National-scale, remotely sensed lake trophic state, 1984-2020

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Abstract (155 of 170 words)

Lake trophic state is a key ecosystem property that integrates a lake's physical, chemical, and biological processes. Despite the importance of trophic state as a gauge of lake water quality, standardized and machine-readable observations are uncommon. Remote sensing presents an opportunity to detect and analyze lake trophic state with reproducible, robust methods across time and space. We used Landsat surface reflectance data to create the first compendium of annual lake trophic state for 55,662 lakes of at least 10 ha in size throughout the contiguous United States from 1984 through 2020. The dataset was constructed with FAIR data principles (Findable, Accessible, Interoperable, and Reproducible) in mind, where data are publicly available, relational keys from parent datasets are retained, and all data wrangling and modeling routines are scripted for future reuse. Together, this resource offers critical data to address basic and applied research questions about lake water quality at a suite of spatial and temporal scales.

Background and Summary

Lakes and reservoirs are of critical importance to society, directly providing drinking water and supporting food production, sanitation, and transportation. Millions of people worldwide face intermittent clean water availability due to climatic and anthropogenic stressors[1]. Current literature suggests that changes in surface water quantity and quality are highly heterogeneous, and trends globally suggest that factors such as ice cover, air temperature, humidity, and lake surface area are likely interacting regionally to affect freshwater ecosystems in synergistic ways[2–7]. To gain a better understanding of the potential threats to freshwater ecosystems, new technologies must be engaged. Satellite-based Earth observations (hereafter "remote sensing") are particularly useful as they can provide information at spatial and temporal scales that are currently impossible to replicate via ground-based observations.

Although remote sensing's usefulness to track changes in water quantity has enabled analyses of water availability from local-to-global scales[8–11], investigations of water quality have historically been more limited in scale and scope. However, remote sensing now offers powerful approaches to assessing patterns and trends in water quality[2,12–15], and data harmonization efforts allow for greater interoperability between *in situ* collections and remote sensing imagery[16,17]. Among studies of remotely sensed metrics of water quality, the majority have centered around specific constituents, such as chlorophyll or suspended sediment, without necessarily offering holistic metrics of ecosystem integrity.

Lake trophic state (LTS) is an example of a metric intended to provide holistic assessments of a lake's aggregate physical (e.g., light attenuation), chemical (e.g., nutrient concentrations), and biological processes (e.g., productivity). Within the limnological community, LTS is a property closely associated with a lake's characteristic autochthonous and allochthonous productivity as well as water color. Broadly speaking, eutrophic lakes are green, oligotrophic lakes are blue, and dystrophic lakes are brown

(Figure 1). From color-trophic state connections, fundamental limnological principles center around linking trophic states to characteristic properties (Figure 1). For example, oligotrophic lakes are usually characterized by having lower phosphorus concentrations, low offshore but comparably higher nearshore productivity, and low colored dissolved organic matter (Figure 1). In contrast, eutrophic lakes have higher phosphorus concentrations and higher phytoplankton biomass (Figure 1).

In a management context, the language of LTS has historically been used to describe conditions relative to nutrient enrichment. For example, following the 1971 announcement of US Federal efforts to limit the use of phosphorus in detergents, the U.S. Environmental Protection Agency (U.S. EPA) and state water resource management agencies launched a National Eutrophication Survey[18]. The survey assessed trophic state, defined as nutrient enrichment, of lakes influenced by wastewater treatment plants. In this case, LTS language was used to focus on and communicate about eutrophication, whereas dystrophication aspects of the framework were not as prominent. These language patterns likely carry over to contemporary uses. Because discussions may have focused on eutrophication in the past, modern tools and frameworks could be enhanced by remotely sensed water quality data that capture aspects of both eutrophication and dystrophication. For example, as climate changes, drinking water utility managers will increasingly face compounding hazards that could negatively impact lakes and reservoirs that supply hundreds of millions of people with drinking water[19]. Data and tools that provide remotely sensed information on LTS could improve the ability to observe multidecadal changes in water quality and save resources by better targeting field monitoring.

Although LTS is often employed as a classification system for characterizing autotrophic production[20], the Nutrient-Color Paradigm (NCP) is an empirically tested framework for discriminating LTS based off two variables: (1) phosphorus concentrations, a proxy for primary productivity; and (2) colored dissolved organic matter or turbidity measured in platinum-cobalt units, both proxies for water transparency[21–23]. By combining characteristic metrics of a lake's primary productivity and optical properties, the NCP presents a powerful system for discriminating LTS, where both autochthonous and allochthonous processes are considered. Leveraging the relationship between LTS, phosphorus concentrations, and water clarity, it is possible to transform remotely sensed lake surface reflectance observations into meaningful limnological and ecosystem properties.

Here, we present the first national-scale compendium of LTS that has been built from remotely sensed lake color (i.e., spectral signatures). The dataset, referred to as LTS-US, is derived from (1) coordinated, continental-scale *in situ* measurements, where LTS has been documented for select lakes and years, and (2) characteristic Landsat surface reflectance values for each lake's Chebyshev center (the point in a polygon furthest from the edge). Using *in situ* LTS, we can build predictive models to associate LTS with characteristic reflectance values, and then apply predictive models to lakes with unknown trophic states. Together, the dataset contains predictions for 55,662 lakes of at least 10 ha in area with annual estimates of LTS from 1984 through 2020. By

coupling satellite-based remote sensing with fundamental limnological principles, the LTS-US dataset provides the means to apply the NCP at the national scale to identify macroscale patterns and trends in LTS. Further, this approach moves beyond remote sensing of individual parameters to provide insights into lakes' physical, chemical, biological, and ecosystem properties.

Methods

The LTS-US dataset is constructed using a four-part pipeline, as shown in Figure 2: (1) aggregate training data, (2) create classification models, (3) apply predictions to lakes outside of the training data, and (4) assess model performance and prediction validity. Individual steps within the pipeline are described below.

Step 1: Identify Parent Datasets

U.S. Environmental Protection Agency National Lakes Assessment

In situ measurements of total phosphorus and true color were compiled from the U.S. EPA's National Lakes Assessment (NLA)[24–28], a synoptic sampling campaign of lakes, ponds, and reservoirs, hereafter collectively referred to as "lakes", conducted in the contiguous U.S. every five years. Lakes used in this analysis were sampled in the summer (June-September) of 2007 (n = 1,028), 2012 (n = 1,038), or 2017 (n = 1,005). Lakes were selected from the National Hydrography Dataset (NHD, https://www.usgs.gov/national-hydrography/national-hydrography-dataset) using a randomized design stratified on aggregated Omernik level III ecoregion[29] and lake surface area. The minimum surface area for inclusion in the 2007 assessment was 4 ha, but, owing to increasing resolution in the NHD, was reduced to 1 ha for the 2012 and 2017 assessments. Natural lakes and reservoirs were treated equally in the site selection process.

To inform internal quality assurance within a campaign, 10% of the lakes were sampled twice within a field season. Approximately 25% of lakes were targeted for resampling in multiple years to examine temporal change. State, Tribal, Federal, and contractor field crews evaluated lakes on site to ensure that selected lakes met criteria for inclusion in the field campaign (e.g., lake ≥1 m deep). A wide set of measurements were collected at each sampled lake, but we only provide details on the variables used in this analysis. Additional details, protocols, and data are available online (https://www.epa.gov/national-aquatic-resource-surveys/nla).

Total phosphorus and true color were collected and processed in the 2007, 2012, and 2017 field campaigns[24–26]. Water samples were collected from a deep (up to 50 m), open water location in natural lakes and at a midpoint in reservoirs. In natural lakes, field crews were asked to sample in a deep area of the lake regardless of whether the sample location was in the geometric center of the system. In reservoirs, field crews were asked to try to find a midpoint in the system that is reasonably lentic, deep (up to 50 m), and away from the dam. Water was collected from the photic zone using a

vertical, depth integrated sampling device. All water samples were placed on ice and shipped overnight to the Willamette Research Station in Corvallis, Oregon for analysis. True color was estimated by visual comparison of filtered water samples to a calibrated glass color disk[30]. Total phosphorus concentrations were measured with manual alkaline persulfate digestion, followed by automated colorimetric analysis (ammonium molybdate and antimony potassium tartrate under acidic conditions, with absorbance at 880 nm) using a flow injection analyzer following standard method 4500-P-E[31]. Detailed descriptions of all water quality analyses are available in the NLA Laboratory Operations Manuals[24,27,28].

HydroLAKES

HydroLAKES (v1.0)[32] is a compendium of more than 1.4 million lake and reservoir shapefiles globally, with surface area of at least 10 ha. For an individual waterbody, HydroLAKES contains its spatial extent and location (using georeferenced polygons), a unique identifier (ranging from 1 to 1,427,688), and its morphological (area, mean depth, elevation, shoreline length etc.), hydrological (e.g., residence time, discharge, and watershed area), and geographical (e.g., name, country, continent) properties. HydroLAKES is a compilation of existing lake databases, with sources from government agencies (e.g., Natural Resources Canada, U.S. Geological Survey, European Environment Agency) and from remote sensing studies (for example, Shuttle Radar Topographic Mission Water Body Data, Global Lakes and Wetlands Database, and Global Reservoir and Dam Database). Most of the lake polygons are sourced from the Shuttle Radar Topographic Mission Water Body Data for regions between 60ºS and 60ºN[33], supplemented by other datasets for higher latitudes and for underrepresented regions. More detailed information on the creation and validation of the HydroLAKES dataset can be found in Messager et al.[32].

LimnoSat

The LimnoSat-US[34] dataset comprises over 22 million remotely sensed observations of lake surface reflectance from 1984 to 2020. Observations cover 55,662 lakes greater than 10 ha[32] aggregated from Landsat 5, 7, and 8 Collection 1 imagery. Each observation was calculated by taking the median surface reflectance within 120 meters of each lake's Chebyshev center, defined as the point farthest from shore and usually located at the lake's deepest point[35]. Extracting reflectance values from the Chebyshev center minimizes signals due to bottom reflectance and adjacent land pixels. For each Landsat observation, non-high confidence water pixels were masked using the Dynamic Surface Water Extent algorithm[36]. Observations were removed if the scene cloud cover was greater than 75%, any snow, ice, cloud, cloud shadow[37], or hillshadow was detected over the lake's Chebyshev center, or if there were fewer than eight high confidence water pixels within the 120 meter buffer of the lake's Chebyshev center. For certain lakes, these filters lead to extended periods (i.e., months to years) with limited observations (see Figure 2 in Topp et al.[2]). Data in LimnoSat-US are presented in a tabular format, where each row reflects a Landsat overpass for a given waterbody, and columns include median Collection 1 surface reflectance values by band extracted from

pixels within 120 m of the Chebyshev center, scene-wide cloud cover, date of imagery acquisition, and number of water pixels within 120 m of the Chebyshev center.

Step 2: Define Lake Trophic State

Many lakes across the United States are experiencing simultaneous changes in their water clarity, with some lakes getting greener due to eutrophication, and others getting browner from increasing terrestrially-derived organic matter, and some are simultaneously 'greening' and 'browning'[23]. Given the need to discriminate between lakes that may be browning and/or greening, the Nutrient Color Paradigm (NCP) is a useful tool to assign LTS based on a lake's characteristic color.

The NCP was initially proposed in the early 20th century, emphasizing that both autochthonous and allochthonous processes are important to understanding LTS[38–40]. Specifically, water color often affects algal biomass and light transparency independent of nutrient availability. Rodhe[41] first assembled the four quadrants of the NCP, placing autochthony on the horizontal axis and allochthony on the vertical axis. This second dimension distinguishes "oligotrophic" (low nutrient, low color) and "eutrophic" (high nutrient, low color) lakes from "dystrophic" (low nutrient, high color) and "mixotrophic" (high nutrient, high color) lakes.

Although metrics such as Trophic State Index[20] gained popularity for providing instantaneous assessments of a lake's autotrophic production, Williamson et al.[21] encouraged a focus on NCP for lake classification given the importance of both nutrients and colored dissolved organic matter to lake structure and function. The NCP's implementation is empirically supported by studies like Webster et al.[22], where an analysis of ~1,600 temperate lakes in North America demonstrated that within lakes grouped by total phosphorus concentration (i.e., oligotrophic, mesotrophic, or eutrophic), those with 'browner' color (indicative of dissolved organic matter) had higher volumetric chlorophyll-a concentrations and shallower Secchi disk depths. A similar pattern was observed by Nürnberg and Shaw[42], which analyzed 600 lakes spanning a latitude of 39°S to 82°N.

Here, we used the thresholds published in Webster et al.[22] to classify lakes in the NLA dataset. Lakes were described as oligotrophic or 'blue' if total phosphorus concentration was less than 30 µg/L and true color was less than 20 platinum cobalt units (PCU), eutrophic or 'green' if total phosphorus was greater than 30 µg/L and true color was less than 20 PCU, dystrophic or 'brown' if total phosphorus was less than 30 µg/L and true color greater than 20 PCU, and mixotrophic or 'murky' if total phosphorus was greater than 30 µg/L and true color greater than 20 PCU (Figure 1). Thresholds for total phosphorus are based on long established and widely accepted ranges affecting primary productivity[43]. True color thresholds are derived from Nürnberg and Shaw[42]. Eutrophic and mixotrophic classifications were combined into a single grouping due to similar spectral characteristics (see Step 3).

Step 3: Create a training dataset

First, to create a dataset of lakes with *in situ* LTS measurements, we aggregated all total phosphorus and true color measurements from the U.S. EPA NLA 2007, 2012, and 2017 data (Figure S1-2, Table S1). Although the NLA includes lakes smaller than 10 ha, we only used lakes of at least 10 ha in area for consistency with the HydroLAKES database. We then assessed the extent to which seasonal shifts in total phosphorus concentrations and true color values may alter interpretation of trophic state for a given lake using the subset of lakes that were sampled intra-annually. For lakes that were sampled multiple times within a U.S. EPA NLA campaign, we calculated the percentage of lakes that transitioned between trophic states within a single year and found that lakes broadly remained in the same NCP trophic state throughout a given summer (85.1% of lakes). Of the lakes that changed trophic state during a sampling season (14.9%), the majority transitioned from oligotrophic (61.5% of changing lakes; 8.7% of all lakes) or dystrophic (15.4% of changing lakes; 2.2% of all lakes) to eutrophic/mixotrophic. Few lakes transitioned from oligotrophic to dystrophic (15.4% of changing lakes; 2.2% of all lakes), and even fewer transitioned to oligotrophic from either dystrophic (3.9% of changing lakes; 0.5% of all lakes) or eutrophic/mixotrophic (3.9% of changing lakes; 0.5% of all lakes). No lakes transitioned from eutrophic/mixotrophic to dystrophic across all three NLA campaigns. Broadly, lakes transitioned between trophic states when lakes were located near a threshold for trophic state delineation (15-45 µg/L total phosphorus or 11-29 PCU). These results mirror those in Leech et al.[23] and suggest that despite some lakes changing trophic states, the majority of lakes do not transition and those that do transition usually fall along an edge of a NCP-determined trophic state. Thus, for lakes sampled twice in one sampling campaign, we averaged total phosphorus and true color estimates.

Second, to match the *in situ* trophic states with remotely sensed imagery, we merged the complete 2007, 2012, and 2017 NLA dataset with the LimnoSat-US dataset[34], where each NLA lake-year had corresponding Landsat spectral data. Because the NLA is designed to describe lakes' summertime conditions, we filtered LimnoSat-US observances for those only occurring in June, July, and August, which we *a priori* defined as the summertime season for the contiguous U.S.; then, to create a characteristic reflectance for a given lake year, we computed each lake-year's median summertime reflectance for red, blue, green, and near-infrared bands. Because LimnoSat-US compiles reflectance values from Landsat 5, 7, and 8, there are differences in the number of images per lake and year. In particular, images from 1984 through 1998 were solely collected from Landsat 5, when lakes averaged 3.04 images per summer (minimum average images: 2.43 images; maximum average images: 3.64 images). From 1999 through 2012, summertime imagery was gathered from Landsat 5 and 7, when lakes averaged 5.64 images per summer (minimum average images: 3.37 images; maximum average images: 6.42 images). From 2013 through 2019, summertime imagery was collected from Landsat 7 and 8, when lakes averaged 5.42 images per summer (minimum average images: 4.87 images; maximum average images: 5.87 images).

Third, to better characterize spectral bands' relative reflectance, we normalized each lake's median summertime reflectance for the red, green, blue, and near-infrared band by the sum of the summertime reflectance values of all four bands. This normalization allowed us to differentiate lakes by trophic state based on their most prominent reflectances. For example, we anticipated that oligotrophic lakes would be dominated by high blue and green reflectances relative to the red and near-infrared bands. In contrast, dystrophic lakes would be dominated by the near-infrared band relative to green and red bands, because dystrophic lakes tend to have exceptionally low primary productivity and elevated dissolved organic matter. When assessing mixotrophic and eutrophic lakes, spectral characteristics were nearly identical, and to be conservative, we combined mixotrophic and eutrophic lakes into one category 'eutrophic/mixotrophic'. These relative reflectances for all three lake trophic states were ultimately intended to discriminate among lakes that were optically similar in the visible spectrum (i.e., oligotrophic and dystrophic lakes). Notably, the decision to use median summertime relative reflectances differed from previous work[2] that focused on the dominant wavelength, which is an aggregation of wavelengths detected in the visible spectrum and has been used to discriminate autotrophic production (i.e., blue vs green lakes), but not dystrophic states. Thus, our methods are better suited towards discriminating between oligotrophic and dystrophic lakes, because the dominant wavelength approach would consider both of these lake types to be "blue".

Step 4: Create Classification Models

To find an optimal performing classifier for lakes with unknown LTS, we employed three classification methods to predict trophic state: multinomial logistic regression[44], extreme gradient boosting regression[45], and a neural network using multilayer perceptrons[46]. Logistic regression is a parametric classification method, whereas gradient boosted regression and multilayer perceptrons are machine learning methods. The methods differ in how they make classifications. Using trophic state as a categorical response variable, logistic regression applies a linear regression of log-odds ratios to model the probability of a given trophic state for each lake. In contrast, gradient boosted regression applies decision trees to iteratively improve its predictions. Multilayer perceptrons apply a type of feedforward artificial neural network in which a backpropagation algorithm is used to subsequently update the individual weights of each neuron unit by comparing modeled predictions to the training data.

