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4 Bioenergetic modeling reveals opposing effects of ocean and terrestrial warming of an intertidal
5 crustacean

6 Running Title: Opposing effects of air and water warming

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18 Keywords: climate change; intertidal; cirripede; thermal ecology; Scope for Growth,
19 bioenergetic model, shore height, *Balanus glandula*

20

21 **Abstract**

22 Organism-level bioenergetics models (OBMs) are an emerging tool for predicting consequences
23 of climate change on organism growth in ecological systems. Global changes in ocean and
24 atmospheric temperature may affect organisms that experience both environments, such as those
25 living in intertidal systems. The acorn barnacle, *Balanus glandula*, is prevalent throughout the
26 intertidal zone in the Eastern Pacific, and laboratory experiments demonstrate that feeding rates
27 and metabolic costs are sensitive to temperature. We ask, based on these thermal responses, do
28 aerial and aquatic warming decrease *B. glandula* growth in a field environment because of
29 increased costs and reduced feeding? We measure environmental conditions (aerial and aquatic
30 temperature) at three intertidal heights over two 6-month intervals and compare growth estimates
31 based on an OBM to observed growth. Initial work indicates that growth is less sensitive to
32 elevation than predicted by lab-based physiological rates alone, so we estimate an elevation-
33 dependent compensation factor (Z) when fitting the model to all three elevations and the two
34 intervals. This full model predicts that, in this environment, aquatic warming will counteract
35 increased costs of aerial warming, by virtue of increased feeding at warmer temperatures. This
36 work advances OBMs by combining the effects of multiple decoupled thermal responses (e.g.
37 feeding, respiration) in multiple contexts (aerial, aquatic), drawing on established model
38 selection and “divide and conquer” techniques, and identifying sources of uncertainty in the
39 model. This work indicates that future intertidal OBMs may benefit from an improved
40 characterization of feeding behavior, including empirical estimates of elevation-dependent
41 feeding compensation.

42

43 **Introduction**

44

45 Organism-level bioenergetic models (OBMs) are emerging tools that predict species' responses
46 to climate change [1–3]. OBMs model an organism's energy needs as a function of abiotic
47 conditions, physiology, and species interactions to predict organismal growth and survival. There
48 are several different OBM frameworks, including Scope for Growth [4], the Wisconsin fish
49 bioenergetics model [5], Dynamic Energy Budget modeling (DEB) [6], the Metabolic Theory of
50 Ecology [7–9], and other agent-based physiological energy budget models (ABMs) [10–12]. As
51 mechanistic models, OBMs are better suited to predicting responses under the novel
52 combinations of environmental conditions that will occur under climate change than simpler,
53 correlational approaches such as species distribution models [1,11]. A successful OBM must
54 accurately capture environmental, physiological, and behavioral processes at the appropriate
55 temporal and spatial scales. The temperatures an organism experiences can vary over hourly,
56 daily, seasonal, and annual timescales [13], and the choice of timescale affects model predictions
57 [14]. Temperature, in turn, affects a wide range of physiological [15,16] and behavioral
58 processes [17–19]. Importantly, each of these processes within an organism may have its own
59 thermal dependence [11,15,20,21].

60

61 To have confidence in a model's climate change predictions, it must be validated against
62 empirical environmental and organismal data [11,22,23]. Model validation in more than one
63 location or against multiple combinations of empirical conditions is essential, since models that
64 perform well at one location or time can fail in other conditions [24,25]. Testing often reveals

65 poor agreement between model predictions and independent data [e.g. 22], but such tests provide
66 an important opportunity to identify mechanistic details that can improve models [23,26]. Model
67 validation should also include testing the sensitivity of the model to parameter uncertainty, as
68 this can identify areas where better empirical estimates of parameters would improve model
69 prediction [5,27,28].

