1	Convergence in simulating global soil organic carbon by structurally different models
2	after data assimilation
3	Feng Tao ^{1, 2*} , Benjamin Z. Houlton ³ , Yuanyuan Huang ⁴ , Ying-Ping Wang ⁵ , Stefano Manzoni ⁶ ,
4	Bernhard Ahrens ⁷ , Umakant Mishra ^{8, 9} , Lifen Jiang ¹⁰ , Xiaomeng Huang ^{2, *} , Yiqi Luo ¹⁰
5	
6	¹ Department of Ecology and Evolutionary Biology, Cornell University, Ithaca, NY, USA
7	² Department of Earth System Science, Ministry of Education Key Laboratory for Earth
8	System Modelling, Institute for Global Change Studies, Tsinghua University, Beijing, China
9	³ Department of Ecology and Evolutionary Biology and Department of Global Development,
10	Cornell University, Ithaca, NY, USA
11	⁴ Key Laboratory of Ecosystem Network Observation and Modeling, Institute of Geographic
12	Sciences and Natural Resources Research, Chinese Academy of Sciences, Beijing, China
13	⁵ CSIRO Environment, Private Bag 10, Clayton South VIC 3169, Australia
14	⁶ Department of Physical Geography and Bolin Centre for Climate Research, Stockholm
15	University, Stockholm, Sweden
16	⁷ Max Planck Institute for Biogeochemistry, Jena, Germany
17	⁸ Computational Biology and Biophysics, Sandia National Laboratories, Livermore, CA, USA
18	⁹ Joint BioEnergy Institute, Lawrence Berkeley National Laboratory, Emeryville, CA, USA
19	¹⁰ School of Integrative Plant Science, Cornell University, Ithaca NY, USA
20	
21	Correspondence to: Feng Tao, <u>phx.tao@gmail.com</u> , Xiaomeng Huang, <u>hxm@tsinghua.edu.cn</u>
22	
23	Statement: This manuscript is a non-peer reviewed preprint submitted to EarthArXiv.
24	

25 Abstract:

The current generation of biogeochemical models produce large uncertainty in carbon-26 climate feedback projections. Structural differences in these models have been identified as a 27 28 major source of inter-model uncertainties when simulating soil organic carbon (SOC) dynamics worldwide. However, parameterization could also play a role, particularly when 29 common observational data are used to constrain model simulations. Here we demonstrate 30 31 the critical role of observational data in reducing model-based uncertainty in global estimates of SOC. We applied the PROcess-guided deep learning and DAta-driven modeling (PRODA) 32 33 approach to constrain both a microbial implicit model based on first-order kinetics (i.e., Community Land Model version 5, CLM5) and a microbial explicit model based on 34 Michaelis-Menten kinetics (i.e., CarbOn cycle and Microbial PArtitioning Soil model, 35 36 COMPAS) with >50,000 globally distributed SOC vertical profiles. Overall, the two 37 constrained models predicted similar carbon transfer efficiency, baseline decomposition rate, and environmental effects on carbon fluxes. These converged model components contributed 38 39 to similar SOC patterns simulated by the two structurally different biogeochemical models. Carbon input allocation and vertical transport were less constrained by SOC profile data and 40 require other data sets to constrain. Moreover, after being constrained by SOC observations, 41 the Michaelis constant in COMPAS tends to be much larger than its corresponding substrate 42 43 concentration in SOC decomposition. Thus, the Michaelis-Menten kinetics in the COMPAS 44 model can be approximated by multiplicative kinetics (i.e., first order with respect to both 45 donor and received pool carbon) in these global scale simulations. Our results highlight the importance of observational data in informing model development and constraining model 46 47 predictions.

48

50 1. Introduction

Soils store more carbon than the atmosphere and vegetation combined (Ciais et al. 2014, 51 Jackson et al. 2017). A small change in soil carbon storage can significantly impact the 52 53 atmospheric carbon dioxide concentration and the climate. Substantial research has been conducted to understand the factors that underly the formation of soil organic carbon (SOC) 54 and its persistence. A conventional paradigm focuses on the balance between plant carbon 55 56 input as the source of SOC and organic matter decomposition that mineralizes SOC with different recalcitrance as CO₂ back to the atmosphere (Schmidt et al. 2011). Recently, 57 58 increasing evidence suggests that soil microorganisms are the key determinant in partitioning 59 soil carbon into accumulation versus loss (Cotrufo et al. 2013, Cotrufo et al. 2015, Bradford et al. 2016, Tao et al. 2023b). These two paradigms are the conceptual foundation of two 60 61 classes of process-based models used to simulate global SOC dynamics. Because these model 62 classes have distinctive structures that reflect different underlying theories and assumptions on soil carbon dynamics, large uncertainties in the simulated SOC emerge among models. 63 64 Diverging simulations of SOC storage and its spatial distributions across the globe hinder a better understanding of the soil carbon cycle and its feedback to climate change (Todd-Brown 65 et al. 2013, Ciais et al. 2014, Luo et al. 2016). 66

In simulating soil carbon dynamics, state-of-the-art process-based models structurally 67 68 differ in classifying soil carbon pools, quantifying SOC decomposition kinetics, and 69 representing carbon transfer processes (Chandel et al. 2023). To represent the heterogeneity 70 of SOC, soil carbon is separated into conceptual pools with different turnover rates that indicate their decomposability. For example, models derived from the Century model (Parton 71 72 et al. 1987) differentiate substrates according to turnover times, with labile substrates that cycle rapidly (i.e., active SOC) and chemically or physically protected pools that cycle 73 74 slowly (i.e., slow and passive SOC). Recently formulated process-based models, carbon

pools are defined as measurable entities that can be validated with field observations

76 (Abramoff et al. 2022) - e.g., microbial biomass, dissolved organic carbon, and mineral-

77 associated organic carbon.

In representing SOC decomposition, a theory dating back to the 1980s portends that 78 organic matter decay in soils follows first-order kinetics: $\frac{dSOC}{dt} = -k \times SOC$, where the loss 79 rate of SOC (i.e., k) is independent of its pool size (i.e., SOC) (Parton et al. 1988) and thus, 80 81 the SOC storage changes over time is proportional to its pool size (Forney and Rothman 82 2012). With increasing evidence pointing to microorganisms as a key factor in soil carbon dynamics, a newer generation of models have explored the possibility of nonlinearity in SOC 83 84 decomposition (Schimel and Weintraub 2003, Allison et al. 2010, Georgiou et al. 2017, Wang et al. 2021). Among various nonlinear structures, the Michaelis-Menten kinetics (i.e., $\frac{dSOC}{dt}$ = 85 $-v \frac{ENZ \times SOC}{K + SOC}$) consider the interplay between the substrate (i.e., SOC) and the extracellular 86 87 enzymes (i.e., ENZ) that catalyze the decomposition of organic matter (Schimel and Weintraub 2003, Wilson and Gerber 2021). Specifically, parameter v specifies the maximum 88 SOC decomposition rate at its saturated content for a given enzyme content. The inverse of 89 the Michaelis-Menten constant (K) specifies the enzyme's affinity for its substrate in a 90 91 catalyzed reaction.

Process-based models also differ in allocating the decomposed carbon to other carbon pools or heterotrophic respiration as CO₂. While soil microbes mineralize SOC into CO₂ through their metabolism, transfers of decomposed carbon from one pool to another could result from either an exclusive effect of microbial processes or an integrative effect of biological, chemical, and physical reactions (i.e., including both microbial and non-microbial transfer). Specifically, when a model explicitly defines a microbial biomass carbon pool, , carbon received by this pool is partitioned according to microbial carbon use efficiency

99 (CUE) – i.e., the ratio of carbon assimilated in new biomass over carbon transferred from the substrate (Gever et al. 2016, Manzoni et al. 2018, Tao et al. 2023b). Correspondingly, carbon 100 transfers among compartments that happen without microbial carbon assimilation can be 101 102 interpreted as results from other biochemical processes (e.g., microbial exudation and mortality) or organo-mineral interactions (Tao et al. 2023b). In contrast, for models without 103 explicit representation of microbial biomass and assimilation processes, carbon transfer 104 105 implicitly integrates the effects of both microbial physiology and other chemical or physical reactions. Depending on the model structure, a range of relations between long-term SOC and 106 107 microbial traits such as CUE or carbon inputs to soil emerge (Wutzler and Reichstein 2008, Georgiou et al. 2017, He et al. 2023). 108

In addition to structural differences among varieties of process-based models, 109 110 parameter values that quantify the strength and represent properties of different processes in the soil carbon cycle also contribute to the uncertainty of model simulations (Luo and Schuur 111 2020). Most current Earth system models adopt the Century-type model structure using first-112 113 order SOC decomposition kinetics. Notwithstanding their structural similarity, varying parameterizations among different models contribute to the divergent estimates of SOC 114 storage both at the site-level and across the globe (Todd-Brown et al. 2013, Luo et al. 2015). 115 Moreover, the same model with different parameterizations could also generate varying 116 117 patterns of SOC and key model components, such as microbial CUE (Tao et al. 2023b) and plant carbon input (Tao et al. 2023a). However, parameterization and model structure are not 118 119 fully independent in affecting model simulation: different model structures can in some cases converge to similar results in the long term via parameter adjustments. For example, the 120 121 Michaelis-Menten kinetics, when the affinity of the enzyme for its substrate is extremely low, such that the Michaelis-Menten constant is much higher than the substrate concentration, the 122

nonlinear decomposition kinetics will converge to linear kinetics with respect to the substrate(Lasaga 1998, Wilson and Gerber 2021).

While simulations by structurally distinctive models with different parameterizations 125 126 present a range of possibilities under specific theories and assumptions, calibrating model simulations against observational data helps identify the most probable mechanistic 127 explanation that fits reality. Data assimilation is a suite of techniques that compare the model 128 129 simulation results with different parameterizations against observed counterparts and adjust the model parameter values to the set with which the process-based model simulations best fit 130 131 observations (Luo et al. 2011). Conventional data assimilation techniques such as Bayesian inference-based Markov Chain Monte Carlo (MCMC) method have been used at the site 132 level to tune process-based models to better simulate soil carbon cycle (Xu et al. 2006, Li et 133 134 al. 2016). Recently, the newly developed PROcess-guided deep learning and DAta-driven 135 modeling (PRODA) approach (Tao and Luo 2022) integrates the site-level MCMC-based data assimilation results with deep learning to optimize the parameterization for global SOC 136 137 simulations and reveals key mechanisms underlying SOC storage (Tao et al. 2020, Tao et al. 2023b). Here we hypothesize that with the same external forcing and observational 138 constraint, simulations on global SOC by two models of different structures (i.e., CLM5 and 139 COMPAS, see Methods below for detailed descriptions) can converge after being optimized 140 141 by the PRODA approach. The convergence in simulating SOC will be achieved from the 142 well-calibrated key processes in the soil carbon cycle despite structural differences among 143 process-based models. Meanwhile, results of PRODA-optimized model simulation can also identify the most probable model structure that best fit observed SOC data across the globe. 144 145

147 2. Materials and Methods

148 2.1. Global vertical soil organic carbon profiles

149 We obtained soil organic carbon data in globally distributed soil profiles from the World Soil Information Service (WoSIS) and other data sources. WoSIS compiled soil data, 150 after quality assessment, from soil profiles distributed across 173 countries (Batjes et al. 151 2020). The 2019 snapshot of the WoSIS dataset consists of 111,380 soil profiles with SOC 152 content information (unit: $g C kg^{-1}$ soil). We estimated the SOC stock ($g C m^{-3}$) by 153 SOC Stock = SOC Content \times BD (Yigini et al. 2018), where BD is the bulk density of soil 154 (g m⁻³). Note that SOC stock were multiplied by $1 - \frac{G}{100}$ to account for the volumetric coarse 155 156 fragment fraction (G, unit: %) at each grid of the global map (data source: SoilGrids, https://soilgrids.org). When the measured bulk density was absent in the dataset, we used a 157 pedo-transfer function to estimate it (Grigal et al. 1989, Yigini et al. 2018): $BD = \alpha + \alpha$ 158 $\beta \times \exp(-\gamma \times OM)$, where OM is organic matter, calculated as SOC×1.724, with SOC 159 160 content in per cent (%); α , β , and γ are fitting parameters. After fitting data of WoSIS (i.e., 78,913 layers from 16,248 profiles that simultaneously recorded bulk density and SOC 161 162 content) to this equation, we obtained $\alpha = 0.32$, $\beta = 1.30$, and $\gamma = 0.0089$. The pedo-transfer function explained 55% of the variation in the bulk density. In addition, we obtained data 163 from a previous study (Mishra et al. 2020) and the Northern Circumpolar Soil Carbon 164 Database (NCSCD) (Hugelius et al. 2013). This dataset contained 2,546 soil profiles with 165 SOC stock (g C m⁻³) information for permafrost regions in North America, northern Eurasia, 166 and Qinghai-Tibet Plateau. In total, we obtained data from 113,926 soil profiles as the raw 167 data. The geographical distributions of all soil profiles are shown in Supplementary Figure 1. 168 Not all the soil profiles are used in this study. We pre-processed the 113,926 SOC 169 170 profiles to ensure the quality of the data before we conducted our analysis. We first excluded SOC profiles with no more than two observation layers or the maximum observation depths 171

of no deeper than 50 cm from this study as such data do not provide enough information onkey processes underlying SOC storage. After that, we obtained 72,377 profiles.

