each sample was
240 extracted by purging directly from the bottle. The effects of these differences are discussed
241 further in Results and Discussion.

242

243 3.2 Data corrections

244 To calibrate each IRMS for scrambling, reference materials prepared as above were run
245 side-by-side on the same day. This was repeated over multiple days. From these runs, a
246 scrambling calibration was obtained as follows: first, each sample peak was compared against a
247 direct injection of pure N₂O from a reference tank (Table 1).⁶⁰ Next, sample peaks were
248 normalized to a peak area of 20 Volt-seconds (Vs) to correct for the effect of peak size on
249 measured isotope ratios. This normalization was performed with a linearity relation specific to
250 each IRMS and purge-and-trap system.³⁷ Then, for each day of analysis, pyisotopomer was used
251 to generate all possible pairings of reference materials on that day. Each pairing of reference
252 materials and the associated size-corrected isotope ratios were used as an input to the Scrambling
253 function of pyisotopomer to calculate a pair of γ and κ , the scrambling coefficients introduced in
254 Eqn. (10). From the resulting array of γ and κ values, γ and κ were averaged to obtain a daily
255 mean. These daily means were further combined into a one-week running average to smooth
256 their variability. The one-week running averages of γ and κ for each system were used to obtain
257 N₂O isotopocules for reference materials and unknowns, using the Isotopomers function of
258 pyisotopomer. Finally, a scale decompression (similar to the two-point offset correction
259 suggested by Mohn et al., 2014) was performed for $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, $\delta^{17}\text{O}_{\text{N}_2\text{O}}$, and $\delta^{18}\text{O}_{\text{N}_2\text{O}}$, based on
260 the mean measured values and calibrated values of the reference materials included in each run.
261 The scale-decompressed values of $\delta^{15}\text{N}^{\text{bulk}}$ and SP were obtained from the scale-decompressed
262 $\delta^{15}\text{N}^\alpha$ and $\delta^{15}\text{N}^\beta$.

263 The data corrections above are described in the README documents associated with
264 pyisotopomer on the Python Package Index. In brief, comparison against the direct N₂O
265 reference injection and normalization to a peak area of 20 Vs can be accomplished with the
266 appropriate IRMS outputs and the pyisotopomer data input template.⁵⁸ The import convention
267 for pyisotopomer is:

```
268 from pyisotopomer import Scrambling, Isotopomers
```

269
270 To calculate scrambling, the only function you need is:

```
271 Scrambling(inputfile="FILENAME.xlsx", ref1="NAME", ref2="NAME", **kwargs)
```

272
273 where “FILENAME.xlsx” is the user-designated file name of the data corrections spreadsheet to
274 be used as the inputfile, “ref1” and “ref2” are the reference materials used to perform the
275 scrambling calibration designated with their user-entered “NAME”s, and “**kwargs” refers to
276 optional additional keyword arguments, such as the initial guess for γ and κ (see section 4.1).
277 Once the scrambling coefficients are determined, the only function needed to calculate
278 isotopomers is:


```
279 Isotopomers(inputfile = "FILENAME.xlsx", scrambling = [0.0..., 0.0...], **kwargs)
```

280
281 where “scrambling” refers to the scrambling coefficients used to calculate isotopomers.
282

283 3.3 Intercalibration

284 To validate the scrambling calibration, samples of unknown isotopic composition were
285 collected from Lake Lugano, Switzerland in July 2020 and analyzed separately by both Lab 1
286 and Lab 2. The samples were collected at depths of 10 and 90 meters, including six replicate
287 bottles at each depth. Samples were collected into 160-mL glass serum bottles (Wheaton),
288 overflowing each bottle twice, closing bubble-free, and removing liquid to form a 10-mL
289 headspace comprised of air. Based on the Northern hemisphere monthly mean N₂O mixing ratio
290 for July, 2020,⁶¹ when the samples were collected, an atmospheric headspace of this volume
291 would have contained 0.13 nmols N₂O, and resulted in a ~1 nM overestimation of the
292 concentration of N₂O in each sample after equilibration with the headspace during storage. Each
293 sample was capped with a gray butyl septum (National Scientific) and sealed with an aluminum
294 crimp seal. Samples were promptly preserved with 100 μL saturated mercuric chloride solution
295 and stored at lab temperature (20-22°C). Given the trace amount of N₂O in the atmosphere,
296 negligible amount of atmospheric N₂O were added with the 100 μL of mercuric chloride, and
297 with complete flushing of the bottle during analysis, the effect of the headspace and N₂O
298 partitioning between the gas and liquid phases falls within the analytical uncertainty for N₂O
299 concentration and isotopocule measurements.³⁷ The six replicate bottles at each depth were split
300 into two groups of three replicate bottles to be measured for N₂O isotopocules by Lab 1 and Lab
301 2, respectively. The scrambling calibration and isotopocule calculations for each set of samples
302 were performed in pyisotopomer, as above.
303

304 4. Results and Discussion

305 4.1 Solver parameters

306 To understand the sensitivity of the Scrambling function of pyisotopomer to the
307 parameters provided to its solver (initial guess, upper bound, lower bound), we performed a
308 series of numerical experiments with said solver. The first of these experiments involved varying
309 the initial guess, x0 (where x0 is a pair of values for γ and κ). Using a range of 50 initial guesses
310 for γ and κ , evenly spaced between $\gamma = \kappa = 0.05$ and $\gamma = \kappa = 0.20$, scrambling was calculated
311 from four reference materials run in December 2020 on the Lab 1 IRMS. These initial guesses
312 for γ and κ were varied in tandem, such that γ and κ were always initialized at the same value.
313 The solver was subsequently used to obtain a range of 50 solutions for γ and κ corresponding to
314 each of the 50 values of x0, holding the lower bounds for the solver constant at $\gamma=0.0$ and $\kappa=0.0$
315 and the upper bounds constant at $\gamma=1.0$ and $\kappa=1.0$. Using the Isotopomers function of
316 pyisotopomer, these 50 solutions for γ and κ were input as scrambling coefficients to obtain a
317 range of 50 values of $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, and SP for each of the four reference materials (Table 2).

318 The range of x0 values tested produced a similar range of solutions for γ and κ (Figure
319 S1), although their solutions exhibited a nearly constant offset of ~0.09 (γ and κ are unitless
320 since they are proportions, but they can be understood as percentages, whereby a γ value of 0.20
321 can be understood to mean that 20% of $^{14}\text{N}^{15}\text{NO}$ undergoing fragmentation yields $^{30}\text{NO}^+$ instead
322 of $^{31}\text{NO}^+$). The solutions for γ and κ increased as x0 increased, such that the initial guess $\gamma=0.05$

323 and $\kappa=0.05$ yielded the solution $\gamma=0.138$ and $\kappa=0.0454$, and the initial guess $\gamma=0.20$ and $\kappa=0.20$
324 yielded the solution $\gamma=0.272$ and $\kappa=0.182$ (Figure S1). Despite the wide range of solutions for γ
325 and κ , when used as scrambling coefficients to calculate isotopocule ratios these solutions
326 yielded nearly constant $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, and SP values for the four test reference materials (Table
327 2). The standard deviations of $\delta^{15}\text{N}^\alpha$ and $\delta^{15}\text{N}^\beta$ were no greater than 0.07‰, and the standard
328 deviation of SP was no greater than 0.13‰ (Table 2). The calculated $\delta^{15}\text{N}^{\text{bulk}}$ and $\delta^{18}\text{O}_{\text{N}_2\text{O}}$ had a
329 standard deviation of 0.0‰ since they are unaffected by scrambling. This indicates that the
330 parameter best constrained by the scrambling calculation is not the absolute value of γ or κ , but
331 rather their relationship to each other, and that it is this relationship that is uniquely determined
332 by each pair of reference materials and IRMS.