70
71 Modeling decisions must be informed by the environment, or ecological niche, experienced by
72 the study species [29]. Warming and increased frequency of marine heat waves are associated
73 with shifts in marine communities at the land-sea interface [30,31]. Intertidal species experience
74 two distinct thermal environments, aquatic and terrestrial, that alternate with the oscillation of
75 tides. Individuals at higher elevations on a shore experience a shorter duration of submerged
76 conditions, producing steep environmental gradients of temperature, desiccation and feeding
77 opportunities [Fig. 1; ,18,25]. They often experience greater acute physiological stress [32,33]
78 and decreased growth and reproduction [34,35]. Given that terrestrial and marine environments
79 are expected to warm at different rates [36,37], an organism's shore height will also affect its
80 overall experience of climate change [38]. Further, individual species may differ in their
81 sensitivity to warming in the two environments [39]. Representing such a dynamic thermal
82 environment, where large shifts in temperature occur multiple times daily [40], and where
83 physiology varies with medium [41], requires a model with high-resolution timesteps [25,42–45]
84 and separate physiological dynamics in each environment (Fig. 2a,b).

85
86 **Figure 1** A) Abiotic conditions vary with the duration of low tide exposure at each shore height.
87 B) Violin plot of daily time emerged (exposed to air) at three different shore heights.

88
89 **Figure 2** Intertidal animals alternate between submergence under water (A) and emergence in air
90 (B), each with a distinct set of physiological conditions and processes. These can be captured in
91 a Numerical Scope for Growth model (C), if timesteps are high enough resolution to capture the
92 tidal fluctuations.

93
94 Scope for Growth (SFG) models are a class of OBMs that predict organismal growth and
95 reproduction by quantifying the effects of environmental conditions and size-scaling
96 relationships on physiology [46–50]. While SFG models were originally designed to calculate
97 energetics and growth from mean conditions over long (e.g. annual) intervals, more recently,
98 Numerical SFG (“NSFG”, Fig. 2C) models have been developed to capture shorter time steps
99 [51,52]. The shorter timesteps allow for energy fluxes to change allometrically as organisms
100 grow [52]. A central advantage of SFG and NSFG models is their use of empirically-derived
101 thermal sensitivities [5,47], rather than assuming a specific theoretically-derived relationship
102 between temperature and physiology [6,7]. This allows for separate physiologies in air and
103 water [e.g., 50] and for separate thermal sensitivities of energy demand and energy acquisition. The
104 latter is ignored in some OBM modeling frameworks [e.g., 6; but see, 51–53], despite strong
105 empirical evidence for its need [20,54,55]. Further, SFG and NSFG models’ rely on independent,
106 empirically-estimated parameters, each with well-characterized uncertainty. This allows for more
107 straightforward sensitivity analyses and model skill assessment than some more complex
108 modeling frameworks [27,56].

109

110 Here we develop an intertidal OBM model of the barnacle, *Balanus glandula*, using a hybrid of
111 NSFG and other modeling approaches. This sessile crustacean is native to the Pacific coast of
112 North America (Mexico to Alaska) [57] with a wide vertical distribution [58]. We model the
113 growth of *B. glandula* across an environmental gradient, shore height. We first develop this
114 model from empirical studies of *B. glandula* growth and physiology. We then validate the model
115 with two six month field observations of barnacle growth at three shore heights. Our objectives
116 are to (1) build the OBM model for *B. glandula* and test it across an environmental gradient, (2)
117 test the sensitivity of the model's predictions to parameter uncertainty, and (3) predict the effects
118 of future warming of air and water temperatures on *B. glandula*'s SFG across its vertical range.
119