To further examine the quality of data along the vertical profiles, we conducted data 174 assimilation for each of the 72,377 SOC vertical profiles with both the Community Land 175 Model version 5 (CLM5) and the CarbOn cycle and Microbial PArtitioning Soil model 176 (COMPAS) using the Markov Chain Monte Carlo (MCMC) method. Model structures of 177 178 CLM5 and COMPAS are described in sections 4.2 and 4.3, respectively. The method of data assimilation is briefly described in section 4.4 below and in detail by Tao et al. (2020). 179 180 We used two statistics (i.e., G-R statistic and Nash-Sutcliffe modelling efficiency coefficient) to ensure the quality of calibration against SOC data along the vertical profiles. 181 We calculated the Gelman-Rubin (G-R) statistic (Gelman et al. 2014) for each of the SOC 182 183 profiles to test the convergence of the site-level data assimilation results after running three independent series of MCMC simulations (see Section 2.6 for details of MCMC). A G-R 184 value approaching 1.0 suggests well converged data assimilation results. A large G-R value, 185 186 in contrast, indicates inconsistent data assimilation results from independent MCMC 187 simulations, and such results may not be trusted. Therefore, we set a threshold of G-R = 1.05and excluded SOC profiles with G-R > 1.05 from our analysis in this study. The remaining 188 66,935 profiles for CLM5 and 59,476 for COMPAS went through the next analysis below. 189 190 We found it was more difficult for independent MCMC simulations to converge when using 191 COMPAS model in data assimilation, probably because of its nonlinearity. Thus, the final 192 adopted profiles for COMPAS are fewer than those for CLM5.

We used the Nash-Sutcliffe modelling efficiency coefficient (Janssen and Heuberger
194 1995) (NSE) to evaluate the effectiveness of retrieving information from observations by
process-based models. NSE is expressed as:

196
$$NSE = 1 - \frac{\sum (obs_i - mod_i)^2}{\sum (obs_i - \overline{obs_i})^2}$$
(1)

197 A value of NSE close to 1 indicates that SOC distributions with depth can be well captured by process-based models so that information contained in the observations can be retrieved to 198 evaluate processes underlying SOC storage. A small value of NSE indicates that the model 199 200 cannot capture the variability in the data, suggesting that such SOC vertical profiles may not offer enough information on the processes underlying SOC storage investigated in this study. 201 We set a threshold NSE = 0.0 to exclude SOC profiles from the analysis. We randomly 202 203 selected a subset of these excluded SOC profiles to visually cross-check their shapes. We found that the thresholds are effective for controlling the quality of data. 204

After all the data pre-processing procedures, we eventually obtained data assimilation results from 62,931 soil profiles for CLM5 and 57,267 soil profiles for COMPAS with which we estimated global SOC storage and its components. Our data pre-processing criteria did not cause significant discrimination against profiles belonging to specific soil orders or ecosystems or different vertical shapes (Tao et al. 2023b). Thus the main conclusions drawn from this study are unlikely influenced by our data pre-processing criteria.

211

212 **2.2. Model structure of CLM5**

CLM5 is the latest version of the land model of the Community Earth System Model version
2 (CESM2) (Lawrence et al. 2018, Lawrence et al. 2019). SOC dynamics in CLM5 can be
expressed in a uniform matrix equation (Huang et al. 2018, Lu et al. 2020, Luo et al. 2022):

216
$$\frac{d\mathbf{X}(t)}{dt} = \mathbf{B}I(t) + \mathbf{A}\xi(t)\mathbf{K}\mathbf{X}(t) + \mathbf{V}(t)\mathbf{X}(t)$$
(2)

This matrix equation has six components (Supplementary Table 1), including plant carbon
inputs (*l(t)*), carbon input allocation to different pools and depths (*B*), substrate
decomposability (or baseline decomposition rates) (*K*), carbon transfer efficiency (*A*),

220 environmental modifier ($\xi(t)$), and vertical transport (V(t)).

CLM5 describes seven carbon pools in the soil, including four litter pools (i.e., coarse woody debris (indicated by subscript CWD), metabolic litter (ML), cellulose litter (CL), and lignin litter (LL)) and three soil organic carbon pools (i.e., active (aSOC), slow (sSOC), and passive (pSOC) soil organic carbon pools). Each of the carbon pools is simulated at 20 layers to a maximum depth of 8.4 m. The state of different carbon pools (i.e., carbon stocks) can be expressed as:

227
$$X(t) = \begin{bmatrix} x_{CWD}(t) \\ x_{ML}(t) \\ x_{CL}(t) \\ x_{LL}(t) \\ x_{aSOC}(t) \\ x_{sSOC}(t) \\ x_{pSOC}(t) \end{bmatrix}$$
(3)

where each of the 7 block elements (i.e., $x_i(t)$) of X(t) has 20 elements to represent the 20 228 229 soil layers. In total, CLM5 simulates carbon transfer among 140 pools. Consequently, there are 140 dimensions for vector \boldsymbol{B} of carbon input allocation, matrix \boldsymbol{K} of substrate 230 231 decomposability, matrix A of carbon transfer from one carbon pool to another, matrix $\xi(t)$ of environmental modifiers, and matrix V(t) of vertical transport. Plant carbon input (I(t)) is a 232 scalar. In this study, parameters (Supplementary Table 1) that generate the above elements in 233 234 the matrix equation will be optimised by the PRODA approach. Specifically, I(t) is allocated to different litter pools in different layers along the soil 235

- 236 profile via the allocation vector B. Organic carbon in pool vector X(t) is decomposed
- 237 following first-order kinetics as described by matrix *K*:

238
$$K = \begin{pmatrix} k_{CWD} \\ k_{ML} \\ k_{CL} \\ k_{LL} \\ k_{aSOC} \\ k_{sSOC} \\ k_{pSOC} \end{pmatrix}$$
(4)

where \mathbf{k}_i is independent from the state of its corresponding substrate $\mathbf{x}_i(t)$. Moreover, we used the environmental modifier (i.e., $\boldsymbol{\xi}(t)$) to account for the effects of environmental conditions on the decomposition processes. $\boldsymbol{\xi}(t)$ is calculated from functions of soil temperature ($\boldsymbol{\xi}_T$), soil water potential ($\boldsymbol{\xi}_W$), nitrogen and oxygen availability ($\boldsymbol{\xi}_{N-0}$), and soil depth ($\boldsymbol{\xi}_D$).

Organic carbon from any carbon pool is further partitioned by either microbial or nonmicrobial processes between a receiver carbon pool and CO₂ released to the atmosphere. All these processes can be summarised in the *A* matrix:

$$248 = \begin{bmatrix} -1 & 0 & 0 & 0 & 0 & 0 & 0 \\ 0 & -1 & 0 & 0 & 0 & 0 & 0 \\ a_{CL,CWD} & 0 & -1 & 0 & 0 & 0 \\ a_{LL,CWD} & 0 & 0 & -1 & 0 & 0 \\ 0 & a_{aSOC,ML} & a_{aSOC,CL} & 0 & -1 & a_{aSOC,SSOC} & a_{aSOC,pSOC} \\ 0 & 0 & 0 & a_{sSOC,LL} & a_{sSOC,aSOC} & -1 & 0 \\ 0 & 0 & 0 & 0 & a_{pSOC,aSOC} & a_{pSOC,sSOC} & -1 \end{bmatrix}$$
(5)

where all the block elements in the *A* matrix ($a_{i,j}$) are diagonal matrices with the dimension of 20. a_{ij} represents the carbon transfer fraction from the donor (*j*) pool to the recipient (*i*) pool (see carbon transfer flows in **Figure 1**). Note that CLM5 does not differentiate carbon transfers mediated by microbial processes from those mediated by non-microbial processes (e.g., organo-mineral interactions). Thus, $a_{i,j}$ in Equation 8 is a integrative value reflecting carbon transfers contributed by both microbial and non-microbial processes.

The transport matrix *V* of CLM5 is a tridiagonal matrix and describes vertical carbon movement between adjacent soil layers within the same carbon pool via bioturbation and cryoturbation. At steady state, the analytical solution of SOC stock by CLM5 was calculated as $X_{steady \ state} = \left[A\overline{\xi(t)}K + \overline{V(t)}\right]^{-1} \left[-B\overline{I(t)}\right]$, where the overbars indicate the mean values of related matrices ($\xi(t)$ and V(t)) and scalar (I(t)) over the period of forcing data. The

- 260 matrix representation for process-based soil carbon cycle models has been described in detail261 by Huang et al. (2018), Lu et al. (2020), and Luo et al. (2022).
- 262

263 2.3. Structure of COMPAS model

The CarbOn cycle and Microbial PArtitioning Soil (COMPAS) model follows the same 264 structure proposed by (Allison et al. 2010) for SOC dynamics, which is further embedded 265 266 within the structure for 20-layered vertical soil profiles. The description of vertical layers was adopted from CLM5. Organic carbon dynamics represented by COMPAS can be expressed 267 268 by the same matrix framework as shown in Equation 2 (Supplementary Table 2). Yet COMPAS structurally differs from CLM5 in classifying soil carbon pools, expressing 269 270 substrate decomposition, and explicitly describing microbial partitioning processes in carbon 271 transfer.

2/1 transfer.

Equation 2 describes COMPAS with 160 dimensions to represent 8 pools in each of

273 the 20 soil layers. Vector X(t) has 8 block elements to represent four litter carbon pools

274 (indicated by subscripts CWD, ML, CL, and LL) and four soil organic carbon pools (i.e.,

275 dissolved organic carbon (DOC), mineral-associated soil organic carbon (mSOC), microbial

276 biomass (MIC), and extracellular enzymes (ENZ)):

277
$$\mathbf{X}(t) = \begin{bmatrix} \mathbf{x}_{CWD}(t) \\ \mathbf{x}_{ML}(t) \\ \mathbf{x}_{CL}(t) \\ \mathbf{x}_{LL}(t) \\ \mathbf{x}_{DOC}(t) \\ \mathbf{x}_{MIC}(t) \\ \mathbf{x}_{ENZ}(t) \\ \mathbf{x}_{mSOC}(t) \end{bmatrix}$$
(6)

Each of the 8 block elements (i.e., $x_i(t)$) of X(t) has 20 elements to represent the 20 soil layers. Similarly, there are 160 dimensions for vector B, matrix K, matrix A, matrix $\xi(t)$, and matrix V(t). Plant carbon input (I(t)) is still a scalar as in CLM5. Parameters (Supplementary Table 2) that generate the above elements in the matrix equation will beoptimised by the PRODA approach.

