

1 **The ostracod clumped-isotope thermometer: A novel tool to**
2 **reconstruct quantitative continental climate changes.**

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12 **STATEMENT**

13 This manuscript has been submitted for publication in Geology. Please note that, despite having
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19 Twitter account: @MaMarchegiano

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30

31 **ABSTRACT**

32 This study presents the ostracod clumped-isotope (Δ_{47}) thermometer, a new tool that
33 provides quantitative temperature and hydrological reconstruction from lacustrine systems, which
34 are among the best archives to reconstruct continental paleotemperature variations. The
35 relationship between Δ_{47} and the temperature at which ostracod shell crystallized, is determined
36 by measuring Δ_{47} on different species grown under controlled temperatures in both natural
37 environments ($4 \pm 2^\circ\text{C}$; $12 \pm 1^\circ\text{C}$) and lab cultures ($23 \pm 0.5^\circ\text{C}$). No consistent offset between the
38 two species originating from the same environment and precipitated at the same temperature is
39 reported, suggesting the absence of a vital effect in ostracod Δ_{47} . In addition, the excellent
40 agreement between the presented ostracod data and the carbonate clumped-isotope unified
41 calibration (Anderson et al., 2021) suggests that the use of the latter can be extent to continental
42 biogenic carbonates that do not present a kinetic effect. The ostracod- Δ_{47} thermometer constitutes

43 a reliable tool for continental palaeoclimate reconstructions that can be widely used in freshwater
44 systems in all climatic belts.

45 **INTRODUCTION**

46 The projected future global temperature increase (Stocker et al., 2013 and IPCC, 2021) will
47 affect continental areas differently depending on their geographic position. Lakes are distributed
48 worldwide, on different climate belts, and from boreal to tropical regions. Lacustrine sedimentary
49 archives constitute one of the most useful recorders of past climate changes (Cohen, 2003). Hence,
50 their study provides the means to better constrain future climate variability and foresee mitigation
51 or adaptation measures to preserve the ecosystem on the continents where human societies thrive
52 (Gornitz, 2009). Because of their smaller size compared to oceans, lakes are more sensitive to
53 environmental variations and thus record even rapid climatic events. In paleoclimatology the
54 reconstruction of past temperatures and hydrology in lake-waters, which reflects atmospheric
55 changes, is of particular interest.

56 Since the work of McCrea (1950) and Urey et al. (1951), stable oxygen isotopic analyses in
57 carbonates constitute an essential component of paleoclimatology. Variations in carbonate $\delta^{18}\text{O}$ in
58 lacustrine settings are commonly attributed to changes in lake water temperature and/or in the
59 isotope composition of the lake. Today, well-constrained lacustrine temperature reconstructions
60 remain challenging as the isotopic composition of water depends on the lake biological
61 productivity and on the hydrological balance of the lake, including the source area of the
62 evaporated water and the precipitation/evaporation ratio (Leng and Marshall, 2004). Therefore, a
63 major limitation of the conventional stable isotope $\delta^{18}\text{O}$ technique for paleotemperature
64 determination is the difficulty to discriminate between the temperature and isotopic composition
65 of the water.

66 Carbonate clumped-isotope (Δ_{47}) paleothermometry (Eiler, 2007) opens the way to more precisely
67 determined lacustrine paleotemperatures. The Δ_{47} technique is based on the temperature-dependent
68 abundance of ^{13}C - ^{18}O bonds in CO_2 . The increased abundance of these bonds in carbonate is
69 associated with decreasing water temperatures, providing a direct insight in carbonate temperature
70 precipitation with an accuracy of $\sim \pm 2$ °C. It is expressed using the Δ_{47} notation, which corresponds
71 to the small overabundance of ^{13}C - ^{18}O bonds relative to a stochastic isotopic distribution.