For each modeling method, we used z-scored, relative red, green, blue, and near-infrared reflectances as predictors. Model performance and potential for overfitting were assessed using a 90:10 train:test data split with spatial-holdout cross-validation. Initial hyperparameters for the gradient boosted regression and multilayer perceptron models were tuned by holding out 20% of each trophic class from the training observations to use for validation and conducting a coarse grid-search across the hyperparameter space. For each combination of hyperparameters, models were trained until validation performance did not increase for 20 consecutive epochs using categorical cross entropy as the objective function. During the multilayer perceptron hyperparameter tuning, we iterated through model fits using all combinations of 5, 10, and 20 hidden layers as well

as a learning rate of 0.01, 0.001, and 0.0005. Multilayer perceptron hyperparameter tuning metrics were optimal for models with 20 hidden units and a learning rate of 0.001. During the gradient boosted regression hyperparameter tuning, we iterated through model fits using all combinations of 2, 3, and 4 maximum tree depths, subsample as well as column samples of 0.5 and 0.8, step sizes of 0.01 and 0.1, as well as a minimum child weight of 1 and 3. Gradient boosted regression hyperparameter tuning metrics were optimal for models with a max depth of 4, subsample of 0.5, column sample of 0.5, step size of 0.01, and minimum child weight of 1. For both multilayer perceptron and gradient boosted regression models, best performing hyperparameter tuning metrics were assessed by having lowest validation loss values.

These hyperparameters were then used in a spatial cross-validation routine[47], where a given lake was held out as test data if it was included in the training data. During the spatial cross-validation routine, training data were divided into five folds, such that lakes within each test partition were not present in remaining training partitions (i.e., test metrics represent performance on unseen lakes). Training data within each fold were then partitioned into a 90:10 split with 10% of each trophic class set aside for an inner-loop fold validation. Models within each fold were trained using an early stopping criteria of 20 epochs to avoid overfitting on the training data. This inner-fold validation was additionally used to hypertune the best number of epochs for the final models. Finally, overall error metrics were calculated based on the mean prediction accuracy of the test partitions withheld from the inner-loop training of each fold. All reported metrics are based on the test partitions from the spatial cross-validation routine while final models were trained on the full dataset using the hyperparameters identified from the grid-search and inner-loop validation routines. We applied the final models to make predictions for all 55,662 lakes in the LimnoSat-US dataset.

Step 5: Assess and Compare Model Performance

To evaluate the final fitted models, we used test data predictions from the spatial-holdout routine to calculate each model's overall and balanced accuracy, receiver-operator-characteristic (ROC) curves, as well as the area under the curve (AUC) of the ROC curve. Overall accuracy was calculated as the sum of true positives and true negatives divided by the total number of LTS predictions. Balanced accuracy was calculated as the sum of a true positive and true negative results for a single lake trophic state. Whereas overall accuracy can be biased towards more prevalent trophic states (i.e., eutrophic and oligotrophic lakes), balanced accuracy is useful to assess a model's capacity to predict more rare trophic states (i.e., dystrophic lakes). As an additional metric of model performance, we calculated the AUC of each model's ROC curve. The ROC curve visually graphs the relationship between the rate of a correct classification with the rate of a false classification. An AUC of 0.5 indicates a false prediction rate increases 1:1 with the rate of a correct prediction. AUCs greater than 0.5 imply a model performing better than random, even when a false positive rate is artificially inflated. Thus, comparing overall and balanced accuracy as well as ROC curves and AUCs allowed us to assess how models performed broadly as well as how robustly models predicted trophic state correctly.

Beyond model performance, we also evaluated whether model coefficients and variable importance for trophic state discrimination reflected NCP groupings. For increased interpretability across all three models, we employed SHAP (SHapley Additive exPlanation) analysis[48–50] to better understand individual feature importance and influence in model predictions. SHAP analysis yields insight into the marginal contribution of a given feature (e.g., near-infrared spectra) on model output - in this case trophic state - and helps decode 'black box' results. Understanding the relative contribution of individual features in trophic state prediction not only helps explain feature roles in model accuracy and misclassification but also quantitatively connects features, such as remotely sensed data, to the biophysical parameters in which LTS prediction is grounded. SHAP feature contribution was calculated for blue, green, red, and near-infrared Landsat spectra. SHAP feature contribution was scored for oligotrophic, dystrophic, and eutrophic/mixotrophic classifications and across each of the three models. This scoring illuminates the relationship among feature values and SHAP contribution for a given trophic state classification and for a given model. Specifically, for classification problems, a positive SHAP value indicates that a given input contributed to a positive classification and a negative value indicates the input contributed to a low probability for a given classification.

Data Records

The LTS-US dataset[51] is structured in a tabular format, where each row is a lake-year combination. The main dataset is contained in "ensemble_predictions.csv" and is structured in a way that provides both categorical LTS predictions as well as probabilities for each LTS prediction. The probabilities reported in "ensemble_predictions.csv" are averaged probabilities generated from each of the three modeling methodologies. An additional tabular dataset ("individual_predictions.csv") contains probabilities generated for each of the three modeling methodologies and can be merged with "*ensemble_predictions*.csv" by the "Hylak_id" and "year" columns.

We provide raw and average predicted LTS probabilities as well as variance among models for a given LTS prediction to allow future users to filter predictions of a certain threshold for their particular analysis. Although many thresholds may exist, reporting probability thresholds used in subsequent analyses will help maintain reproducibility and synthesis across studies. Below, we detail column names and metadata for each of the core datasets contained within the LTS-US data product.

"ensemble_predictions.csv"

Hylak_id
HydroLAKES unique identifier of lake. Preserved from HydroLAKES input data to enable future merging with HydroLAKES attributes.

year
Year, spans 1984 through 2020.

categorical_ts
Categorical predicted lake trophic state (i.e., oligo, eu/mixo, dys). Categorical prediction is based on the highest probability among *mean_prob_dys*, *mean_prob_eumixo*, and *mean_prob_oligo*.

mean_prob_dys
Probability that a lake-year combination is dystrophic. Probability is calculated by averaging probabilities from all three modeling methods.

mean_prob_eumixo
Probability that a lake-year combination is eutrophic or mixotrophic. Probability is calculated by averaging probabilities from all three modeling methods.

mean_prob_oligo
Probability that a lake-year combination is oligotrophic. Probability is calculated by averaging probabilities from all three modeling methods.

var_prob_dys
Variance in probabilities among all three modeling methods that a given lake-year is dystrophic.

var_prob_eumixo
Variance in probabilities among all three modeling methods that a given lake-year is eutrophic/mixotrophic.

var_prob_oligo
Variance in probabilities among all three modeling methods that a given lake-year is oligotrophic.

"individual_predictions.csv"

Hylak_id
HydroLAKES unique identifier of lake. Preserved from HydroLAKES input data to enable future merge with HydroLAKES attributes.

year
Year, spans 1984 through 2020.

prob_dys_mlr
Probability that a lake-year combination is dystrophic. Probability is calculated by multinomial, multiple logistic regression.

prob_eumixo_mlr
Probability that a lake-year combination is eutrophic or mixotrophic. Probability is calculated by multinomial, multiple logistic regression.

prob_oligo_mlr
Probability that a lake-year combination is oligotrophic. Probability is calculated by multinomial, multiple logistic regression.

prob_dys_mlp

Probability that a lake-year combination is dystrophic. Probability is calculated by multilayer perceptron.

prob_eumixo_mlp
Probability that a lake-year combination is eutrophic or mixotrophic. Probability is calculated by multilayer perceptron.

prob_oligo_mlp
Probability that a lake-year combination is oligotrophic. Probability is calculated by multilayer perceptron.

prob_dys_xgb
Probability that a lake-year combination is dystrophic. Probability is calculated by a gradient-boosted regression.

prob_eumixo_xgb
Probability that a lake-year combination is eutrophic or mixotrophic. Probability is calculated by a gradient-boosted regression.

prob_oligo_xgb
Probability that a lake-year combination is oligotrophic. Probability is calculated by a gradient-boosted regression.

Technical Validation

Model performance diagnostics

To assess how each model correctly classified training data, we compared the model accuracies, balanced accuracies, and AUC of ROC curves. Overall and balanced model accuracies were similar, where all models had accuracies ranging from 72.4 to 72.9% and balanced accuracies ranging from 69.9 to 71.5%. AUCs of ROCs were likewise similar across all three model techniques, ranging from 0.88 to 0.90 (Figure S3). These combined metrics suggest that all three modeling approaches performed similarly, when assessing model performance with global metrics.

Although models performed similarly at high levels, they varied more in their robustness to classify dystrophic lakes (Figure 3). Machine learning-based methods, such as multilayer perceptron (60%) and gradient boosted regression (58%), had higher balanced accuracies, whereas distribution-based methods, such as logistic regression (55%), had lower balanced accuracies. These differences were largely driven by deviations in true positive rates (47.5-50.6%), whereas true negative rates were higher (91.8-92.7%). This difference in true negative and true positive rates is likely due to spectral similarities between oligotrophic and dystrophic lakes, where both are characterized by low primary production in comparison to eutrophic/mixotrophic lakes. Although these differences only span 5%, they may be important, given that dystrophic lakes tend to be uncommon relative to oligotrophic and eutrophic lakes[23]. Such differences imply variation in each model's robustness to predict rarer trophic states, but

our overall metrics of model performance highlight exceptional congruence across all three modeling techniques.

Spatial patterns in lake classification

To evaluate spatio-temporal patterns in trophic state classification, we created spatial confusion matrices, where predictions and reference sites were plotted across the entire United States. We *a priori* hypothesized that when misclassifications result from lake-specific deviations, misclassifications should be distributed throughout the United States without any clear spatial patterns. In the event that spatial clustering of misclassified lakes occurred, these patterns should be more pronounced where high densities of a given lake trophic state are located. In cases when lake clustering appears in an unexpected area, these patterns should be more attributed to place-based irregularities in spectral data.

Confusion between oligotrophic and eutrophic/mixotrophic lakes were spatially distributed throughout the entire continental United States, with no evidence of spatial clustering (Figures S4-S6). In contrast, dystrophic misclassifications were broadly isolated to the Upper Midwest and Upper Northeast regions. Consistent with our hypotheses, these regions are associated with increased densities of dystrophic lakes, suggesting that optical similarities between oligotrophic and dystrophic lakes in these regions may lead to increased misclassification. Notably, dystrophic lakes tended to be misclassified as oligotrophic, whereas oligotrophic lakes tended to not be misclassified as dystrophic, meaning that our predictions should be conservative with assigning an individual lake as dystrophic.

Assessing patterns in lake classification

Given that lake trophic state classifications may be a product of a lake's limnological, morphological, and geographic properties, we performed a series of analyses of variance (ANOVA) to test for significant differences (i.e., p-value < 0.05) in lake classification accuracy. For each ANOVA, a lake property was the response variable, and predictors were lake trophic state, model correctness (i.e., correct or incorrect classification), and model type. All response variables were log-transformed to approximate a normal distribution. Because each analysis had an unbalanced sample size, we calculated Type II Sum-of-Squares[52]. Residuals for each model were assessed for normality and homogeneity of variance.

The main goal of each ANOVA was to assess whether variation in a lake parameter could be associated with variation in model methodologies, model correctness, or trophic states themselves. Consequently, our ANOVAs do not include interaction terms, as most interactions would not be helpful for understanding patterns in how our classification models performed.

NCP patterns in lake classification

To assess how a lake's misclassification may be related to its position in the NCP, we assessed where correctly and incorrectly classified lakes were located in the NCP. Lakes that were incorrectly classified tended to be located near total phosphorus (30 ± 15 µg/L) and color (20 ± 9 PCU) thresholds, with a large portion at the nexus of the total phosphorus and color thresholds (Figure 4). Across all modeling techniques, correctly classified lakes spanned a wider range across both axes, especially total phosphorus. Median total phosphorus concentration for misclassified lakes was 24 µg/L (range: 1-4,772 µg/L), whereas median total phosphorus concentration for correctly classified lakes was 36 µg/L (range: 0.24-4,144 µg/L). Similarly, median PCU for correctly (14 PCU; range: 0-724 PCU) and incorrectly (16 PCU; range: 0-350 PCU) classified lakes were along the edge of the color threshold of 20 PCU. When assessing total phosphorus and color independently, ANOVA suggested that total phosphorus concentrations were significantly different for correctly and incorrectly classified lakes, whereas differences in color were not significantly different across correctly and incorrectly classified lakes (Table 1; Figure 5).

Beyond total phosphorus and color patterns influencing lake classification, our analyses of lakes that transitioned trophic states within a summer suggest that lakes along a NCP boundary (i.e., near total phosphorus or color threshold) are more prone to misclassification. Among lakes that transitioned within a summer, the most frequent change in lake trophic state was among lakes switching from oligotrophic to eutrophic. Considering that both total phosphorus concentrations as well as summertime lake phenologies are associated with algal production that can optically obfuscate whether a lake is oligotrophic or eutrophic, our results of NCP patterns are not surprising. Rather, confusion along the total phosphorus axis of the NCP, an axis that corresponds with autotrophic productivity, is concordant with the idea that a lake can experience moments of eutrophy - e.g., a pulse of nutrients or algal growth - while otherwise being oligotrophic for the majority of the summer. Therefore, classifications made for lakes at the boundary of trophic states can be challenging, and our validation analyses describe total phosphorus and color conditions where misclassifications may be more common.

Morphological and locational patterns in lake classification

At the spatial resolution of Landsat's sensors, there is a risk of "mixed pixels", where a pixel includes water with fractions of adjacent bare land or vegetation. Given the difference in optical contrast between water and other features, even minor differences can lead to large errors in estimating surface reflectance. A major source of uncertainty in lake optical water quality estimation is the separation of water and atmospheric effects[53]. The latter increases in severity near land and this adjacency effect can extend several kilometers, depending on the state of the atmosphere.

Before assessing how edge and lakebed effects may influence model classifications, we first ensured that spectral differences between each trophic state in our dataset were greater than differences within a trophic state when accounting for lake area and depth. To evaluate how edge and lakebed effects may be present within our training and test data, we used lake area and average depth data from HydroLAKES[32] as well as

maximum depth from the GloBathy dataset[54]. While evaluating lake area, we noticed that smaller lakes tended to have higher near-infrared relative reflectance values, and relative near-infrared reflectance generally decreased with increasing lake area (Figures S7-S8). Because LimnoSat-US aggregates reflectance data at the lake's Chebyshev center, the point in the lake farthest away from shore, smaller lakes would likely have Chebyshev centers that are closer to the shoreline. As terrestrial near-infrared reflectances tend to be higher than aquatic near-infrared reflectances, smaller lakes with Chebyshev centers closer to the shoreline may be associated with increased near-infrared signatures. Similarly, relative blue reflectance increased with increasing lake surface area, which would likewise be expected, as larger lakes likely have a Chebyshev center that is farther from shore, and therefore, less influenced by shoreline effects. With respect to lakebed effects, the shallowest lakes tended to have slightly elevated relative green reflectance, which would be consistent with increased primary production. Across all trophic states, lakes with average depths of 1-10 m were also associated with increased relative near-infrared reflectance, suggesting that these lakes may have the highest near-infrared reflectance due to reflectance signatures of lakebed substrate or increased benthic algal production.

To evaluate how models might misclassify lakes in response to morphological and geographic characteristics, we examined how lake depth, elevation, and surface area may correspond to correct and incorrect classifications. Average and maximum lake depth can be used to evaluate a lake's potential for lakebed effects, where reflectance from benthic algae or sediment may confound signals for the actual surface of the lake. Assessing classification differences across elevation ranges can be important for understanding atmospheric effects on reflectance data, where higher elevations may have fewer aerosols, and therefore contain fewer misclassifications. Lastly, examining misclassifications across lake sizes can reveal potential for adjacency effects, where surrounding geologies or vegetation may obscure surface reflectances observed over the lake.

ANOVA results suggested that average depth, maximum depth, and elevation differed significantly across correct and incorrect misclassifications (Table 2), although differences based on average and maximum depth were more visually apparent than those observed for elevation (Figure 6). In contrast, lake area did not differ significantly across correct and incorrect classifications (Table 2).

Together, these analyses suggest that lakebed reflectance may lead to lake trophic state misclassification, whereas edge effects are likely not consequential for inaccurate lake trophic state classifications. In particular, shallower oligotrophic lakes (i.e., average depth < 5 m and maximum depth < 15 m; Figure 6) and deeper eutrophic lakes (i.e., average depth > 5 m and maximum depth > 15 m; Figure 6) tended to be misclassified. We speculate that these differences may stem from shallower, oligotrophic lakes having pronounced benthic algal growth[55] that can produce a strong green signal. Conversely, deeper eutrophic lakes may have less concentrated algal growth in the water column, thereby creating a stronger blue reflectance relative to green reflectance and increasing chances for misclassification (see *Optical patterns in lake classification*). Overall, these results correspond with our NCP validation analyses, where total phosphorus

concentrations were associated with greater misclassifications of oligotrophic lakes as eutrophic. Given the potential for lakes to be misclassified because of issues with lakebed reflectance, considering whether depth could alter results and building analytical workflows to assess sensitivity to interference from lakebed reflectance (see *SHAP Analysis* for more detail) could improve model lake classifications.

Optical patterns in lake classification

To evaluate how models might misclassify lakes based on reflectance values, we assessed how z-scored relative red, green, blue, and near-infrared reflectance values differed between correctly and incorrectly predicted lake trophic state. Because we used relative reflectances that are inherently interdependent, and thus violate ANOVA assumptions, we elected to forgo significance tests for whether band ranges differed across modeling methods.

For dystrophic lakes, incorrectly classified lakes, compared to correctly classified lakes, tended to have lower z-scored near-infrared and blue band values as well as higher green and red values (Figure 7). For eutrophic/mixotrophic lakes, misclassified lakes tended to have lower values for red and green bands as well as higher values for blue bands (Figure 7). For oligotrophic lakes, incorrectly classified lakes tended to have higher red and lower blue band values (Figure 7).

These inconsistencies in LTS classification correspond with variation that can be present in natural systems. Dystrophic lakes are generally characterized as having low primary productivity and high dissolved organic matter, which should result in low green band values as well as higher near-infrared values, yet misclassified dystrophic lakes tended to have low near-infrared as well as high red and green bands. Eutrophic and mixotrophic lakes are generally characterized as having high productivity, which should result in high green values, yet misclassified eutrophic and mixotrophic lakes tended to have low green and red as well as high blue bands. Oligotrophic lakes should be characterized as having high blue bands, yet misclassified lakes tended to have low blue and high red bands, which may be a product of bottom reflectance. Together, these misclassifications likely represent lakes that are not characteristic of LTS classifications. For example, a more productive oligotrophic lake could produce a stronger red and green signature and, therefore, be classified as eutrophic. Likewise, less productive eutrophic lakes may be optically more similar to oligotrophic lakes and, therefore, be characterized by lower red and green bands.

