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Improving Predictions of Potential Soil Respiration Across the Continental United States with Soil Organic Matter Composition

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21 Abstract

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Despite the importance of microbial respiration of soil organic matter (SOM) in regulating 23 carbon flux between soils and the atmosphere, soil carbon (C) cycling models remain primarily 24 based on climate and soil properties, leading to large uncertainty in their predictions. Molecular 25 data have long been proposed as a promising avenue for resolving modeling errors, but evidence 26 for improved predictions of soil C cycles with high-resolution measurements remains mixed. 27 With data from the 1000 Soils Pilot of the Molecular Observation Network (MONet), we 28 29 analyzed the molecular composition of water-extractable SOM from 66 soil cores across the United States to address this knowledge gap. Our innovation lies in using machine learning (ML) 30 to distill the thousands of SOM formula that we detected per sample into tractable units. Then, 31 we compared ML predictions of measured potential soil respiration using (1) a suite of standard 32 33 soil physicochemical data, (2) ultrahigh-resolution SOM composition independently, and (3) in combination with physicochemistry to assess the added value of molecular information to predict 34 soil respiration. In surface soils (0-10 cm), water-extractable SOM chemistry alone provided 35 better estimates of potential soil respiration than soil physicochemical factors alone, and using 36 37 the combined sets of predictors yielded the highest explanatory power of soil respiration rates. In contrast, in subsoils (>10 cm), SOM composition did not improve ML-based respiration model 38 performance, possibly due to the greater importance of mineral-associated SOM below the 39 surface layer. Our results underscore the potential of integrating SOM composition into carbon 40 cycle modeling for enhanced predictions of terrestrial-atmosphere climate feedback. 41

42 Introduction

Soil respiration is estimated to release 60-100 Gt of C to the atmosphere per year, [1, 2] six to ten 43 times as much C as released by fossil fuel combustion (~10 Gt C[3]). Microbial respiration of 44 soil organic matter (SOM) is one of the most important contributors to soil carbon dioxide (CO_2) 45 emissions and a critical link in the global C cycle.[4] With increasing temperatures under climate 46 change, soil C repositories are vulnerable to increased rates of microbial respiration, [5-7] which 47 can lead to positive feedbacks in global CO₂ emissions and temperature rises.[8] Despite decades 48 49 of research, soil C fluxes remain one of the largest uncertainties in global climate predictions.[9-14] Novel molecular measurements have recently been applied to identify SOM composition in 50 an effort to understand molecular-scale processes that could improve model predictions of CO₂ 51 fluxes.[15-18] Despite these efforts, our attempts to improve soil C model predictions by refining 52 53 chemical pools have yielded mixed results.[19-21] 54 The interplay of factors such as soil moisture, pH, nutrients, mineralogy, and SOM concentration 55 and chemistry governs microbially-derived transformations of SOM;[22-27] but these 56 57 relationships are difficult to constrain.[4, 28] The most commonly used modeling approaches are based on Raich's model, which estimates respiration primarily as a function of temperature and 58 59 water availability.[29][,][30] Newer process-based model formulations use an additional suite of physical and biogeochemical measurements to represent microbial and mineral processes. They 60 incorporate SOM chemistry either through several discrete pools or through their thermodynamic 61 properties.[21, 31-34] With large spatiotemporal heterogeneity and limited availability of 62 comprehensive and standardized measurements at regional-to-continental scales, accurate 63 predictions of microbial SOM decomposition across different ecosystems remain 64 challenging.[35] 65

66

A better understanding of SOM concentration, composition, and bioavailability may enhance our ability to predict soil C cycling processes through their controls on soil respiration and related enzymatic activities.[21, 31-34] Variations in the bioavailability of chemical classes of SOM are mediated by geochemical conditions and biophysical constraints, such as microbial biomass and necromass, reactive metals and minerals, organic and mineral horizon thickness, and other climate-related variables.[36] For example, coarse-textured soil is more conducive to decomposition of chemically labile litter-derived C potentially due to higher fungal activity in
organic-rich horizons.[37, 38] In addition, the interface between fresh litter inputs and soil
minerals can serve as a hotspot for microbial breakdown of C found in the litter, resulting in the
formation of soil aggregates and organo-mineral associations.[39] This variability underlines the
essential need to identify unique subsets of SOM formula that contribute more to soil respiration
among different ecosystems and soil depths.