72 The Δ_{47} presents three main advantages over conventional stable isotope $\delta^{18}\text{O}$ technique:

- 73 • Δ_{47} is only temperature-dependent, and not influenced by the water isotopic composition
74 (Schauble et al., 2006).
- 75 • Δ_{47} combined with the $\delta^{18}\text{O}$ of carbonate, reconstructs the isotopic composition of the water
76 from which the carbonate precipitated ($\delta^{18}\text{O}_w$) and thereby changes in the precipitation
77 over evaporation relationship.
- 78 • Δ_{47} values are not affected by a so-called “vital effect” (i.e., disequilibrium between the
79 water isotopic signal and the one recorded by the organisms) for several organisms such as
80 foraminifera and molluscs (Tripathi et al., 2010; Grauel et al., 2013; Peral et al., 2018;
81 Meinicke et al., 2020; Huyghe et al., 2022; de Winter et al., 2022).

82 The clumped paleothermometer is not commonly applied to lacustrine carbonates because the
83 traditional micro- and macrofossils used for marine paleoclimate Δ_{47} studies (e.g., foraminifera,
84 gastropods, and bivalves) are rare or absent in lakes. Δ_{47} technique has been performed on fossil
85 ostracods, common in lacustrine sediments, by Song et al., 2022 and Yue et al., 2022 to reconstruct
86 paleotemperatures and paleoelevation. Yet these authors did not explicitly test the robustness of
87 this paleothermometry technique and the absence of a vital effect prior to its application. In this

88 study, we fill this scientific need by carrying out Δ_{47} analyses on living ostracods that precipitated
89 their shells under known temperatures.

90 Ostracods are small aquatic crustaceans (mostly 0.3-5 mm) with a stable low-Mg calcite shell
91 mineralogy, which makes them ideally suited for targeted geochemical analyses (Holmes and De
92 Deckker, 2012). They secrete their shell relatively rapidly (Chivas et al., 1983) starting from ions
93 present in the host water at the time of valve calcification (Turpen and Angell, 1971). Ostracods
94 grow by moulting and undergo eight juvenile stages (designated in descending order from A-1
95 with A-8 being the first instar) in their development before reaching adulthood (A). They occur in
96 almost all aquatic environments and have an excellent and abundant fossil record. Because of their
97 high sensitivity to the variation of several ecological variables (i.e., pH, salinity, temperature, water
98 level, and oxygen saturation), ostracods are valuable proxies to reconstruct past environmental and
99 climatic changes (Ruiz et al., 2013; Marchegiano et al., 2018, 2019, 2020, Marchegiano and John,
100 2022).

101 The unified Δ_{47} calibration recently published by Anderson et al., (2021) may be applied across a
102 wide range of natural carbonate material precipitated at temperatures between 0.5 to 1100°C. It
103 includes exclusively authigenic continental carbonates (i.e., tufa, travertines, speleothems) and
104 synthetic carbonates. However, the good correspondence with previous published calibrations on
105 foraminifera (e.g., Meinicke et al., 2020; Peral et al., 2018) and marine bivalves data (Huyghe et
106 al., 2022; de Winter et al., 2022) confirm its validity also in biogenic carbonate that do not present
107 any isotopic disequilibrium. Yet, temperatures data from continental biogenic carbonates remain
108 missing to date.

109 This study improves the current understanding of the application of Δ_{47} to lacustrine environments
110 with a particular focus on ostracod shells and it provides a new tool to reconstruct continental
111 climate changes using lake sediments. This article aims to:

- 112 • Determine, for the first time, the relationship between Δ_{47} and precipitated temperature of
113 freshwater ostracod shells by using ostracods grown under known temperatures (from lab
114 cultures and natural environments)
- 115 • Investigate the potential so-called “vital-effects”. This test relies on measuring of different
116 ostracod species growing in the same environment and at the same temperatures. The
117 selected ostracod species are among the most widespread and easily recovered in both
118 recent and fossil records.