SHAP Analysis

To evaluate the influence of remote sensing reflectance inputs on final predictions, we assessed the distribution of SHAP values calculated for each predictor and for each trophic state. In general, SHAP values can be useful for decoding how machine learning and parametric methods may assign relative importance to a given predictor, thereby increasing interpretability of a model. In an instance where models are classifying lakes based on *a priori* hypothesized relationships, SHAP values across predictors should

correspond to the *a priori* hypothesized relationships. For example, oligotrophic lakes are generally characterized as having high blue reflectance relative to red and green, and in a case where models reflect this understanding, SHAP values should attribute an oligotrophic classification to high values in the relative blue reflectance. Consistently high attributions for blue reflectances should subsequently result in high overall feature importance when discriminating oligotrophic lakes.

When evaluating feature importance across trophic states, measured as the mean absolute SHAP value of a given feature, all models agreed on the most influential features for classification (Figure S9). Furthermore, the distribution of SHAP values reflected limnological understanding of each trophic state's inherent properties. For dystrophic lakes, SHAP values indicate that models relied on low green and high near-infrared and red band values, corroborating the idea that dystrophic lakes should have lower primary production and increased CDOM[21,56] (Figure S9). Predictions for eutrophic and mixotrophic lakes were attributed to high red and low blue band values, corresponding with the idea that eutrophic and mixotrophic lakes should have higher algal production[23] (Figure S9). Conversely, SHAP values for oligotrophic lakes attributed predictions to low red and high blue band values, agreeing with the idea that oligotrophic lakes should have lower algal production[23] (Figure S9). Beyond each individual trophic state's most important predictors, our SHAP analysis mirrored the logic of NCP analyses, where lakes with lower true color values (i.e., oligotrophic and eutrophic) were discriminated more effectively by bands associated with autotrophic capacity, whereas lakes with higher true color values (i.e., dystrophic) were discriminated more effectively by bands suggesting decreased autotrophic production and increased colored dissolved organic carbon.

SHAP values can also provide insight on what drives models to misclassify certain lakes. Specifically, when examining smaller, shallower oligotrophic lakes that could potentially be influenced by bottom reflectance or adjacency effects, we observed that some misclassifications were attributable to models relying on low relative blue reflectance and high relative near-infrared reflectance (Figures S10-S15). These patterns indicate that in certain lakes, the models were unable to distinguish the spectral signatures that are potentially attributable to sediment or benthic algae as well as shoreline vegetation and soil. The spectral similarity between shallow oligotrophic and deep eutrophic lakes is relevant to active research trajectories in limnology, particularly those examining the relatively high contributions of benthic algal communities to whole lake productivity in oligotrophic lakes[55,57–60]. Given the potential for lakebed effects to alter classifications, research questions could consider the influence of depth-related misclassifications.

Comparing predicted and NLA spatial patterns

To independently validate the LTS-US dataset's robustness in capturing macroscale and multi-year changes in lake trophic state, we replicated analyses similar to Leech et al.[23] and compared statistics from the NLA with those from the LTS-US dataset. We first merged the lake trophic state classifications from the 2007, 2012, and 2017 NLA

campaigns as well as the LTS-US dataset with the U.S. EPA Level I Ecoregions[29]. We then calculated the proportion of each trophic state occurring within each ecoregion in a given year. To compare the NLA and the LTS-US dataset, we calculated the absolute difference between predicted and estimated proportions for each trophic state within each year and ecoregion.

Predicted and measured proportions were broadly consistent across all three years. Visually, all three years and trophic states followed consistent trends across all ecoregions (Figure 8). For example, our models generally captured increasing dystrophic and decreasing oligotrophic lakes in northern forested regions, a pattern consistent with Leech et al.[23]. Absolute differences between estimated and predicted proportions across ecoregions were likewise congruent across all three years. Eutrophic/mixotrophic lakes tended to have the smallest differences (mean = -5.3%, sd = 19%), indicating that our models may overestimate relative abundance of eutrophic and mixotrophic lakes (Figure S16). In contrast, dystrophic (mean = 7.0%, sd = 6.7%) and oligotrophic (mean = 7.6%, sd = 22.4%) relative abundance tended to be underestimated (Figure S16). Larger standard deviation values were caused by some ecoregions having few lakes overall, thereby increasing proportions of a given trophic state within an ecoregion. When filtering for ecoregions that contained at least 10 lakes, we noticed similar patterns of eutrophic and mixotrophic lakes being slightly overestimated (mean = -7.9%, sd = 11.3%), as well as dystrophic (mean = 9.1%, sd = 6.7%) and oligotrophic (mean = 4.6%, sd = 13.8%) lakes being underestimated; yet the standard deviation in absolute differences decreased.

Together, these analyses demonstrate that though the LTS-US dataset does contain biases towards eutrophic/mixotrophic classification, its overall congruence with the NLA highlights its robustness. These biases may stem from our models attempting to classify lake ecosystem properties based on optically visible (i.e., color) and optically invisible (i.e., phosphorus) properties, where the exceptionally oligotrophic, dystrophic, and eutrophic/mixotrophic lakes are more consistently discriminated. In contrast, the NLA may likewise contain biases due to site selection, whereas our methods select for all lakes of at least 10 ha in area. Regardless of the biases in the LTS-US and NLA datasets, the congruence between the two is even more notable considering that our modeling approaches and the NLA use independent methods for classifying lake trophic state. The NLA uses *in situ* total phosphorus and true color measurements, whereas our methods use lake red, green, blue, and near-infrared reflectance. Furthermore, despite not including temporal or spatial predictors, our models reproduce NLA spatial and temporal trends in lake trophic state at the spatial and temporal scales.

Given both the potential biases and robustness of the LTS-US data product, cross-referencing the LTS-US dataset with known trends in an area of interest, especially in areas where lakes may be less abundant could enhance regional and local analyses. In instances where the LTS-US dataset may be more biased, reproducing the LTS-US dataset using both our existing code and particular predictors of interest for a region, such as average depth, lake area, or watershed area could offer particular insights into why a given region may be more prone to misclassificaitons. Creating tailored versions

of the core LTS-US dataset can promote further understanding of multi-scale features that may be important for assessing lake trophic state with remotely sensed surface reflectance data.

Manual Quality Control

To ensure integrity of lake classifications across all steps of our pipeline, we subsampled 250 lakes from the final dataset and manually cross-referenced their predicted trophic state with independent sources. The random subsample only included lakes that had associated names in the HydroLAKES dataset and was stratified by lake surface area, where surface areas were binned by orders of magnitude (i.e., < 1 km^2, (1, 10] km^2, (10, 100] km^2, (100, 1,000] km^2, (1,000, 10,000] km^2, $> 10{,}000$ km^2). We filtered specifically for lakes with names because we assumed that named lakes within the HydroLAKES database would likely have more publicly available information about their water quality and would likely be easier to find within managerial reports and scientific publications.

To minimize bias, persons engaged in manual checking only received lake latitude and longitude, name, and the U.S. state where the lake was located. All persons engaged in manual checking were not involved in model and prediction development and were, therefore, blind to individual lake predictions. When possible, persons identified trophic states for multiple years, although many sources only referenced a lake's trophic state in an individual year or broadly across multiple years. In either case, LTS was reported for the lake and years that independent sources reported.

Of the 250 target lakes, we were able to find verified trophic state data on 93 lakes (38%). For the 93 lakes that had independent lake trophic state data, our models corroborated independent, *in situ* observations 74% of the time, which is consistent with our models' overall accuracy against testing data from the U.S. EPA NLA. We did not observe any apparent spatial patterns with model misclassification, which complements our spatial confusion validation (Figure S17). Together, these results demonstrate that our manual checking procedure returned similar results as our evaluation procedures against our testing data, giving confidence that our modeling pipeline and evaluation procedures are both robust and able to capture natural processes occurring in lakes.

Effects of processor heterogeneity

When recreating lake trophic state predictions *de novo*, care should be taken to ensure that effects from heterogeneous processors are minimized. When creating the LTS-US dataset from the original LimnoSat-US dataset[34], we specified seeds for each modeling framework, which enabled us to reproduce results between model runs. Final dataset production occurred on one machine using an Intel(R) Xeon(R) W-10885M processor with eight cores, however, slight differences may arise due to differences in a user's hardware float precision.

If users recreate or update LimnoSat-US prior to recreation of the LTS-US predictions, care should be taken as Google Earth Engine[61] uses a heterogeneous processor framework, where individual processors cannot currently be specified. Meyer et al.[9] quantified the effect of Google Earth Engine's processor heterogeneity on various lake surface area and basin-level climatological estimations, and effects of processor heterogeneity were likely inconsequential (e.g., differences of 10^{-12}), although these differences may result in slightly different trophic state predictions. The extent to which these values would influence results or conclusions of other studies will depend on the level of precision required and scope of research question.

Usage Notes

The LTS-US dataset was constructed to be an accessible and interoperable product for a range of basic and applied research questions related to water quality and ecological integrity at national scales. Here, we detail several options for application of the LTS-US dataset and associated pipeline.

First, the LTS-US dataset can be joined with water quantity and quality datasets to assess how changes in LTS, and therefore ecosystem integrity, may be influenced by watershed processes, climate, and human population. At the local scale, the LTS-US dataset can be merged with *in situ* sampling data or modeled data from individual lakes to assess how hydrodynamic, climatic, physicochemical, and biological processes may be associated with interannual variation in LTS. As demonstrated here, local *in situ* observations are important for providing validation of the LTS data, and potentially, refinement of methods for deriving LTS predictions. Similarly, the LTS-US dataset can be merged with data from research coordination networks, such as the National Ecological Observatory Network (www.neonscience.org) or the Global Lake Ecological Observatory Network[62], to enable upscaling highly localized processes to regional and national scales. Beyond watershed-specific processes, the LTS-US dataset can likewise be useful for synthetic questions focused on macroscale water quality trends. For example, in cases where users may wish to synthesize changes in lake ecosystem metabolism with trends in lake water quantity, climate, and human population, the LTS-US dataset can be merged with the GLCP (Global Lake area, Climate, and Population)[9] or LakeATLAS[63], thereby enabling users to assess how changes in seasonal and permanent lake surface area may correlate with changes in lake trophic state. The LTS-US dataset offers a valuable resource for addressing a broad spectrum of basic and applied research questions from local and regional to continental scales.

Second, the LTS-US dataset provides a tool for using remote sensing products with the NCP, a framework increasingly used by limnologists, to understand lake water quality at macroscales. Although previous studies have remotely sensed lake trophic state index[64], our data product is the first to incorporate NCP with remote sensing reflectance data. Where TSI focuses exclusively on eutrophication patterns (also known as greening) associated with nutrient-driven primary production, the LTS-US dataset enables investigations of the spatial extent and temporal trends of lake dystrophication (also known as lake browning). This difference between TSI and NCP is important for

assessing long term and spatially extensive changes in lake browning, as well as "murkification" (i.e. simultaneous browning and greening), which has been associated with complex, often non-linear changes in temperature, pH, dissolved oxygen, and food web structure[23,56]. Further, national-scale sampling campaigns, such as the U.S. EPA NLA, have helped reveal that the proportion of dystrophic lakes has been increasing nationally since 2007[23]. The U.S. EPA NLA is one of the most extensive, structured, and coordinated lake sampling efforts at the national scale, and the LTS-US dataset can complement these *in situ* data by providing finer temporal information at comparable spatial scales. When data from successive NLA sampling campaigns become available, the LTS-US dataset can be updated and further benefit from additional training data. Together, the use of remote sensing imagery with extensive sampling campaigns, like the NLA, can be useful for identifying broadscale changes in water quality.

Third, although our reflectance data are spatially aggregated to represent each lake's characteristic summertime reflectances, our data pipeline and modeling frameworks are amenable to numerous data aggregations, thereby enabling investigation of lakes' intra- and inter-annual phenologies. For example, many oligotrophic lakes experience summertime greening, due to increased algal growth throughout the summer. Although algal succession tends to follow similar temporal and community compositional patterns[65,66], users may be interested in understanding how greening events may shift temporally in response to climatic and anthropogenic disturbances. Similarly, end users may be interested in understanding intra-lake heterogeneities, where embayments or nearshore areas may differ in trophic state from the offshore. In both cases, users could adapt our data, modeling, and validation pipeline, where temporal and spatial resolution are more finely resolved. Operationally, end users could modify the aggregation scripts ("1_aggregate.R" and "aggregate_utils.R")[51] and LimnoSat codes[34] to accommodate input data that aggregate at monthly or fortnightly timesteps as well as on a per-pixel basis. Because our data pipeline allows for automated re-running of all harmonization, modeling, and quality control routines, users are able to build off of the existing infrastructure to tailor the LTS-US dataset to their particular research questions without high computational overhead or the need to build new workflows *de novo*.

Beyond any specific research question, the LTS-US dataset is a streamlined resource for many end users looking to incorporate remote sensing and its derived products into their analyses. Because of the dataset's interoperability and flexible structure, the LTS-US dataset serves as a powerful resource for evaluating and contextualizing aquatic ecosystem change at local-to-national spatial as well as annual-to-decadal temporal scales.

Code Availability

All data harmonization, modeling, and validation procedures for the LTS-US dataset were scripted in the R Statistical Environment[67], using the tidyverse[68], lubridate[69], data.table[70], sf[71], keras[72], tensorflow[73], caret[74], CAST[75], yaml[76], reticulate[77], xgboost[78], nnet[44], viridis[79], trend[80], multiROC[81], ggpubr[82], fastshap[83], maps[84], ggtext[85], and ggforce[86] packages.

To enhance reproducibility, all scripts are designed to work within a single pipeline that uses the targets package[87]. The targets pipeline is divided into four main components: “1_aggregate”, “2_train”, “3_predict”, and “4_qc”. Each component corresponds to one of the steps presented above and can be customized by users to fit their specific needs. The associated pipeline setup and user guide can be found on the Environmental Data Initiative[51], where the “README_targets.pdf” file details directory architecture and how to execute the pipeline.

To ensure reproducibility across operating platforms, all scripts for the pipeline can be executed within a container. Running the pipeline within the container allows users to execute the entire pipeline without the need to make small, yet important, edits to the code, or to configure their own operating environment to conform to the pipeline’s requirements. For example, recent versions of the sf package default to using the s2 spherical geometry engine instead of the Graphic Environment Operating System (GEOS), which assumes planar coordinates. End users on a system with one version of the sf library might need to adjust the code to use the correct geometry engine, whereas users with another version might be able to run the pipeline without any adjustments. The container crystallizes a known-working set of libraries, both at the system level (e.g. GEOS, GDAL, PROJ) and at the R level (e.g. sf), so that anybody can run the code without reconfiguring their own environment. This also provides future proofing by ensuring that the inevitable changes to other libraries over time do not lead to errors. To help end users, who are less familiar with running containerized code, a tutorial for installing and executing the pipeline within the container is located in the Environmental Data Initiative repository as a compressed entity (see “README_container.pdf”)[51].

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Author Contributions

MFM, SNT, TVK, JRE, and MRVR conceived the idea of the manuscript. MFM, TVK, SEH, and DML designed the manuscript. MFM provided leadership for the project and also performed all data harmonization. MFM, SNT, RL, JCR, and XY contributed to model development. MFM and SNT performed high-level validation checks for the data and models. TVK, RL, RMP, JRE, and JR conducted manual quality control. IAO, JCR, MRVR, RIW, and MRB reproduced coding routines. MFM, SNT, RMP, HAD, IAO, and RIW drafted figures and/or tables. MFM, SNT, RL, RMP, HAD, SEH, DML, IAO, JCR, RIW, XY, KCF, JCP, and AIP wrote original parts of the manuscript. All authors performed critical review and editing of the manuscript. All authors read and approved the final manuscript.

Competing Interests

The authors declare no competing interests.

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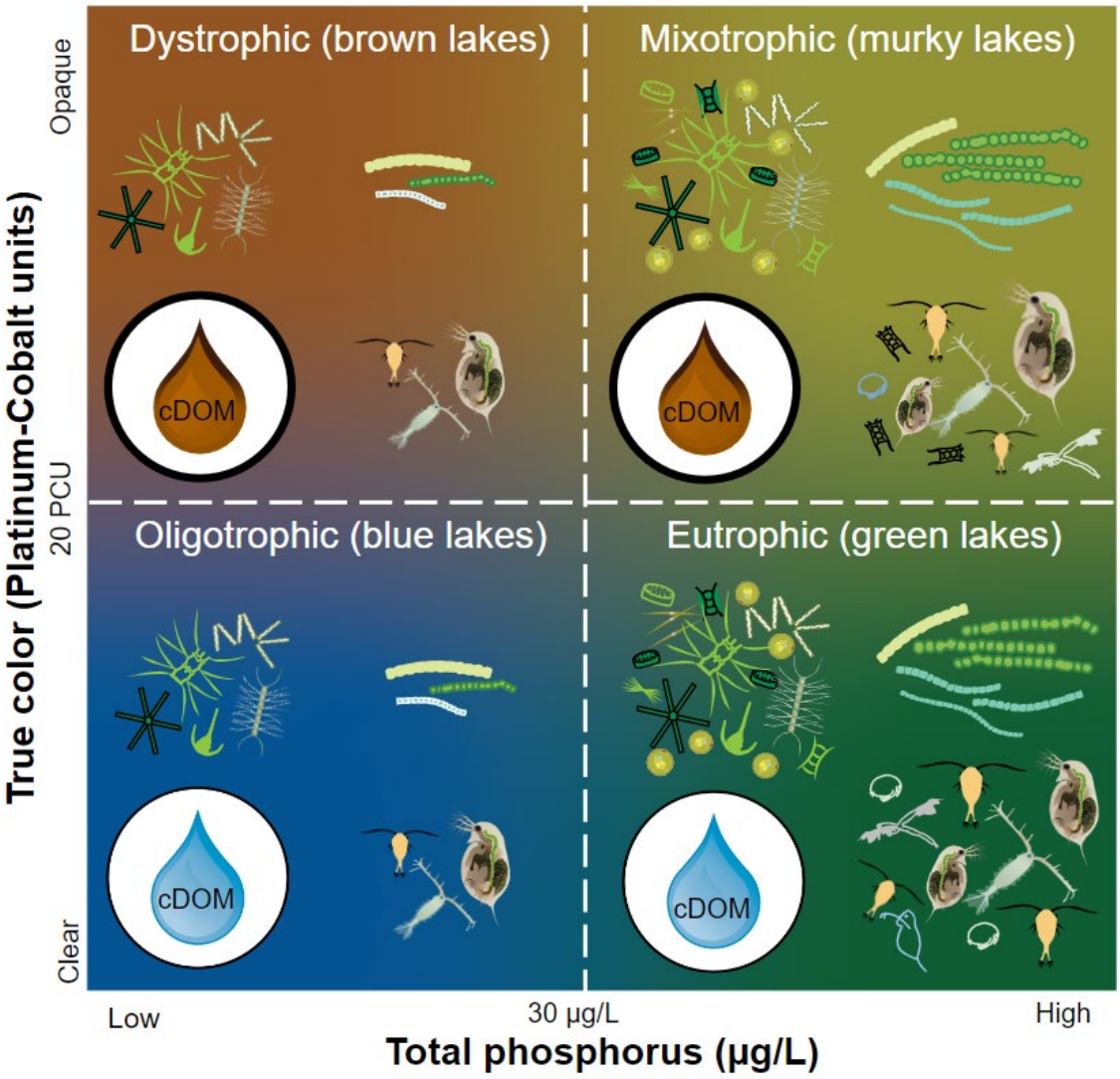

Figure 1: Nutrient-Color Paradigm (NCP) scheme for classifying oligotrophic, eutrophic, dystrophic, and mixotrophic lakes. Schematic is adapted from Williamson et al.[21] and Webster et al.[22], and characteristic lake descriptions broadly stem from results presented in Leech et al.[23] and Oleksy et al.[88] Within each NCP-quadrant, there are a suite of physical, chemical, and biological characteristics that distinguish each type of lake: colored Dissolved Organic Matter (cDOM), primary production, cyanobacterial abundance, and higher order production. Lower cDOM concentrations (blue water drops) are characteristic in oligotrophic and eutrophic lakes. When cDOM is low, light can transmit through the water column more deeply, allowing for primary producers to undergo photosynthesis and zooplankton to consume primary producers (oligotrophic). When nutrients, such as phosphorus, are at higher concentrations and cDOM is low (eutrophic), primary production, cyanobacterial abundance, and higher order production can all increase, resulting in increased biomass. When cDOM concentrations are high (brown water drop) and nutrient concentrations are low (dystrophic), the increased light attenuation can result in decreased primary production, which can reciprocally cause decreased higher order production. Lastly, when nutrient and cDOM concentrations are

both high (mixotrophic), primary production, cyanobacterial abundance, and higher order production can exceed values observed when solely cDOM or nutrient concentrations alone are higher. Phytoplankton and filled-in zooplankton cartoons were downloaded from the University of Maryland Center for Environmental Science Integration and Application Network (https://ian.umces.edu/media-library/). Phytoplankton were designed by Tracey Saxby of the Integration and Application Network, Dieter Tracey of the Water and Rivers Commission, Kim Kraeer and Lucy Van Essen-Fishman of the Integration and Application Network. Transparent crustacean zooplankton and rotifer cartoons were drawn by Stephanie E. Hampton.