283 Different from CLM5, organic carbon pools in vector X(t) of COMPAS can be 284 transferred to recipient pools either through microbial- or enzyme-mediated kinetics, or 285 without going through microbial metabolism. These transfers are described by the baseline 286 decomposition matrix *K*:

287
$$K = diag \begin{pmatrix} k_{CWD} \\ k_{ML} \\ k_{CL} \\ k_{LL} \\ k_{DOC}(x_{DOC}, x_{MIC}) \\ k_{MIC} \\ k_{ENZ} \\ k_{mSOC}(x_{mSOC}, x_{ENZ}) \end{pmatrix}$$
(7)

288 While all the litter organic carbon pools and two mineral organic carbon pools (i.e., MIC and 289 ENZ) are decomposed following first-order kinetics where their baseline decomposition rates 290 are constants, the baseline decomposition rates of DOC and mSOC are functions of carbon pool states. Specifically, the baseline decomposition rate of DOC (a.k.a. the baseline rate of 291 microbial assimilation of DOC) is: $k_{DOC}(x_{DOC}, x_{MIC}) = \frac{v_{max,assim}x_{MIC}}{K_{m,assim}\xi + x_{DOC}}$; the baseline 292 decomposition rate of mSOC is: $\mathbf{k}_{mSOC}(\mathbf{x}_{mSOC}, \mathbf{x}_{ENZ}) = \frac{v_{max,decom}\mathbf{x}_{ENZ}}{K_{m \, decom} \boldsymbol{\xi} + \mathbf{x}_{mSOC}}$. Parameters 293 $v_{max,assim}$ and $v_{max,decom}$ represent the maximum DOC assimilation and mSOC 294 295 decomposition rates, respectively. $K_{m,assim}$ and $K_{m,decom}$ are the Michaelis constants for 296 DOC assimilation and mSOC decomposition, respectively. 297 The COMPAS model also explicitly differentiates carbon transfers in microbial 298 processes from those in non-microbial processes. The decomposed organic carbon is either 299 partitioned by microorganisms to microbial biomass growth versus respiration (i.e., according to the microbial CUE), or alternatively, transferred to other carbon pools with a fraction that 300

301 is not mediated by microbial processes (i.e., non-microbial carbon transfer). All these

302 processes are summarised in the A matrix:

$$303 \quad A = \begin{bmatrix} -1 & 0 & 0 & 0 & 0 & 0 & 0 & 0 \\ 0 & -1 & 0 & 0 & 0 & 0 & 0 & 0 \\ a_{CL,CWD} & 0 & -1 & 0 & 0 & 0 & 0 \\ a_{LL,CWD} & 0 & 0 & -1 & 0 & 0 & 0 \\ 0 & a_{DOC,ML} & a_{DOC,CL} & 0 & -1 & a_{DOC,MIC} & 1 & a_{DOC,MSOC} \\ 0 & a_{MIC,ML} & a_{MIC,CL} & a_{MIC,LL} & a_{MIC,DOC} & -1 & 0 & 0 \\ 0 & 0 & 0 & 0 & 0 & a_{ENZ,MIC} & -1 & 0 \\ 0 & 0 & 0 & 0 & a_{mSOC,LL} & 0 & a_{mSOC,MIC} & 0 & -1 \end{bmatrix}$$

304 Because DOC is always assimilated by the microbes with release of CO₂ (Figure 1), the microbial CUE for DOC (η_{DOC}) equals $a_{MIC,DOC}$. In contrast, organic carbon in the 305 metabolic, cellulose, and lignin litter pools is decomposed by microbes following first-order 306 kinetics to generate CO₂ and grow biomass while a fraction of litter organic carbon is broken 307 down without going through microbial metabolism and, thus, directly transferred to DOC or 308 309 mSOC. In this case, the microbial CUE for the three litter carbon pools can still be expressed as: $\eta_{ML} = \frac{a_{MIC,ML}}{1 - a_{DOC,ML}}$, $\eta_{CL} = \frac{a_{MIC,CL}}{1 - a_{DOC,CL}}$, and $\eta_{LL} = \frac{a_{MIC,LL}}{1 - a_{mSOC,LL}}$, respectively. 310 COMPAS applies the same approach to simulate carbon input allocation (*B*), 311

environmental modifier (i.e., $\xi(t)$) and transport matrix V as those used in CLM5. It should 312 be noted that while COMPAS and CLM5 use the same scheme to simulate $B, \xi(t)$, and V, 313 parameter values (Supplementary Tables 1-2) that were used to calculate the above elements 314 in the matrix equation were estimated independently by the PRODA approach. 315

In calculating the steady state of different carbon pools by COMPAS, Equation 2 can 316 be separated into two equations: one for litter carbon cycle and the other for mineral SOC 317 cycle, because there is no carbon transfer from mineral soil carbon pools to litter carbon 318

pools (i.e., $a_{litter pool, soil pool} = 0$ in the A matrix). Since A, K, $\xi(t)$, and V are all 319

independent from litter carbon pool states (i.e., X), the analytical solution of litter carbon 320

stock at the steady state (SS) can be calculated as $X_{litter,SS} = \left[A_{litter}\overline{\xi(t)_{litter}}K_{litter} + \right]$ 321

3)

322 $\overline{V(t)}_{lutter}]^{-1} [-B_{litter}\overline{I(t)}_{lutter}]$. For the soil organic carbon pools, the related *K* matrix is 323 carbon pool state dependent (see Equation 7). We assumed there is no vertical transport for 324 soil organic carbon pools such that litter is added to different soil layers and transported 325 vertically, and then it is transferred to soil pools that are immobile in that layer. According to 326 a method reported by Georgiou et al. (2017), the steady-state solutions for soil organic carbon 327 pools are:

$$328 \quad \boldsymbol{X}_{soil,SS} = \begin{bmatrix} \boldsymbol{x}_{DOC,SS} \\ \boldsymbol{x}_{MIC,SS} \\ \boldsymbol{x}_{ENZ,SS} \\ \boldsymbol{x}_{mSOC,SS} \end{bmatrix} = \begin{bmatrix} \frac{\boldsymbol{k}_{MIC} \boldsymbol{\xi} \boldsymbol{K}_{m,assim} \boldsymbol{\xi} \boldsymbol{x}_{MIC,SS} - \boldsymbol{u}_{MIC} \boldsymbol{K}_{m,assim} \boldsymbol{\xi}}{(\eta_{DOC} \boldsymbol{v}_{max,assim} - \boldsymbol{k}_{MIC}) \boldsymbol{\xi} \boldsymbol{x}_{MIC,SS} + \boldsymbol{u}_{MIC}} \\ \frac{\boldsymbol{u}_{MIC} + \eta_{DO} \quad (\boldsymbol{u}_{mSOC} + \boldsymbol{u}_{DOC})}{(1 - \eta_{DOC}) \boldsymbol{k}_{MIC} \boldsymbol{\xi}} \\ \frac{\boldsymbol{a}_{ENZ,MIC} \boldsymbol{k}_{MIC,SS}}{\boldsymbol{k}_{ENZ}} \\ \frac{(\boldsymbol{u}_{mSOC} + \boldsymbol{a}_{mSOC,MIC} \boldsymbol{k}_{MIC} \boldsymbol{\xi} \boldsymbol{x}_{MIC,SS}) \boldsymbol{K}_{m,decom} \boldsymbol{\xi}}{(\boldsymbol{v}_{max,decom} \boldsymbol{\xi} \boldsymbol{x}_{ENZ,SS} - \boldsymbol{a}_{mSOC,MIC} \boldsymbol{k}_{MIC} \boldsymbol{\xi} \boldsymbol{x}_{MIC,SS} - \boldsymbol{u}_{mSOC})} \end{bmatrix}$$
(9)

where \boldsymbol{u}_{S_i} is the carbon input from litter pools (L_j) to a mineral soil carbon pool $(S_i, \text{ see}$ Extended Data Fig. 3 for corresponding carbon flows for each mineral soil carbon pool) and is expressed as $\sum_{L_j} \left(\boldsymbol{a}_{S_i,L_j} \boldsymbol{k}_{L_j} \boldsymbol{\xi} \boldsymbol{x}_{L_j} \right)$. Note that all the elements with bold font indicate vectors of the corresponding variables or parameters for the 20 soil layers. All the multiplications shown in Equation 12 are element-wise operations.

334

335 2.4. Inputs and environmental conditions

For both CLM5 and COMPAS, the carbon input for the litter carbon pools (i.e., net primary
productivity, NPP) and environmental forcings (e.g., soil temperature and moisture) are from
20 years of monthly model outputs (Supplementary Table 3) by CLM5 at the steady state
using a preindustrial forcing (i.e., I1850Clm50Bgc, from year 1901 to 1920) at 0.5-degree
resolution. We used the 20-year annual mean values of different components in Equation 2 to

341 calculate total soil organic carbon stock at steady state.

342

343 **2.5.** Default (*ad hoc*) parameterizations

We compared the model simulation results of CLM5 and COMPAS when applying default 344 parameterization and the parameterization optimised by the PRODA approach. For CLM5, 345 we applied the parameter values used in its current version (Lawrence et al. 2019). In the 346 default scheme, most of the selected 21 parameters of CLM5 are constants across the globe, 347 348 except two carbon transfers that are sand content dependent, and the parameter controlling plant carbon input allocation that depends on plant functional types (Supplementary Table 349 350 1). For COMPAS, it is a newly constructed model and thus does not have well-tuned parametrization for global simulation. We applied the global mean values of the selected 23 351 parameters after site-level data assimilation as the default parameterization for COMPAS to 352 353 drive the global simulation.

354

2.6. PROcess-guided deep learning and DAta-driven modelling (PRODA)

356 The PRODA approach integrates big data with Bayesian data assimilation and deep learning 357 to optimize soil carbon cycle simulation with process-based models (Tao and Luo 2022). We used the PRODA approach to optimize both CLM5 and COMPAS at the global scale. Data 358 assimilation was first applied at each SOC profile to estimate parameter values. 21 359 360 parameters for CLM5 and 23 parameters for COMPAS were optimised for each SOC profile 361 so that the process-based model simulations can best fit local observations. Because we 362 conducted data assimilation independently at each observational site, optimised values of the same parameter vary across space. We further used a neural network to generalise those 363 364 estimated parameter values after the site-level data assimilation to the global scale by a neural network. The global parameter maps predicted by the neural network were then used in the 365

process-based models to simulate global SOC storage and retrieve the spatial patterns ofrelated model components over the globe.

We conducted Bayesian data assimilation by using the MCMC method for each of the SOC profiles to estimate the parameter values of the process-based models that best fit model simulations with SOC observations. Because the soil profile data collected from field measurement of soil organic carbon includes all components of the organic matter (e.g., microbial biomass carbon), we used the sum of modeled mineral soil carbon pools classified in CLM5 and COMPAS to be compared with soil profile data.

374 Specifically, at site-level data assimilation, for each SOC profile, we applied an adaptive Metropolis algorithm (Haario et al. 2001) to generate the posterior distributions of a 375 total of 21 parameters of CLM5 (Supplementary Table 1) and 23 parameters of COMPAS 376 377 (Supplementary Table 2) related to six model components with two phases of simulations (i.e., a test run and a formal run). We first conducted a test run assuming uniform 378 distributions for each of the preselected parameters as the proposal distributions (i.e., prior 379 380 knowledge). The prior ranges of the uniform distributions for each parameter are shown in Supplementary Tables 1-2. The proposal distributions continuously generated a set of 381 parameter values for the process-based models to simulate SOC storage. We then judged 382 whether the proposed parameter values should be accepted or not by comparing their model 383 384 simulation results with SOC observations. In the formal run, we used the accepted sets of 385 parameter values obtained in the test run as the proposal distributions and assumed that all the 386 target parameters are multivariate Gaussian distributed. We proposed new sets of parameter values and judged them to be accepted or not following the same rule in the test run. Unlike 387 388 the test run, the proposal distributions in the formal run were continuously adjusted according to the newly accepted sets of parameters. 389

390 We set 20,000 iterations for the test run and 50,000 iterations for the formal run. Eventually, we controlled the acceptance ratio (i.e., the ratio of accepted sets of parameters 391 out of the total number of iterations) of the formal run between 10% and 50%. We set the 392 393 burn-in coefficient as 50%, where the first half of the accepted parameter values in the formal run was discarded, and the second half was used to generate the posterior distributions of 394 parameters. We calculated the mean values of the posterior distributions of parameters as the 395 396 final point estimates. We ran three independent series of MCMC for each SOC profile and 397 calculated the G-R statistic to test the convergence of data assimilation results. The mean G-398 R values of the target parameters were further calculated as the holistic performance of MCMC for each SOC profile. The mathematical foundations of Bayesian data assimilation 399 and technical details of the MCMC method were documented by Tao et al. (2020). 400

401 It should be noted that the data assimilation was conducted under the assumption that SOC profiles are at steady state (i.e., $\frac{d\mathbf{X}(t)}{dt} = 0$). This assumption makes data assimilation 402 403 computationally more feasible than that under non-steady state (see the non-steady-state data assimilation in Zhou et al. (2023) and Zhou et al. (2015)). While soil carbon stocks in some 404 ecosystems (e.g., agricultural soils) may not be at the steady state because of the concurrent 405 406 climate change and human activities, previous research demonstrated that such disequilibrium component of the transient carbon cycle dynamics, especially in SOC pools, is 407 408 minor in comparison with the amount of SOC storage that was developed over thousands of 409 years (Lu et al. 2018).