333 Additional numerical experiments were conducted to test the sensitivity of the scrambling
334 solver to the upper and lower bounds provided to it. The lower bounds for both γ and κ were
335 varied from $\gamma=0.0$ and $\kappa=0.0$ to $\gamma=0.08$ and $\kappa=0.08$, to yield a range of 50 lower bound inputs.
336 For these experiments, the initial guesses for γ and κ were held constant at $\gamma=0.17$ and $\kappa=0.08$
337 (standard values for Lab 1) across the range of tested lower bound values. Likewise, the upper
338 bounds for γ and κ were held constant at $\gamma=1.0$ and $\kappa=1.0$ for the lower bound tests. Varying the
339 lower bounds input to the solver resulted in a much smaller range of solutions than varying x_0 :
340 with the lower bounds set to $\gamma=0.0$ and $\kappa=0.0$, the solver yielded $\gamma=0.172$ and $\kappa=0.0798$; with the
341 lower bounds set to $\gamma=0.08$ and $\kappa=0.08$, the solver yielded $\gamma=0.172$ and $\kappa=0.0800$. Again, the
342 difference between γ and $\gamma - \kappa$ was consistently ~ 0.09 . It should be noted here that the range of
343 lower bounds was smaller than the range of x_0 in the analogous experiment above, because the
344 lower bounds for γ and κ cannot, by definition, exceed the lowest value at which either
345 coefficient is initialized — in this case, 0.08. Using the resulting range of solutions for γ and κ to
346 calculate isotopocules for the four test reference materials, we found that the standard deviations
347 of $\delta^{15}\text{N}^\alpha$ and $\delta^{15}\text{N}^\beta$ were no greater than 0.0002‰, and the standard deviation of SP was no
348 greater than 0.0004‰ (Table 2).

349 Repeating this procedure to test the sensitivity of the scrambling solver to its upper
350 bounds yielded similar results. Holding x_0 constant at $\gamma=0.17$ and $\kappa=0.08$ and the lower bounds
351 constant at $\gamma=0.0$ and $\kappa=0.0$, the upper bounds were varied from $\gamma=0.4$ and $\kappa=0.4$ to $\gamma=1.0$ and
352 $\kappa=1.0$ across an array of 50 upper bound inputs. The upper bounds $\gamma=0.4$ and $\kappa=0.4$ yielded the
353 solutions $\gamma=0.171$ and $\kappa=0.0795$, and the upper bounds $\gamma=1.0$ and $\kappa=1.0$ yielded the solutions
354 $\gamma=0.172$ and $\kappa=0.0798$. From these results, it becomes apparent that the solutions for γ and κ
355 relate more closely to the x_0 values $\gamma=0.17$ and $\kappa=0.08$ than they relate to either the lower or
356 upper bounds input to the solver. In this case, the upper bounds are varied from 0.4 to 1.0 to
357 represent a reasonable range of values — unsurprisingly, the edge case wherein the upper bounds
358 are equal or very close to the initial guesses for γ and κ yields solutions for each coefficient that
359 converge at the upper boundary (not shown). Using the resulting range of γ and κ from varying
360 the upper bounds to calculate isotopocules for the four test reference materials, we found that the
361 standard deviations of $\delta^{15}\text{N}^\alpha$ and $\delta^{15}\text{N}^\beta$ were no greater than 0.002‰, and the standard deviation
362 of SP was no greater than 0.005‰ (Table 2). The higher standard deviations in this experiment,
363 compared to the lower bounds experiment, can be attributed to the wider range of input values.

364 From this set of experiments, we can provide three recommendations. Firstly, it may be
365 useful to iterate through the scrambling calculation twice if scrambling coefficients have never
366 been obtained for the given system before. The solution from the first iteration may be used as
367 the initial guess for subsequent iterations, such that the initial guesses for γ and κ are as close to

368 their solved values as possible. This can be accomplished with the “initialguess” argument to the
369 Scrambling function of pyisotopomer:

```
370 Scrambling(inputfile="FILENAME.xlsx", ref1="NAME", ref2="NAME", initialguess=[0.0...,  
371 0.0...])
```

372
373 Secondly, we recommend setting the lower bounds for the scrambling solver to $\gamma=\kappa=0.0$,
374 representing 0% scrambling for $^{14}\text{N}^{15}\text{NO}$ and $^{15}\text{N}^{14}\text{NO}$, respectively. Likewise, we recommend
375 setting the upper bounds to $\gamma=\kappa=1.0$, representing 100% scrambling for $^{14}\text{N}^{15}\text{NO}$ and $^{15}\text{N}^{14}\text{NO}$,
376 respectively. These are the defaults for pyisotopomer. We recommend these as the bounds to
377 allow the scrambling solver to search for solutions across the widest possible range of plausible
378 solutions (it wouldn't make sense to have negative scrambling coefficients, and neither would it
379 makes sense to have scrambling coefficients greater than 1) and thus to avoid converging at a
380 boundary. Our third and final recommendation is to carefully consider the bounds of $^{15}\text{R}^\alpha$ and
381 $^{15}\text{R}^\beta$ provided to the Isotopomers solver. The default upper bounds are $^{15}\text{R}^\alpha=^{15}\text{R}^\beta=1.00$, but they
382 can be set to lower ratios such as $^{15}\text{R}^\alpha=^{15}\text{R}^\beta=0.005$ in the example below, corresponding to
383 $\delta^{15}\text{N}^\alpha=\delta^{15}\text{N}^\beta=360\%$. This is a reasonable ceiling for natural abundance N_2O measurements, but
384 may artificially constrain $^{15}\text{R}^\alpha$ and $^{15}\text{R}^\beta$ in ^{15}N -labelled samples. Thus, is important to consider
385 the context and type of sample when changing default arguments.

```
386 Isotopomers(inputfile = "FILENAME.xlsx", scrambling = [0.0..., 0.0...], lowerbounds=[0.002,  
387 0.002], upperbounds=[0.005, 0.005])
```

388
389

390 4.2 Reference injection scrambling

391 To aid in the optimization process, it is assumed that the flat-topped reference peak could
392 have a slightly different γ and κ from the triangular sample peak. Using Eqn. (10), constant
393 values of γ and κ are used to calculate the ^{31}R of the pure N_2O reference gas from its calibrated
394 $\delta^{15}\text{N}^\alpha$ and $\delta^{15}\text{N}^\beta$ (Table 1) and $\delta^{17}\text{O}$ values. This ^{31}R of the pure N_2O reference gas is used as the
395 “C” parameter in Eqns. (13) and (14) and is assumed to be constant. To test the sensitivity of
396 pyisotopomer to this assumption, the ^{31}R of the pure N_2O reference gas was re-calculated across
397 a range of γ and κ using Eqn. (10), then substituted into the “C” parameter in equations 13 and
398 14. This range of “C” was combined with the measured isotope ratios for one atmosphere-
399 equilibrated seawater and one S2 reference gas run on the Lab 1 IRMS to calculate γ and κ with
400 the Scrambling function of pyisotopomer. Through this sensitivity test, each theoretical γ and κ
401 for the pure N_2O reference gas (C) was paired with a calculated γ and κ for the instrument based
402 on the paired reference materials (Figure 1).

403 The calculated γ and κ varied linearly with the theoretical γ and κ for the direct reference
404 injection (Figure 1). Varying the reference injection γ from 0.17 to 0.18 and holding the
405 reference injection κ constant at 0.080 (white points in Figure 1) resulted in γ ranging from 0.172
406 to 0.181 (Figure 1A) and κ ranging from 0.080 to 0.079 (Figure 1B). Likewise, holding the

407 reference injection γ constant at 0.17 and varying the reference injection κ from 0.080 to 0.070
408 (gray points in Figure 1) also resulted in γ ranging from 0.172 to 0.181 and κ ranging from 0.080
409 to 0.079. Varying the reference injection γ from 0.17 to 0.18 and varying the reference injection
410 κ from 0.080 to 0.070 (black points in Figure 1) resulted in γ ranging from 0.172 to 0.190 and κ
411 ranging from 0.080 to 0.078.

412 Next, the isotopocules for each reference material were calculated from each pairing of
413 theoretical and calculated γ and κ . Despite the variations in the calculated γ and κ shown in
414 Figure 1, the resulting isotopocule ratios varied by less than 0.3‰ (Table 2). The standard
415 deviations of $\delta^{15}\text{N}^\alpha$ and $\delta^{15}\text{N}^\beta$ were 0.13‰ for both the atmosphere-equilibrated seawater and S2
416 reference materials, and the standard deviation of SP was 0.27‰ for both reference materials
417 (Table 2). This result indicates that the γ and κ solutions output by pyisotopomer compensate
418 implicitly for the γ and κ assumed for the direct reference injection. Thus, we conclude that the
419 assumption of different scrambling for the flat-topped reference peak and triangular sample peak
420 has a small effect on the calculated isotopocules.