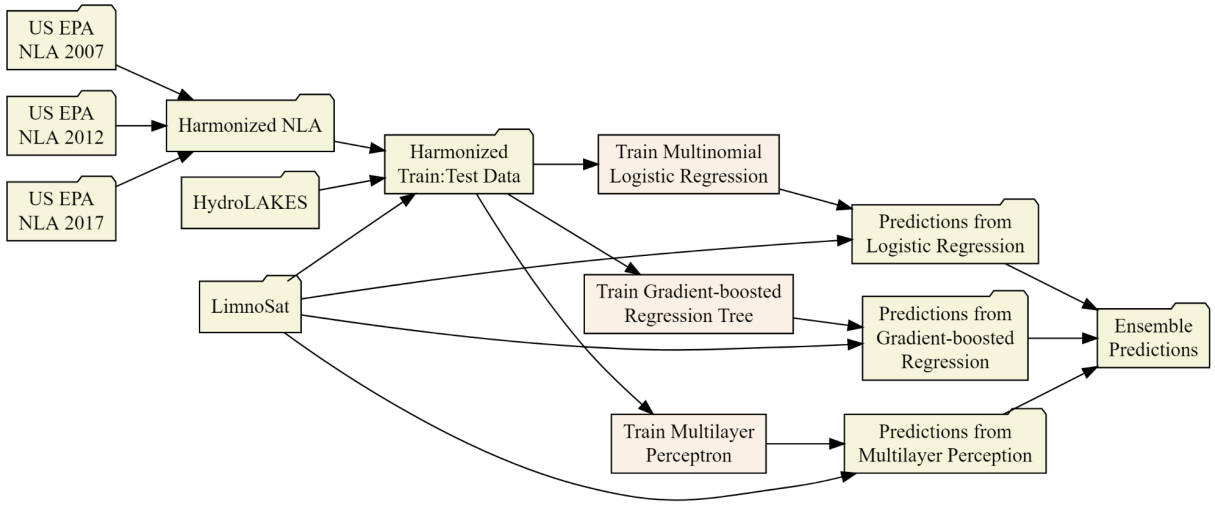

Figure 2: Flowchart for data harmonization, modeling, and prediction steps of the LTS-US dataset pipeline. Steps shaped as a file-folder correspond to an intermediary data product, and rectangles correspond to an intermediary model. Data aggregation combines data from the U.S. EPA's National Lakes Assessment, HydroLAKES, and LimnoSat-US to create a single, harmonized dataset of *in situ* lake trophic states with paired remotely sensed surface reflectances. Model training steps create multinomial logistic regression, multilayer perceptron, and extreme gradient boosted regression tree models. Each fitted model is then applied to the entire LimnoSat-US data, where national-scale predictions are made for each modeling method. Probabilistic predictions are then averaged to create ensemble predictions of lake trophic state. Quality control steps (described in "Technical Validation") use both the ensemble and individual model predictions to assess model performance. Each of these four components correspond to a piece of the overall data production pipeline: data aggregation functions are described in "1_aggregate"; model training functions are described in "2_train"; national-scale prediction functions are described in "3_predict"; quality control procedures are described in "4_qc". Flowchart was designed with the "DiagrammeR" package[89].

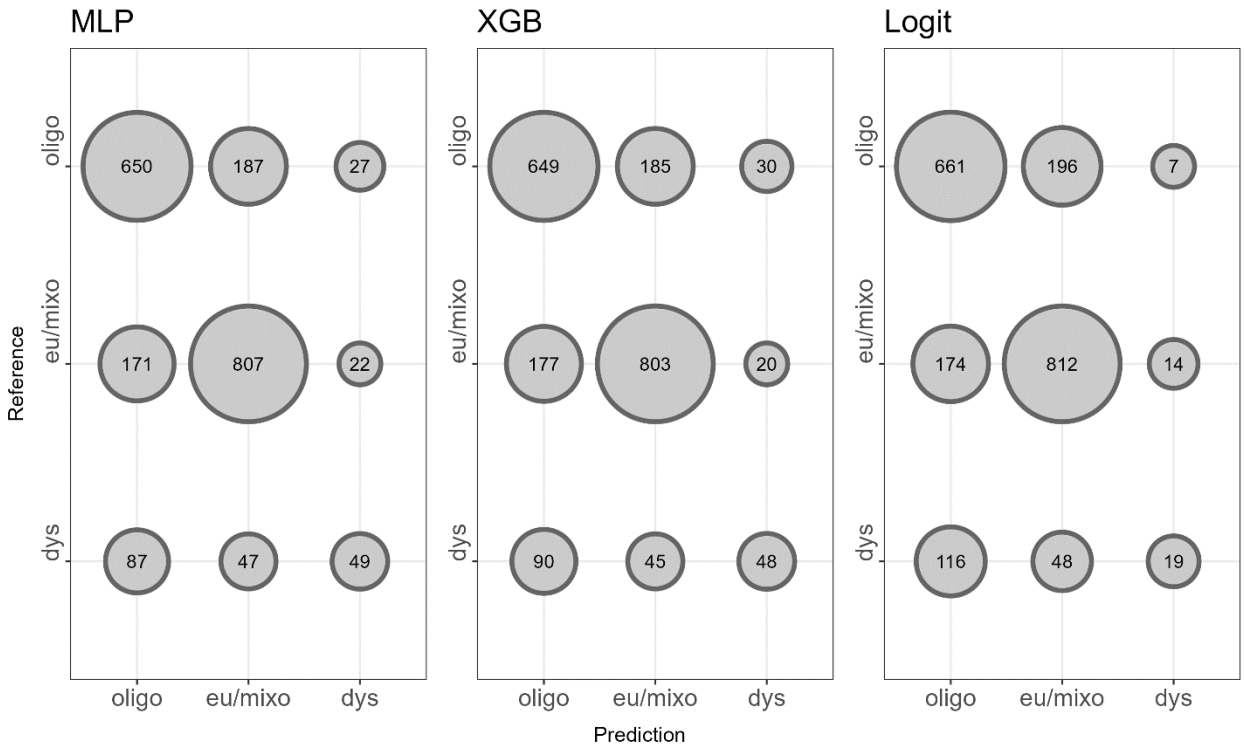

Figure 3: Confusion matrices from each modeling approach. Confusion matrices were generated using the test partitions for each spatial-holdout cross-validation. Circle size is scaled by the number of lakes falling within each category. Trophic states for model predictions and reference data correspond to the acronyms “dys” for “dystrophic”, “eu/mixo” for “eutrophic/mixotrophic”, and “oligo” for “oligotrophic”. Model acronyms are located as the title for each confusion matrix, where “MLP” signifies “Multilayer Perceptron”, “XGB” signifies “Gradient Boosted Regression”, and “Logit” signifies “Multinomial Logistic Regression”.

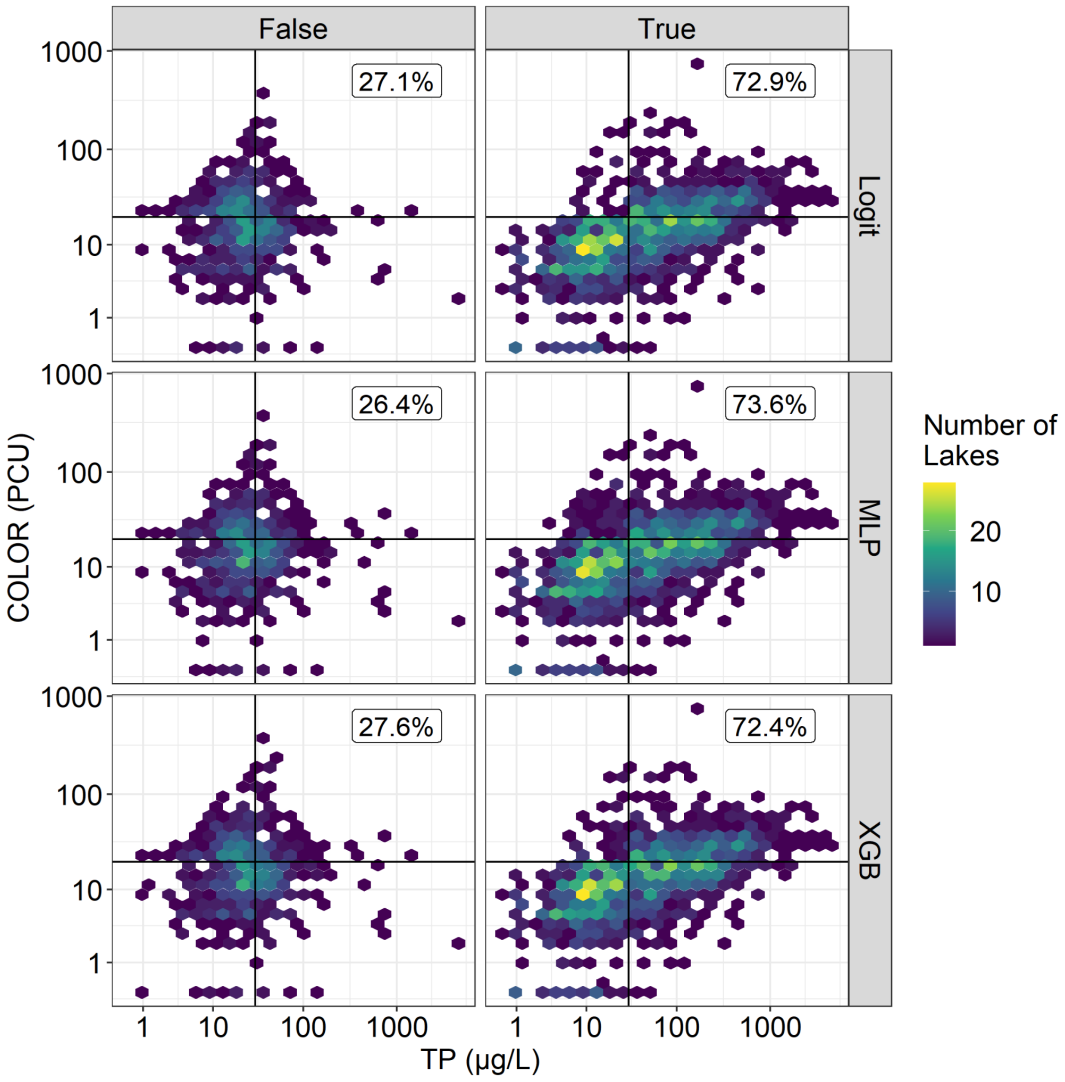

Figure 4: True and false classifications from testing data displayed on the NCP axes. Hexbins are colored by the number of lakes they contain. Labels reflect the percentage of lakes correctly or incorrectly predicted within a given modeling technique. Incorrect LTS predictions tend to be located at the nexus trophic state groupings. Correct predictions tend to more accurately reflect the breadth of ranges that can be observed within each of the LTS groupings.

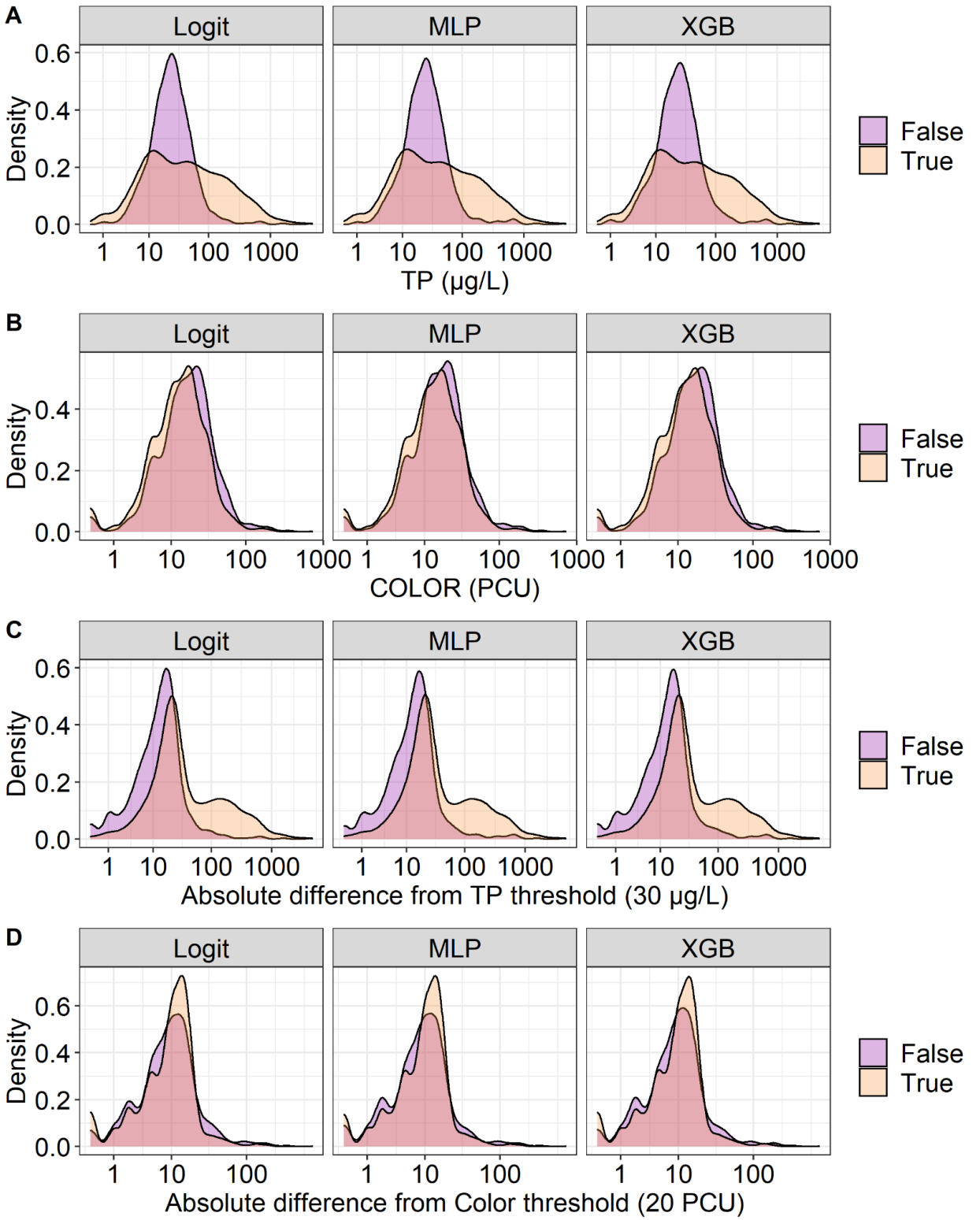

Figure 5: Density plots for total phosphorus (A) and true color (B) values as well as absolute differences from NCP thresholds for total phosphorus (C) and true color (D) among correctly (i.e., True; orange) and incorrectly (i.e., False; purple) classified trophic states. Because misclassifications appeared to increase in frequency near threshold

values for trophic state classification, we also assessed classification accuracies across absolute differences for each variable and threshold value. Across all models, we noticed that misclassifications tended to be highest near NCP thresholds for total phosphorus and color. Total phosphorus concentrations of 15-45 µg/L tended to be associated with false classifications. True color concentrations of 11-29 PCU tended to be associated with a false classification.