We trained a fully connected multilayer neural network to predict the site-level parameter values estimated from data assimilation with a suite of 60 environmental variables (Supplementary Table 4). To achieve better training effectiveness, we first normalized all the environmental variables and parameters to the interval of [0, 1] according to their maximum and minimum values. We then conducted a set of pre-experiments to determine the best 415 configuration setting of the neural network. The neural network used in the final training
416 consisted of four hidden layers. The node numbers for each hidden layer were 256, 512, 512,
417 and 256, respectively. We used a rectified linear unit (ReLU) as the activation function and a
418 gradient descent optimization algorithm (adadelta) as the optimizer. The loss function was

419 designed as the multiplication of L1 (i.e., ratio loss:
$$RL = \frac{\sum_{l=1}^{N} \left| \frac{para_{i,true} - para_{i,pred}}{para_{i,true}} \right|}{N}$$
) and L2
420 (i.e., mean squared error: $MSE = \frac{\sum_{l=1}^{N} (para_{i,true} - para_{i,pred})^2}{N}$) errors, where $para_{i,true}$ is the
421 *i*th parameter value optimized in the site-level data assimilation, $para_{i,pred}$ is the *i*th
422 parameter predicted by the neural network, and *N* is the total number of parameters of the
423 process-based models to be predicted by the neural network ($N =$ training size × 23 for
424 COMPAS and training size × 21 for CLM5).. We decided to use this composite $L1 \times L2$ loss
425 function because training with either L1 or L2 loss alone did not yield sufficient prediction
426 accuracy. The batch size for each iteration of optimization was 32. We set a maximum of
427 6,000 epochs to train the neural network and selected the model with the lowest validation
428 loss as the final training result. To avoid overfitting in training the neural network, we set a
429 drop-out ratio of 20% for each of the hidden layers.

430

431 2.7. Global maps of SOC, residence time, and related model components

Global maps of parameters that were predicted by the best-guess neural network using the
gridded environmental variables were applied to the two process-based models to generate
global maps of SOC storage and its related components (i.e., 57,267 sets of site-level data
assimilation results for COMPAS and 62,931 for CLM5).

We retrieved the system-level carbon transfer efficiency (CTE), plant carbon inputs,
allocation of input carbon to different soil layers, substrate decomposability, environmental
modifications, and vertical transport from the optimized parameters of COMPAS and CLM5

439 (Supplementary Tables 1-2) via the PRODA approach. All the six model components referred
440 to in this study are ensembles of processes that were represented by different parameters in
441 the process-based model. Note that all the system-level components discussed in this study
442 are for the soil system that integrates both litter organic carbon and mineral soil organic
443 carbon.

444 Specifically, we calculated the system level carbon transfer efficiency as the sum of 445 carbon transfer coefficients along each carbon transformation pathway (i.e., a_{ij} in Equation 446 3) weighted by the carbon fluxes over all the pathways in the soil system:

447
$$CTE_{system} = \sum_{ij} a_{ij} \frac{\sum_{z} x_{j,z} k_{j} \xi_{z} \Delta z}{\sum_{j} \sum_{z} x_{j,z} k_{j} \xi_{z} \Delta z}$$
(10)

where a_{ij} represents the carbon transfer fraction from the donor (j) pool to the recipient (i) 448 pool; $x_{j,z}$ is the carbon pool size at depth z (g C m⁻³); k_j is the depth-independent baseline 449 decomposition rate (yr⁻¹) of the corresponding carbon pool; ξ_z represents the environmental 450 modifier at depth z; and Δz is the thickness of zth soil layer. Note that CTE along the carbon 451 452 transfer pathway to recipient pool i from donor pool j (i.e., a_{ij}) is weighted by the flux size from donor pool *j* (i.e., $\sum_{z} x_{i,z} k_i \xi_z \Delta z$), which measures the amount of decomposed carbon 453 along the *j* to *i* transfer pathway, normalized by the total flux in the soil system (i.e., 454 $\sum_{j} \sum_{z} x_{j,z} k_{j} \xi_{z} \Delta z$). A higher CTE value indicates a larger amount of carbon remaining in the 455 recipient soil pool after organic carbon is decomposed or transformed by biological and/or 456 chemical and physical reactions, which, by definition, also associates with less CO₂ released 457 458 back to the atmosphere.

The baseline decomposition rate (unit: yr^{-1}) expresses the rate of organic carbon decomposition at optimal soil temperature and water conditions. We calculated the systemlevel baseline decomposition rate (K_{system} , unit: yr^{-1}) by weighting the baseline decomposition rate of SOC pools by their carbon pool sizes:

463
$$K_{bulk} = \sum_{i} k_i \frac{x_i}{\sum x_i} \qquad (11)$$

464

Similarly, we weighted the vertical transport rate (yr⁻¹) and environmental modifiers (unitless) at different soil depths (z) by their corresponding sizes of SOC stock (i.e., x_z , with unit of g C m⁻²):

468
$$V_{system} = \sum_{z} \left(v_z \frac{x_z}{\sum x_z} \right) \qquad (12)$$

469
$$\xi_{system} = \sum_{z} \left(\xi_{T,z} \xi_{W,z} \xi_{D,z} \frac{x_z}{\sum x_z} \right)$$
(13)

470

471 Carbon input is distributed vertically according to the distribution of root biomass at different
472 soil depths (Jackson et al. 1996). Therefore, to quantify how effectively the input allocation
473 process distributes litterfall and root exudation to different soil depths, we calculated the
474 fraction of carbon input allocated to soils layers below 5 cm as the system-level index for
475 plant carbon input allocation:

476
$$B_{system} = \left[\frac{\sum_{z} exp\left[\frac{ln(1-Y_{z})}{D_{z}}\right]}{n}\right]^{5}$$
(14)

477 where Y_z is the cumulative fraction of input carbon at soil depth of D_z ; *n* is the number of soil 478 layers. A larger system-level input allocation index indicates that more carbon from litterfall 479 and root exudation will be allocated to deeper soils. This index differs between models 480 because the parameters describing the vertical distribution of carbon inputs are optimized 481 independently in the two models, even if we used the simulated total litterfall (equivalent to 482 the net primary productivity, NPP) in CLM5 as the plant carbon input for both models.



- 485 Figure 1. Distinctive model structures of CLM5 (a) and COMPAS (b).

488 **3. Results**

Process-based models with different structures and *ad hoc* parameterizations present 489 diverging results in representing global SOC storage and spatial patterns. With its default 490 491 parameterization, CLM5 simulates much more SOC in the boreal regions than tropics. In East Siberia and Alaska, SOC storage is more than 50 kg C m⁻² for the first meter, whereas, in the 492 Amazon and Congo basins and Indonesia, the average SOC storage is less than 10 kg C m⁻² 493 (Figure 2a, c). As COMPAS does not have well-tuned default parameter values at the global 494 scale, we used the global mean values of the selected parameters after site-level data 495 496 assimilation as the default parameterization. COMPAS with such ad hoc parameterization 497 simulates distinctively different SOC patterns from CLM5 across latitudes. Tropical regions with the highest carbon input are simulated to store the largest amount of SOC. The average 498 SOC storage declines from more than 20 kg C m⁻² in Amazon, Congo, and Indonesia to less 499 than 5 kg C m⁻² in boreal regions (Figure 2b-c). The correlation between the simulated 500 501 spatial patterns of SOC by CLM5 and COMPAS is -0.026 (logarithmically transformed SOC 502 values, d.f. = 45,213, P < 0.0001). Despite the contrasting spatial patterns, both of the models reasonably estimate the total global SOC storage with their ad hoc parameterizations. CLM5 503 and COMPAS simulate 1281 Pg C and 1308 Pg C preserved as SOC for the first meter soils 504 across the globe, respectively. 505



Figure 2. Diverging SOC simulation by structurally different models with *ad hoc*parameterization. (a) SOC estimated by CLM model, (b) SOC estimated by COMPAS, (c)
latitudinal variation in estimated SOC by the two models.

511

507

After being constrained by the same SOC data using the PRODA approach, the two 512 structurally different models simulate similar SOC storage and spatial patterns. At site-level, 513 514 we found that posterior distributions of selected parameters after data assimilation could be very different from their default values (Supplementary Figure 2) and also differ from site 515 to site. We further used PRODA to generalise the emerging spatial heterogeneity of optimised 516 517 parameter values in site-level data assimilation to the global scale and found similar SOC 518 simulations by CLM5 and COMPAS. Simulations by CLM5 continue to show higher SOC 519 storage in the boreal regions than in the tropics. In addition to simulating higher SOC in East 520 Siberian and Alaska, PRODA-optimised CLM5 also identifies western Siberian lowlands as 521 areas holding high SOC storage (Figure 3a, c). Meanwhile, the simulated SOC storage in 522 tropical regions, after being constrained by observations, is increased to an average value of

more than 10 kg C m⁻² (Figure 3b-c). Simulation results by COMPAS after PRODA 523 524 optimization now follow a pattern similar to that shown by CLM5. The correlation between simulations by COMPAS and CLM5 is 0.51 (logarithmically transformed SOC values, d.f. = 525 526 45,213, P < 0.0001). Notably, differences still exist in simulating sub-continental patterns by these two models. While both of the models simulate the highest SOC storage in western 527 Siberian lowlands, Alaska, and Canadian Shield, COMPAS simulates more SOC in tropics 528 but less SOC in East Siberian than CLM5. The total SOC storage simulated by COMPAS is 529 slightly higher than that by CLM5. Globally, the total SOC storage in 1m depth estimated by 530 PRODA-optimised CLM5 and COMPAS is 1469 Pg C and 1507 Pg C, respectively. 531

532





539 Simulations of key components related to SOC storage also converge after the two structurally different models are constrained by the same set of SOC data (Figure 4). We 540 assessed the spatial patterns of six components simulated by the two models: carbon transfer 541 542 efficiency, baseline decomposition, environmental modifier, carbon input allocation, vertical transport rate, and plant carbon input. The carbon transfer efficiency quantifies the ratio of 543 decomposed carbon being transferred from one carbon pool to another. CLM5 and COMPAS 544 545 represent the carbon transfer efficiency differently (Figure 1). COMPAS explicitly describes microbial CUE that partitions the metabolized organic carbon into microbial biomass 546 547 accumulation versus respiration and the non-microbial carbon transfer related to the transformation of carbon from one carbon pool to another via organo-mineral interactions 548 549 (Figure 1b). In contrast, CLM5 fuses microbial CUE and non-microbial carbon transfer in its 550 structure, such that the related parameters do not differentiate these two processes but 551 integrate their effects together in simulations (Figure 1a). Despite the difference in structure, CLM5 and COMPAS simulate similar patterns of system-level carbon transfer efficiency 552 553 (Figure 4c, Pearson correlation coefficient = 0.52, d.f. = 45,228, P < 0.001) after being constrained by the same observed SOC dataset. Both models show higher carbon transfer 554 555 efficiency in boreal regions than in the tropics (Figure 4a-b), which indicates that in boreal regions, more carbon is maintained in the soil system after SOC is decomposed or 556 557 transformed by biological and/or chemical and physical reactions instead of being released 558 back to the atmosphere as CO₂.