119 **MATERIALS AND METHODS**

120 **Ostracod shells**

121 The ostracod species *Heterocypris incongruens* (Ramdohr, 1808) was cultured at the Royal
122 Belgian Institute of Natural Science (Brussels-Belgium) from September to December 2021 under
123 controlled temperature at 23°C (\pm 0.5°C) (Fig.1). Living ostracods were collected in their natural
124 environments during two field campaigns in January (Ghent-Belgium) and February (Granada-
125 Spain) 2022 (Fig.1). The selected species have one generation per year, and they live only in the
126 months during which they were collected. *Eucypris virens* (Jurine, 1820) and *Bradleystrandesia*
127 *fuscata* (Urine, 1820) were sampled in a monitored temporary pond (Ghent-Belgium) and
128 precipitated their shells between end of December and end of January, at a temperature of 4°C (\pm
129 2°C). *Herpetocypris brevicaudata* (Kaufmann, 1900) specimens were collected in a small river
130 fed by groundwater in the Padul lake area (Granada-Spain). The collected adult precipitated their
131 shells between November and beginning of February a period during which the water temperature

132 is constant at 12°C ($\pm 1^\circ\text{C}$). Water depths at both sites were lower than 10 cm. Adult specimens
133 were selected to prevent a potential size effect in the samples since water isotopic composition and
134 temperature could have changed from the juvenile stage, which can commonly last several weeks,
135 to the adult stage. All the collected ostracods were euthanized and preserved in ethanol (see in
136 Supplementary material for more details)

137 **Clumped isotope analyses**

138 The four samples, representing four different ostracod species that precipitated their shells
139 at three different temperatures (see Tab1), are used for clumped isotope (Δ_{47}) analyses. To avoid
140 contamination, each ostracod is cleaned to remove the organic matter. The two ostracod valves are
141 gently opened to remove the body using two needles and each shell accurately cleaned with a thin
142 brush and ethanol under the microscope. A total of 40 to 60 valves per replicate is needed to assure
143 enough material (0.500 - 0.600 mg per replicate), whereby the exact number depends on the species
144 (i.e., size and thickness of the shells). Each sample is replicated 5 to 11 times (300 - 700 valves
145 per samples in total). The Δ_{47} analyses of ostracod shells are carried out at the Analytical,
146 Environmental & Geo-Chemistry (AMGC) clumped isotope lab of the Vrije Universiteit Brussel
147 (VUB), using a Nu Instruments Perspective-IS stable isotope ratio mass spectrometer (SIRMS) in
148 conjunction with a Nu-Carb carbonate sample preparation system, as described in detail in De
149 Vleeschouwer et al. (2022). The analyses are performed between October 2021 and November
150 2022. The carbonate standard ETH-2 is systematically measured and compared to InterCarb values
151 (Bernasconi et al., 2021) to ensure the measurements quality control. The raw measured Δ_{47} values
152 are processed using the IUPAC Brand's isotopic parameters (Daëron et al., 2016) and converted
153 to the ICDES 90°C scale, using the most recent values for the ETH-1, ETH-3, and ETH-4
154 carbonate reference materials (Bernasconi et al., 2021) within the ClumpyCrunch software. Nu

155 Instruments does not require a background check. Both analytical and calibration uncertainties are
156 propagated to calculate the final uncertainties on temperatures.

157

158 **RESULTS**

159 The Δ_{47} values of the ostracod samples range from 0.603 to 0.6648 with a SE from 0.0104
160 to 0.0148 ‰ (Fig.2 and Tab.1). The availability of sample material directly determined the number
161 of replicated measurements carried out per sample. The large number of replicates per specimen
162 ensures the robustness of the results. The repeatability along the 9 sessions of the standards used
163 to standardize the results and the ostracod samples is of 31.7 ppm and 30.0 ppm respectively. The
164 ETH2 standard, used to ensure and control the quality of the analyses, presents a Δ_{47} value of
165 0.2117 and a standard error (SE) of 0.0057 for 97 replicates. The difference between our measured
166 value of ETH2 and the official one from Bernasconi et al., (2021) (ETH2- Δ_{47} value of $0.2086 \pm$
167 0.0015) is negligible (Δ_{47} of 0.0031) and falls within the calculated SE. This confirms both the
168 accuracy and precision of the Δ_{47} analyses presented in this study. The homogeneity in the $\delta^{18}\text{O}$
169 and $\delta^{13}\text{C}$ values (supplementary material S1) indicate that the non-linear mixing effect (Deflies
170 and Lohmann, 2015) did not affected our measurements.