421

422 **4.3 Choosing reference material pairings**

423 To understand the effect of different pairings of reference materials on the outcome of the
424 scrambling calibration, γ and κ were calculated from a collection of reference pairings run over
425 two days. On the Lab 1 IRMS, these pairings included the “S2” and “B6” reference materials, as
426 well as atmosphere-equilibrated seawater N_2O (Table 1), the calibrated isotopocule ratios of
427 which fall within the range of values produced in culture^{34,62} and nature.^{42,46} The mean and
428 standard deviation of γ and κ were calculated from each of three possible pairings of reference
429 materials: atmosphere-equilibrated seawater and S2 (n=8), atmosphere-equilibrated seawater and
430 B6 (n=6), and S2 and B6 (n=6). Reference material isotopocules were then re-calculated with the
431 mean γ and κ from each pairing. Finally, a pooled standard deviation, calculated as the square
432 root of the average of the squared standard deviations,⁶³ was calculated from the three sets of
433 scrambling coefficients and isotopocule values (Table S1). The same procedure was performed
434 with three sets of N_2O reference materials (“EMPA1”, “EMPA2”, and “EMPA3”) prepared and
435 run on the Lab 2 IRMS. Unlike the reference materials run in Lab 1, however, one of the Lab 2
436 reference materials (“EMPA2”) had a $\delta^{15}\text{N}^\beta$ value (94.44‰) much higher than what is found in
437 culture^{34,62} or nature.^{42,46} Again, the mean and standard deviation of γ and κ were calculated from
438 each of three possible pairings: EMPA1 and EMPA2 (n=37), EMPA1 and EMPA3 (n=37), and
439 EMPA2 and EMPA3 (n=34). The greater number of pairings is due to a greater number of
440 reference materials run per day. Pooled standard deviations were then calculated for γ and κ as
441 well as isotopocule values (Table S1).

442 The pooled standard deviations of γ and κ calculated from the Lab 1 reference materials
443 were 0.0007 and 0.0001, respectively, which correspond to relative uncertainties of 0.39% and
444 0.16% (Table S1). T-tests between pairings of reference materials yielded no significant
445 differences in γ and κ . The pooled standard deviations of $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, and SP were all less than
446 1‰ (Table S1). The pooled standard deviations of γ and κ calculated from the Lab 2 reference
447 materials were larger, corresponding to relative uncertainties of 0.77% and 1.17% in γ and κ ,
448 respectively. This was mostly due to the inclusion of the EMPA2 reference gas — calculating
449 scrambling from just the EMPA1 and EMPA3 reference gases, which did not have extreme
450 values, resulted in relative uncertainties in γ and κ of 0.15% and 0.61%, respectively. T-tests
451 between pairings of reference materials yielded significant differences between the scrambling

452 coefficients calculated from pairings including the EMPA2 reference gas and those without. The
453 inclusion of the EMPA2 reference gas resulted in pooled standard deviations of $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$ of
454 greater than 2‰ and a pooled standard deviation of SP greater than 4‰ (Table S1).

455 Based on these results, we suggest that calculated γ and κ have a dependence on the
456 reference materials used to solve for these parameters, the magnitude of which increases with
457 increasingly extreme values of $\delta^{15}\text{N}^\alpha$ and $\delta^{15}\text{N}^\beta$. Thus, we caution against using reference
458 materials with values that go far beyond bracketing the unknowns to be analyzed to calculate
459 scrambling coefficients. Instead, we recommend calculating γ and κ from reference materials that
460 bracket the range of $\delta^{15}\text{N}^\alpha$ and $\delta^{15}\text{N}^\beta$ expected for the unknowns. We also recommend that a
461 scale decompression, if applied, brackets the range of unknowns but excludes reference materials
462 such as EMPA2 that lie far outside the expected ranges of $\delta^{15}\text{N}^\alpha$ and $\delta^{15}\text{N}^\beta$.

463

464 **4.4 MATLAB vs. Python**

465 To compare the performance of the Python and MATLAB versions of pyisotopomer, a
466 week's worth of paired reference materials (atmosphere-equilibrated seawater, S2 reference gas,
467 and B6 reference gas) from December 2020 was processed in both versions of the software. The
468 resulting mean scrambling coefficients were used to re-calculate isotopocules for each reference
469 material. The differences between MATLAB and Python results for γ and κ were both ~ 0.001 ,
470 which represents a significant difference (see section 4.6). Using the MATLAB scrambling
471 coefficients in the MATLAB isotopomers solver, and the Python scrambling coefficients in the
472 Python isotopomers solver, the resulting $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, and $\delta^{18}\text{O}_{\text{N}_2\text{O}}$ (and thus site preference,
473 $\delta^{15}\text{N}^{\text{bulk}}$ and $\delta^{17}\text{O}_{\text{N}_2\text{O}}$), showed extremely similar values (Table S1, Figure 2). Results from
474 MATLAB and Python diverged in the second decimal place for $\delta^{15}\text{N}^\alpha$ and $\delta^{15}\text{N}^\beta$ (Table S1,
475 Figure 2), and differences in the fourth decimal place for $\delta^{18}\text{O}_{\text{N}_2\text{O}}$ (not shown). The final output
476 isotopocule values from MATLAB and Python had pooled standard deviations of 0.028‰,
477 0.026‰, and 0.054‰ for $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, and SP, respectively. Thus, we recommend that if the
478 MATLAB version of pyisotopomer is used to calculate scrambling, the MATLAB version
479 should also be used to calculate isotopocules, and likewise if the Python version of pyisotopomer
480 is used to calculate scrambling, the Python version should also be used to calculate isotopocules.

481

482 **4.5 Variability in fragmentation behavior**

483 To examine the change in the fragmentation behavior of a single IRMS over time, the
484 scrambling coefficients for the Lab 1 IRMS were compiled from 2018-2021 (Figure 3). A
485 running average was calculated using a window size of 20, equivalent to 20 pairs of reference
486 materials. High volatility in γ and κ in March-April 2019 (sample pair numbers 20-40) coincided
487 with a period where the lab temperature was poorly controlled, and thus exhibited strong day-
488 night variation. Before and after this period, the fragmentation behavior of the instrument
489 exhibited smaller variations (Figure 3). The rolling standard deviation of γ and κ (not shown)
490 confirms that the highest standard deviation of a 20-sample window occurred during April 2019,
491 with the rolling standard deviation in γ equal to 0.0018, or a relative uncertainty of 1.03%, and
492 the rolling standard deviation in κ equal to 0.0025, or a relative uncertainty of 3.40%. The
493 standard deviation of γ across the full dataset of scrambling over time was 0.0019, and the
494 standard deviation of κ across the full dataset was 0.0018.

495 There are several reasons why the scrambling behavior of the ion source might change
496 over time. The NO^+ fragment ion can be produced by one of several routes from N_2O^+ .^{64,65} The

497 pathways and associated isotope effects for the formation of fragment ions are affected by
498 collision frequency, the distribution of excited states, and the time spent in the ion source, which
499 suggests that ion source conditions such as vapor pressure, ionizing energy, and accelerating
500 voltage may all influence the fragmentation behavior of an IRMS system at a given time.^{50,64-67}
501 For these reasons, performing the scrambling calibration only once is not sufficient to obtain
502 high-quality N₂O isotopocule data. Instead, it is important to recalibrate an IRMS system for
503 scrambling on a regular basis since ion source conditions may change with time and can shift
504 abruptly with events such as filament changes. We recommend using a running average of γ and
505 κ over a window of 20 sample pairs, equal to 5 runs of samples with four potential sample pairs
506 per run, rather than calibrating based on day-to-day variation in γ and κ . If there is high volatility
507 in γ and κ , as seen above in March-April 2019, it may be necessary to shorten this window, at the
508 likely expense of accuracy in isotopocule measurements.

509

510 **4.6 Sensitivity of isotopomers to uncertainty in scrambling coefficients**

511 To quantify the effect of uncertainty in γ and κ on $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, and SP, a Monte Carlo
512 simulation was used to introduce random uncertainty in the γ and κ values used to calculate these
513 isotopomers from the measured isotope ratios of three reference materials run on December 7th,
514 2020. Based on past instrument performance (see section 4.5), we modeled γ as a random
515 number centered around $\gamma=0.1722$ with a standard deviation of 0.00192, which is the cumulative
516 standard deviation of all values of γ for the Lab 1 IRMS from 2018-2021 and corresponds to a
517 relative uncertainty of 1.11%. Likewise, we modeled κ as a random number centered around
518 $\kappa=0.0797$ with a standard deviation of 0.00184, which is the cumulative standard deviation of all
519 values of κ for the Lab 1 IRMS from 2018-2021 and corresponds to a relative uncertainty of
520 2.31%. We sampled 1,000 pairs of γ and κ from normal distributions with these means and
521 standard deviations, and then used these values of γ and κ to calculate the 1,000 simulated values
522 of $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, and SP for the three measured reference materials (“S2”, “B6”, and atmosphere-
523 equilibrated seawater; Table 1). We determined the resulting uncertainties in each isotopomer by
524 taking a pooled standard deviation across the three reference materials.