The rate of SOC decomposition is determined by the substrate decomposability (as indicated by the baseline decomposition) and modified by surrounding environmental factors (i.e., soil temperature and water). A high baseline decomposition indicates the organic substrate is chemically and physically more accessible to soil microorganisms (e.g., simpler chemical compounds or weaker interactions with the soil mineral matrix), whereas a lower 564 environmental modifier value suggests the SOC decomposition is more restricted by either low temperature or too much or little soil water. In this study, CLM5 and COMPAS, 565 respectively, assume first-order and Michaelis-Menten kinetics in representing SOC 566 567 decomposition. Notwithstanding their difference in kinetic assumptions, PRODA-optimised CLM5 and COMPAS agree on the highest baseline decomposition rates and the lowest 568 environmental modifier values in boreal regions across the globe (Figure 4d-i). The 569 570 correlation coefficients between the simulations by the two models are 0.55 (d.f. = 45,228, P < 0.001) for baseline decomposition and 0.80 (d.f. = 45,228, P < 0.001) for environmental 571 572 modifier.

573 However, not all components investigated in this study show convergence after data assimilation. Vertical transport quantifies the rate of organic carbon moving from the surface 574 575 to deeper soil layers. The plant carbon allocation represents how the vertical distribution of 576 carbon inputs. While CLM5 and COMPAS adopt identical mathematical functions to describe these two processes, no agreement was reached on simulated spatial patterns after 577 578 the related parameters of the two models were optimized by the PRODA approach (Figure 579 4j-0). Moreover, it should be noted that the retrieved model components using CLM5 and 580 COMPAS are usually outside the 1:1 line even when they are well correlated. While the two models agree well on the magnitude of the simulated environmental modifier (Figure 4i), the 581 582 linear CLM5 simulates higher carbon transfer efficiency values (Figure 4c) but lower 583 baseline decomposition rates (Figure 4f) than the nonlinear COMPAS. This pattern may 584 occur because parameters related to carbon transfer efficiency and baseline decomposition compensate each other in CLM5 and COMPAS for a similar SOC storage simulation. Even 585 586 though we used the same plant carbon input (i.e., the total amount of carbon from plant to litter) from CESM2 outputs in simulating SOC storage by the two models (Figure 4p-r), 587 588 COMPAS and CLM5 simulated differently how carbon transfers from litter to mineral soils

(Figure 1), as quantified by the ratio between the amount of carbon transferred from litter to
mineral soils and the total carbon input. COMPAS simulates larger amounts of litter carbon to
be transferred to mineral soils than CLM5 (Supplementary Figure 3), which requires higher
baseline decomposition rates of COMAS than CLM5 to reach similar SOC storage in SOC
storage simulation, as shown in Figure 4d-f.



596 Figure 4. Spatial patterns of different model components retrieved by CLM (left

column) and COMPAS (central column) models using the PRODA approach. The right
column shows comparisons between the model components retrieved from the two models.
The model components were: (a-c) carbon transfer efficiency, (d-f) baseline decomposition,
(g-i) environmental modifier, (j-l) carbon input allocation, (m-o) vertical transport rate, and
(p-r) plant carbon input.

602 The nonlinear decomposition kinetics in COMPAS can be approximated as first-order kinetics with respect to both donor and receiver carbon pools, after being constrained by 603 604 observed SOC data. Compared with the linear first-order kinetics used in CLM5, COMPAS specifies the decomposition of SOC and assimilation of DOC as nonlinear Michaelis-Menten 605 kinetics. Thus, the decomposition of substrates is determined by both the catalyst (i.e., 606 607 microbes for DOC assimilation and enzyme for mSOC decomposition) and the substrate concentration (i.e., DOC for DOC assimilation and mSOC for mSOC decomposition). 608 Mathematically, when the Michaelis constants (i.e., $K_{m,decom}$ and $K_{m,assim}$) are much larger 609 (e.g., 100 times larger) than their corresponding substrate concentrations, the Michaelis-610 611 Menten kinetics becomes first-order kinetics with respect to DOC or SOC (but also with 612 respect to MIC or ENZ, so that the rate equations are still nonlinear). After data assimilation at each SOC profile using COMPAS, we found that both $k_{m,decom}$ and $k_{m,assim}$ in the 613 Michaelis-Menten equation are more than 100 times that of their substrate concentrations 614 615 (i.e., SOC and DOC concentrations) for most of the soil profiles (Figure 5). Thus, the 616 nonlinear kinetics for enzyme-based mSOC decomposition and microbe-based DOC assimilation can be approximated by first-order kinetics with respect to mSOC and DOC after 617 COMPAS is constrained by globally distributed SOC vertical profiles. While losing the 618 619 nonlinear character of the donor pool effect, these kinetics laws still retain the effect of 620 microbial biomass or enzyme carbon, resulting in multiplicative kinetics.



624 Figure 5. Relationship between Michaelis-Menten constants and their corresponding

625 substrate content in COMPAS after being constrained by observational SOC profiles.

626 For decomposition, 'Substrate' is mineral-associated organic carbon (mSOC) and $K_m =$

 $K_{m,decom}$; for assimilation, 'Substrate' is dissolved organic carbon (DOC) and $K_m =$

 $K_{m,assim}$.

632 **4. Discussion**

4.1. Data assimilation enables converged SOC simulations by structurally differentmodels

635 The divergent simulations by process-based models with different structures and *ad hoc* parameterizations reflect large uncertainties in current understanding of soil carbon dynamics 636 with different theories and assumptions. In this study, CLM5 and COMPAS structurally differ 637 638 in classifying soil carbon pools, quantifying SOC decomposition kinetics, and representing carbon transfer processes. The structural differences between these two models contributed to 639 640 the contradictory spatial patterns in simulating SOC storage across the globe (Figure 2). Differences in parameter values further cause divergent simulation results between these two 641 642 models. Parameter values in process-based models quantify the strength, or represent the 643 properties, of different processes in regulating the soil carbon cycle (Luo and Schuur 2020). 644 Previous studies have demonstrated that models sharing the same first-order kinetics for SOC decomposition estimated contrasting soil carbon residence time (Zhou et al. 2018, Wei et al. 645 646 2022) and age (He et al. 2016, Shi et al. 2020) due to their different parameterizations. These differences resulted in large uncertainties in simulating global SOC storage (Todd-Brown et 647 al. 2013). While all these simulations are, to some degree, plausible under given assumptions 648 and theories, we need to identify the most probable ones to better understand how the soil 649 650 carbon cycle responds to a changing climate.

Data assimilation enables converging simulations of global SOC storage by constraining key components in the soil carbon cycle in structurally different process-based models. Regardless of their difference in structure, our results show well-converged global SOC simulations by CLM5 and COMPAS after being optimized by the PRODA approach with the same soil carbon observations. The convergence in SOC simulations derives from the fact that the PRODA approach effectively retrieves the spatial patterns of parameters of

process-based models from observational data. Parameters in CLM5 and COMPAS are both 657 conceptually and functionally different from each other due to their structural dissimilarity 658 (e.g., the turnover time values for conceptually different carbon pools and the carbon transfer 659 660 coefficients in CLM5 and COMPAS, see Figure 1 and Methods for details). However, the spatial distributions of parameters aggregated into six model components defined in the same 661 way for both models exhibit some agreement between the models. Carbon transfer efficiency, 662 663 baseline decomposition rate, and environmental modifiers have been identified as determinants in explaining the spatial patterns of global SOC storage by process-based 664 665 models (Tao et al. 2023b). In this study, these components show converged spatial patterns despite structurally different models after being informed by observations. In contrast, other 666 model components that are less important for determining global SOC storage (e.g., carbon 667 668 input allocation and vertical transport) did not converge in the simulations by CLM5 and COMPAS. This difference is probably caused by lack of sufficient information in the data to 669 constrain parameters underlying these specific components (more discussion on this issue in 670 671 Section 4.3).

The converged simulations of SOC and its related components demonstrate the fact 672 that although it is impossible to include all the processes in the soil carbon cycle into one 673 process-based model, unresolved processes can be well accounted for in model 674 675 parameterization at resolved scales after data assimilation (Luo and Schuur 2020). In this 676 study, COMPAS explicitly describes the microbial CUE that represents the carbon 677 partitioning process in microbial physiology and non-microbial carbon transfer that relates to other biological, chemical and physical reactions driving organic matter transformations in 678 679 soils. CLM5, however, does not differentiate these two processes in its structure but represents them through aggregated carbon transfer coefficients (see Methods). After being 680 681 optimized by the PRODA approach, CLM5 simulates similar spatial patterns of the carbon

transfer index as COMPAS (Figure 4). Similarly, a previous study reported that a processbased model that does not explicitly couple nitrogen-related processes with the soil carbon
cycle can still well represent nitrogen limitation after its parameters were constrained by data
(Wang et al. 2022).

686

4.2. Data assimilation identifies probable decomposition kinetics at investigated scales 687 688 Organic carbon decomposition in soils has been debated for decades. In this study, we compared two possible SOC decomposition kinetics at the global scale, namely a linear first-689 690 order kinetic model as represented by CLM5 and a nonlinear Michaelis-Menten kinetic model as represented by COMPAS. Our data assimilation results suggest that first-order 691 692 kinetics may be the simplest and effective mechanism in explaining global SOC storage and 693 its spatial patterns. After PRODA optimization, CLM5 and COMPAS show similar 694 performance in explaining the spatial variability of SOC across the globe. A linear model such as CLM5 that adequately considers the spatial heterogeneity of its parameters can 695 696 generate sufficient variability in simulating the soil carbon cycle. Indeed, notwithstanding its 697 simplicity, the linear relationship between the decomposition rate and the substrate concentration has been observed from macroscopic litter and soil organic carbon 698 decomposition experiments (Zhang et al. 2008, Schädel et al. 2014, Xu et al. 2016, Cai et al. 699 700 2018, Luo 2022).

Microorganism-centric kinetics (e.g., Michaelis-Menten kinetics) that considers enzymatic depolymerization has been advocated in recent years to account for the nonlinearity in organic carbon decomposition such that the decomposition rate is a function of both the substrate and the enzyme concentrations. Nonlinear kinetics can help capturing spatial variability of soil carbon dynamics (Wieder et al. 2013) and is necessary for understanding lignin decomposition (Liao et al. 2022) and priming effects (Wutzler and Reichstein 2008). In this study, our data assimilation results show that, at the global scale,
nonlinearity in COMPAS does not necessarily lead to more accurate quantification of SOC
storage than CLM5. Moreover, after being informed by data constraints, Michaelis constants
in COMPAS were much larger than their corresponding substrate concentrations (Figure 5).
In such a case, the Michaelis-Menten kinetics can be mathematically approximated by a
linear structure with respect to its corresponding substrate, but also including a first order
effect of the receiver pool, resulting in a multiplicative kinetics.

It should be noted that diversity in model structures is still necessary for a better 714 715 understanding of the soil carbon cycle at different spatial and temporal scales. Microbial 716 models with nonlinear structures can be useful for studying complex carbon dynamics at 717 small scales that cannot be explained by linear models (Manzoni and Porporato 2007, Liao et 718 al. 2022). Also the microbial responses to environmental fluctuations are highly nonlinear 719 and can be captured only by modelling specific microbial processes (Brangarí et al. 2020). 720 Moreover, models of SOC storage with different structures can perform differently across 721 subregions, suggesting that some structures are more suitable for certain pedoclimatic 722 conditions. We have observed different patterns of SOC storage simulated by CLM5 and 723 COMPAS in boreal (e.g., East Siberia) and tropical regions (e.g., Amazon and Congo Basins), even though both of the models were constrained by observational SOC data. 724 725 Moreover, the Michaelis-Menten kinetics investigated in this study is only one possibility 726 from an array of theories. How other nonlinear kinetics (e.g., reverse Michaelis Menten kinetics (Tang and Riley 2019) perform in simulating SOC at different scales in comparison 727 with linear models requires more studies in the future. 728

729

730 **4.3 More data required to diminish prediction uncertainty**

731 Uncertainty still exists in predicting SOC storage by structurally different models after 732 PRODA optimization (Supplementary Figure 2). The PRODA approach used in this study reveals the spatial heterogeneity of model parameters after site-level data assimilation. Thus, 733 734 at the global scale, PRODA optimizes about 1.41 million parameter values (21 selected parameters for each of the 66,935 vertical SOC profiles) for CLM5 and 1.37 million 735 parameter values (23 selected parameters for each of the 59,476 vertical SOC profiles) for 736 737 COMPAS across observational sites. The posterior distributions of different parameters showed substantial uncertainties after data assimilation at site-level. In an example of data 738 739 assimilation at one site (Supplementary Figure 2), while a few parameters can be well constrained by vertical SOC profile data, resulting in narrower posterior distributions than the 740 741 priors, more than half of the selected parameters had weak identifiability to the observations 742 such that their posterior distribution showed flat shapes within the prior ranges.