171 **DISCUSSION**

172 **Relationship between ostracod- Δ_{47} and temperature**

173 The Δ_{47} – T relationship for ostracod shells is provided by measuring clumped isotopes on
174 four species that precipitated their shells at three different temperatures. Mean measured Δ_{47} values
175 follow the principle of clumped-isotope technique (Eiler et al., 2017) by decreasing with an
176 increase in temperatures (Fig.2 and Tab.1). Indeed, they strongly correlate with the known
177 calcification temperature. The use of a large number of valves (between 300 and 700) per sample

178 ensures the consistency of the analyses. Fig.2, shows the ostracod data compared with the unified
179 calibration and those from recent studies on both biogenic carbonates and inorganic speleothems
180 (see references in Fig.2). All the measured ostracod samples fall within the 95% confidence
181 interval of the regression line, suggesting that clumped-isotope thermometry can be applied to
182 ostracods shells.

183 **Vital effect in ostracods- Δ_{47}**

184 The potential ostracod vital effect was assessed by measuring Δ_{47} on two different species
185 that lived at the same time and environment and precipitated their shells at the same temperature.
186 The long-lasting problem of the species-dependent vital effect, which currently limits
187 unambiguous paleotemperature reconstructions based on $\delta^{18}\text{O}$, Mg/Ca and other commonly
188 applied proxies, remains an active field of research. A vital effect exists on the $\delta^{18}\text{O}$ signal of
189 several ostracod species (e.g., Grafenstein et al., 1999; Holmes & Chivas, 2002) but so far no study
190 has investigated whether it also affects their clumped-isotope composition. The absence of a
191 consistent offset ($\Delta_{47} 0.002 \pm 0.01\text{‰}$, Tab.1), within the analytical uncertainty, between the species
192 *E. virens* and *B. fuscata* suggests the absence of a vital effect at the genus and species level. This
193 is reinforced by the observation that all four species, coming from lab culture and different natural
194 environments (i.e., river and temporary pond) record the corresponding temperatures.
195 These findings show that the ostracod- Δ_{47} thermometer is fully applicable throughout geological
196 time, independently of the ostracod species, and in all kinds of freshwater environments where
197 ostracods live, irrespective of the geographical area.

198

199 **Implication for palaeotemperature reconstructions**

200 Ostracods show the same $T-\Delta_{47}$ relationship as all other carbonates, supporting a common
201 temperature relationship between biogenic and authigenic carbonates from both marine and
202 continental environments (Fig.2). The strong agreement between all the datapoints from Fig.2 and
203 the unified calibration (Anderson et al., 2021) supports that ostracods are not affected by isotopic
204 disequilibrium effects. Consequently, the unified calibration is applied here to convert ostracod-
205 Δ_{47} values in temperatures (Tab.1). The difference between the calculated temperatures and the
206 temperature measured in situ is between 0.5 and 1 °C, which falls within the uncertainties of the
207 measurements (SE from 0.8 to 1.7 °C). De Winter et al., (2022), suggests that for lower earth
208 surface T, the combined foraminifera-based calibration from Meinicke et al., 2020 is more suitable.
209 However, our data (Tab.1) appear to be in better agreement with the unified calibration. This
210 difference may result from a more robust constraint on temperatures of calcification rather than
211 using isotopically estimated temperatures as done in Meinicke et al. (2021).