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80 The distillation of multidimensional SOM composition profiles into a tractable set of formula that influence soil respiration is a key challenge in soil ecology. [15, 28, 40-45] Unsupervised 81 machine learning models that summarize large data into a small number of significant features 82 have been widely used to study microbial communities, SOM composition, and other 83 84 environmental problems with multidimensional data.[46] Dimensionality reduction such as Principal Component Analysis (PCA)[47-49] and clustering methods such as hierarchical 85 86 clustering analysis [50-52] are the most common tools to explore large molecular datasets. Although these tools are beginning to be applied to determine the relationship between SOM 87 88 composition and soil physicochemistry, [49] it is still challenging to extract a subset of SOM substrates associated with specific processes, like soil respiration. 89 90

Although ultrahigh mass resolution measurements can provide unprecedented characterization of 91 the thousands of individual formulae that comprise SOM, the interpretation of these data types 92 largely remains guided by coarse chemical and ecological groupings. Here, we develop models 93 using semi-supervised machine learning (non-negative matrix factorization with custom k-means 94 clustering, NMFk) to reduce the complexity of molecular information into k distinct signatures of 95 water-extractable SOM chemistry at two depths in cores collected across the continental United 96 97 States. We then explore the extent to which these signatures and NMFk-enabled feature set can provide additional insight into rates of soil respiration beyond variables that are more routinely 98 collected. By examining a multitude of physicochemical and SOM signatures, our goal is to 99 elucidate the specific subset of SOM substrates that may be vital to understanding and predicting 100 below-ground C storage. 101

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104	Methods
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106	Soil sampling and characterization.
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108	As part of the 1000 Soils Pilot study for the Molecular Observation Network (MONet) program,
109	we collected 66 soils from across the continental US using standardized sampling procedures
110	described by Bowman et al.[53] (Figure S1). Two long cores (30 cm) and three short cores (10
111	cm) were collected at each site. We also conducted field measurements, including soil
112	temperature, volumetric water content, vegetation type, and weather conditions. Cores were
113	shipped on ice overnight to the Pacific Northwest Laboratory for further analysis. A full
114	description of sampling and analytical methodologies is available in Supporting Information and
115	Bowman et al.[53]
116	
117	Water extractable SOM characterization.
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119	We extracted water-soluble SOM from soils using solid phase extraction and analyzed using a
120	Bruker 7-T Fourier transform ion cyclotron resonance mass spectrometry (FTICR MS) at the
121	Environmental Molecular Sciences Laboratory (EMSL) in Richland, WA. More details on SOM
122	extraction methods and FTICR MS analysis are in Supporting Information and Bowman et
123	al.[53]
124	
125	Raw FTICR MS data was processed with CoreMS (Python package, installed on
126	2022/11/22),[54] including signal processing, peak detection, and molecular formula assignment.
127	Noise thresholding was performed with signal-to-noise threshold (5 std.), mass error (0.3 ppm),
128	and stoichiometric limits from domain knowledge (supporting information). Suwannee River
129	fulvic acid (SRFA) standards were used to set a calibration threshold for all soils in the same
130	batch. Molecular formula was assigned based on both accurate mass and filtered by their
131	confidence score from CoreMS. After calibration and formulae assignment, we filtered the
132	assigned peaks by m/z between 200 to 1,000, present in at least 2 out of 3 replicates, not present
133	in two or more lab blanks, and with formulae confidence scores (combines m/z error and isotopic
134	pattern)[54] above 0.7. We predicted compound classes of the filtered formulae based on O/C

and H/C ratios of van Krevelen classes.[55, 56] The suffix "-like" in chemical classes indicates
the uncertainty of the van Krevelen classification method.[56] We converted the peak intensity
values to present/absent (1/0) and separated the final dataset by soil depth (surface vs. subsoil)
for statistical analysis. Alpha diversity was calculated as the total number of SOM formulae
identified in each sample.