743 The identifiability of different parameters is associated with the convergence of their corresponding model components by structurally different models and further affects the final 744 745 global SOC simulations (Luo et al. 2009). For parameters that are well constrained by vertical SOC profiles in data assimilation, their corresponding model components (e.g., 746 747 carbon transfer efficiency, baseline decomposition, and environmental modifiers) also showed similar spatial patterns by CLM5 and COMPAS despite differences in model 748 749 structures. The revealed spatial patterns of these model components further presented high 750 explanatory power to predict model-simulated SOC spatial patterns across the globe (Tao et 751 al. 2023b). In contrast, for parameters that are less identifiable to observational data in data assimilation, different choices of optimized parameter value could lead to similar simulation 752 753 results on SOC storage, causing the so-called equifinality problem. Thus, the spatial pattern 754 of their corresponding components, such as vertical transport and carbon input allocation, did

not agree well after data assimilation in different models. Their spatial variability was alsoless responsible for improved global SOC simulations.

757 The equifinality problem (or weak identifiability of parameters) imposes challenges to 758 using the optimised models to predict future SOC changes under climate change. In this study, we found that the spatial patterns of vertical transport and carbon input allocation may 759 be less consequential to simulating SOC storage at the steady state at the global scale. 760 761 However, both these processes can influence the physical disconnection of SOC from decomposers, so they could regulate the transient dynamics of SOC in response to climate 762 763 change, warranting further investigations. Moreover, despite reasonable correlations between 764 results retrieved from the two structurally different models, carbon transfer efficiency and 765 baseline decomposition simulated by CLM5 and COMPAS are numerically different (i.e., not 766 on the 1:1 line in Figure 4). Whether structurally different models after PRODA optimization 767 can also predict converged SOC changes at different temporal scales is still an open question. Broader inclusion of various kinds of observational data related to soil carbon cycle at 768 769 different spatial-temporal scales is the key to resolving the equifinality problem and better 770 predictions of SOC dynamics. While this study only used SOC content as the constraint to 771 process-based models, our results clearly demonstrated that applying PRODA approach with observational constraints can effectively realize converged simulations of SOC storage by 772 773 structurally different models, even if they could generate contrasting simulation results before 774 PRODA optimization.

Beyond SOC content data, an array of measurements could be used in the PRODA approach to further improve model predictive ability and inform model development. First, decomposition data of different soil carbon pools and soil radiocarbon data could help better understand decomposition kinetics. While the decomposition of litter chemical fractions is well characterized, a better understanding of contrasting nutrient limitation mechanisms still 780 needs data beyond time series of litter total carbon, total nitrogen, and lignin (Manzoni et al. 781 2021). Meanwhile, measured carbon pools with clear physical meanings, such as particulate and mineral-associated organic carbon can help to constrain their conceptual counterparts in 782 783 models (Abramoff et al. 2022, Guo et al. 2022). Second, heterotrophic respiration time series could also be useful in transient conditions, as they vary more than SOC stocks. Third, 784 besides pool and flux data, microbial trait data can inform some of the model parameters, or 785 offer avenues for testing emerging properties such as CUE. For example, data related to 786 microbial carbon use efficiency can potentially constrain carbon transfer related parameters, 787 but only if measurements are representative of in situ conditions (e.g., using the ¹⁸O 788 incorporation method instead of adding labile ¹³C sources) (Geyer et al. 2019). Moreover, 789 790 including observations related to vegetation and hydrology dynamics in data assimilation 791 may be more effective in understanding the spatial patterns of carbon input allocation and 792 vertical transport.

793

795 **5.** Conclusion

796 This study highlights the importance of data in constraining model development and 797 simulations. While diverse model structures and varying parameterizations generate an array 798 of possibilities in simulating SOC storage under different assumptions and theories, data 799 assimilation identifies the most probable ones that best explain the observations. The PRODA approach in this study optimizes the parameters of a model based on first-order kinetics (i.e., 800 801 CLM5) and one based on Michaelis-Menten kinetics (i.e., COMPAS). Optimised parameters lead to convergence in simulated SOC storage and its related key components (i.e., the main 802 803 contributing mechanisms), such as carbon transfer and baseline decomposition. Meanwhile, our PRODA approach identifies the first-order kinetics as an equally effective explanation 804 805 with the Michaelis-Menten kinetics for global SOC storage. In the future, it is still critical to 806 explore various processes of the soil carbon cycle at different scales by developing 807 structurally different models to be tested with new data sets. A tool such as PRODA will be 808 critical in reconciling field observations and theoretical reasoning in modelling. New findings 809 and patterns revealed by the PRODA approach will further stimulate new data acquisition and 810 improvement of models.

813 References

- Abramoff, R. Z., B. Guenet, H. Zhang, K. Georgiou, X. Xu, R. A. V. Rossel, W. Yuan, and P.
 Ciais. 2022. Improved global-scale predictions of soil carbon stocks with Millennial
 Version 2. Soil Biology and Biochemistry 164:108466.
- Allison, S. D., M. D. Wallenstein, and M. A. Bradford. 2010. Soil-carbon response to
 warming dependent on microbial physiology. Nature Geoscience 3:336-340.
- Batjes, N. H., E. Ribeiro, and A. Van Oostrum. 2020. Standardised soil profile data to support
 global mapping and modelling (WoSIS snapshot 2019). Earth System Science Data
 12:299-320.
- Beck, H. E., N. E. Zimmermann, T. R. McVicar, N. Vergopolan, A. Berg, and E. F. Wood.
 2018. Present and future Köppen-Geiger climate classification maps at 1-km
 resolution. Scientific data 5:180214.
- Bradford, M. A., W. R. Wieder, G. B. Bonan, N. Fierer, P. A. Raymond, and T. W. Crowther.
 2016. Managing uncertainty in soil carbon feedbacks to climate change. Nature
 Climate Change 6:751-758.
- Brangarí, A. C., S. Manzoni, and J. Rousk. 2020. A soil microbial model to analyze
 decoupled microbial growth and respiration during soil drying and rewetting. Soil
 Biology and Biochemistry 148:107871.
- Cai, A., G. Liang, X. Zhang, W. Zhang, L. Li, Y. Rui, M. Xu, and Y. Luo. 2018. Long-term
 straw decomposition in agro-ecosystems described by a unified three-exponentiation
 equation with thermal time. Science of the Total Environment 636:699-708.
- Chandel, A., L. Jiang, and Y. Luo. 2023. Microbial Models for Simulating Soil Carbon
 Dynamics: A Review. Journal of Geophysical Research Biogeosciences.
- Ciais, P., C. Sabine, G. Bala, L. Bopp, V. Brovkin, J. Canadell, A. Chhabra, R. DeFries, J.
 Galloway, M. Heimann, C. Jones, C. L. Quéré, R. Myneni, S. Piao, P. Thornton, N.
 Metzl, and R. Wania. 2014. Carbon and other biogeochemical cycles. Pages 465-570
 Climate change 2013: the physical science basis. Contribution of Working Group I to
 the Fifth Assessment Report of the Intergovernmental Panel on Climate Change.
 Cambridge University Press.
- Cotrufo, M. F., J. L. Soong, A. J. Horton, E. E. Campbell, M. L. Haddix, D. H. Wall, and W.
 J. Parton. 2015. Formation of soil organic matter via biochemical and physical
 pathways of litter mass loss. Nature Geoscience 8:776-779.
- Cotrufo, M. F., M. D. Wallenstein, C. M. Boot, K. Denef, and E. Paul. 2013. The Microbial
 Efficiency-Matrix Stabilization (MEMS) framework integrates plant litter
 decomposition with soil organic matter stabilization: do labile plant inputs form stable
 soil organic matter? Global change biology 19:988-995.
- Fick, S. E., and R. J. Hijmans. 2017. WorldClim 2: new 1-km spatial resolution climate
 surfaces for global land areas. International Journal of Climatology 37:4302-4315.
- Forney, D. C., and D. H. Rothman. 2012. Common structure in the heterogeneity of plantmatter decay. Journal of The Royal Society Interface 9:2255-2267.
- Gelman, A., J. B. Carlin, H. S. Stern, D. B. Dunson, A. Vehtari, and D. B. Rubin. 2014.
 Bayesian data analysis. CRC press Boca Raton, FL.
- Georgiou, K., R. Z. Abramoff, J. Harte, W. J. Riley, and M. S. Torn. 2017. Microbial
 community-level regulation explains soil carbon responses to long-term litter
 manipulations. Nature communications 8:1-10.

- Geyer, K. M., P. Dijkstra, R. Sinsabaugh, and S. D. Frey. 2019. Clarifying the interpretation
 of carbon use efficiency in soil through methods comparison. Soil Biology and
 Biochemistry 128:79-88.
- Geyer, K. M., E. Kyker-Snowman, A. S. Grandy, and S. D. Frey. 2016. Microbial carbon use
 efficiency: accounting for population, community, and ecosystem-scale controls over
 the fate of metabolized organic matter. Biogeochemistry 127:173-188.
- Grigal, D., S. Brovold, W. Nord, and L. Ohmann. 1989. Bulk density of surface soils and peat
 in the north central United States. Canadian Journal of Soil Science 69:895-900.
- Guo, X., R. A. Viscarra Rossel, G. Wang, L. Xiao, M. Wang, S. Zhang, and Z. Luo. 2022.
 Particulate and mineral-associated organic carbon turnover revealed by modelling
 their long-term dynamics. Soil Biology and Biochemistry 173:108780.
- Haario, H., E. Saksman, and J. Tamminen. 2001. An adaptive Metropolis algorithm.
 Bernoulli 7:223-242.
- He, X., R. Abramoff, E. Abs, K. Georgiou, H. Zhang, and D. S. Goll. 2023. Contribution of
 carbon inputs to soil carbon accumulation cannot be neglected. bioRxiv:2023.2007.
 2017.549330.
- He, Y., S. E. Trumbore, M. S. Torn, J. W. Harden, L. J. Vaughn, S. D. Allison, and J. T.
 Randerson. 2016. Radiocarbon constraints imply reduced carbon uptake by soils
 during the 21st century. science 353:1419-1424.
- Hengl, T., J. M. de Jesus, G. B. Heuvelink, M. R. Gonzalez, M. Kilibarda, A. Blagotić, W.
 Shangguan, M. N. Wright, X. Geng, and B. Bauer-Marschallinger. 2017.
 SoilGrids250m: Global gridded soil information based on machine learning. PLoS
 One 12:e0169748.
- Huang, Y., X. Lu, Z. Shi, D. Lawrence, C. D. Koven, J. Xia, Z. Du, E. Kluzek, and Y. Luo.
 2018. Matrix approach to land carbon cycle modeling: A case study with the
 Community Land Model. Global change biology 24:1394-1404.
- Hugelius, G., C. Tarnocai, G. Broll, J. Canadell, P. Kuhry, and D. Swanson. 2013. The
 Northern Circumpolar Soil Carbon Database: spatially distributed datasets of soil
 coverage and soil carbon storage in the northern permafrost regions. Earth System
 Science Data 5:3-13.
- Jackson, R., J. Canadell, J. R. Ehleringer, H. Mooney, O. Sala, and E. Schulze. 1996. A global
 analysis of root distributions for terrestrial biomes. Oecologia 108:389-411.
- Jackson, R. B., K. Lajtha, S. E. Crow, G. Hugelius, M. G. Kramer, and G. Piñeiro. 2017. The
 ecology of soil carbon: pools, vulnerabilities, and biotic and abiotic controls. Annual
 Review of Ecology, Evolution, and Systematics 48:419-445.
- Janssen, P., and P. Heuberger. 1995. Calibration of process-oriented models. Ecological
 modelling 83:55-66.
- Lasaga, A. C. 1998. Kinetic theory in the earth sciences. Princeton university press.
- Lawrence, D., R. Fisher, C. Koven, K. Oleson, S. Swenson, M. Vertenstein, B. Andre, G.
 Bonan, B. Ghimire, L. van Kampenhout, D. Kennedy, E. Kluzek, R. Knox, P.
- 898 Lawrence, F. Li, H. Li, D. Lombardozzi, Y. Lu, J. Perket, W. Riley, W. Sacks, M. Shi,
- W. Wieder, C. Xu, A. Ali, A. Badger, G. Bisht, P. Broxton, M. Brunke, J. Buzan, M.
 Clark, T. Craig, K. Dahlin, B. Drewniak, L. Emmons, J. Fisher, M. Flanner, P.
- 901 Gentine, J. Lenaerts, S. Levis, L. R. Leung, W. Lipscomb, J. Pelletier, D. M. Ricciuto,
- 902 B. Sanderson, J. Shuman, A. Slater, Z. Subin, J. Tang, A. Tawfik, Q. Thomas, S.
- 903Tilmes, F. Vitt, and X. Zeng. 2018. Technical Description of version 5.0 of the904Community Land Model (CLM).
- Lawrence, D. M., R. A. Fisher, C. D. Koven, K. W. Oleson, S. C. Swenson, G. Bonan, N.
 Collier, B. Ghimire, L. van Kampenhout, and D. Kennedy. 2019. The Community