525 This analysis showed that a small relative uncertainty in each scrambling coefficient can
526 lead to per mil-level errors in $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, and SP. Specifically, relative uncertainties of 1.11%
527 and 2.31% in γ and κ , respectively, resulted in pooled standard deviations of 3.61‰ in $\delta^{15}\text{N}^\alpha$ and
528 $\delta^{15}\text{N}^\beta$ (the pooled standard deviations for $\delta^{15}\text{N}^\alpha$ and $\delta^{15}\text{N}^\beta$ were equivalent), and 7.23‰ in SP
529 (Figure 4). Modeling γ and κ with relative uncertainties 0.39% and 0.16%, instead, which were
530 the uncertainties in each parameter based on reference materials run over two days (see section
531 4.3), led to pooled standard deviations of 0.98‰ in $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, and 1.95‰ in SP (Figure S2).

532

533 Performing this exercise along a range of modeled uncertainties in γ and κ produced a
534 response curve of the expected standard deviations in each isotopomer for a given level of
535 uncertainty in γ and κ (Figure S3). Fitting a linear regression through each curve, we obtain
536 equations of form $y=mx+b$ for SP and $\delta^{15}\text{N}^\alpha$ (the response curves for $\delta^{15}\text{N}^\alpha$ and $\delta^{15}\text{N}^\beta$ were
537 identical). Solving for the uncertainty in γ and κ needed to achieve 1‰ pooled standard deviation
538 for SP gives $x=0.197\%$. In other words, to obtain site preference values with an uncertainty of
539 1‰, the relative uncertainties in γ and κ should be reduced to less than $\sim 0.2\%$. Averaging
540 scrambling coefficients over a greater number of reference material pairings under the same ion
541 source conditions should reduce their uncertainty; thus, we recommend calculating γ and κ from

542 a ~1-week moving average, instead of daily. Using a one-week moving average of each
543 scrambling coefficient for the intercalibration exercise below (see section 4.7), we found relative
544 uncertainties in γ and κ of 0.4% and 0.2%, respectively — which are close to this target. We do
545 *not* recommend calculating γ and κ from a cumulative average or cumulative moving average
546 that includes longer term changes in ion source conditions, which can affect γ and κ .

547

548 **4.7 Intercalibration**

549 The application of pyisotopomer was tested through an intercalibration including four
550 reference materials and two Lake Lugano samples measured by two IRMS laboratories (Table
551 3). Pyisotopomer was used to perform the scrambling calibration for each laboratory and to
552 obtain isotopocule ratios. Afterwards, a scale decompression was applied, as described in the
553 Methods. The root mean square deviation (RMSD) for each reference material was calculated by
554 comparison to the calibrated values provided by a previous intercalibration effort⁵² (atmosphere-
555 equilibrated seawater) and J. Mohn (EMPA1, EMPA2, and EMPA3).

556 The $\delta^{15}\text{N}^{\text{bulk}}$ measured by the two labs displayed good agreement for three out of the four
557 reference materials, as well as the lake water samples. The $\delta^{15}\text{N}^{\text{bulk}}$ RMSDs for atmosphere-
558 equilibrated seawater and reference material EMPA3 were 0.71‰ and 0.50‰, respectively, both
559 of which represent an improvement upon the intercalibration presented by Mohn et al. (2014). In
560 contrast, the RMSD for EMPA1 was 2.46‰, larger than the 0.8‰ presented for IRMS labs by
561 Mohn et al., 2014 (Table 3). The RMSD for EMPA2 was highest at 3.59‰, but this is to be
562 expected, given that EMPA2 was excluded from the scrambling calculation and scale
563 decompression conducted in this study due to its extreme values (see section 4.3). For the lake
564 water sample taken at 10 m depth, the $\delta^{15}\text{N}^{\text{bulk}}$ values measured by Lab 1 and Lab 2 were
565 statistically indistinguishable (Table 3; Figure S4). For the lake water sample taken at 90 m
566 depth, the $\delta^{15}\text{N}^{\text{bulk}}$ values measured by Lab 1 and Lab 2 were -5.00 ± 0.08 ‰ and -6.29 ± 1.06 ‰,
567 respectively (Table 3; Figure S4).

568 The $\delta^{15}\text{N}^{\alpha}$ measured by the two labs also showed good agreement for reference materials
569 EMPA1, EMPA3, and atmosphere-equilibrated seawater: in each case, the combined RMSD was
570 less than 2.60‰ (Table 3). This is similar to the data presented in Mohn et al. (2014), who find
571 an RMSD for $\delta^{15}\text{N}^{\alpha}$ for IRMS laboratories of 2.47‰. The values of $\delta^{15}\text{N}^{\alpha}$ measured by the two
572 labs for the lake water unknowns differed by ~2-3‰, but no consistent offset emerged between
573 the two labs, and neither did such an offset emerge in the reference materials (i.e., the values for
574 some samples were higher in Lab 1, and others are higher for Lab 2; Figure S4). For $\delta^{15}\text{N}^{\beta}$, the
575 RMSD for atmosphere-equilibrated seawater, EMPA1, and EMPA3 were slightly larger, and
576 only EMPA3 represents an improvement upon the data presented in previous intercalibrations
577 (Table 3). The $\delta^{15}\text{N}^{\beta}$ measured by Lab 1 for the lake water unknowns was 1-2‰ different from
578 that measured by Lab 2, but again, no consistent offset emerged (Figure S4).

579 The SP values measured by the two laboratories showed larger standard deviations than
580 the $\delta^{15}\text{N}^{\alpha}$ and $\delta^{15}\text{N}^{\beta}$ individually, which is to be expected, since SP is a measure of difference
581 between the latter two parameters. The RMSD values, however, were all less than 3‰ for
582 atmosphere-equilibrated seawater, EMPA1, and EMPA3 (Table 3). This represents an
583 improvement on Mohn et al. (2014), who find an RMSD of 4.29‰ for SP measured by IRMS
584 laboratories. The Lake Lugano unknowns showed larger offsets than the reference materials
585 (Figure S4). The lake water sample from 10 m depth showed an especially large difference in SP
586 between Lab 1 and Lab 2: Lab 1 measured a mean SP of 16.43 ± 1.35 ‰ at this depth, while Lab 2

587 measured a mean SP of $20.77 \pm 2.67\%$ (Table 3). At 90 m depth, Lab 1 measured a mean SP of
588 $54.40 \pm 1.23\%$, and Lab 2 measured a mean SP of $55.09 \pm 1.69\%$.

589 For $\delta^{18}\text{O}_{\text{N}_2\text{O}}$, Lab 1 obtained consistently higher values than Lab 2, despite the application
590 of a scale decompression, which should compensate for such offsets.⁵² The only exception to this
591 rule was the lake water unknown from 10 m depth, for which the $\delta^{18}\text{O}_{\text{N}_2\text{O}}$ values measured by
592 the two labs were statistically indistinguishable (Table 3; Figure S4). For the other samples and
593 reference materials, the magnitude of this offset varied from 2.81‰ (EMPA3) to 6.80‰ (lake
594 water unknown from 90 m). For reference materials EMPA1 and EMPA3, the $\delta^{18}\text{O}_{\text{N}_2\text{O}}$ values
595 measured by Lab 1 were 2-3‰ higher than the calibrated $\delta^{18}\text{O}_{\text{N}_2\text{O}}$ (Tables 1 & 3), while the
596 $\delta^{18}\text{O}_{\text{N}_2\text{O}}$ values measured by Lab 2 did not exhibit such an offset. For atmosphere-equilibrated
597 seawater, the $\delta^{18}\text{O}_{\text{N}_2\text{O}}$ values measured by Lab 2 were $\sim 3\%$ lower than the calibrated $\delta^{18}\text{O}_{\text{N}_2\text{O}}$
598 (Tables 1 & 3), while the $\delta^{18}\text{O}_{\text{N}_2\text{O}}$ values measured by Lab 1 did not exhibit such an offset. This
599 offset did not exhibit any dependence on N_2O concentration — rather, the variation in $\delta^{18}\text{O}_{\text{N}_2\text{O}}$
600 values tended to be larger for lower peak areas, resulting in greater offsets, but without any
601 measurable trend (cite figure or table?). Similarly, the linearity correction could not correct for
602 the variation at lower peak areas, because this variation was random.