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141 Data analysis and machine learning methods.

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We used linear regression models to evaluate the relationship between soil potential respiration and soil physicochemical variables. To avoid the impacts of different magnitudes of the data that might lead to biased relationships, we performed log₁₀ transformation on potential respiration rates, total C, total N, total sulfur, and Mn concentration. *stats.linregress* function from *scipy* package (v 1.11.4) in Python (v 3.7.1) was applied to calculate the fitted line, r² value (*rvalue*², Pearson correlation), and p-value (*pvalue*). Pairwise plots with regression fitting were generated by the pairplot function from the *seaborn* package (v 0.12.1) in Python.

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We used non-negative matrix factorization (NMF)[57] with custom k-means clustering 151 152 (NMFk)[58] to identify signature components from the 7312 and 5515 SOM molecular formula (for surface and subsoil, respectively) we detected (i.e., N formulae in m soils) with pyNMFk153 154 package (Python, https://github.com/lanl/pyDNMFk, Figure 1). More details on NMFk assumptions, model settings, and model robustness are in the Supporting Information. Briefly, 155 NMFk tends to be more successful at extracting explainable basis or signatures from large 156 multivariate datasets, compared to other dimensionality reduction tools such as principal 157 158 component analysis.[57, 59] As applied here, NMFk summarizes data into discrete signatures 159 that contain weights for each SOM formulae detected by FTICR-MS for each soil layer independently (i.e., a separate set of signatures was generated to summarize surface versus 160 subsoils, allowing us to explore depth-specific relationships with potential soil respiration). The 161 optimal number of signatures was determined from silhouette coefficients of different NMFk 162 models. A W-matrix with the weights of each SOM formulae (N) to each extracted signature (k), 163 and an H-matrix with the contribution of each signature (k) to each soil sample (m) were 164 generated from NMFk. To visualize the composition of each NMFk signatures (W-matrix), we 165

166 generated a heatmap of SOM formula with normalized weights (0-1) > 0.5 in at least one NMFk,

167 clustered by van Krevelen class assignment (clustermap function from seaborn package). Within

168 each inferred chemical class of SOM formula, we further clustered formula using the "linkage"

169 method from the *scipy* package ("ward" method with "Euclidean" distance) to illustrate the

- 170 difference between NMFk signatures.
- 171

To define groups of soils with high, medium, or low rates of potential respiration, we used k-172 173 means clustering on potential soil respiration with the elbow method to select the number of groups (*KMeans* from *scikit-learn* package).[60] Then, we mapped the extracted k signatures to 174 soil respiration using supervised machine learning. To evaluate the potential value of NMFk-175 extracted SOM signatures for explaining soil respiration, we conducted three sets of machine 176 learning models: (1) selected environmental parameters alone (i.e., variables with $R^2 > 0.2$ in 177 178 individual regressions, Figure 2, Table S1), (2) SOM composition alone (NMFk weights from H-179 matrix), and (3) environmental and SOM composition in combination. All machine learning models were built using gradient boosting regression (GBR) from scikit-learn package (v. 0.24, 180 181 Python). More details in model training, testing and validation are in Supporting Information.