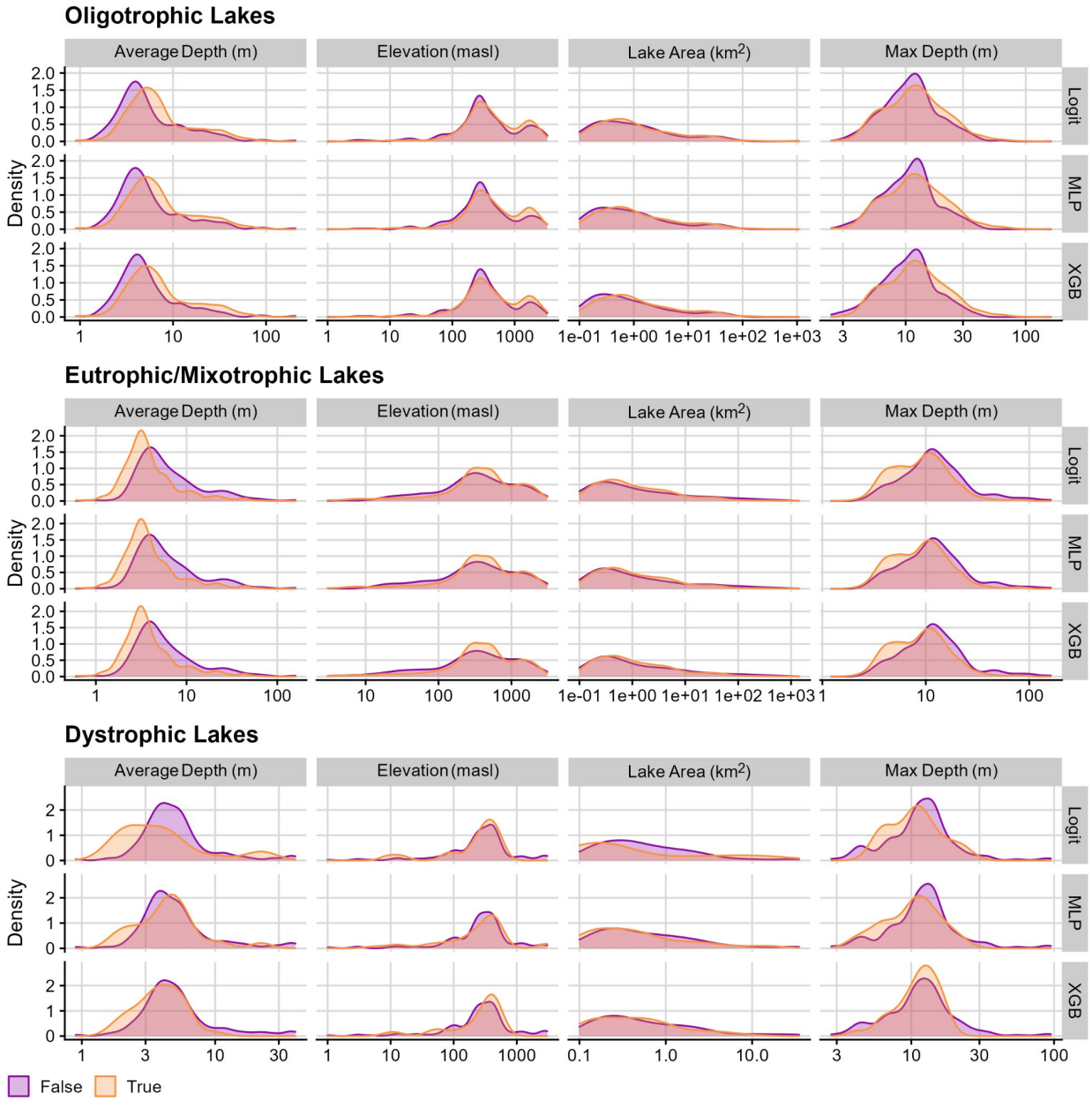

Figure 6: Density distributions for each lake's average depth, elevation, area, and maximum depth values across true (orange) and false (purple) classifications. Values are log-transformed to show characteristic density distributions over a wide range in value magnitudes. In general, patterns across true and false classifications were consistent across all three types of models. Depth was the primary morphometric characteristic for misclassified oligotrophic and eutrophic/mixotrophic lakes, where shallower oligotrophic and deeper eutrophic/mixotrophic lakes tended to be misclassified.

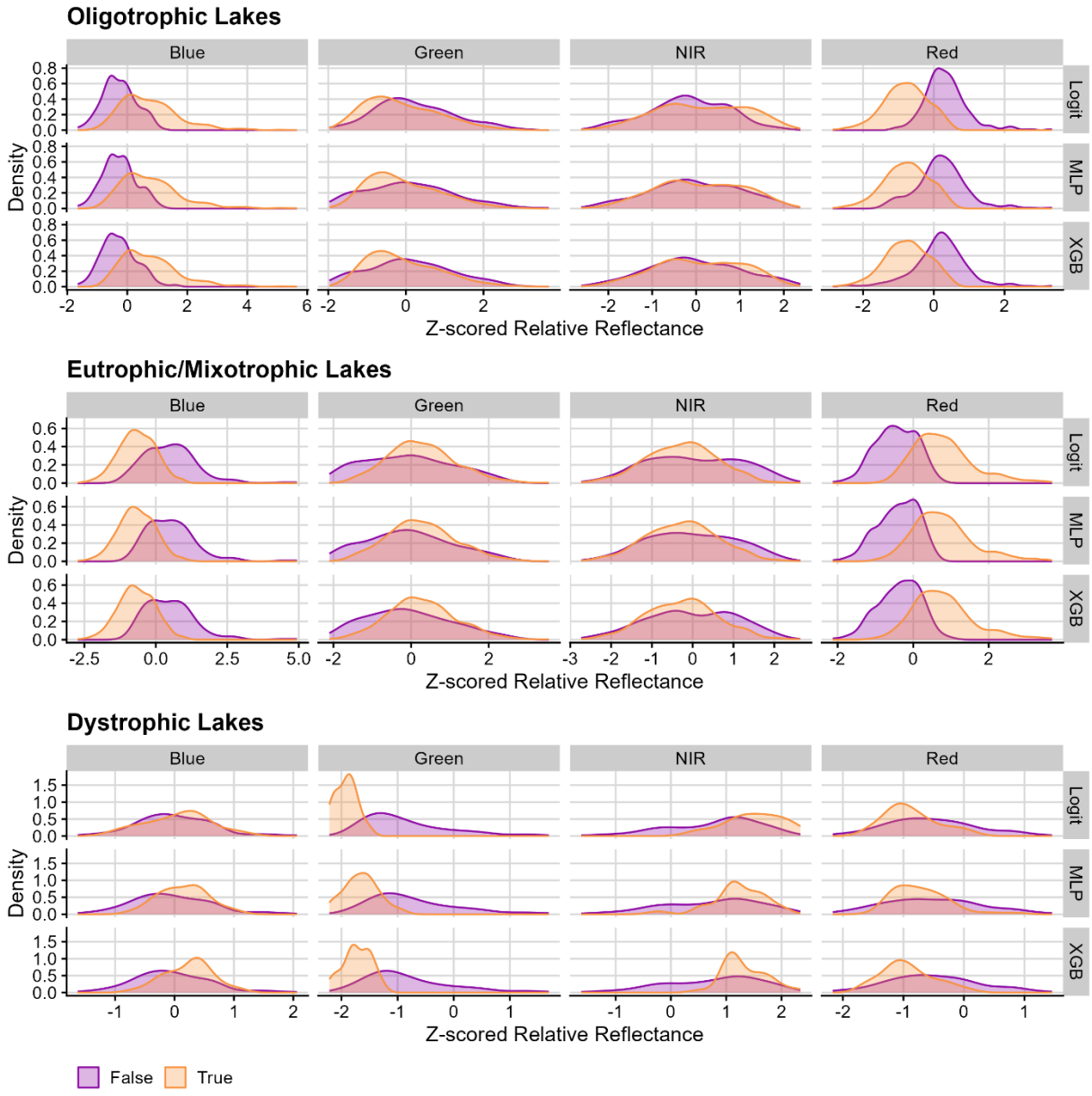

Figure 7: Density distributions for each Landsat band's z-scored, relative reflectance value across true (orange) and false (purple) classifications. In general, patterns across true and false classifications were consistent across all three types of models. Oligotrophic lakes tended to be misclassified when red bands were high and blue bands were low. Conversely, eutrophic/mixotrophic lakes tended to be misclassified when blue bands were high, and red bands were low. Dystrophic lakes tended to be misclassified when near infra-red bands were low and when green bands were high.

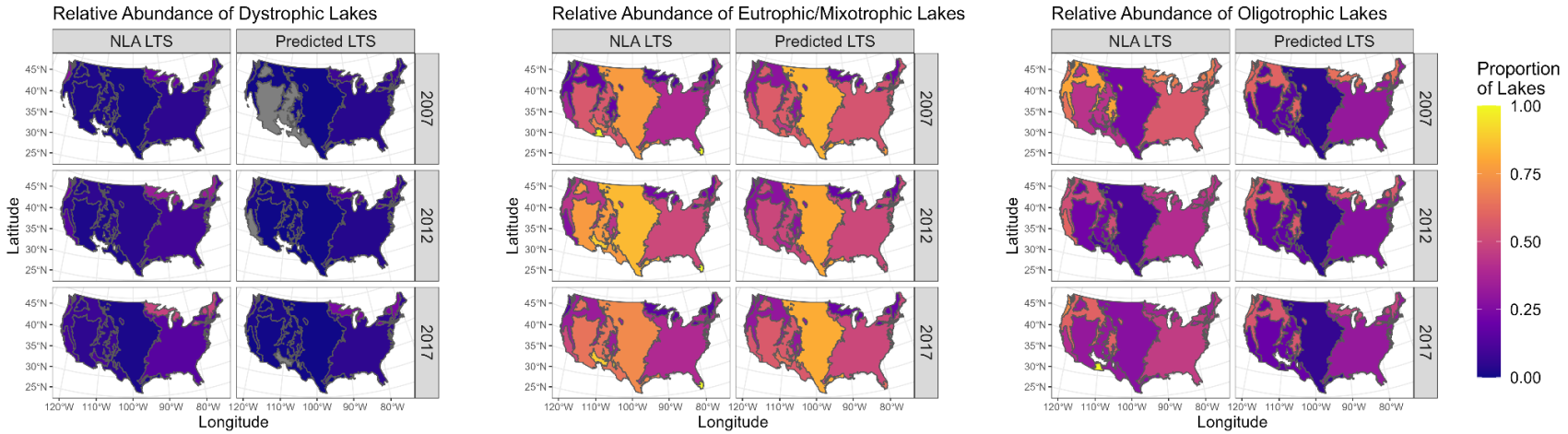

Figure 8: National-scale maps of U.S. Environmental Protection Agency Level I Ecoregions colored by proportion of lakes occurring in that ecoregion. For each trophic state, we compare estimated trophic state relative abundance from the NLA with predicted proportions from the ensemble LTS-US dataset.

Table 1: ANOVA table for total phosphorus and true color measurements in response to model type, model correctness, and trophic state. ANOVAs were assessed with Type II Sum-of-Squares to account for unbalanced sample sizes. To approximate a normal distribution, both total phosphorus and color were log-transformed. A p-value threshold of 0.05 was used to assess significance for each predictor.

(A) Total Phosphorus

	Sum-of-Squares	Degrees of Freedom	F-value	P-value
Model	0	2	0.003	0.997
Correct	5.62	1	43.64	< 0.001
Trophic State	1,335.42	2	5,183.4	< 0.001

(B) Color

	Sum-of-Squares	Degrees of Freedom	F-value	P-value
Model	0	2	0	1
Correct	0	1	0.02	0.89
Trophic State	286.62	2	1521.80	< 0.001

1302

Table 2: ANOVA table for lake morphological and locational properties in response to model type, model correctness, and trophic state. ANOVAs were assessed with Type II Sum-of-Squares to account for unbalanced sample sizes. To approximate a normal distribution, all response variables were log-transformed. A p-value threshold of 0.05 was used to assess significance for each predictor.

(A) Lake area

	Sum-of-Squares	Degrees of Freedom	F-value	P-value
Model	0	2	< 0.001	1
Correct	0.4	1	0.76	0.36
Trophic State	35.2	2	33.17	< 0.001

(B) Average Depth

	Sum-of-Squares	Degrees of Freedom	F-value	P-value
Model	0	2	0.001	1
Correct	1.61	1	14.94	< 0.001
Trophic State	54.32	2	251.53	< 0.001

(C) Maximum Depth

	Sum-of-Squares	Degrees of Freedom	F-value	P-value
Model	0	2	0.001	1
Correct	1.2	1	16.86	< 0.001
Trophic State	14.17	2	99.53	< 0.001

(D) Elevation

	Sum-of-Squares	Degrees of Freedom	F-value	P-value
Model	0	2	0.001	1
Correct	3.06	1	10.61	0.001

Trophic State	15.39	2	26.63	< 0.001

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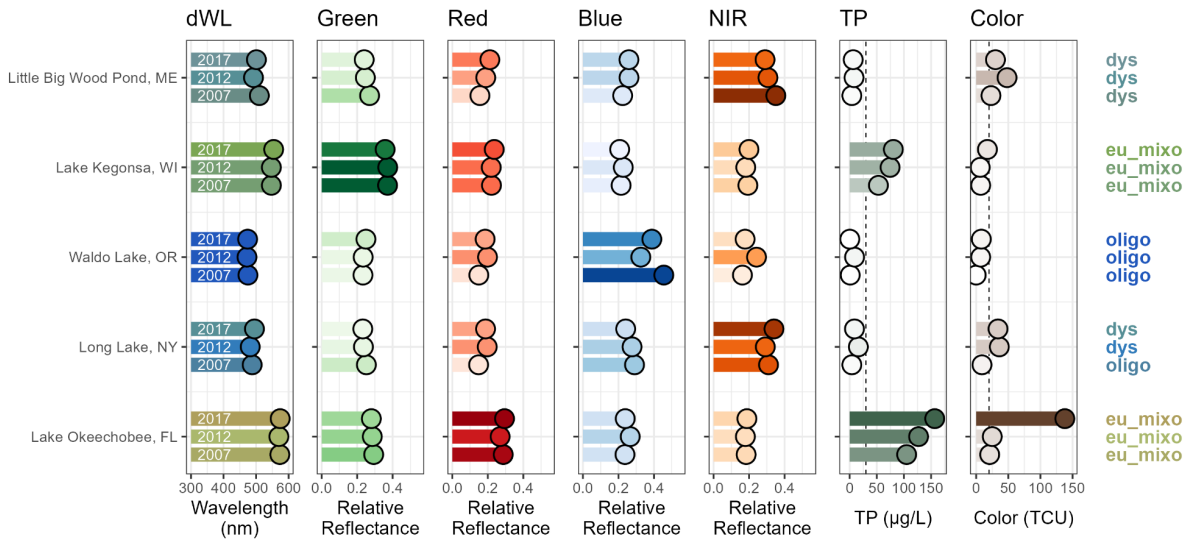

Figure S1: Example comparative summary of five lakes that were sampled in all three U.S. EPA NLA campaigns[1–3]. In general, variation between lakes is visually greater than within a lake[4]. Colors of a lake's summertime median dominant wavelength (dWL) are represented as the color of the bar and point. All remaining variables are colored by a variable's value, where a darker bar and point refers to a higher variable value.

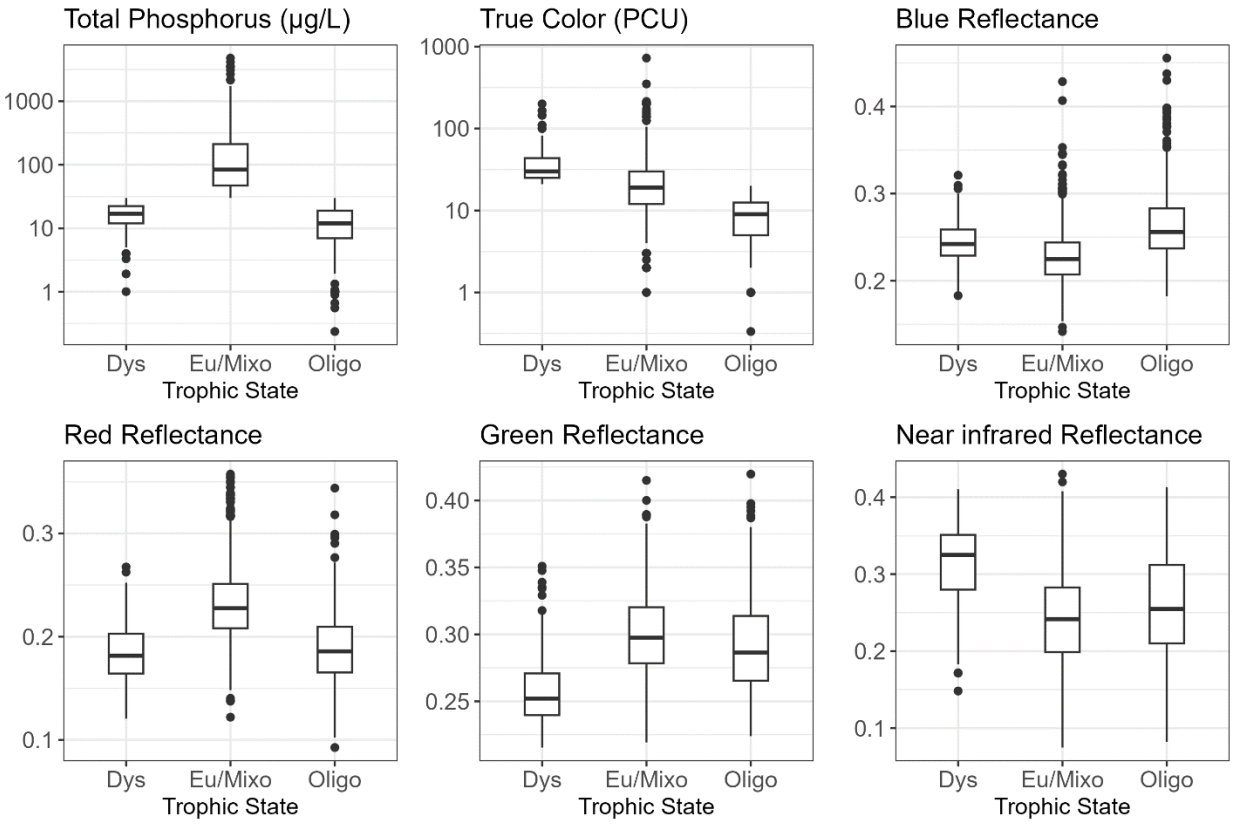

Figure S2: Boxplots of all variables used to define trophic state. Total Phosphorus and True Color data are shown on a log-scale axis to accommodate multiple orders of magnitude.

Table S1: Summary table of training data used for LTS-US Dataset creation[1–4]. These data are used for creating training and test data for each of the three modeling techniques described in the main text. Data are presented as means with standard deviations in parentheses.