603

604 **5. Conclusion: How to obtain high-quality N_2O isotopocule data using pyisotopomer**

605 Using pyisotopomer and at least two reference materials (three characterized gases,
606 assuming the N_2O reference tank is also calibrated), one can calculate scrambling for a given
607 IRMS and apply those scrambling coefficients to calculate the isotopocule values of unknown
608 samples. To ensure high-quality results from these calculations, we provide the following
609 recommendations. Firstly, if scrambling has never been calculated for the IRMS or current
610 filament, iterate through the scrambling calculation twice. Use the solution from the first
611 iteration as the initial guess for subsequent calculations. This can be achieved with the
612 “initialguess” argument to the Scrambling function of pyisotopomer. In a similar vein, use the
613 default settings for the lower and upper bounds for both the Scrambling and Isotopomers solvers.
614 These settings allow the solvers to search for solutions across the widest possible range of
615 values. Secondly, to calculate scrambling, where possible, choose pairings of reference materials
616 that bracket the range of unknowns but do not have $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, or $\delta^{18}\text{O}_{\text{N}_2\text{O}}$ that lie far outside
617 of this range. The same is true for scale decompression. Third, if the MATLAB version of
618 pyisotopomer is used to calculate scrambling, the MATLAB version should also be used to
619 calculate isotopocules. Finally, it is necessary to run paired reference materials daily to obtain
620 accurate running estimates of γ and κ . It is recommended to calculate convert these daily
621 estimates to a 1-week running average and use that average to calculate isotopocules. For a 1‰
622 precision in SP, the standard deviation of this running average should represent no more than a
623 0.2% uncertainty in γ and κ .

624 Using pyisotopomer in an intercalibration exercise and implementing the above
625 recommendations, we find good agreement between the isotopocules measured by two different
626 IRMS labs for both reference materials and natural lake samples. We conclude that while the
627 intercalibration results demonstrate potential for further improvement in both precision and
628 accuracy, the intercalibration of SP using a uniform scrambling calculation (pyisotopomer)
629 presented here represents an improvement upon previous N_2O intercalibrations.

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632 **Data availability statement**

633 The manuscript is prepared to comply with the RCMS data policy. The latest version of
634 pyisotopomer is available for installation via the Python Package index
635 (pypi.org/project/pyisotopomer). The first release of pyisotopomer is also available via Zenodo
636 for both Python (doi.org/10.5281/zenodo.5031218) and MATLAB
637 (doi.org/10.5281/zenodo.5031237). This research was supported by U.S.-NSF grant OCE-
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639 The authors declare no competing financial interests.

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838 **Table 1:** Reference materials for N₂O isotopic analysis and intercalibration. The laboratories
 839 participating in the intercalibration exercise were at Stanford University (“Lab 1”) and the
 840 University of Basel (“Lab 2”).
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Reference material	$\delta^{15}\text{N}^{\alpha}$	$\delta^{15}\text{N}^{\beta}$	SP	$\delta^{15}\text{N}^{\text{bulk}}$	$\delta^{18}\text{O}_{\text{N}_2\text{O}}$	Calibration by
	<i>(‰ vs. air N₂)</i>			<i>(‰ vs. VSMOW)</i>		
S2 reference gas	5.55	-12.87	18.42	-3.66	32.73	S. Toyoda, Tokyo Tech
B6 reference gas	-0.40	-0.15	-0.26	-0.28	41.95	Lab 1 internal standard
Atmosphere-equilibrated seawater	15.70	-3.30	19.00	6.20	44.30	Mohn et al. (2014)
EMPA1 reference gas (CA06261)	-22.21	-49.28	27.07	-35.75	26.94	J. Mohn, EMPA
EMPA2 reference gas (CA08214)	1.71	94.44	-92.73	48.08	36.01	J. Mohn, EMPA
EMPA3 reference gas (53504)	17.11	-3.43	20.54	6.84	35.39	J. Mohn, EMPA
	³¹ R	⁴⁵ R	⁴⁶ R			
Lab 1 pure N ₂ O direct injection	0.003733763	0.007741025	0.002101295			S. Toyoda, Tokyo Tech
Lab 2 pure N ₂ O direct injection	0.004049069	0.007738762	0.002100262			J. Mohn, EMPA

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845 **Table 2:** Performance tests of pyisotopomer. A) The initial guess x_0 used to solve for γ and κ
846 was varied from $\gamma=\kappa=0.05$ to $\gamma=\kappa=0.20$. B) The lower bounds used to solve for γ and κ were
847 varied from $\gamma=\kappa=0.0$ to $\gamma=\kappa=0.08$. C) The upper bounds used to solve for γ and κ was varied
848 from $\gamma=\kappa=0.4$ to $\gamma=\kappa=1.0$. D) The γ and κ used to calculated ^{31}R for the direct reference injection
849 were varied. For each test, the resulting range of solutions for γ and κ were used to calculate
850 isotopocule values for test reference materials. Reported values are the means and standard
851 deviations of the range of 50 solutions.
852

Sample ID	$\delta^{15}\text{N}^\alpha$	$\delta^{15}\text{N}^\beta$	SP	$\delta^{15}\text{N}^{\text{bulk}}$	$\delta^{18}\text{O}_{\text{N}_2\text{O}}$
	(‰ vs. air N_2)				(‰ vs. VSMOW)
A. Varying initial guess x_0					
Atmosphere-equilibrated seawater #1	15.45±0.02	-3.59±0.02	19.03±0.03	5.93±0.00	45.39±0.00
Atmosphere-equilibrated seawater #2	14.85±0.04	-3.22±0.04	18.07±0.08	5.82±0.00	46.45±0.00
S2 reference gas #1	5.18±0.07	-12.28±0.07	17.46±0.13	-3.55±0.00	33.30±0.00
S2 reference gas #2	5.19±0.05	-12.52±0.05	17.71±0.10	-3.66±0.00	34.31±0.00
B. Varying lower bounds					
Atmosphere-equilibrated seawater #1	15.51±0.00	-3.66±0.00	19.17±0.00	5.93±0.00	45.39±0.00
Atmosphere-equilibrated seawater #2	14.97±0.00	-3.34±0.00	18.30±0.00	5.82±0.00	46.45±0.00
S2 reference gas #1	5.32±0.00	-12.42±0.00	17.74±0.00	-3.55±0.00	33.30±0.00
S2 reference gas #2	5.32±0.00	-12.64±0.00	17.96±0.00	-3.66±0.00	34.31±0.00
C. Varying upper bounds					
Atmosphere-equilibrated seawater #1	15.51±0.00	-3.65±0.00	19.16±0.01	5.93±0.00	45.39±0.00
Atmosphere-equilibrated seawater #2	14.97±0.00	-3.33±0.00	18.30±0.00	5.82±0.00	46.45±0.00
S2 reference gas #1	5.31±0.00	-12.42±0.00	17.73±0.00	-3.55±0.00	33.30±0.00
S2 reference gas #2	5.32±0.00	-12.64±0.00	17.96±0.00	-3.66±0.00	34.31±0.00
D. Varying ref. injection scrambling					
Atmosphere-equilibrated seawater #1	15.40±0.13	-3.54±0.13	18.94±0.27	5.93±0.00	45.39±0.00
S2 reference gas #1	5.20±0.13	-12.30±0.13	17.50±0.27	-3.55±0.00	33.30±0.00