182

183 **Results**

184 Soil physicochemistry and potential respiration

185

Overall, many soil parameters, including potential soil respiration, tended to be higher in surface soils than in subsoils. Significant differences (p<0.05) between surface soils and subsoils in total C, total N, total sulfur, C/N ratio, and other factors are shown Figure S3. In particular, surface soils had higher potential respiration rates (median: 72.6 ug CO₂/g soil/day) than subsoils (median: 21.9 ug CO₂/g soil/day) (Mann–Whitney U = 3022.5, N_{surface} = 63, N_{subsoil} = 61, P <

191 0.05).

192

193 We grouped potential soil respiration into 3 levels corresponding to low, medium, and high

respiration in each soil layer using k-means clustering (Figure S2). For both surface and subsoils,

soil with high potential respiration tended to be sourced from the Midwestern and Northeastern

196 United States. (Figure 3, Figure S5). In surface soil, high potential respiration was associated

with five soils collected in Utah, Wyoming, and Virginia (within temperate conifer forest and
temperate broadleaf & mixed forest biomes, Figure 3, Figure S1). In subsoils, high respiration
was associated with three soils from Utah and Maryland (temperate conifer forests and broadleaf
& mixed forests biomes). Desert soils had the lowest respiration in both layers (Figure S1).

201

We found relationships between soil respiration and many variables that supported prevailing 202 paradigms. A full correlation table of associations between different soil properties is available in 203 the SI (Table S1). Briefly, potential respiration rates in both surface and subsoils were positively 204 correlated with gravimetric water content (GWC) (r^2 : 0.246 and 0.225, p<0.05) and cation 205 exchange capacity (CEC, r^2 : 0.405 and 0.354, p<0.05, Figure 2). They were also positively 206 correlated with total C and total N content, with stronger relationships in surface soils (r²: 0.487 207 v.s. 0.268 for total C, r²: 0.439 v.s. 0.248 for total N, p<0.05). Total bases and magnesium (Mg) 208 concentrations had a higher correlation to respiration in subsoils than surface soils (r^2 : 0.227 v.s. 209 0.146 and 0.287 v.s. 0.160, p<0.05, Figure 2), while manganese (Mn) concentrations were 210 correlated to respiration in surface soils (r^2 : 0.324, p<0.05, Figure 2). 211

212

213 SOM composition and NMFk partitioning of SOM.

214

Across all soils, the most common chemical classes of SOM were lignin-, condensed 215 216 hydrocarbon-, and tannin-like formula. Most formula in these classes were present in both surface and subsoils (i.e., 'shared' formula). However, surface soils contained more unique 217 formula than subsoils for all compound classes (Figure 3b). In particular, many protein-, amino 218 sugar-, and lipid-like compounds were identified in surface soils only, with very few compounds 219 220 in these classes being unique to subsoils. Because SOM consists of thousands of different 221 compounds, we also used alpha diversity to represent the SOM richness per sample (Figure 3). Soils from the Midwestern U.S. and the West Coast had relatively higher alpha diversity than 222 soils from other regions. 223

224

We used NMF*k* to summarize SOM composition into 7 and 5 NMF*k* signatures, respectively, for

surface and subsoils (Figure 4). Geographic patterns in SOM signatures are displayed in Figure

227 S6-7, with more geographic clustering of NMFs in surface soils than in subsoils. For surface

soils, NMF3 presented as the largest relative contributor to SOM composition in 20 soils across 228 all biomes (i.e., highest weighting in H-matrix, hereafter, 'dominant signature', Figure S6). 229 NMF2, NMF5, and NMF7 served as the dominant signature in at least 9 soils each. For subsoils, 230 NMF5 and NMF2 were the dominant signature in 27 soils and 16 soils respectively distributed 231 across all biomes in the continental United States. There was no single NMF signature that could 232 exclusively represent SOM composition of all sites in the same region for either surface or 233 subsoils, suggesting that SOM composition at local sites is best summarized by a combination of 234 235 multiple NMFs.