120 **Methods**

121 *The traditional Scope for Growth model*

122 Scope for Growth (SFG) models [59,60] convert physiological rates into energy equivalents and
123 use the following equation:

$$124 \quad C = P + R + U + F \quad \text{eq. 1}$$

125 Where C is the total consumption of energy from food, P is the production of both somatic and
126 gonadal tissues, and R is the energy expenditure measured in terms of respiration. Additional
127 energy is lost in the excretion of nitrogen waste (U) and feces (F). We assumed U was zero
128 because Wu and Levings [50] found very little energy lost to excretion in *B. glandula*. The
129 production of tissue (P) may then be estimated as the difference between energy assimilated
130 from the food ($A = C - F$) and energy expenditure via respiration (R):

$$131 \quad P = A - R \quad \text{eq. 2}$$

132 This estimate (P) is known as the Scope for Growth (SFG).

133

134 *The Numerical Scope for Growth model*

135 We built a NSFG model (Table 1) in which energy fluxes (*i.e.*, feeding and respiration) were
136 calculated every 15 minutes. The energy flux estimates were then used to calculate a daily net
137 SFG and change in body size [52, Fig. 2]. We estimated daily SFG as the difference between
138 consumption and respiration costs within each day:

$$139 \quad \text{SFG}_{\text{day}} = A_{\text{day}} - R_{\text{day}} \quad \text{eq. 3}$$

140

141 Daily SFG was used to calculate barnacle opercular length (L_{day}) over the course of each 6-
142 month experiment. First, we calculated the daily mass at the start of the following day ($M_{\text{day}+1}$) as
143 the prior day's mass plus the change in mass (ΔM_{day}). We calculated ΔM_{day} by dividing the
144 SFG_{day} by the energy density of barnacle tissue (E_D , J / mg AFDW, Table 1) multiplied by 1.4 to
145 incorporate overhead costs of new tissue growth [6].

$$146 \quad \Delta M_{\text{day}} = \frac{\text{SFG}_{\text{day}}}{1.4 \times E_D} \quad \text{eq. 4}$$

147 Daily length was then calculated as a function of mass estimated from *B. glandula* from the same
148 location in the Pacific Northwest (<3mm opercular length, n=89, Palmer, 1980):

$$149 \quad L_{\text{day}} = 2.055921 M_{\text{day}} + 0.7912 \quad \text{eq. 5}$$

150

151

152

153

154 Table 1. Summary of model parameters and their sources.

Parameters from temperature experiments and the literature				
Symbol	Parameter name	Mean and SE	Unit	Source, equations
E_D	Energy density	22.8 ± 0.7	J / mg AFDW	Wu and Levings [50], June – Aug, 1 cal = 4.184J
Temperature parameters				Supplemental Information S1, Ober et al. (2019), Ober et al. <i>unpublished data</i>
a_I	Ingestion exponent	0.05936 ± 0.02853	unitless	
c_I	Ingestion coefficient	0.00056 ± 0.00022	prey / min	
$a_{R_{aq}}$	Aquatic resp. exponent	0.08031 ± 0.04077	unitless	
$c_{R_{aq}}$	Aquatic resp. coefficient	0.00003 ± 0.00002	$\mu\text{mol O}_2/15 \text{ min}$	
$a_{R_{EXP}}$	Aerial expos. exponent	0.04562 ± 0.01156	unitless	
$c_{R_{EXP}}$	Aerial expos. coefficient	0.00105 ± 0.00030	$\mu\text{mol O}_2/15 \text{ min}$	
$a_{R_{REC}}$	Aerial recov. exponent	0.02533 ± 0.00743	unitless	
$c_{R_{REC}}$	Aerial recov. coefficient	0.02677 ± 0.00473	$\mu\text{mol O}_2/15 \text{ min}$	
Size-scaling exponents				Supplemental Methods, estimated from Wu and Levings (1978)
d_I	Feeding size-scaling exponent	1.88 ± 0.10	unitless	
$d_{R_{aq}}$	Aquatic resp. size-scaling exponent	1.88 ± 0.10	unitless	
$d_{R_{aer}}$	Aerial resp. size-scaling exponent	2.32 ± 0.10	unitless	
Mass vs. length				
-	AFDW size-scaling exponent (used in temperature experiment equations)	2.83 ± 0.26	unitless	Gilman et al. [62]
-	AFDW size-scaling exponent (used for small experimental barnacles, 3mm operculum length and below)	0.4864	unitless	Palmer (1980)
-	AFDW size-scaling coefficient (used for small experimental barnacles, 3mm operculum length and below)	0.7912	unitless	Palmer [61]
Model parameters estimated from the optimization				
F_H	Chlorophyll half max	0.13 ± 0.04	Chlorophyll ($\mu\text{g} / \text{L}$)	This text
z	Feeding compensation exponent	1.16 ± 0.04	unitless	This text
p_{max}	Max assimilation rate relative to size and food concentration f (“a” in SFG)	2.04 ± 0.11	J / tissue mg AFDW	Calculated from Nishizaki and Carrington [63]