907 Land Model version 5: Description of new features, benchmarking, and impact of forcing uncertainty. Journal of Advances in Modeling Earth Systems 11:4245-4287. 908 Li, O., J. Xia, Z. Shi, K. Huang, Z. Du, G. Lin, and Y. Luo. 2016. Variation of parameters in a 909 910 Flux-Based Ecosystem Model across 12 sites of terrestrial ecosystems in the conterminous USA. Ecological modelling 336:57-69. 911 Liao, C., X. Lu, Y. Huang, D. Lawrence, E. Kluzek, K. Oleson, W. Wieder, and Y. Luo. 2022. 912 913 Accelerated spin-up of Community Land Model version 5 (CLM5) with coupled terrestrial carbon and nitrogen cycles. in prep. 914 Lu, X., Z. Du, Y. Huang, D. Lawrence, E. Kluzek, N. Collier, D. Lombardozzi, N. Sobhani, 915 916 E. A. Schuur, and Y. Luo. 2020. Full Implementation of Matrix Approach to Biogeochemistry Module of CLM5. Journal of Advances in Modeling Earth Systems 917 918 12:e2020MS002105. Lu, X., Y.-P. Wang, Y. Luo, and L. Jiang. 2018. Ecosystem carbon transit versus turnover 919 920 times in response to climate warming and rising atmospheric CO 2 concentration. 921 Biogeosciences 15:6559-6572. 922 Luo, Y. 2022. Theoretical foundation of the land carbon cycle and matrix approach. Land 923 Carbon Cycle Modeling: Matrix Approach, Data Assimilation, & Ecological Forecasting. CPC Press, Taylor & Francis Group, Boca Raton, Florida. 924 Luo, Y., A. Ahlström, S. D. Allison, N. H. Batjes, V. Brovkin, N. Carvalhais, A. Chappell, P. 925 Ciais, E. A. Davidson, and A. Finzi. 2016. Toward more realistic projections of soil 926 carbon dynamics by Earth system models. Global biogeochemical cycles 30:40-56. 927 Luo, Y., Y. Huang, C. A. Sierra, J. Xia, A. Ahlström, Y. Chen, O. Hararuk, E. Hou, L. Jiang, 928 929 C. Liao, X. Lu, Z. Shi, B. Smith, F. Tao, and Y.-P. Wang. 2022. Matrix approach to 930 land carbon cycle modeling. Journal of Advances in Modeling Earth Systems:e2022MS003008. 931 932 Luo, Y., T. F. Keenan, and M. Smith. 2015. Predictability of the terrestrial carbon cycle. 933 Global change biology **21**:1737-1751. Luo, Y., K. Ogle, C. Tucker, S. Fei, C. Gao, S. LaDeau, J. S. Clark, and D. S. Schimel. 2011. 934 935 Ecological forecasting and data assimilation in a data-rich era. Ecological Applications **21**:1429-1442. 936 937 Luo, Y., and E. A. Schuur. 2020. Model parameterization to represent processes at unresolved 938 scales and changing properties of evolving systems. Global change biology 26:1109-939 1117. Luo, Y., E. Weng, X. Wu, C. Gao, X. Zhou, and L. Zhang. 2009. Parameter identifiability, 940 941 constraint, and equifinality in data assimilation with ecosystem models. Ecological 942 Applications 19:571-574. Manzoni, S., P. Čapek, P. Porada, M. Thurner, M. Winterdahl, C. Beer, V. Brüchert, J. Frouz, 943 944 A. M. Herrmann, and B. D. Lindahl. 2018. Reviews and syntheses: Carbon use 945 efficiency from organisms to ecosystems-definitions, theories, and empirical 946 evidence. Biogeosciences 15:5929-5949. Manzoni, S., A. Chakrawal, M. Spohn, and B. D. Lindahl. 2021. Modeling microbial 947 948 adaptations to nutrient limitation during litter decomposition. Frontiers in Forests and Global Change 4:686945. 949 950 Manzoni, S., and A. Porporato. 2007. A theoretical analysis of nonlinearities and feedbacks in soil carbon and nitrogen cycles. Soil Biology and Biochemistry 39:1542-1556. 951 Mishra, U., S. Gautam, W. Riley, and F. M. Hoffman. 2020. Ensemble machine learning 952 approach improves predicted spatial variation of surface soil organic carbon stocks in 953 954 data-limited northern circumpolar region. Frontiers in Big Data 3:40.

- Parton, W., D. S. Schimel, C. Cole, and D. Ojima. 1987. Analysis of factors controlling soil
 organic matter levels in great plains grasslands1. Soil science society of America
 journal 51:1173-1179.
- Parton, W. J., J. W. Stewart, and C. V. Cole. 1988. Dynamics of C, N, P and S in grassland
 soils: a model. Biogeochemistry 5:109-131.
- Schädel, C., E. A. Schuur, R. Bracho, B. Elberling, C. Knoblauch, H. Lee, Y. Luo, G. R.
 Shaver, and M. R. Turetsky. 2014. Circumpolar assessment of permafrost C quality
 and its vulnerability over time using long-term incubation data. Global change
 biology 20:641-652.
- Schimel, J. P., and M. N. Weintraub. 2003. The implications of exoenzyme activity on
 microbial carbon and nitrogen limitation in soil: a theoretical model. Soil Biology and
 Biochemistry 35:549-563.
- Schmidt, M. W., M. S. Torn, S. Abiven, T. Dittmar, G. Guggenberger, I. A. Janssens, M.
 Kleber, I. Kögel-Knabner, J. Lehmann, and D. A. Manning. 2011. Persistence of soil organic matter as an ecosystem property. Nature 478:49.
- Shi, Z., S. D. Allison, Y. He, P. A. Levine, A. M. Hoyt, J. Beem-Miller, Q. Zhu, W. R. Wieder,
 S. Trumbore, and J. T. Randerson. 2020. The age distribution of global soil carbon
 inferred from radiocarbon measurements. Nature Geoscience 13:555-559.
- Tang, J., and W. J. Riley. 2019. Competitor and substrate sizes and diffusion together define
 enzymatic depolymerization and microbial substrate uptake rates. Soil Biology and
 Biochemistry 139:107624.
- Tao, F., B. Z. Houlton, S. D. Frey, J. Lehmann, S. Manzoni, Y. Huang, L. Jiang, U. Mishra, B.
 A. Hungate, and M. W. Schmidt. 2023a. Reply to: Contribution of carbon inputs to soil carbon accumulation cannot be neglected. bioRxiv:2023.2008. 2020.552557.
- Tao, F., Y. Huang, B. A. Hungate, S. Manzoni, S. D. Frey, M. W. I. Schmidt, M. Reichstein,
 N. Carvalhais, P. Ciais, L. Jiang, J. Lehmann, Y.-P. Wang, B. Z. Houlton, B. Ahrens,
 U. Mishra, G. Hugelius, T. D. Hocking, X. Lu, Z. Shi, K. Viatkin, R. Vargas, Y.
 Yigini, C. Omuto, A. A. Malik, G. Peralta, R. Cuevas-Corona, L. E. Di Paolo, I.
 Luotto, C. Liao, Y.-S. Liang, V. S. Saynes, X. Huang, and Y. Luo. 2023b. Microbial
 carbon use efficiency promotes global soil carbon storage. Nature 618:981-985.
- Tao, F., and Y. Luo. 2022. PROcess-guided deep learning and DAta-driven modelling
 (PRODA).*in* Y. Luo and B. Smith, editors. Land Carbon Cycle Modeling: Matrix
 Approach, Data Assimilation, and Ecological Forecasting. Taylor and Francis.
- Tao, F., Z. Zhou, Y. Huang, Q. Li, X. Lu, S. Ma, X. Huang, Y. Liang, G. Hugelius, L. Jiang,
 R. Doughty, Z. Ren, and Y. Luo. 2020. Deep Learning Optimizes Data-Driven
 Representation of Soil Organic Carbon in Earth System Model Over the
 Conterminous United States. Frontiers in Big Data 3.
- 992 Todd-Brown, K., J. Randerson, W. Post, F. Hoffman, C. Tarnocai, E. Schuur, and S. Allison.
 993 2013. Causes of variation in soil carbon simulations from CMIP5 Earth system
 994 models and comparison with observations. Biogeosciences 10:1717-1736.
- Wang, S., Y. Luo, and S. Niu. 2022. Reparameterization Required After Model Structure
 Changes From Carbon Only to Carbon-Nitrogen Coupling. Journal of Advances in
 Modeling Earth Systems 14:e2021MS002798.
- Wang, Y. p., H. Zhang, P. Ciais, D. Goll, Y. Huang, J. D. Wood, S. V. Ollinger, X. Tang, and
 A. k. Prescher. 2021. Microbial activity and root carbon inputs are more important
 than soil carbon diffusion in simulating soil carbon profiles. Journal of Geophysical
 Research: Biogeosciences 126:e2020JG006205.
- Wei, N., J. Xia, J. Zhou, L. Jiang, E. Cui, J. Ping, and Y. Luo. 2022. Evolution of uncertainty
 in terrestrial carbon storage in earth system models from CMIP5 to CMIP6. Journal of
 Climate 35:5483-5499.

- Wieder, W. R., G. B. Bonan, and S. D. Allison. 2013. Global soil carbon projections are
 improved by modelling microbial processes. Nature Climate Change 3:909-912.
- Wilson, C. H., and S. Gerber. 2021. Theoretical insights from upscaling Michaelis–Menten
 microbial dynamics in biogeochemical models: a dimensionless approach.
 Biogeosciences 18:5669-5679.
- Wutzler, T., and M. Reichstein. 2008. Colimitation of decomposition by substrate and
 decomposers–a comparison of model formulations. Biogeosciences 5:749-759.
- Xu, T., L. White, D. Hui, and Y. Luo. 2006. Probabilistic inversion of a terrestrial ecosystem
 model: Analysis of uncertainty in parameter estimation and model prediction. Global
 biogeochemical cycles 20.
- Xu, X., Z. Shi, D. Li, A. Rey, H. Ruan, J. M. Craine, J. Liang, J. Zhou, and Y. Luo. 2016. Soil
 properties control decomposition of soil organic carbon: Results from data assimilation analysis. Geoderma 262:235-242.
- Yigini, Y., G. Olmedo, S. Reiter, R. Baritz, K. Viatkin, and R. Vargas. 2018. Soil organic
 carbon mapping: cookbook.
- Zhang, D., D. Hui, Y. Luo, and G. Zhou. 2008. Rates of litter decomposition in terrestrial
 ecosystems: global patterns and controlling factors. Journal of Plant Ecology 1:85-93.
- Zhou, S., J. Liang, X. Lu, Q. Li, L. Jiang, Y. Zhang, C. R. Schwalm, J. B. Fisher, J. Tjiputra,
 and S. Sitch. 2018. Sources of uncertainty in modeled land carbon storage within and
 across three MIPs: Diagnosis with three new techniques. Journal of Climate 31:28332851.
- 1026 Zhou, T., P. Shi, G. Jia, Y. Dai, X. Zhao, W. Shangguan, L. Du, H. Wu, and Y. Luo. 2015.
 1027 Age-dependent forest carbon sink: Estimation via inverse modeling. Journal of 1028 Geophysical Research: Biogeosciences 120:2473-2492.
- Zhou, T., P. Shi, G. Jia, and Y. Luo. 2013. Nonsteady state carbon sequestration in forest
 ecosystems of China estimated by data assimilation. Journal of Geophysical Research:
 Biogeosciences 118:1369-1384.
- 1032

1034 Acknowledgement:

- 1035 F.T. is supported by the Eric and Wendy Schmidt AI in Science Postdoctoral Fellowship, a
- 1036 Schmidt Futures program. S.M. has received funding from the European Research Council
- 1037 (ERC) under the European Union's Horizon 2020 Research and Innovation Programme
- 1038 (grant agreement no 101001608).