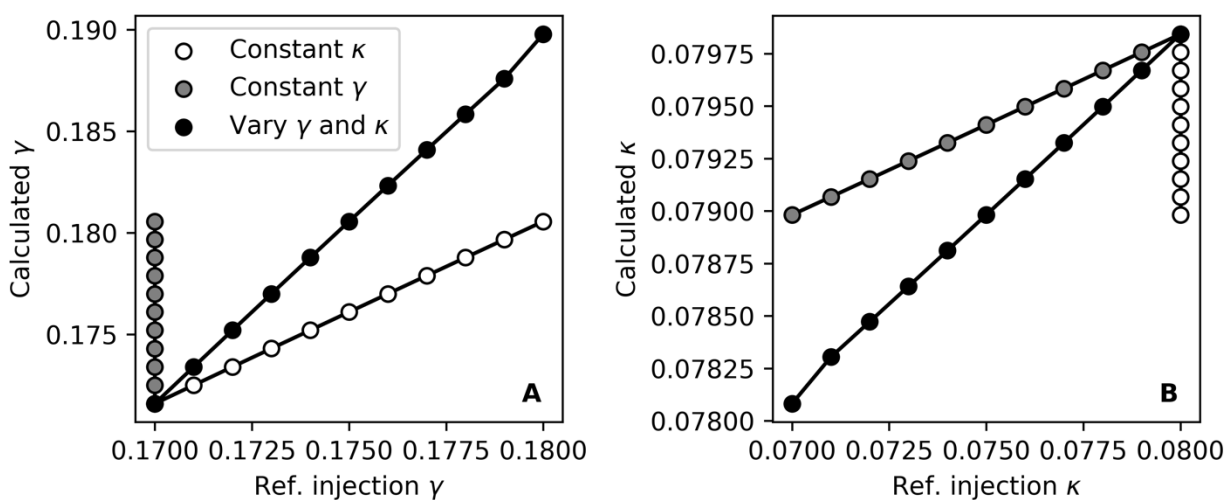
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856 **Table 3:** N₂O isotopic composition of four reference materials and two unknowns analyzed by
 857 two IRMS laboratories. $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, SP, and $\delta^{15}\text{N}^{\text{bulk}}$ are reported in ‰ vs. Air N₂, and $\delta^{18}\text{O}_{\text{N}_2\text{O}}$
 858 is reported in ‰ vs. VSMOW. Uncertainties are standard deviations of replicate bottles and do
 859 not include the propagation of calibration uncertainties. The root-mean square deviation (RMSD)
 860 was calculated with respect to the calibration data presented in Table 1.
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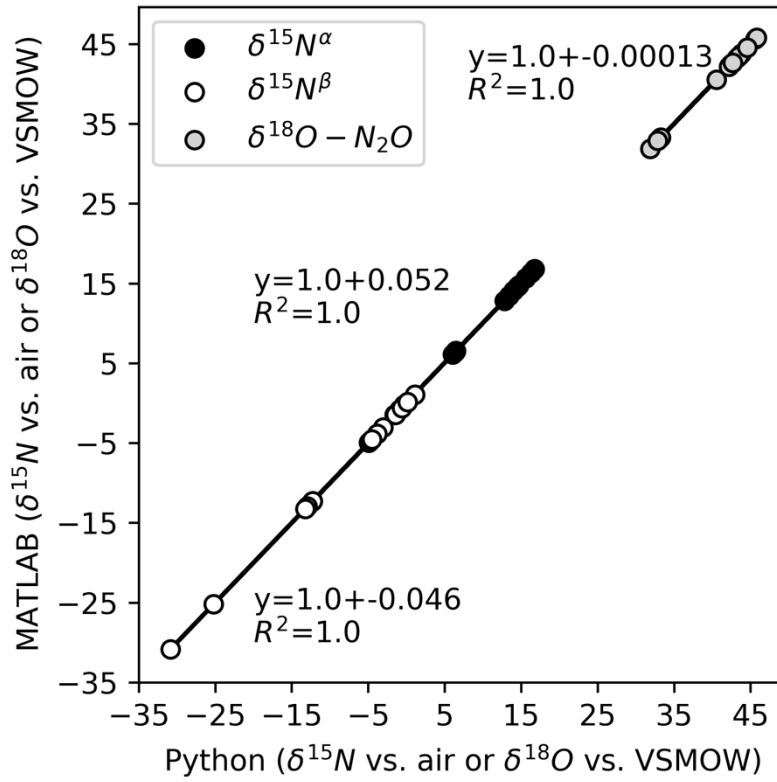
Reference gas		$\delta^{15}\text{N}^\alpha$	$\delta^{15}\text{N}^\beta$	SP	$\delta^{15}\text{N}^{\text{bulk}}$	$\delta^{18}\text{O}_{\text{N}_2\text{O}}$
		(‰ vs. air N ₂)				(‰ vs. VSMOW)
Atmosphere-equilibrated seawater	mean (Lab 1)	15.46±0.72	-3.12±2.00	18.58±2.43	6.17±0.89	44.67±2.08
	mean (Lab 2)	16.88±1.82	-4.21±1.75	21.09±3.51	6.34±0.33	40.81±1.04
	RMSD (Labs 1 & 2)	1.20	1.81	2.73	0.71	6.49
EMPA1	mean (Lab 1)	-17.77±1.69	-43.88±1.89	26.12±3.55	-30.83±0.23	30.31±1.27
	mean (Lab 2)	-22.41±1.55	-49.19±2.42	26.78±3.71	-35.80±0.83	27.01±1.68
	RMSD (Labs 1 & 2)	2.57	3.10	2.89	2.46	2.04
EMPA2*	mean (Lab 1)	2.49±1.33	82.89±1.69	-80.40±2.39	42.69±0.94	39.36±0.79
	mean (Lab 2)	-10.43±1.44	102.16±3.86	-112.60±4.46	45.87±1.88	35.44±1.35
	RMSD (Labs 1 & 2)	10.96	9.82	19.53	3.59	2.07
EMPA3	mean (Lab 1)	16.55±1.75	-2.90±0.95	19.99±2.49	7.10±0.66	37.85±1.25
	mean (Lab 2)	17.11±1.31	-3.51±1.30	20.62±2.28	6.80±0.64	35.04±3.34
	RMSD (Labs 1 & 2)	1.31	1.04	2.13	0.50	2.32
Unknown #1 (Lake Lugano, 10m depth)	mean (Lab 1)	11.93±0.29	-4.51±1.05	16.43±1.35	3.71±0.38	44.56±1.16
	mean (Lab 2)	14.49±1.34	-6.28±1.47	20.77±2.67	4.10±0.44	44.90±1.64
Unknown #2 (Lake Lugano, 90m depth)	mean (Lab 1)	22.20±0.55	-32.20±0.68	54.40±1.23	-5.00±0.08	60.68±0.04
	mean (Lab 2)	21.26±1.91	-33.81±0.22	55.09±1.69	-6.26±1.06	53.88±4.69

*Large errors are due to exclusion from scrambling calibration

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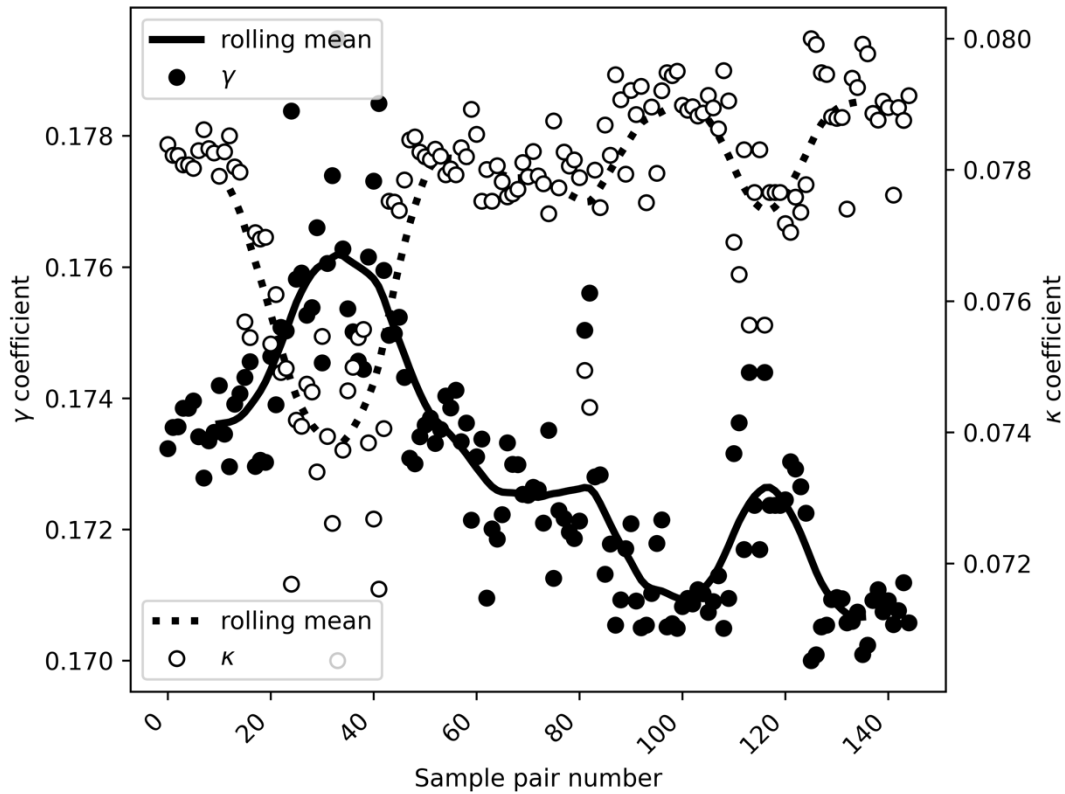
864
 865 **Figure 1.** Solutions for γ (A) and κ (B) calculated across a range of theoretical γ and κ for the
 866 direct reference injection: changing the reference injection γ and holding the reference injection
 867 κ constant (white), changing the reference injection κ and holding the reference injection γ
 868 constant (gray), changing both reference injection γ and κ (black).