236

The most important formula contributing to the composition of each NMF (i.e., formula with 237 normalized weights >0.5 in W-matrix) are shown in Figure 4a-b. For surface soils, NMF1, 4, 6, 238 239 and 7 had a relatively higher number of important compounds identified as lignin-like. NMF6 and 7 had larger contributions of condensed hydrocarbon-like formula. NMF1 had higher 240 241 contribution from protein-like and amino sugar-like compounds, while NMF3 and 5 had the lowest contribution from protein-like, amino sugar-like, and lipid-like compounds, suggesting 242 243 their low microbial activities. NMF4 had the largest number of lipid-like compounds as important features. In subsoil samples, important formula for all NMFs tended to be classified as 244 245 lignin-, tannin-, and/or condensed hydrocarbon-like. NMF1 and NMF5 had more features identified as lignin-like and some tannin-like compounds. NMF2 had the largest fraction of 246 247 condensed hydrocarbon-like compounds. NMF4 had larger contributions of protein-like and amino sugar-like formula (Figure S8). We also compared if formula contributing to NMF 248 signatures tended to be similar among surface and subsoils by assessing shared vs unique 249 formula. NMF-selected formula (weights >0.5 in W-matrix) followed the same general patterns 250 251 as the overall SOM pool but showed amplified relationships (Figure 4c).

252

We also observed differences in dominant NMF signatures across high-, medium-, and lowrespiration soils, particularly in surface soils (Figure 4d-e). High respiration surface soils were characterized by five NMF signatures (1, 2, 3, 6, and 7), with the largest contribution from NMF6. Low respiration surface soils, in contrast, uniquely contained NMF5, and they did not have any contribution from NMF6. In subsoils, high respiration soils consisted of NMF 1, 2 and 4, while low respiration soils consisted of NMF1, 2, and 5. NMF5 had a larger contribution in low-respiration soils from both temperate forests and grasslands/shrublands. NMF5 had higher
weights in low-respiration soils, and NMF3 and NMF4 had lower weights in low-respiration
soils.

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Relative importance of physicochemistry and SOM composition in potential soil respiration
 models

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We developed gradient-boosting regression models to predict potential soil respiration with (1) physicochemical variables, (2) SOM composition represented by NMF signatures, and (3) both of them combined. Model performances are summarized in Table 1 and Figure 5.

269

270 Selected physicochemical variables (consisting of total C, total N, CEC, moisture, Mn (surface),

total base (subsoil), and Mg (subsoil) concentration) had significant independent Pearson's

272 correlation to respiration w/ p < 0.05 and $r^2 > 0.2$ (Table S1). Physicochemical variables

predicted potential respiration rates in surface and subsoils equally well ($R^2 = 0.44$ and 0.43

respectively for testing data). In surface soils, total C, total N, and cation exchange capacity

275 (CEC) were identified as the top 3 most important predictors, followed by Mn concentration and

soil moisture (Figure 4). In subsoils, CEC, total N, and soil moisture were the most important

277 predictor, and total C was the least important predictor (Figure S9).

278

Using SOM composition (NMF signatures) as predictors, we had better model performance in surface soils than in subsoils (testing $R^2 = 0.54$ vs. 0.08), and SOM composition alone predicted

281 more slightly variation in potential respiration rates than physicochemical variables alone in

surface soils (testing $R^2 = 0.54$ vs. 0.44), even when controlling for an equal number of

predictors (testing $R^2 = 0.48$ vs. 0.44). NMF3, NMF5, and NMF2 were the most important SOM

composition variables for explaining soil respiration in surface soils (Figure 5).

285

286 When we combined both physicochemical variables and SOM composition into a single

predictor set, we obtained better respiration model performance ($R^2 = 0.62$) compared to models

with environmental variables or SOM composition in surface soils only. However, the model

describing potential respiration rates in subsoil was worse ($R^2 = 0.36$) when compared to models

based on physicochemical variables only. In surface soils, the 3 most important variables were

the same as the physicochemical model (Figure 5). NMF6 was identified as the most important

SOM variable, followed by NMF3, NMF2, and NMF5 (Figure 5).