1039

1042





Supplementary Figure 1. Geographic distributions of vertical SOC profiles used in this

1045 study.



1047

Supplementary Figure 2. Posterior distributions of parameters after data assimilation at 1048 one site (98.27W, 55.90N) using CLM5 (a) and COMPAS (b). Violin plots present the 1049 shapes of posterior distributions. The lower, middle, and upper hinges of boxplots show the 1050 1051 first, median, and third quartiles of the distribution. Whiskers in the boxplot represent the 1.5 times the interquartile range from the hinges. Red stars indicate the default (ad hoc) 1052 parameter values used in global simulations. Parameter values are scaled by their prior ranges 1053 (Supplementary Tables 1 - 2). Note that for parameters that are set to vary across space in 1054 the original CLM5, there is an absence of red stars. Moreover, default CLM5 set the default 1055 efolding value as 10. In this study, we set the prior range of efolding as [0, 1]. Thus, the 1056 default value of efolding in CLM5 is not shown in panel a. 1057



1059

1060 Supplementary Figure 3. Spatial patterns of the ratio of litter carbon transferred to

- 1061 mineral soil over the total carbon input, as simulated by (a) CLM5 and (b) COMPAS
- 1062 after PRODA optimization. (c) Comparison of the same ratio as simulated by the two
- 1063 models.
- 1064

1065 Supplementary Table 1 | Parameters in CLM5 that were optimized in the profile-level

1066 data assimilation.

No.	Name	Matrix term	Corresponding mechanism	Description	Default values	Unit	Prior range in profile- level data assimilation
1	fs3s1	А		Transfer fraction, fast SOC to passive SOC	Sand dependent	unitless	[0 0.05]
2	fs3s2	А		Transfer fraction, slow SOC to passive SOC	0.03	unitless	[0 0.1]
3	fs2s1	Α		Transfer fraction, fast SOC to slow SOC	Sand dependent	unitless	[0 0.4]
4	fs2l3	А		Transfer fraction, lignin litter to slow SOC	0.5	unitless	[0.2 0.8]
5	fs112	Α	Microbial carbon use	Transfer fraction, cellulose litter to fast SOC	0.5	unitless	[0.2 0.8]
6	fs111	Α	efficiency (CUE)	Transfer fraction, metabolic litter to fast SOC	0.45	unitless	[0.1 0.8]
7	fs1s2	А		Transfer fraction, slow SOC to fast SOC	0.42	unitless	[0.1 0.74]
8	fs1s3	Α		Transfer fraction, passive SOC to fast SOC	0.45	unitless	[0 0.9]
9	fl2cwd	А		Transfer fraction, coarse woody debris to cellulose litter	0.786	unitless	[0.5 1]
10	tau4s2	К		Turnover time of slow SOC	5	year	[1 50]
11	tau4s3	К		Turnover time of passive SOC	222.222	year	[200 1000]
12	tau4s1	К	Substrate	Turnover time of fast SOC	0.1370	year	[0 1]
13	tau411	К	decomposability	Turnover time of metabolic litter	0.0541	year	[0 0.11]
14	tau4cwd	К		Turnover time of coarse woody debris	3.33	year	[1 6]
15	tau4l2	К		Turnover time of cellulose and lignin litter	0.2041	year	[0.1 0.3]
16	w-scaling	ξ		Scaling factor to soil water scalar	1	unitless	[0 5]
17	q10	ξ	Environmental modifiers	Temperature sensitivity	1.5	unitless	[1.2 3]
18	efolding	ξ		E-folding parameter to calculate depth scalar	10	metre	[0, 1]
19	cryo	v		Cryoturbation rate	0.0005	m ² yr ⁻¹	[3×10 ⁻⁵ 16×10 ⁻⁴]
20	diffus	V	Vertical transport	Bioturbation rate	0.0001	m ² yr ⁻¹	[3×10 ⁻⁵ 5×10 ⁻⁴]
21	Ь	I	Carbon input allocation	Parameter controlling vertical distribution of carbon input to litter pools	PFT dependent	unitless	[0.5 1]

1067

1069 Supplementary Table 2 | Parameters in COMPAS that were optimized in the profile-

1070 level data assimilation.

No.	Name	Related components	Description	Default values	Unit	Prior range
1	η_{DOC}		Microbial CUE for DOC assimilation	0.27	unitless	[0.01 0.7]
2	η_{ML}		Microbial CUE for metabolic litter assimilation	0.66	unitless	[0.4, 0.9]
3	η_{CL-LL}	Microbial carbon use efficiency	Microbial CUE for cellulose/lignin litter assimilation	0.16	unitless	[0, 0.4]
4	K _{m,assim}		Concentration of DOC for half max DOC assimilation reaction	1.8×10 ³	gCm ⁻³	[300 3000]
5	τ_{assim}		Inverse of $v_{max,assim}$ in DOC assimilation	0.015	year	[0.03 0.001]
6	τ_{decom}		Inverse of $v_{max,decom}$ in SOC decomposition	1.62×10 ⁻⁴	year	[0 3×10 ⁻⁴]
7	K _{m,decom}		Concentration of SOC for half max SOC decomposition reaction	5.65×10 ⁵	gCm ⁻³	$[10^5 \ 10^6]$
8	$ au_{ENZ,prod}$		Turnover time for enzyme production	22	year	[15 30]
9	$ au_{ML}$	Decomposition	Turnover time of metabolic litter	0.049	year	[0 0.1]
10	τ_{CWD}		Turnover time of coarse woody debris	3.5	year	[1 6]
11	τ_{CL-LL}		Turnover time of cellulose and lignin litter	0.2	year	[0.1 0.3]
12	$\tau_{ENZ,decay}$		Turnover time for enzyme decay	0.04	year	[0.001 1]
13	$ au_{MIC}$		Turnover time for microbial mortality	1.1	year	[0 2]
14	а _{SOC,MIC}		Fraction of microbial necromass that is stabilized as SOC	0.47	year	[0 1]
15	a _{CL,CWD}		Fraction of decomposed CWD that goes to cellulose litter	0.75	unitless	[0.5, 1]
16	$a_{DOC,ML}$	Carbon transfer fraction	Fraction of total decomposed metabolic litter that goes to DOC	0.05	unitless	[0 0.1]
17	a _{DOC,CL}		Fraction of total decomposed cellulose litter that goes to DOC	0.17	unitless	[0.05 0.3]
18	a _{SOC,LL}		Fraction of total decomposed lignin litter that goes to SOC	0.78	unitless	[0.6 0.95]
19	w-scaling	Environmental	Scaling factor to soil water scalar	2.9	unitless	[0 5]
20	q10	modification	Temperature sensitivity	2.1	unitless	[1.2 3]
21	cryo		Cryoturbation rate	0.0008	m ² yr ⁻¹	[3×10 ⁻⁵ 16×10 ⁻⁴]
22	diffus	Vertical transport	Bioturbation rate	0.00026	m ² yr ⁻¹	[3×10 ⁻⁵ 5×10 ⁻⁴]
23	b	Carbon input allocation	Parameter controlling vertical distribution of carbon input to litter pools	0.72	unitless	[0.5 1]

1075 Supplementary Table 3 | Forcing variables for driving simulations of SOC by process-

1076 based models.

Variable Names	Full Description	Resolution	
nbedrock	Soil layer number that reaches the bedrock		
ALTMAX	Maximum active layer depth of current year		
ALTMAX_LASTYEAR	Maximum active layer depth of last year		
CELLSAND	Sand content		
NPP	Net primary productivity	0.5 degree, monthly record of 20-yea simulation after the system reaches the steady state.	
SOILPSI	Soil water potential		
TSOI	Soil temperature		
O_SCALAR	Oxygen scalar for decomposition		
FPI_vr	Nitrogen scalar for decomposition		

1078

1077

1080 Supplementary Table 4 | Environmental variables used in predicting optimized

1081 parameter values of process-based models by the deep learning model. Note that

1082 information of some of the variables (e.g., clay content) was reported at different depths (i.e.,

1083 0cm, 30cm, and 100cm).

No.	Variable Name	Data Source	Category	Description
1	Longitude	WoSIS		
2	Latitude	WoSIS		
3	Elevation	NOAA	Geography	Available at https://www.ngdc.noaa.gov/mgg/global/
4	Absolute Depth to Bedrock	(Hengl et al. 2017)		
5	Bedrock Depth	CLM5 simulation		
6	Koppen Climate Classification	(Beck et al. 2018)		
7	Annual Mean Temperature			
8	Mean Diurnal Range Temperature			
9	Isothermality			
10	Temperature Seasonality			
11	Max Temperature of Warmest Month			
12	Min Temperature of Coldest Month			
13	Temperature Annual Range			
14	Mean Temperature of Wettest Quarter			
15	Mean Temperature of Driest Quarter		Climate	
16	Mean Temperature of Warmest Quarter	(Fick and Hijmans 2017)	Cliniate	
17	Mean Temperature of Coldest Quarter			
18	Annual Precipitation			
19	Precipitation of Wettest Month			
20	Precipitation of Driest Month			
21	Precipitation Seasonality			
22	Precipitation of Wettest Quarter			
23	Precipitation of Driest Quarter			
24	Precipitation of Warmest Quarter			
25	Precipitation of Coldest Quarter			
26	USDA 2014 Suborder Classes			
27	WRB 2006 Subgroup Classes			
28	Coarse Fragments Volumetric			Three depths were included, which are 0cm, 30cm and 100cm, respectively
29	Clay Content			Three depths were included, which are 0cm, 30cm and 100cm, respectively
30	Silt Content	(Hengl et al. 2017)	Soil Texture	Three depths were included, which are 0cm, 30cm and 100cm, respectively
31	Texture Classes			Three depths were included, which are 0cm, 30cm and 100cm, respectively
32	Sand Content			Three depths were included, which are 0cm, 30cm and 100cm,
33	Bulk Density			Three depths were included, which are 0cm, 30cm and 100cm, respectively
34	Soil Water Capacity			Three depths were included, which are 0cm, 30cm and 100cm,
35	Soil pH in H ₂ O			Three depths were included, which are 0cm, 30cm and 100cm,
36	Soil pH	(Hengl et al. 2017)	Soil Chemical Properties	Three depths were included, which are 0cm, 30cm and 100cm,
37	Cation Exchange Capacity		risperues	Three depths were included, which are 0cm, 30cm and 100cm,
38	Grade of a Sub-Soil Being Acid			respectively
39	ESA Land Cover	ESA. Land Cover	Vegetation	Available at: <u>maps.elie.ucl.ac.be/CCI/viewer/download/ESACCI-LC-</u> Ph2-PLIGv2 2.0 pdf

40	NPP	Guide Version 2. Tech. Rep. (2017).	Mean value of 20-year simulation after the system reaches the steady
41	Standard deviation of NPP	CLM5 simulation	Standard deviation of 20-year simulation after the system reaches the steady state
42	Vegetation Carbon Stock		Mean value of 20-year simulation after the system reaches the steady state
084			