212 **CONCLUSIONS**

213 Ostracod- Δ_{47} is a new freshwater paleothermometer to reconstruct temperature with a
214 precision of around $\pm 2^\circ\text{C}$. The absence of a vital effect enables the use of the paleothermometer
215 with all ostracod species in different lacustrine systems and throughout geological time. This
216 makes the ostracod- Δ_{47} thermometer a powerful tool to reconstruct past continental climate
217 change. Because of the good agreement between the ostracod data and the unified calibration
218 (Anderson et al., 2021), the latter can be used for future application of clumped isotope in
219 ostracods. The establishment of this new lacustrine proxy opens the door to new high-resolution
220 continental paleoclimate and paleoenvironmental reconstructions and therefore has the potential
221 to become a key tool in future lacustrine research. The ostracod- Δ_{47} thermometer is of particular
222 importance in the upcoming global warming as the understanding of its effect on specific

223 continental areas remains limited and its reconstruction challenging because of the large
224 environmental variability on the continents.

225 Furthermore, to reconstruct reliable paleotemperature is important to know the life history
226 (i.e., calcification season) of the used ostracod species. It is, thus, preferable to use only adult
227 valves to avoid erroneous species identification and to reduce the inter sample temperature
228 variability that could be caused by the different timing of shell calcification among the 8 moulting.

229

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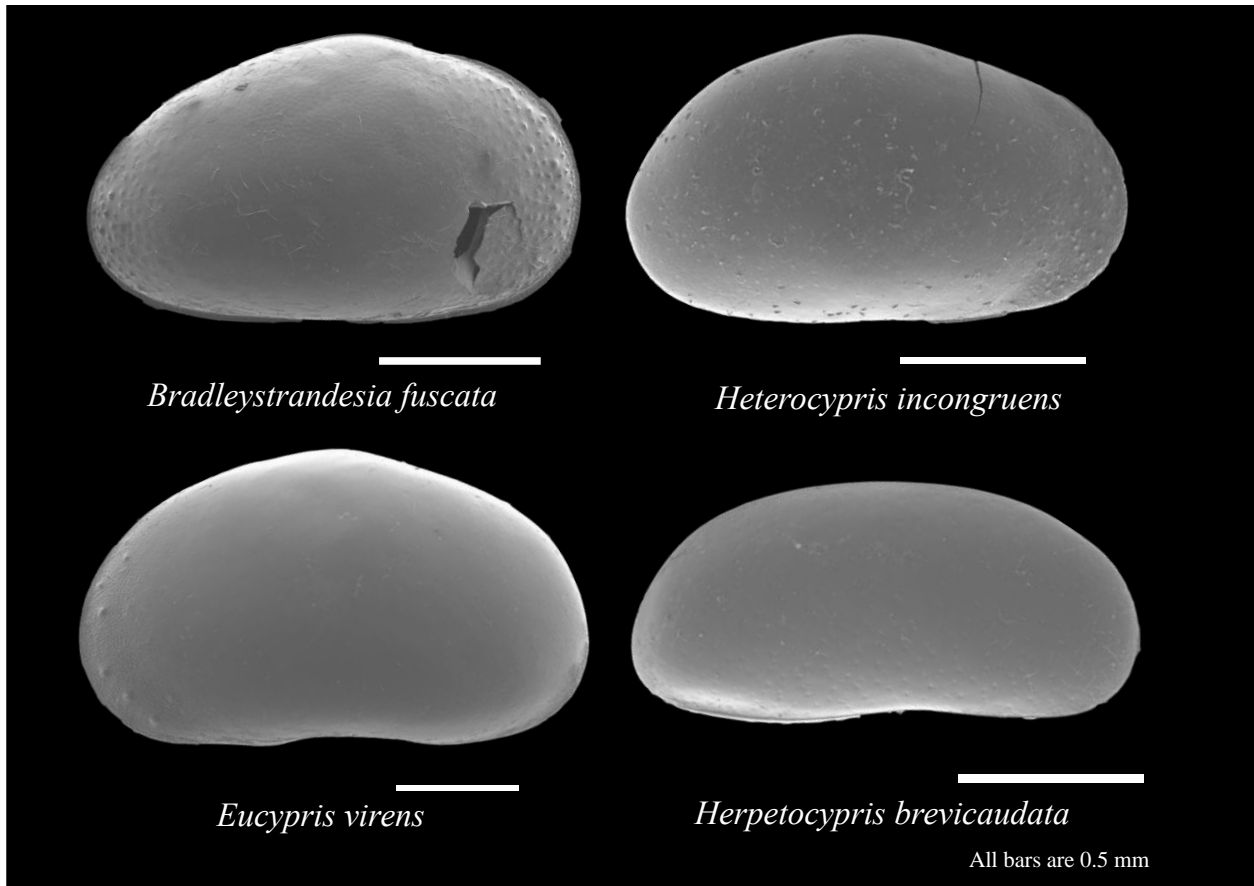
336 FIGURES AND TABLES