293

294 Discussion

Depth partitioning in relationships between SOM composition and potential soil respiration 295 Given that not all chemical constituents of SOM contribute to soil respiration and that surface 296 and subsoils differ substantially in mineralogy and structure, we hypothesized that distinct 297 subsets of SOM would contribute to respiration in surface vs. subsoils. There was no single NMF 298 299 that dominated low- vs. high-potential respiration soils in either layer, however, NMF weightings varied substantially across soils with different rates of potential respiration in both layers (Figure 300 4d-e). This suggests that different subsets of SOM were disproportionately associated with soils 301 exhibiting high vs. low potential respiration rates. While patterns in SOM chemical across 302 303 geographic regions were difficult to disentangle, the spatial distribution of NMF types suggested local similarity in SOM composition in both layers (Figure S6-7), likely reflecting similar 304 305 underlying chemistry, mineralogy, and/or biogeochemical processes.[61]

306

307 The SOM formula within NMFs that correspond to changes in soil respiration may represent a key step forward in understanding the chemical bioavailability of water-extractable organic 308 309 matter in soils. In surface soils, NMF6 displayed a dramatic increase in weightings from low-tohigh respiration soils. It contained a diverse suite of compounds including protein-, (soluble) 310 311 lipid-, and amino sugar-like formula that can be rapidly used as microbial substrate. Proteins and amino sugars can fuel microbial metabolism of SOM, [62, 63] thus the prevalence of these 312 compounds within NMF6 may support high potential rates of soil respiration. NMF1 and NMF7 313 in surface soils contained a diverse mixture of compounds and also increased from low-to-high 314 respiration soils, supporting a possible relationship between SOM pool diversity and microbial 315 respiration (see previous section). In contrast, surface NMF2, NMF3 and NMF5 decreased in 316 importance from low-to-high respiration soils and primarily consisted of a small but unique 317 subset of lignin- and tannin-like compounds (Figure 4a). This is consistent with low 318 bioavailability of its chemical constituents suppressing microbial respiration.[64, 65] It suggests 319

that despite the often-inferred high bioavailability of water-extractable SOM, [41, 66] there may 320 be a significant fraction of water-extractable SOM that is chemically protected from microbial 321 decomposition.[40, 41, 63] Interestingly, NMF4 in surface soils — which contained the greatest 322 number of lipid-like formula (Figure 4a) and had a comparatively large fraction of protein-like 323 formula —was not present in any high-respiration soils. We therefore suggest that NMF4 may be 324 an indicator of non-living microbial biomass (i.e., necromass) which is disproportionately 325 comprised of lipids (microbial cell wall remnants) and amino sugars and proteins (the basis of 326 intracellular materials).[67, 68] The comparatively weak relationship between subsoil water-327 extractable SOM and potential soil respiration as compared to surface soils highlights recent 328 work emphasizing the importance of mineral-associated organic matter in soil C storage.[69-71] 329 330

Relative importance of physicochemistry and SOM composition in predicting potential soil
 respiration

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By developing machine learning models to predict respiration with soil physicochemistry and 334 335 SOM composition (NMFs) separately and in combination, we were able to distinguish the contributions of each set of factors for predicting soil potential respiration. The models based on 336 337 physicochemistry alone explained a modest amount of variation in soil respiration (44% and 43% in surface and subsoils, respectively), in line with the range of explanatory power observed 338 339 in other works.[72, 73] For surface soils, models based on SOM composition alone (54% variation explained) and both physiocochemical factors and SOM composition combined (62% 340 variation explained) suggest that SOM composition (1) can predict soil respiration at least as 341 well as commonly measured physiocochemical variables and (2) explains some portion of soil 342 343 respiration that is not captured by physiocochemistry. In models based on SOM composition 344 alone, NMF3 (which was mainly in low-respiration soil and was comprised of lignin- and tanninlike formula, see previous sections) was the strongest predictor of soil respiration followed by 345 NMF2 and NMF5. The relative chemical recalcitrance of the most important predictors of 346 respiration may suggest that the proportion of thermodynamically unfavorable formula in water-347 extractable SOM has a direct inhibitory effect on soil metabolism. Indeed, thermodynamic 348 regulation of organic C composition can be a key control for the rate of respiration in 349

ecosystems.[40, 41] Therefore, the inclusion of SOM composition in more mechanistic modeling
 approaches may be able to improve predictions of soil respiration rates.