869

870 **Figure 2.** Comparison of isotopocule results for Python and MATLAB versions of
 871 pyisotopomer.

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873

874 **Figure 3.** Scrambling coefficients for the Lab 1 IRMS from October 2018 to January 2021.

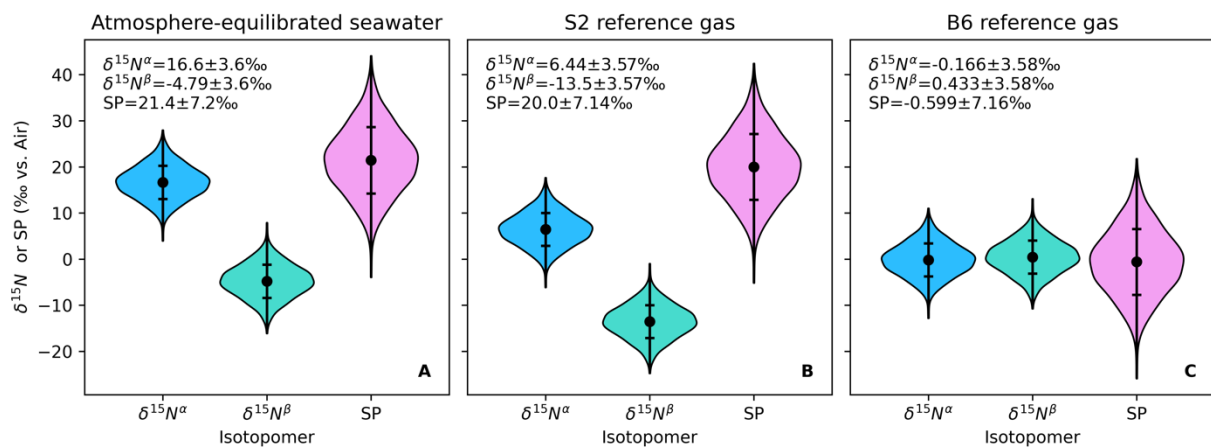
875 Individual pairs of scrambling coefficients, calculated from individual pairs of reference

876 materials, are shown as a scatter plot. A rolling mean with a 20-value window (roughly

877 equivalent to one week of analysis) is plotted for each coefficient. The x-axis is sample pair

878 number, rather than date, to better visualize short-term variability.

879



880

881 **Figure 4:** Isotopocule values and error associated with an uncertainty of ± 0.00192 in γ and

882 ± 0.00184 in κ , based on Monte Carlo simulation results. The violin plots are based on a kernel

883 density estimate of the distribution and the values plotted and reported on each figure show the
884 mean value $\pm 1\sigma$.
885

Supplementary material

pyisotopomer: A Python package for obtaining nitrous oxide isotopocules from isotope ratio mass spectrometry

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Table S1: Different pairings of reference materials. Pooled standard deviations of γ and κ calculated from different pairings of reference materials, and the corresponding pooled standard deviations of $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, and SP, are shown for tests performed on the Lab 1 and Lab 2 IRMS. Pooled standard deviations are also shown for scrambling and isotopomer calculations done in the MATLAB and Python versions of the software.

Test	Pooled standard deviations				
	γ (% uncertainty*)	κ (% uncertainty*)	$\delta^{15}\text{N}^\alpha$ (% vs. <i>AirN</i> ₂)	$\delta^{15}\text{N}^\beta$ (% vs. <i>AirN</i> ₂)	SP (% vs. <i>AirN</i> ₂)
Lab 1 three ref. pairings, no extreme values	0.39	0.16	0.47	0.44	0.91
Lab 2 three ref. pairings, incl. extreme values	0.77	1.17	2.54	2.29	4.83
Lab 2 one ref. pairing, excl. extreme values	0.15	0.61	(see section 4.7 for precision between replicates)		
MATLAB vs. Python	0.44	0.93	0.028	0.026	0.054

*Percent uncertainties represent pooled SD as a percentage of the mean

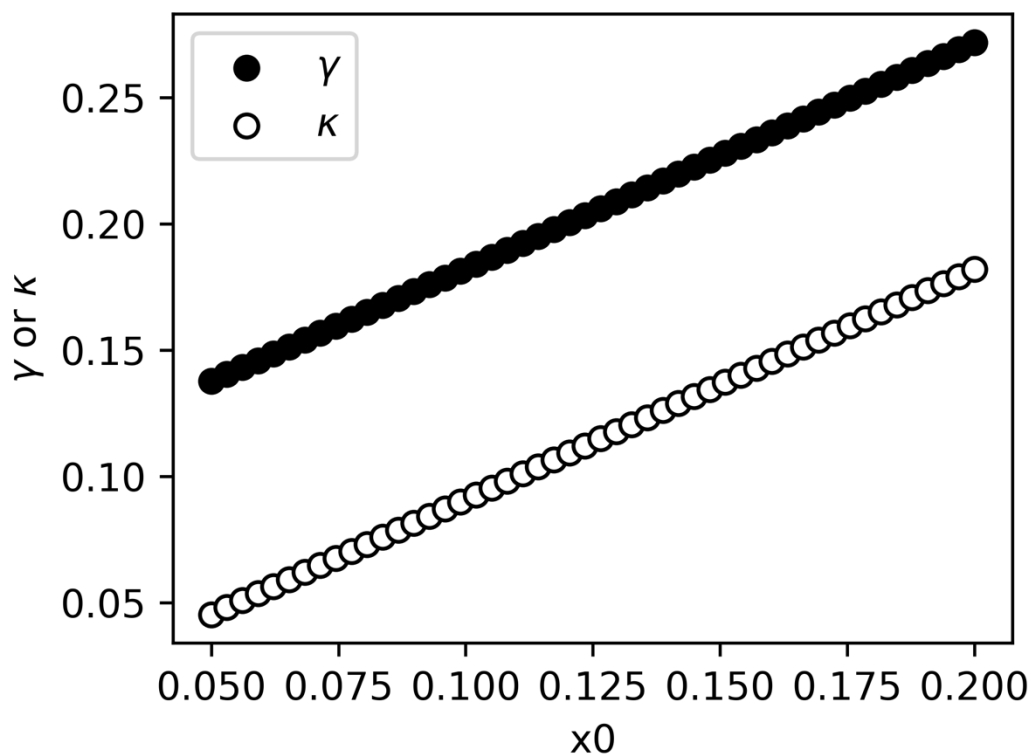


Figure S1. Solutions for γ and κ calculated across a range of initial guesses (x_0) for the least squares solver function at the core of pyisotopomer. The default values for x_0 are $\gamma=0.17$ and $\kappa=0.08$, based on the performance of the ThermoFinnigan Delta V mass spectrometer at Lab 1.