337

| Samples name | Species | Environmental T (°C) | N | Δ_{47} (‰) I CDES | 1SE | T(°C) Anderson et al., 2021 | T(°C) Meinicke et al., 2020 |
|--------------|------------------------|----------------------|----|--------------------------|--------|-----------------------------|-----------------------------|
| OSTRA-1 | <i>E. virens</i> | 4 (± 2) | 11 | 0.6629 | 0.0104 | 4 (± 1.7) | 5.5 (± 1.7) |
| OSTRA-2 | <i>H. brevicaudata</i> | 12 (± 1) | 10 | 0.6364 | 0.0112 | 11.5 (± 2.1) | 13 (± 2.1) |
| OSTRA-3 | <i>B. fuscata</i> | 4 (± 2) | 9 | 0.6648 | 0.0109 | 3.5 (± 1.8) | 5 (± 1.8) |
| OSTRA-4 | <i>H. incongruens</i> | 23 (± 0.5) | 5 | 0.603 | 0.0148 | 22 (± 3.4) | 23 (± 3.4) |

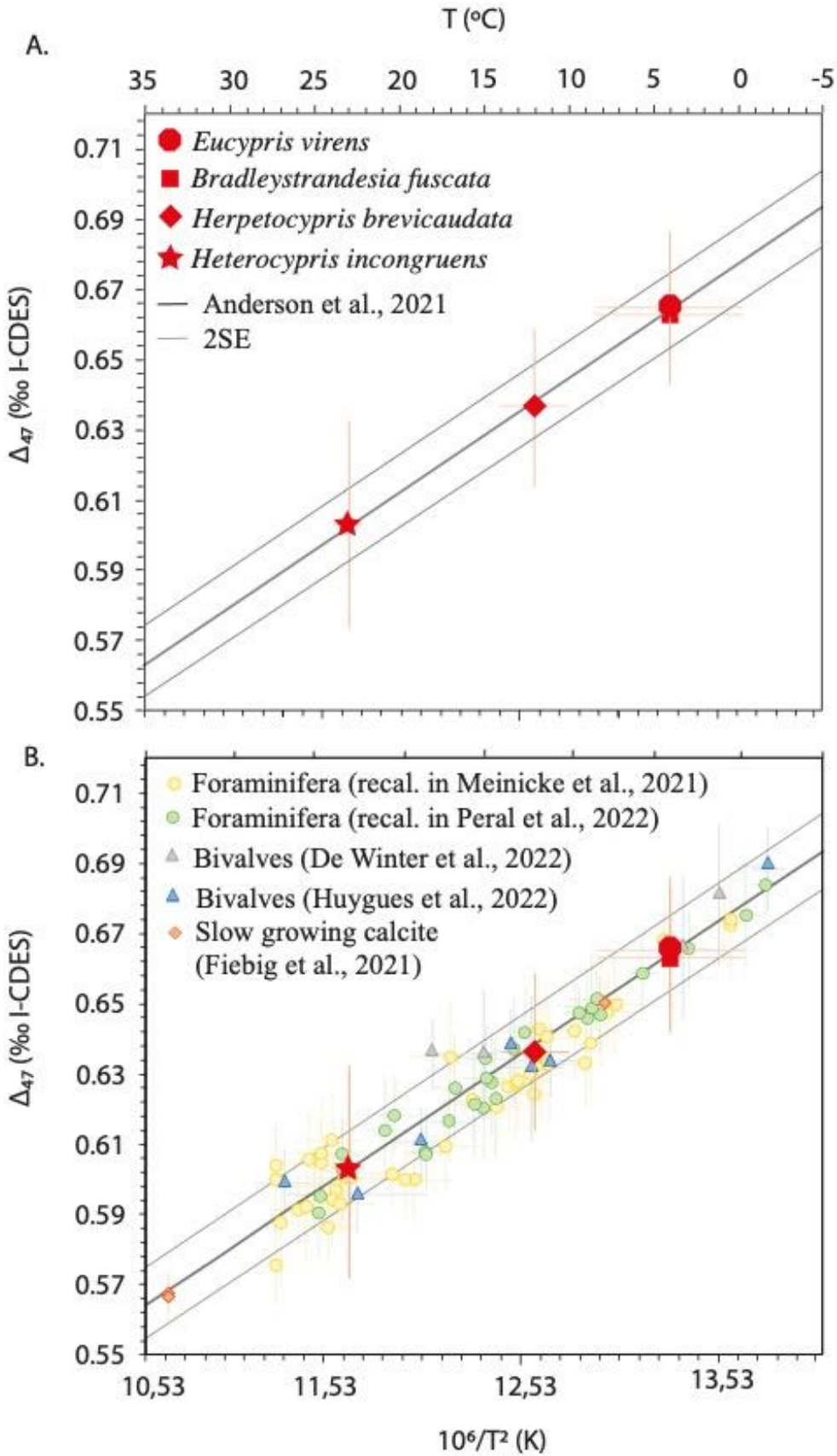
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339 Table 1: Ostracod clumped results. N = number of replicates per each sample.



340

341 Figure 1: SEM pictures of ostracod shells



343 Figure 2: A. Clumped isotope (Δ_{47}) measured on living ostracods shells are presented against the
344 known precipitated temperatures. The ostracod- Δ_{47} values are compared to the unified calibration
345 of Anderson et al., 2021 (black line; grey lines indicate the 2SE confidence interval). B. Ostracod
346 clumped-isotope data against precipitated temperatures compared with clumped-isotope data from
347 marine biogenic carbonate and continental slow grown calcite speleothems. All Δ_{47} errors are
348 expressed at 2SE. The horizontal lines indicate the variations of the temperature measured in the
349 field per each sample.

350

351 **SUPPLEMENTARY MATERIAL**

352

353 **LIVING OSTRACODS FROM LAB AND NATURAL ENVIRONMENTS**

354

355 **Lab Grown *Heterocypris incongruens***

356

357 The ostracod species *Heterocypris incongruens* (Ramdohr, 1808) was cultured at the Royal
358 Belgian Institute of Natural Science (Brussels-Belgium) from September to December 2021.

359 This species is cosmopolitan and very common in shallow seasonal ponds and small permanent
360 water bodies (Meisch, 2000). It tolerates a wide range of air temperatures, in the summer from 5
361 to 28 °C and in winter from -15 to 18°C (Horne and Mezquita, 2008). The length of its valves can
362 vary from 1.2 to 1.9 mm for female and from 1.2 to 1.3 for male specimens.