352

However, models for subsoils displayed different dynamics. In the subsoil model based on 353 physicochemical variables alone, total C was the least important predictor (vs. the most 354 important predictor for surface soils), and the model containing SOM composition did not yield 355 high predictive power. The marginal effect in partial dependence of surface soil respiration to 356 total C was stronger than the effect of subsoil respiration (Figure S10), supporting a stronger 357 association between total C and potential respiration in surface soil vs. subsoil. The low 358 predictive power of total C relative to other physicochemical factors could explain why SOM 359 composition did not add predictive power to potential respiration in subsoils. Since more total 360 361 and organic C is stored in surface soils, resolution into the water-extractable SOM pool (reflected here by NMFs) might be a more significant factor for predicting surface soil respiration than in 362 subsoils that are characterized by lower total C and more mineral-associated SOM.[74] 363

364

Our results suggest that NMF-extracted signatures of SOM composition are able to improve 365 surface soil model performance by integrating fundamental molecular information into soil 366 respiration models across very different soil ecosystems at the continental scale. NMF6, which 367 was the most important NMF signature in combined models of surface respiration, consisted of 368 369 diverse chemically-bioavailable compounds, and it mainly existed in high-respiration soils (see previous sections).[65] We therefore suggest that chemically-bioavailable compounds in water-370 extractable SOM pools may provide the greatest complementary explanatory power to 371 physicochemical factors in respiration predictions. Because SOM pools vary tremendously at the 372 373 continental-scale, refined regional or local studies that encompass lower-variability parameter 374 spaces may yield even more value of SOM molecular data to soil C modeling.

375

We note that physicochemical predictors were stronger predictors of soil respiration than SOM composition in the combined surface soil models. However, the inclusion of SOM composition improved physicochemistry-only models by 18%, indicating that it may significantly impact our ability to predict the rate of soil C cycling processes. Future modeling with carefully applied machine learning approaches may open up new avenues for further extracting the relevantportions of SOM pools for inclusion in climate models.

382

383 Conclusion

Leveraging molecular information of SOM chemistry to improve conceptualizations and models 384 of soil C cycling is a pressing challenge for global biogeochemical and climate predictions. In 385 this study, we use machine learning (NMFk) to distill the thousands of SOM molecules detected 386 by ultrahigh resolution mass spectrometry in soil cores across the continental United States into 387 tractable units. These signatures of SOM composition represent different subsets of SOM 388 composition that contribute differently to soils exhibiting low versus high rates of potential 389 respiration. We then disentangle the SOM formula from each NMFk-extracted signatures and 390 validate their chemical recalcitrance in microbial respiration in surface soils. Additionally, SOM 391 chemistry (as summarized by NMFk) explained a greater proportion of potential soil respiration 392 than commonly measured physicochemical factors, and provided additional explanatory power 393 beyond these factors in combined models. Our results provide a basis for molecular information 394 395 to spur the development of new process-based modeling of soil C cycles and underscore the role of specific chemical constituents within the water-extractable SOM as a determinant of soil 396 397 respiration.

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Figure 1. Proposed workflow: Machine learning models summarize molecular data to predict soil

respiration. Non-negative matrix factorization (NMF*k*) extracts key SOM signatures from high

resolution mass spectrometry measurements of SOM. Gradient boosting regression predicts soil

respiration with physicochemistry, SOM signatures, and physicochemistry combined with SOM
 signatures.