Trophic State	Year	Total Phos	Color	Blue	Green	Red	Near Infrared	Number of lakes
Dys	2007	16.05 (6.86)	36.03 (24.12)	0.24 (0.03)	0.26 (0.02)	0.19 (0.03)	0.31 (0.05)	40
Dys	2012	19.36 (7.58)	37.45 (24.21)	0.24 (0.02)	0.26 (0.03)	0.19 (0.04)	0.31 (0.06)	62
Dys	2017	15.78 (6.79)	42.39 (29.96)	0.25 (0.02)	0.26 (0.03)	0.18 (0.02)	0.32 (0.05)	81
Eu/Mixo	2007	233.3 (401.7)	19.87 (14.75)	0.22 (0.03)	0.3 (0.03)	0.23 (0.04)	0.25 (0.06)	362
Eu/Mixo	2012	196.61 (362.4)	26.82 (41.71)	0.23 (0.03)	0.3 (0.03)	0.23 (0.04)	0.24 (0.06)	386
Eu/Mixo	2017	169.01 (332.2)	28.12 (35.53)	0.23 (0.03)	0.3 (0.03)	0.23 (0.03)	0.24 (0.06)	252
Oligo	2007	10.85 (7.45)	8.01 (4.91)	0.26 (0.04)	0.29 (0.03)	0.19 (0.03)	0.26 (0.07)	411
Oligo	2012	16.25 (7.5)	10.89 (4.85)	0.26 (0.04)	0.29 (0.04)	0.18 (0.03)	0.26 (0.07)	229
Oligo	2017	14.01 (6.96)	8.58 (6.03)	0.27 (0.04)	0.29 (0.03)	0.19 (0.03)	0.26 (0.06)	224

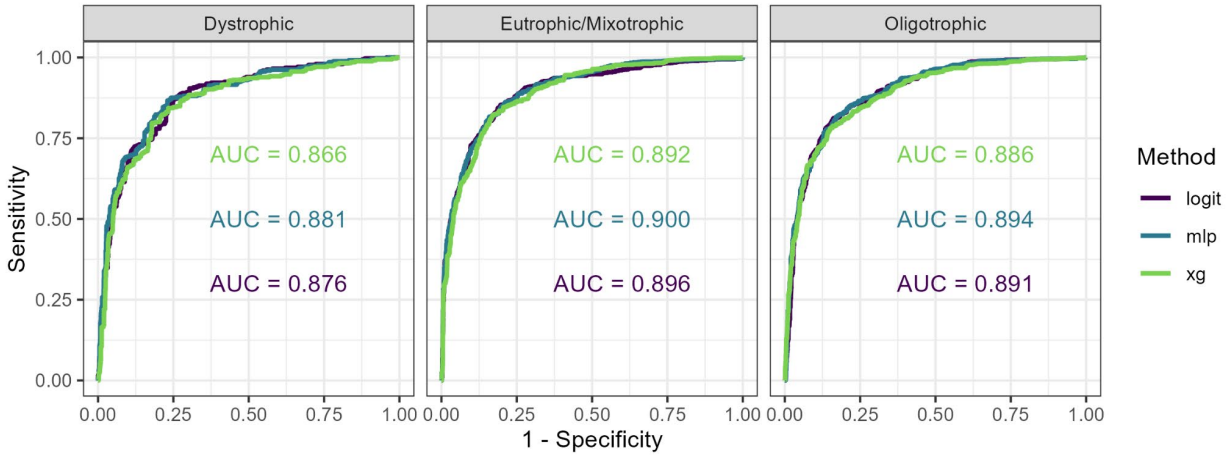

Figure S3: Receiver-Operator-Characteristic (ROC) curves for each trophic status prediction and model method. Area under the Curve (AUC) is reported for each ROC curve. AUC is a metric that generally reflects model fit, where the ROC curve details a model's capacity to give a true result as the false positive rate is artificially inflated. Across all LTS and modeling methods, ROC curves and resulting AUCs are exceptionally similar, suggesting overall congruence among modeling methodologies.

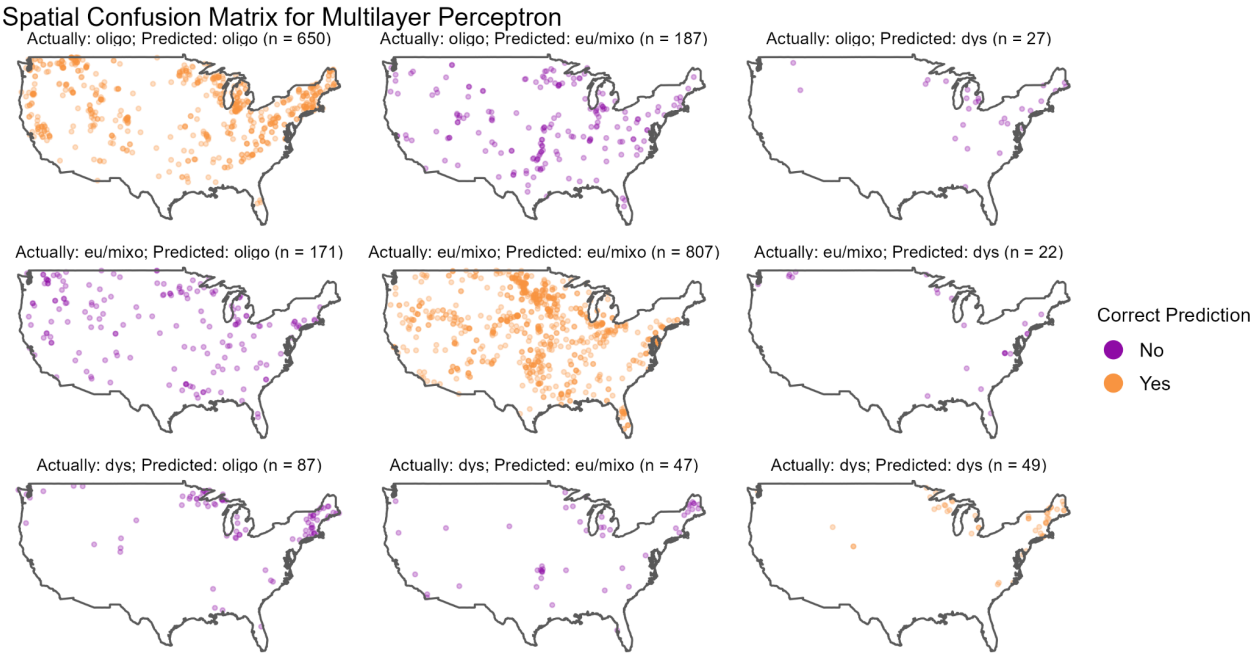

Figure S4: Spatial confusion matrix for multilayer perceptron model. In general, the multilayer perceptron model did not classify or misclassify lakes in a spatial pattern, giving confidence that models were likely misclassifying due to differences other than locational biases at the continental scale.

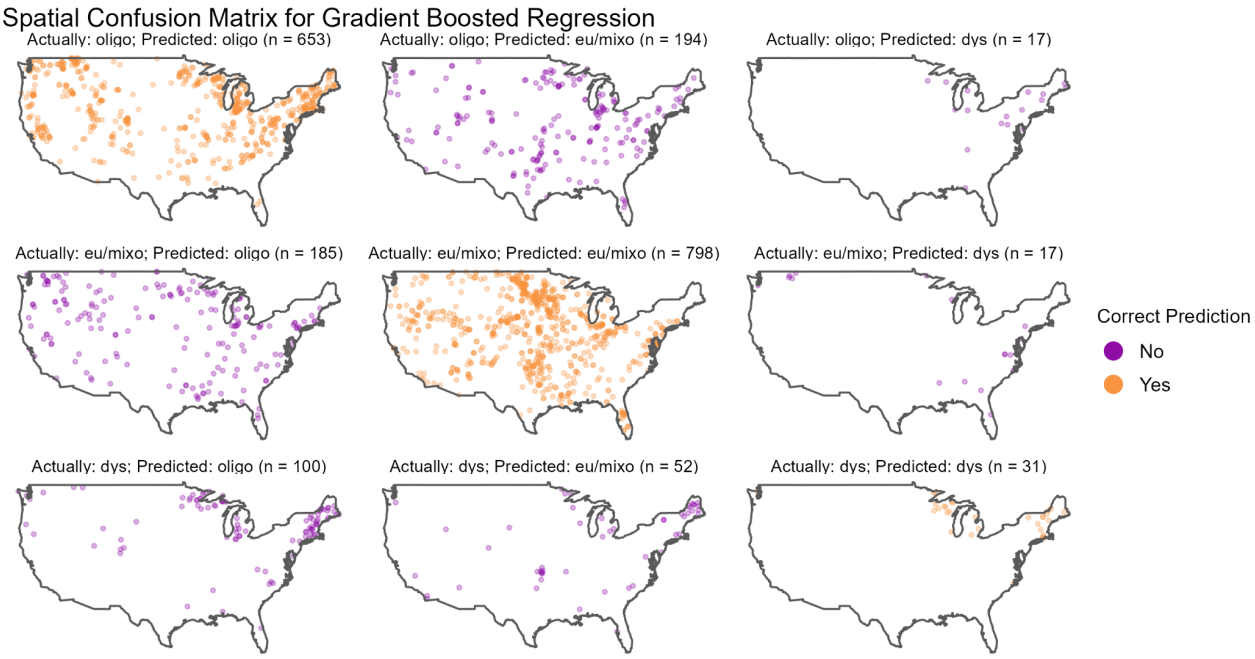

Figure S5: Spatial confusion matrix for gradient boosted regression model. In general, the model did not classify or misclassify lakes in a spatial pattern, giving confidence that models were likely misclassifying due to differences other than locational biases at the continental scale.

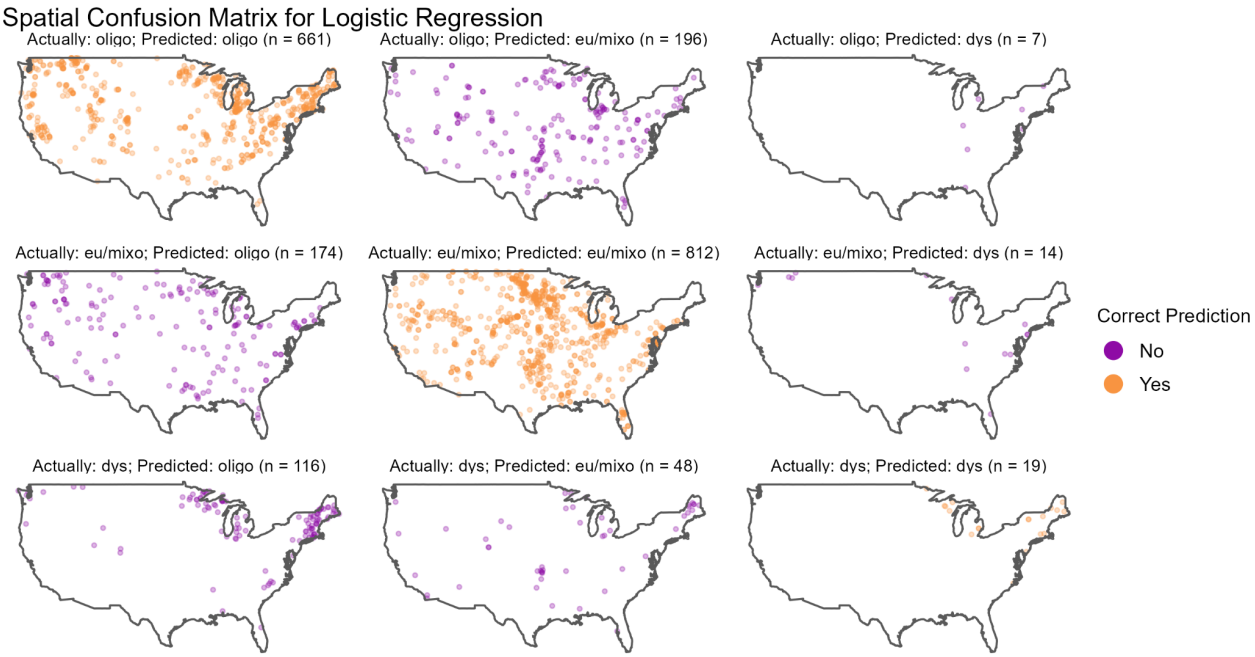

Figure S6: Spatial confusion matrix for Logit model. In general, the Logit model did not classify or misclassify lakes in a spatial pattern, giving confidence that models were likely misclassifying due to differences other than locational biases at the continental scale.

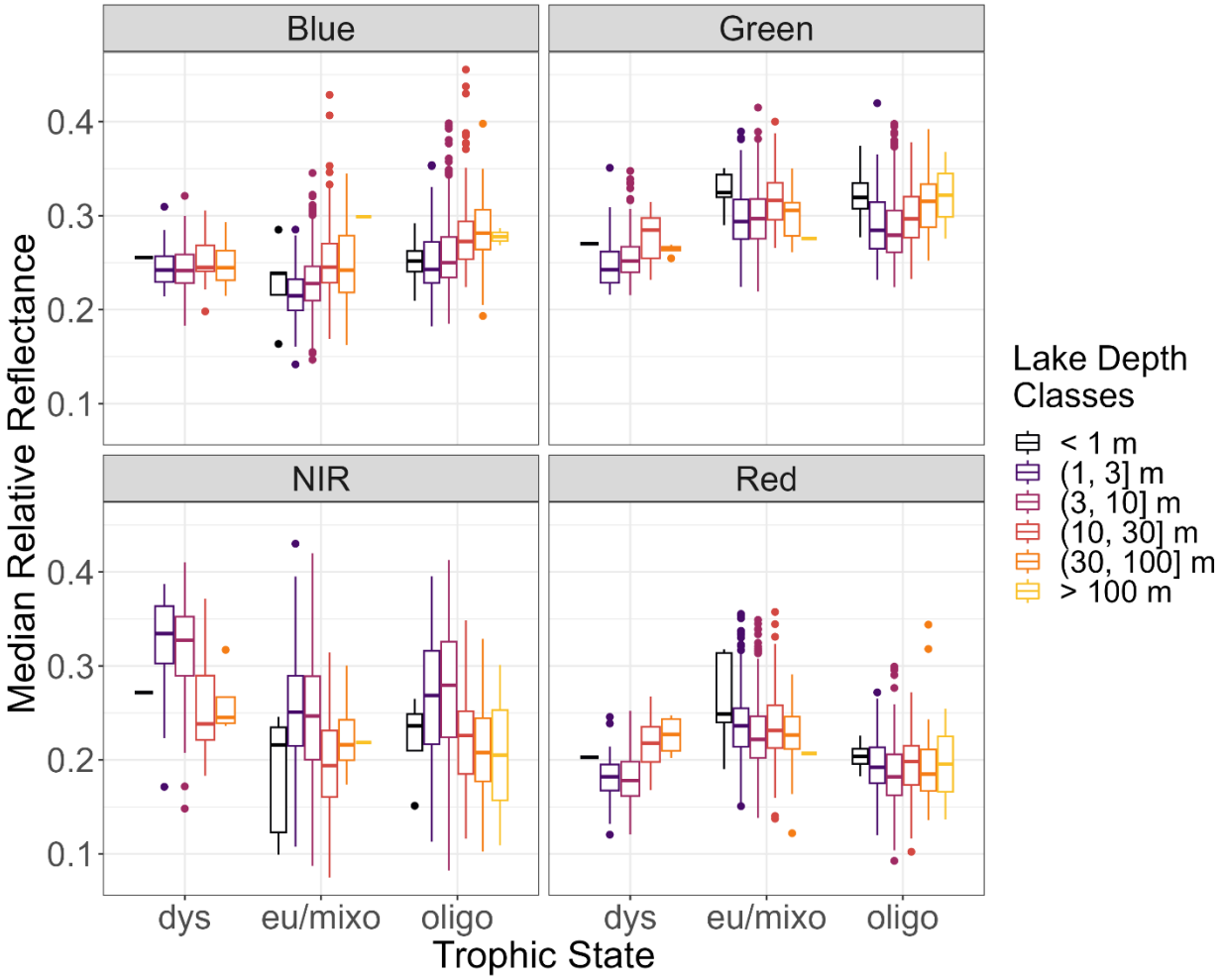

Figure S7: Boxplots of median summertime relative reflectance values for each Landsat band and NCP-defined trophic state divided by lake depth classes. Water quality data are aggregated from the 2007, 2012, and 2017 U.S. EPA NLA campaigns. Reflectance data are aggregated from LimnoSat-US. Relative reflectance is defined as the value of a given band's reflectance divided by the sum of all four bands. Summertime median relative reflectances are defined as the median of all relative reflectance values from June through August in a given year.

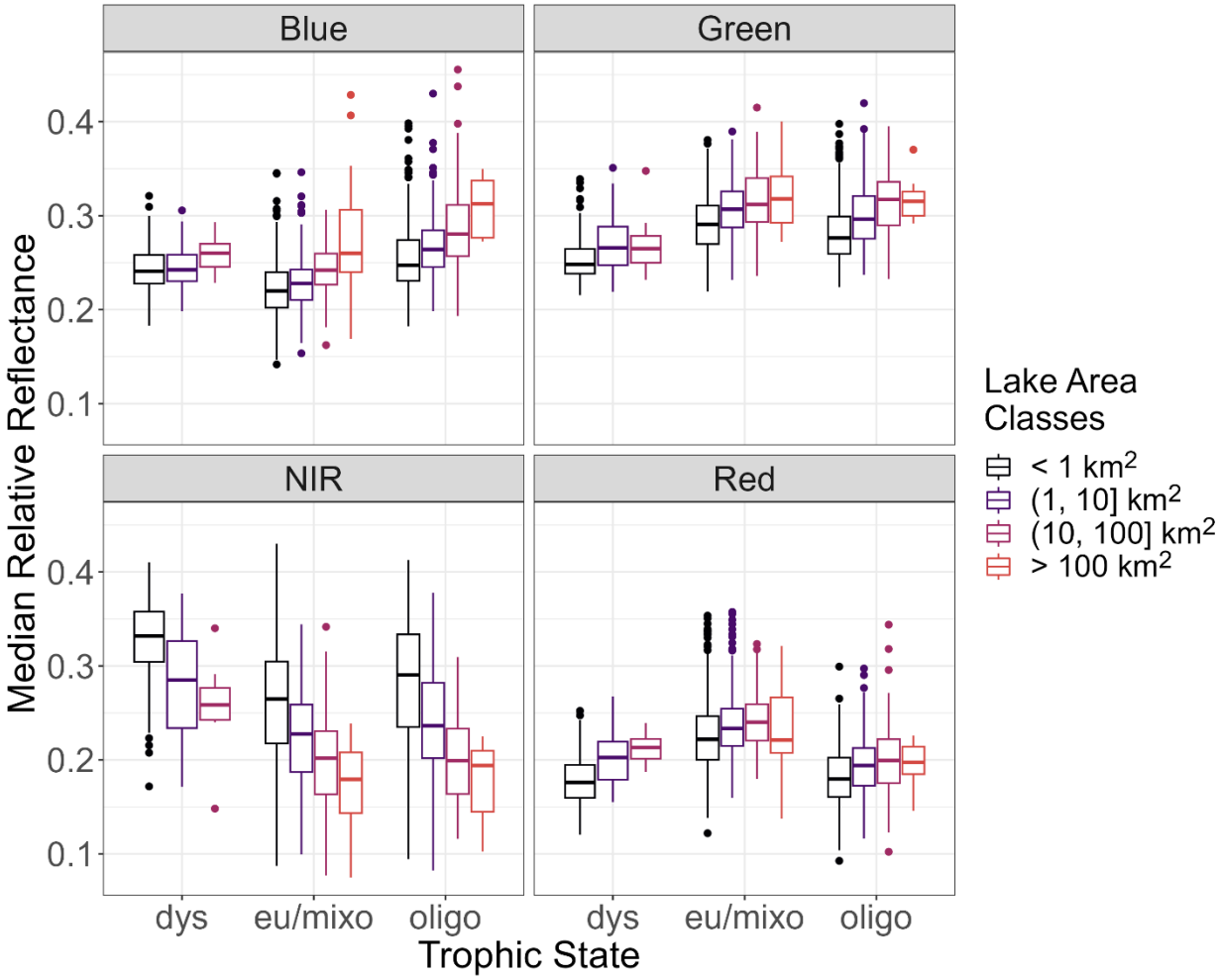

Figure S8: Boxplots of median summertime relative reflectance values for each Landsat band and NCP-defined trophic state divided by lake area classes. Water quality data are aggregated from the 2007, 2012, and 2017 U.S. EPA NLA campaigns. Reflectance data are aggregated from LimnoSat-US. Relative reflectance is defined as the value of a given band's reflectance divided by the sum of all four bands. Summertime median relative reflectances are defined as the median of all relative reflectance values from June through August in a given year.