363 Living specimens of *H. incongruens* were sampled using a rectangular hand-net (120 μ m mesh
364 size) in a temporary pond in a nature area called “Drongengoed” (Ursel, Belgium - 51.151N,
365 3.474E). The main population was kept alive in an aquarium filled with fine sand sediments (ca.

366 2cm thickness at the bottom) and water coming from their natural environment. Every three days
367 they were fed with *Spirulina* algae. For this study, juvenile specimens were selected from the last
368 three stages (A-1, A-2, A-3 valves). These specimens were isolated to make satellite populations
369 in a (self-made) incubator system under controlled temperature at 23°C (\pm 0.5°C) and 50-50 Light-
370 Dark diurnal circle. Deceased adult *H. incongruens* specimens were removed from the isolated
371 temperature-controlled aquarium and their valves were used for Δ_{47} analysis.

372

373

374 **Ostracod collected in natural environments**

375

376 Living ostracods were collected in their natural environments using a hand-net (120 μ m mesh size)
377 during two field campaigns in January (Ghent-Belgium) and February (Granada-Spain) 2022.
378 *Eucypris virens* (Jurine, 1820) and *Bradleystrandesia fuscata* (Jurine, 1820) come from a
379 monitored temporary pond (Ghent-Belgium) only a few cm deep. *Eucypris virens* is a winter-early
380 spring species (December/April) widely distributed (Europe, Greenland, Azores, North Africa,
381 Central Asia, China, North America and Middle East) that prefers to live in temporary habitats,
382 mostly ponds, which dry up before summer, as well as in streams, rivers, and rice fields (Meisch
383 2000). It tolerates a wide range of air temperatures (summer from 13 to 28 °C (July) and winter
384 from -8 to 14°C (January), (Horne and Mezquita 2008)). The length of its valves varies from 1.4
385 to 2.4 mm in females and from 1.4 to 2.1 in males. Adults have a life span of about 2 months and
386 die shortly after egg production (Meisch 2000). The collected adult specimens precipitated their
387 shells between December and beginning of January 2022 under temperature of 4°C (\pm 2°C).

388 *Bradleystrandesia fuscata* is a species characteristic of seasonal temporary ponds, mainly from
389 autumn to spring and very rare during summer. In winter ponds larvae develop faster than in those
390 in other seasons and they live only for a few weeks (Meisch 2000). The species tolerates a large
391 range of air temperatures (summer from 6 to 26 °C and winter from -15 to 12°C, Horne and
392 Mezquita 2008). It has been found in Europe, Turkey, Middle East and North America (Meisch
393 2000), in the Neotropics (Coviaga et al., 2018) and as invasive species also in Western Australia
394 (Koenders et al. 2012, 2016). Female valve length varies from 1.1 to 1.5 mm. The collected adult
395 specimens precipitated their shells between December and the beginning of January under
396 temperatures of 4°C ($\pm 2^\circ\text{C}$).

397 *Herpetocypris brevicaudata* (Kaufmann, 1900) specimens were collected in a small and very
398 shallow river (less than 20 cm deep), with continuous running water, that is fed by groundwater in
399 the Padul lake area (Granada-Spain). This species was found in Europe and North Africa and
400 prefers springs, slowly flowing streams, and the littoral zone of lakes. It tolerates a large range of
401 air temperatures (summer from 16 to 33 °C (July) and winter from -1 to 16°C (January) (Horne
402 and Mezquita 2008)). Female valve length varies from 1.3 to 2.2 mm. Little is known about the
403 life cycle of *H. brevicaudata* but it is supposedly similar to that of its closest congener, *H. reptans*
404 (Baird, 1835), which produce two generations per year (April/August and November/overwinter).
405 The collected adults precipitated their shells between November and the beginning of February,
406 which is a period during which the water temperature is constant at 12°C ($\pm 1^\circ\text{C}$).

407 All the collected ostracods were euthanized and preserved in 70% ethanol.

408

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428 The raw data are available upon request and will be deposited after publication in EarthChem.

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