Figure 2. The relationship between soil characteristics and potential respiration. (a-h) show [Manganese(Mn), Magnesium(Mg), Total Bases, CEC, Total C, Total N, GWC, Soil Temperature], respectively. Orange represents surface soils and blue represents subsoils. Lines denote the fitted linear regression function. Numbers on each panel are r^2 value from linear regression, the stars behind represents statistical significance (*** (p ≤ 0.001), **(p ≤ 0.01), ns (p > 0.05)).



Figure 3. (a) Spatial distribution of soil respiration levels (labeled by colors) and alpha diversity of each sample (sizes). Soil respiration levels are determined by *k*-means clustering on soil respiration rates (ug CO2/g soil/day). Soils from temperate conifer forests and temperate grasslands, savannas & shrublands have relatively higher respiration rates compared to other biomes (Figure S1). (b) The number of shared and unique SOM compound classes identified between surface and subsoils. The classes were suggested by van-Krevelen plot. (c) The difference of alpha diversity in surface and subsoil soils (p < 0.05 from ANOVA, *: p<0.05 from Tukey's HSD test) (d) the difference of alpha diversity in surface of alpha diversity in surface of alpha diversity in subsoils with different levels of potential respiration (p < 0.05 from ANOVA, *: p<0.05 from Tukey's HSD test) (e) the difference of alpha diversity in subsoils with different levels of potential respiration (p < 0.05 from ANOVA, *: p<0.05 from Tukey's HSD test) (e) the difference of alpha diversity in subsoils with different levels of potential respiration (p < 0.05 from ANOVA, *: p<0.05 from Tukey's HSD test) (e) the difference of alpha diversity in subsoils with different levels of potential respiration (p < 0.05 from ANOVA, *: p<0.05 from ANOVA).



Figure 4. NMF*k* partitioning of SOM composition. (a-b) Relative contribution of organic formula to each SOM signatures identified by NMF*k* in a) surface and b) subsoils. The color in each cell represents the normalized (0 to 1) relative contribution for each SOM feature (row) to each NMF*k* signature (column), red indicates the most important contributor, and blue indicates the least. The side bar indicates the compound class of each SOM feature. (c) The number of shared and unique formula identified as important (normalized weights >0.5) by NMF*k* in surface and subsoils. (d-e) The relative contribution of NMF*k* signatures to each level of respiration rates in both d) surface and e) subsoils. Surface soils: low respiration level (N = 44), medium respiration level (N = 14), high respiration level (N = 5, UT12, UT23, UT24, WY03, Temperate Conifer Forests, SCBI Temperate Broadleaf & Mixed Forests). Subsoils: low respiration level (N = 48), medium respiration level (N = 10), high respiration level (N = 3, T12, UT19, Temperate Conifer Forests, WLLO, Temperate Broadleaf & Mixed Forests).



Figure 5. Relative importance of each predictor in surface soil potential respiration machine learning models. a) Physicochemistry model, with physicochemical variables only. b) SOM model, with SOM signatures represented by NMFs only. c) Physicochemistry & SOM model with both physicochemical variables and SOM signatures.

Table 1. Model performance for predictions of potential soil respiration with physicochemical variables (Physiochemistry model), SOM by NMF*k* signatures (SOM_model), and combined physicochemical variables and SOM variables (Physiochemistry &SOM_model) for average 5-fold cross-validation accuracies (training soils, RMSE), and testing sample accuracies (RMSE, R2).

	Physiochemistry Model	SOM_model	Physicochemistry &SOM_model
Surface_CV	0.80	1.05	0.82
Surface_test	0.98	0.89	0.82
Surface_test (R2)	0.44	0.54	0.62
Subsoil_CV	0.60	0.82	0.67
Subsoil_test	0.46	0.80	0.49
Subsoil_test (R2)	0.43	0.08	0.36