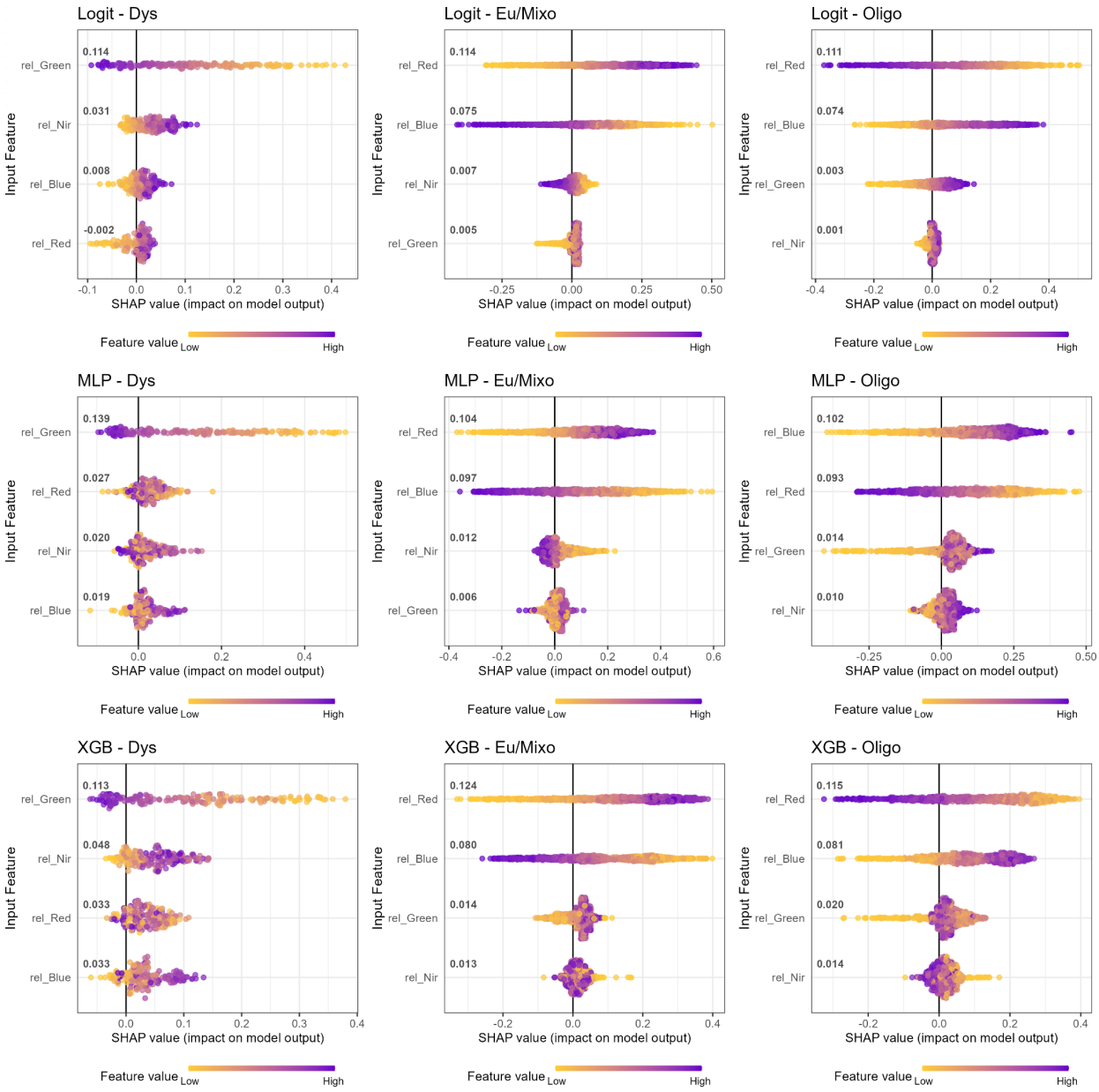

Figure S9: Summary plots from SHAP analysis with SHAP values arranged by model type and lake trophic state. SHAP values include those from correct and incorrect classifications. Importance scores are located next to each feature on the left side of each plot panel. Features are arranged on the y-axis of each plot in order of relative importance, where most important features are at the top of the plot and decrease in relative importance towards the bottom of the plot. Across all modeling types, features were comparable in importance. In all cases, the top two features for each modeling technique and trophic state were identical. Further, the top two features also corresponded to limnological and ecological understanding of each lake type. Dystrophic lakes were most influenced by green and near-infrared bands, which corresponds to these lakes being characterized by increased sediment and dissolved organic carbon as well as decreased primary production. Eutrophic/mixotrophic and

oligotrophic lakes were most influenced by red and blue bands, which corresponds to these lakes as being most characterized by primary production.

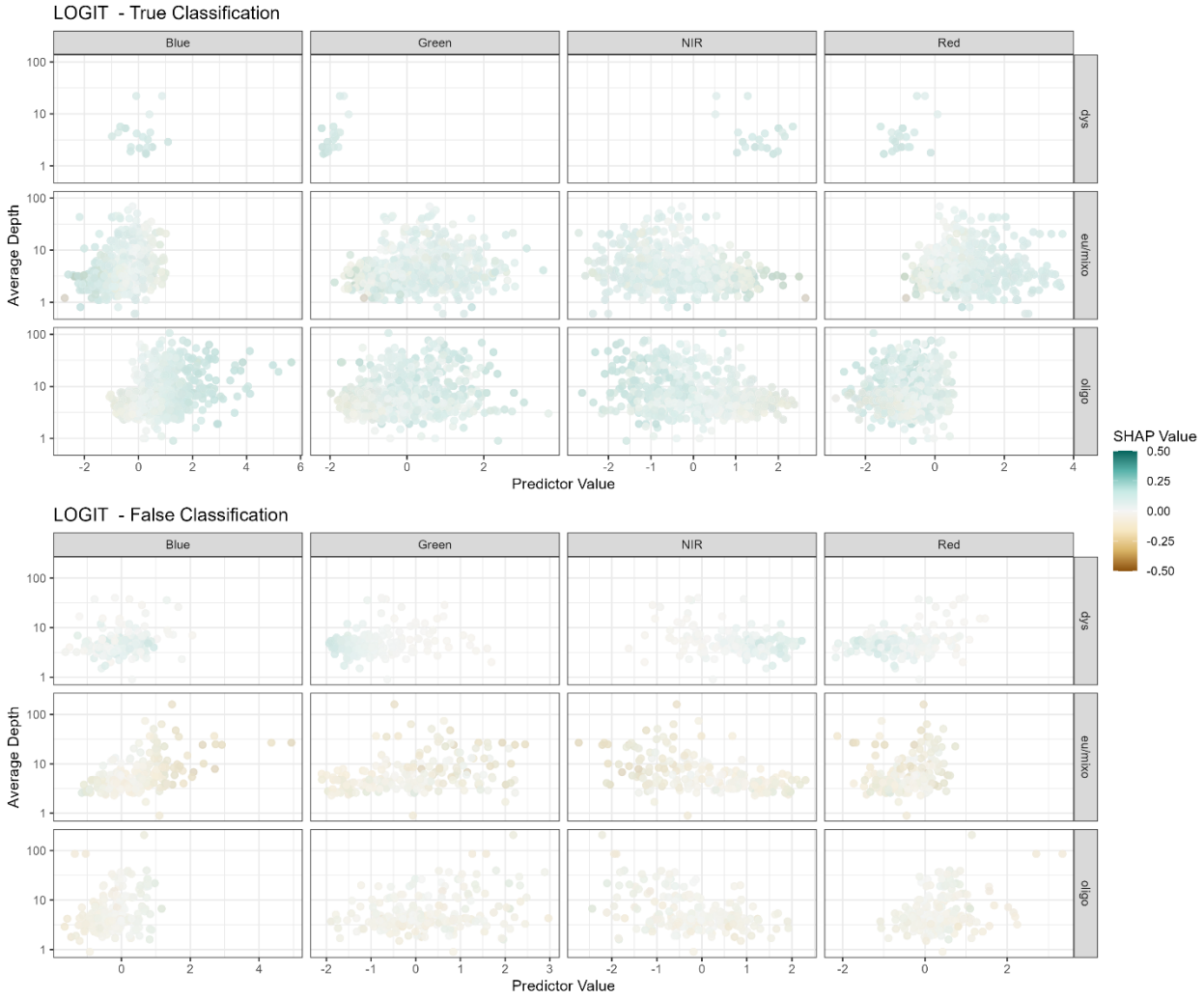

Figure S10: SHAP value analysis by each trophic state's band value and average depth from logistic regression models. While correct and incorrect classifications generally occupied the same parameter space for band values and average depths, greatest incongruence between correct and incorrect classifications occurred in blue and red bands for eutrophic and oligotrophic lakes. In particular, shallow oligotrophic lakes tended to have lower blue reflectances, which corresponded to a lower SHAP value; shallow eutrophic/mixotrophic lakes likewise had low blue reflectances, but these bands had high SHAP values. Conversely, deeper oligotrophic lakes tended to have lower red band values, which were associated with higher SHAP values; deeper eutrophic/mixotrophic lakes tended to have higher red reflectances, which also had a higher SHAP value. Together, this analysis suggests that lakebed effects may influence classification. For example, benthic algal production in oligotrophic lakes may produce reflectance values similar to eutrophic lakes, leading to model confusion. This same result is implied throughout all analysis steps, where depth appears to be the major issue for correct trophic state classification.

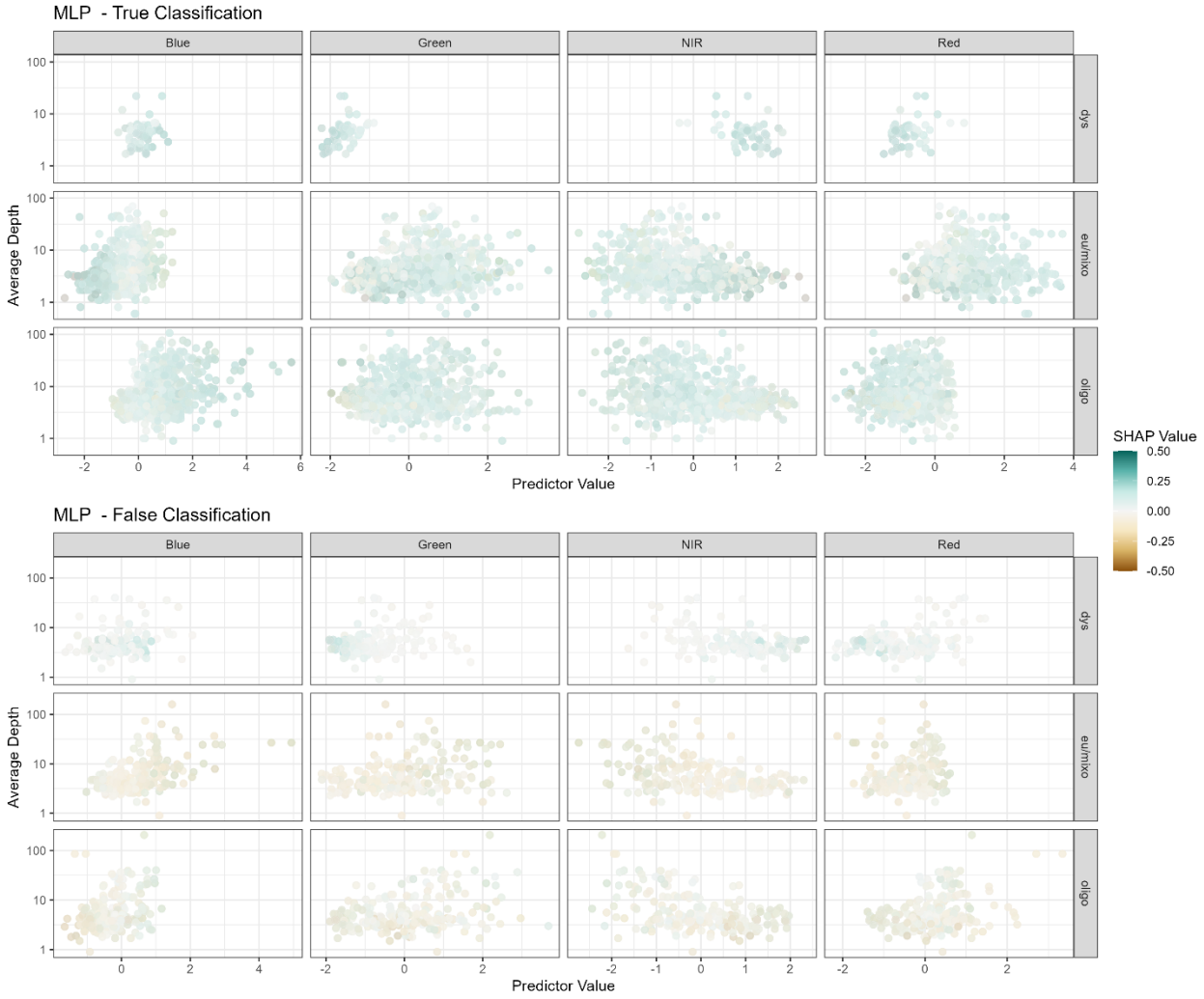

Figure S11: SHAP value analysis by each trophic state's band value and average depth from multilayer perceptron models. While correct and incorrect classifications generally occupied the same parameter space for band values and average depths, greatest incongruence between correct and incorrect classifications occurred in blue and red bands for eutrophic and oligotrophic lakes. In particular, shallow oligotrophic lakes tended to have lower blue reflectances, which corresponded to a lower SHAP value; shallow eutrophic/mixotrophic lakes likewise had low blue reflectances, but these bands had high SHAP values. Conversely, deeper oligotrophic lakes tended to have lower red band values, which were associated with higher SHAP values; deeper eutrophic/mixotrophic lakes tended to have higher red reflectances, which also had a higher SHAP value. Together, this analysis suggests that lakebed effects may influence classification. For example, benthic algal production in oligotrophic lakes may produce reflectance values similar to eutrophic lakes, leading to model confusion.

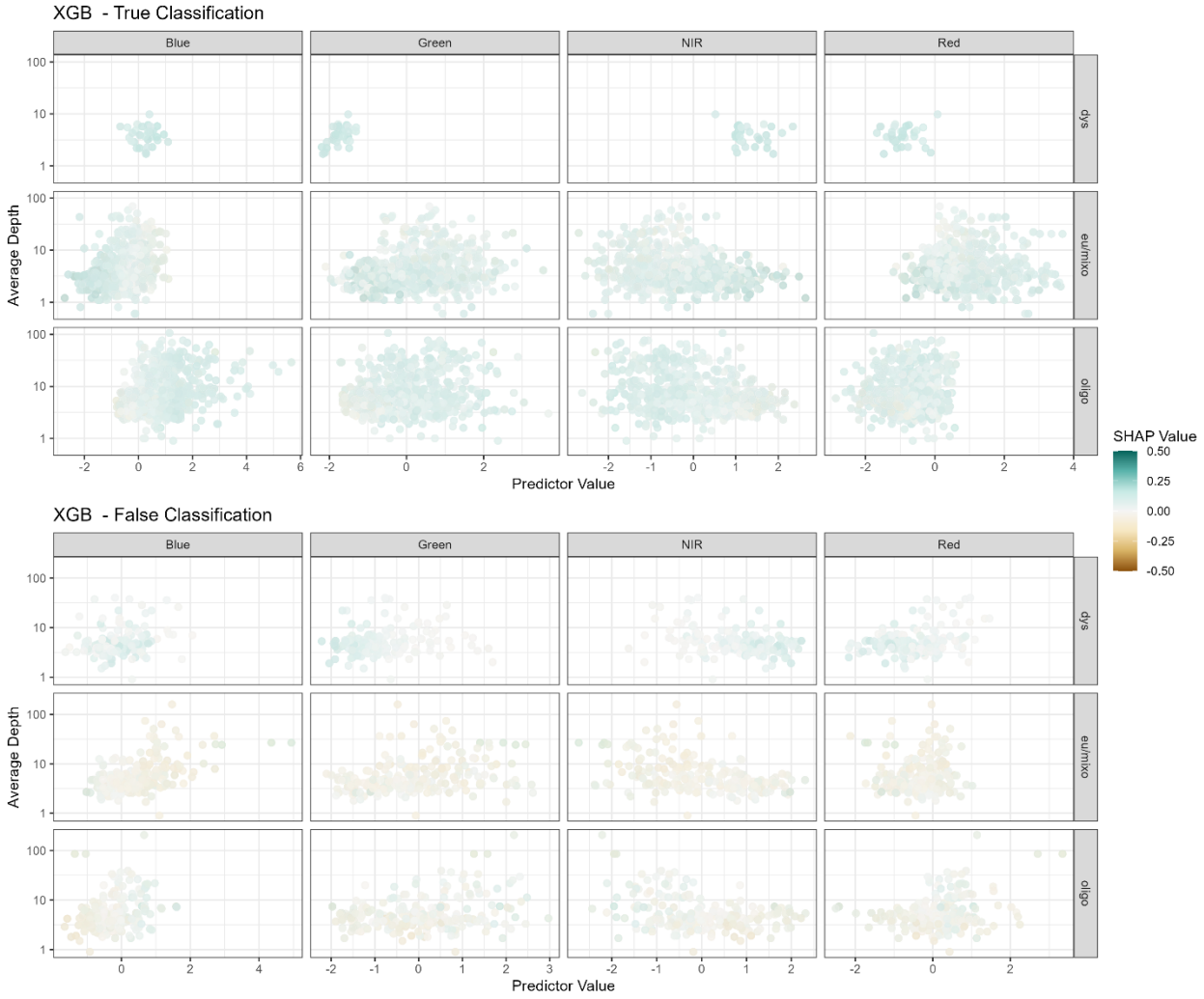

Figure S12: SHAP value analysis by each trophic state's band value and average depth from gradient boosted regression models. While correct and incorrect classifications generally occupied the same parameter space for band values and average depths, greatest incongruence between correct and incorrect classifications occurred in blue and red bands for eutrophic and oligotrophic lakes. In particular, shallow oligotrophic lakes tended to have lower blue reflectances, which corresponded to a lower SHAP value; shallow eutrophic/mixotrophic lakes likewise had low blue reflectances, but these bands had high SHAP values. Conversely, deeper oligotrophic lakes tended to have lower red band values, which were associated with higher SHAP values; deeper eutrophic/mixotrophic lakes tended to have higher red reflectances, which also had a higher SHAP value. Together, this analysis suggests that lakebed effects may influence classification. For example, benthic algal production in oligotrophic lakes may produce reflectance values similar to eutrophic lakes, leading to model confusion.