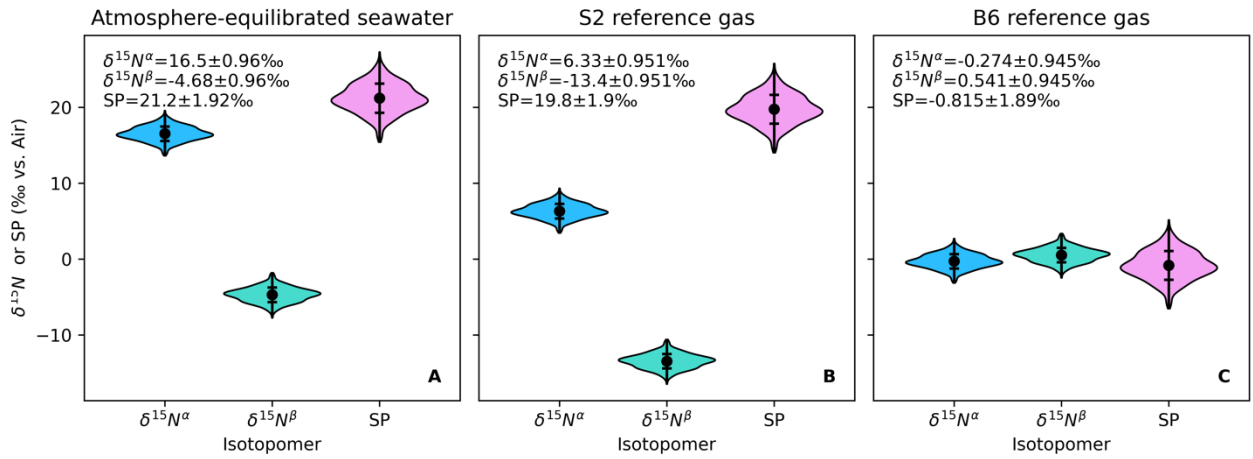


Figure S2: Isotopocule values and error associated with an uncertainty of $\pm 6.83 \cdot 10^{-4}$ in γ and $\pm 1.48 \cdot 10^{-4}$ in κ , based on Monte Carlo simulation results. The violin plots are based on a kernel density estimate of the distribution and show the mean value $\pm 1\sigma$.

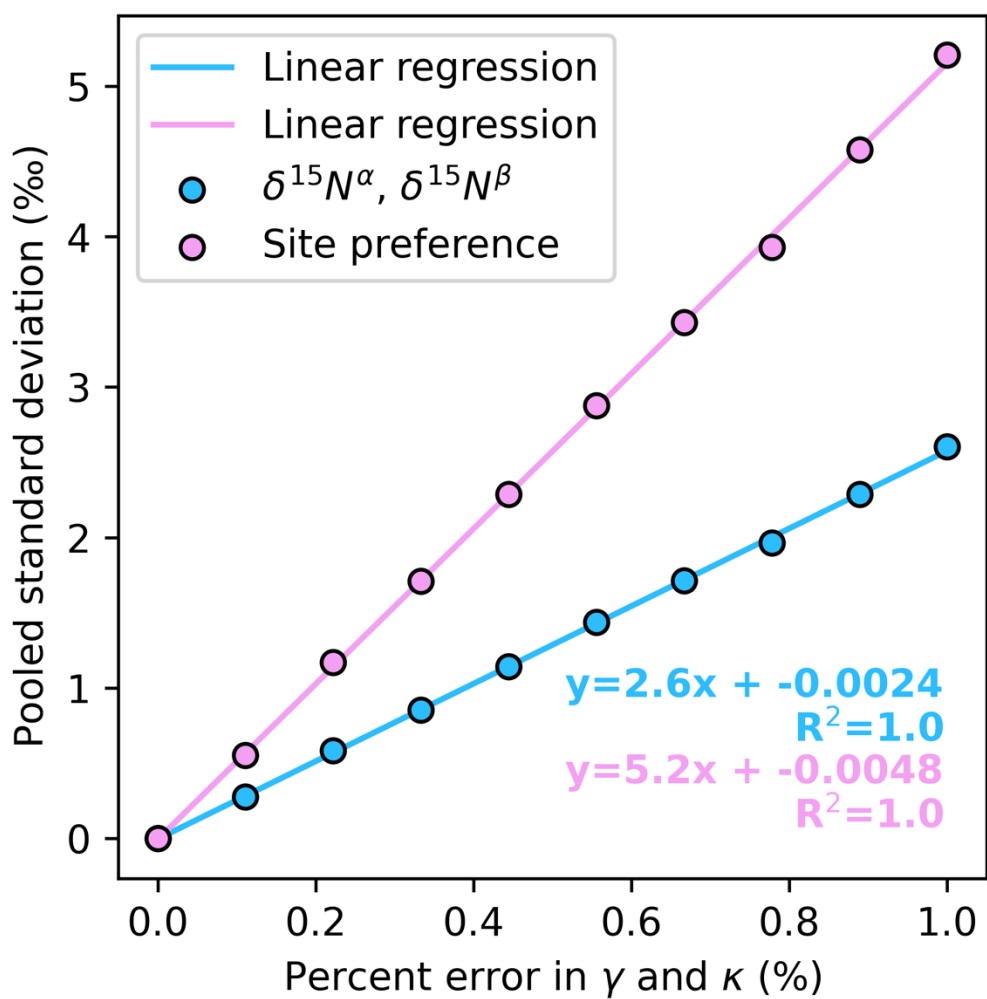


Figure S3: Pooled standard deviation of $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, and SP resulting from increasing relative uncertainties in γ and κ . Each point corresponds to 1,000 values of γ and κ sampled from a distribution with standard deviation equal to the given level of uncertainty, and the corresponding pooled standard deviation of 1,000 simulated values of $\delta^{15}\text{N}^\alpha$, $\delta^{15}\text{N}^\beta$, and SP.

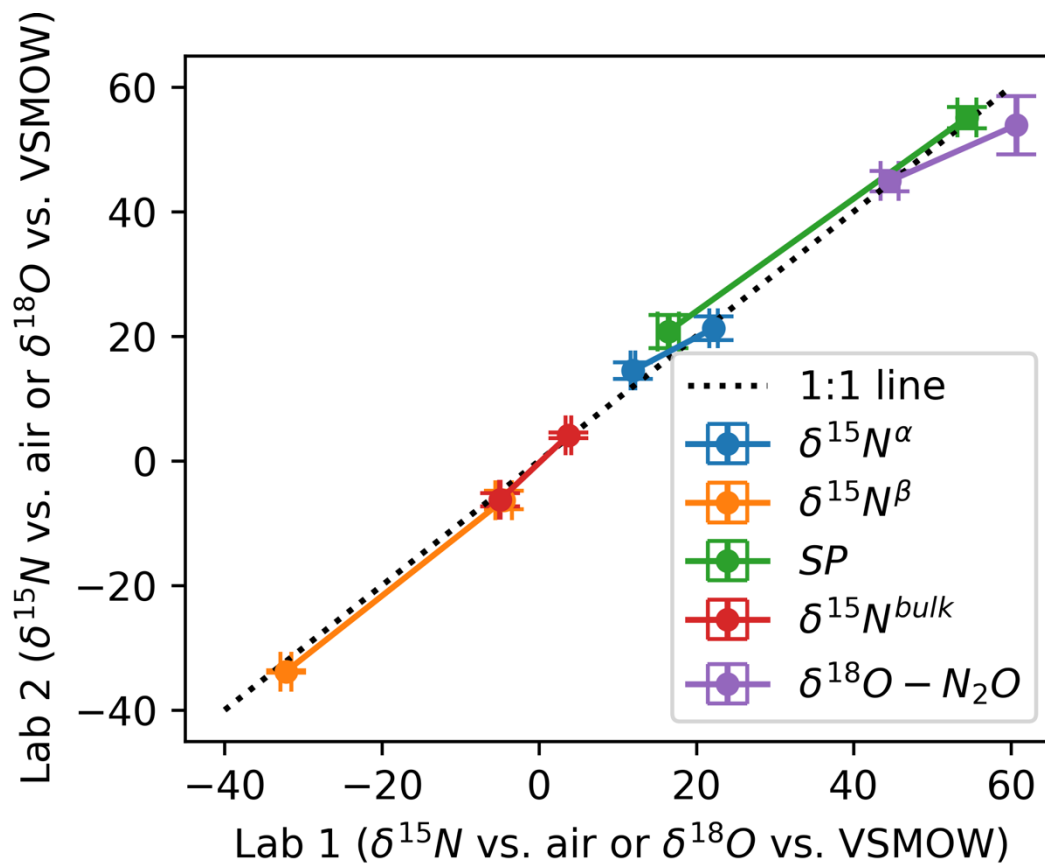


Figure S4: Intercalibration results for lake water unknowns taken at 10 meters and 90 meters depth from lake Lugano in Switzerland. Independent measurements of the isotopic composition of N_2O in each sample, performed by Lab 1 and Lab 2, respectively, are plotted against each other. Data points represent the mean from replicate bottles taken at the same depth ($n=2-5$) and error bars represent one standard deviation. A one-to-one line (black dashed line) is plotted for comparison.