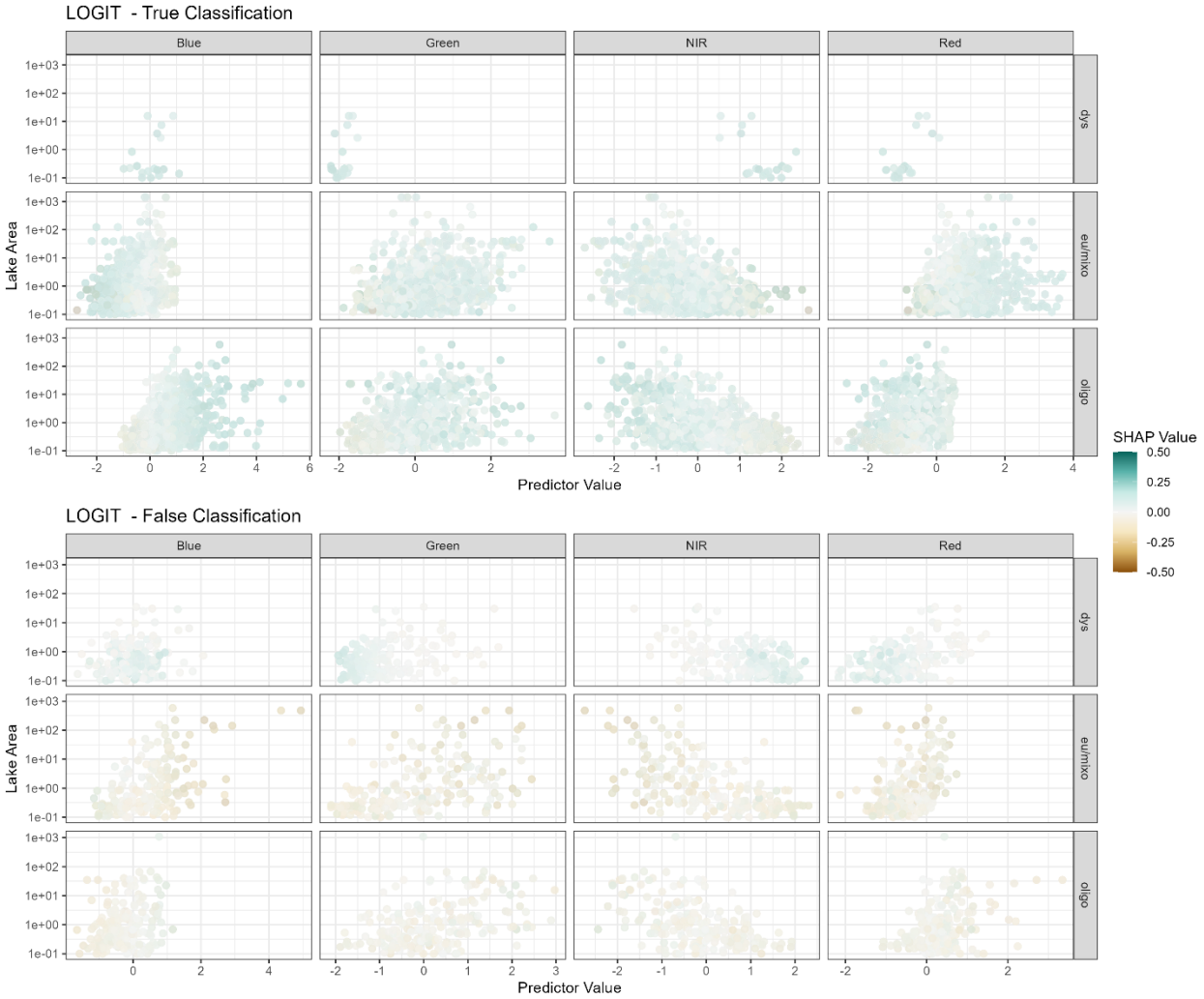

Figure S13: SHAP value analysis by each trophic state's band value and surface area from logistic regression models. Visually, SHAP and reflectance values as well as lake areas all occupied the same parameter space, implying that lake area, a proxy for adjacency effects, is likely not consequential for feature importance and correct classification. This general result is likewise observed in lake areas being generally consistent across correctly and incorrectly classified lakes.

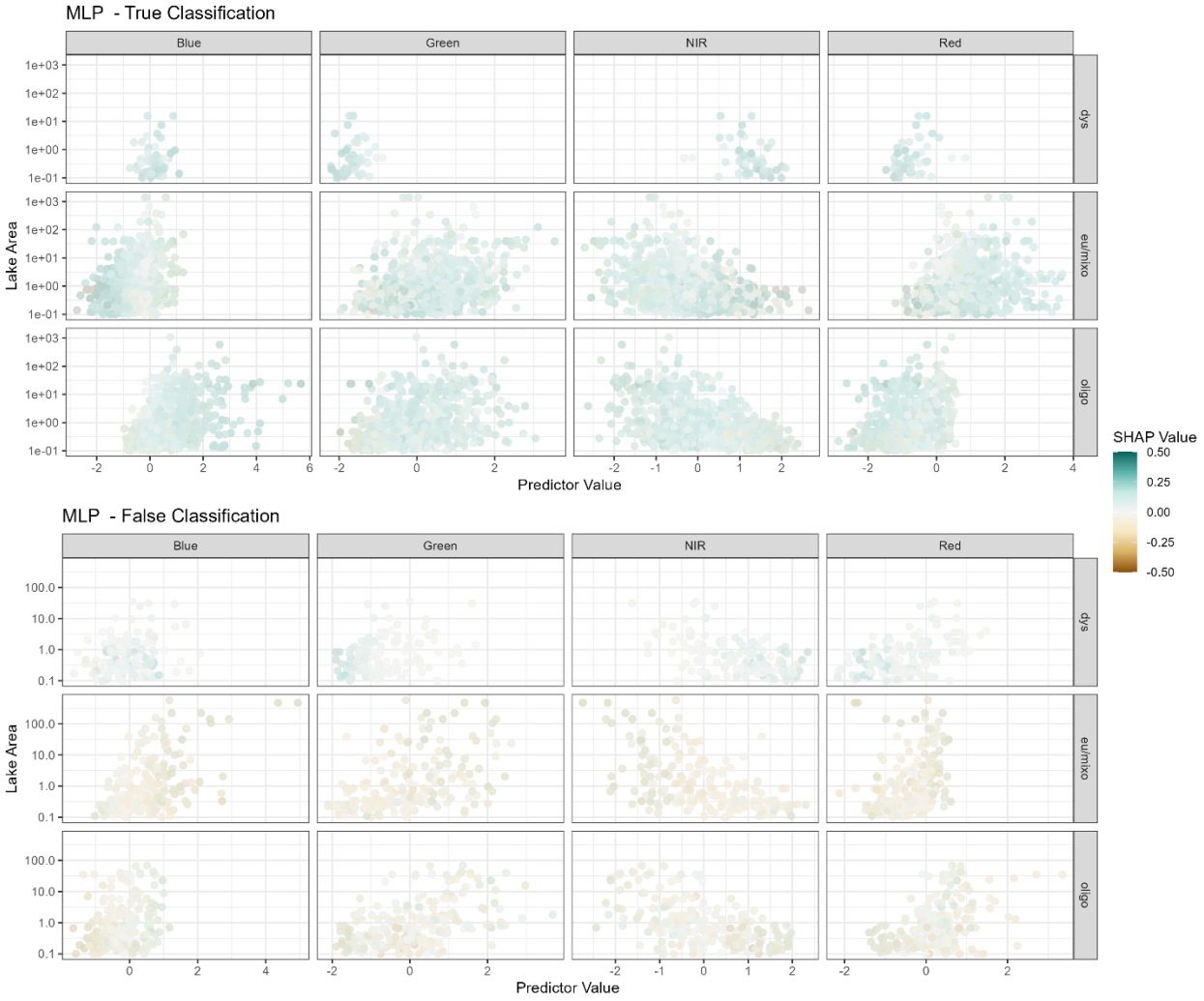

Figure S14: SHAP value analysis by each trophic state's band value and surface area from multilayer perceptron models. Visually, SHAP and reflectance values as well as lake areas all occupied the same parameter space, implying that lake area, a proxy for adjacency effects, is likely not consequential for feature importance and correct classification. This general result is likewise observed in lake areas being generally consistent across correctly and incorrectly classified lakes.

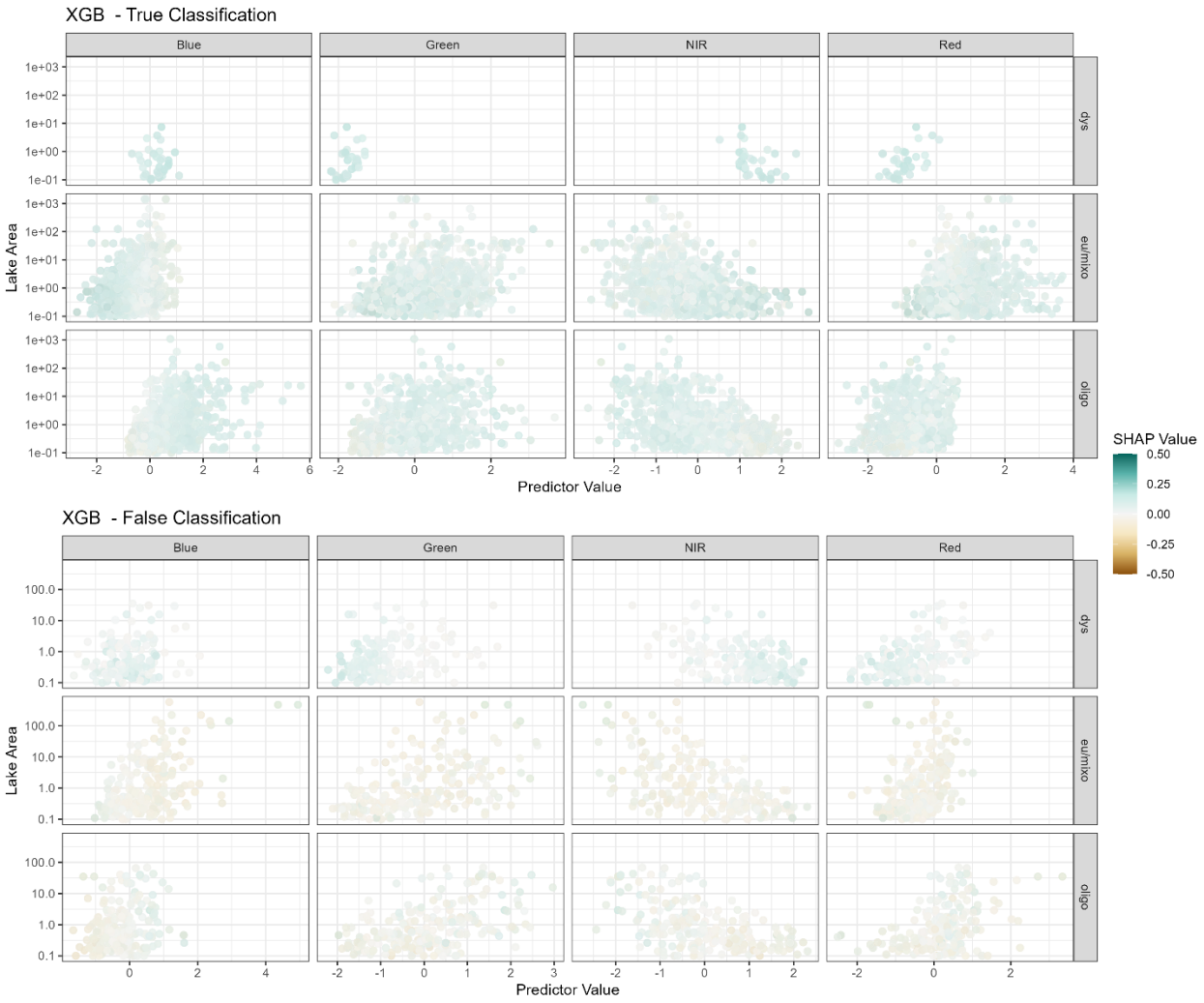

Figure S15: SHAP value analysis by each trophic state's band value and surface area from gradient boosted regression models. Visually, SHAP and reflectance values as well as lake areas all occupied the same parameter space, implying that lake area, a proxy for adjacency effects, is likely not consequential for feature importance and correct classification. This general result is likewise observed in lake areas being generally consistent across correctly and incorrectly classified lakes.

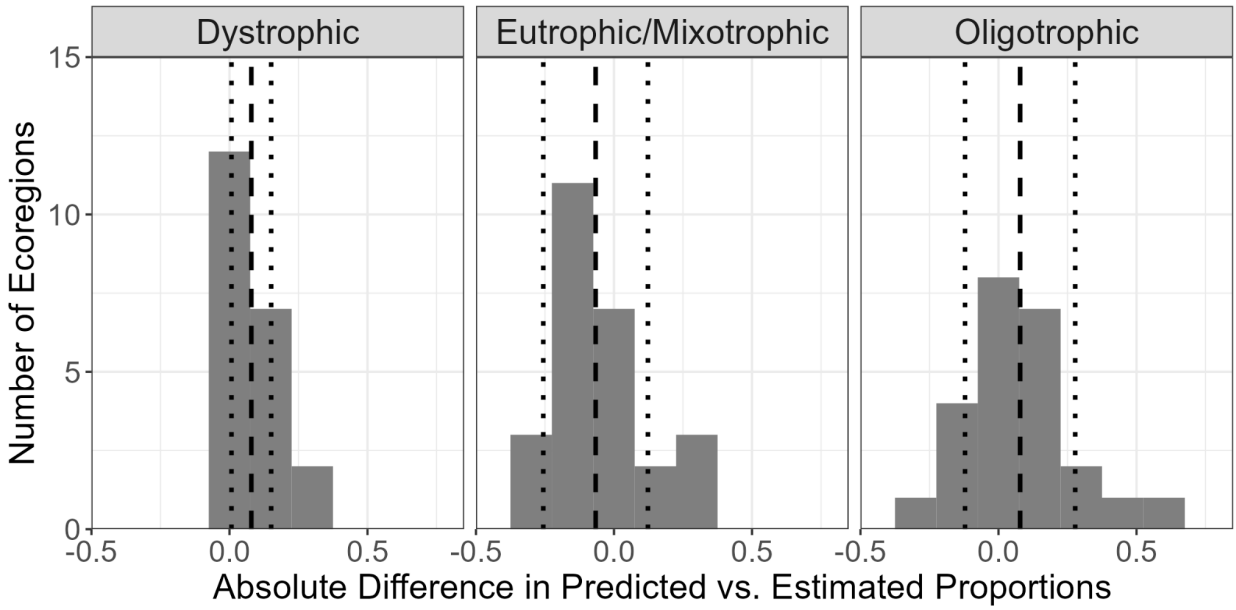

Figure S16: Histograms of absolute difference (Estimated - Predicted) in predicted and estimated proportions of each lake trophic state across U.S. EPA Level I Ecoregions. Vertical dashed lines reflect the mean, and vertical, dotted lines reflect one standard deviation from the mean. For all trophic states, distributions approximately center around zero. Oligotrophic and dystrophic lakes tend to be slightly underpredicted, whereas eutrophic and mixotrophic lakes tend to be slightly overpredicted.

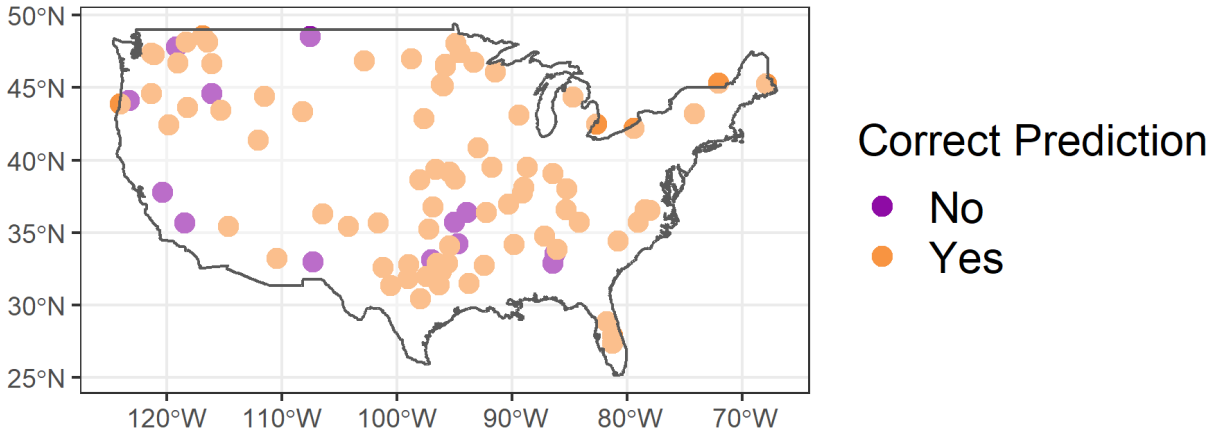

Figure S17: National-scale map of correct and incorrect trophic state classifications as assessed by manual checking of lake trophic state predictions against independent sources. Among lakes where independent sources could be identified, 73.5% of lakes were correctly predicted, which is notably similar to accuracies assessed from the NLA sampling campaign data. Additionally, correct and incorrect classifications did not follow apparent spatial patterns, implying that our models were not influenced by geographical or locational differences.