155

156

157 *15-minute energy flux calculations*

158 We modeled the temperature and size-dependence of ingestion and respiration separately for
159 immersion and emersion periods. All fluxes followed the same general function, $b(T, L)$:

160
$$b_{x,i}(T_i, L_i) = c_x \times e^{a_x T_i} \times L_i^d, \quad \text{eq. 6}$$

161 where x refers to a specific energy flux in either an air or water medium: ingestion (b_I) or
162 respiration (b_{R_AER} , b_{R_Recov} , b_{R_AQ}), and i refers to the 15-minute interval. T is the temperature, L
163 is the body size (operculum length, mm), d is an allometric length-scaling exponent, and a_x and
164 c_x are constants for the individual flux equations, fit from laboratory data (Table 1). We fit four
165 separate flux equations (Ingestion, Aquatic Respiration, Aerial Respiration, Aerial Recovery,
166 Table 1) to laboratory collected data (Supplemental Methods) [64]. The size scaling parameters
167 (d) were sourced from the published literature (Table 1, Supplemental Methods). Exponential
168 temperature scaling equations were selected (eq. 6) after first comparing the fit of 5 different
169 thermal scaling equations to the laboratory-collected data (Supplemental Methods) [65].

170

171 We calculated energy assimilation (A_i) at each time point from the ingestion flux ($b_{I,i}$),

172
$$A_i = p_{max} \times \frac{F_i}{F_i - F_H} \times b_{I,i}(T_i, L_i) \quad \text{eq. 7}$$

173 Where p_{max} is the maximum ingestion rate, and $b_{I,i}$ is the ingestion of the organism as a function
174 of T_i and L_i (eq. 6). Food availability ($f = \frac{F_i}{F_i - F_H}$, unitless), is a Holling's Type II functional
175 response where F_i is chlorophyll concentration at time i , and F_H is the chlorophyll concentration
176 at half of the maximum ingestion rate [6].

177

178 *Additional modifications for an intertidal species*

179 Additional modifications of the NSFG model were needed to accurately reflect intertidal
180 dynamics. First, we used separate energy demand equations during immersion and emersion that
181 reflected their differing thermal sensitivities. We further calculated the metabolic cost of aerial
182 exposure as the sum of measured respiration during emersion (b_{R_EXP}) plus additional recovery
183 costs (“oxygen debt”) upon resubmersion [64, b_{R_REC} 66]. Daily energy demand values were then
184 calculated as a daily sum of the 15-minute timesteps:

$$185 \quad R_d = \sum_{i=1}^{96} R_i \quad R_i = \begin{cases} WL_i \geq elev & b_{R_AQ,i} \\ WL_i < elev & b_{R_EXP,i} + b_{R_REC,i} \end{cases} \quad \text{eq. 8}$$

186 where WL_i is the water level at timestep i and $elev$ is the elevation or height on the shore of the
187 modeled barnacles.

188

189 Second, assimilation was calculated under immersion using Eq. 7, but during emersion
190 assimilation was considered to be negligible. Daily assimilation values (A_d) were then
191 determined from the summation of assimilation estimates at 15-minute timestep:

$$192 \quad A_d = \sum_{i=1}^{96} A_i \quad A_i = \begin{cases} WL_i \geq elev & A_i \\ WL_i < elev & 0 \end{cases} \quad \text{eq. 9}$$

193 Last, we added a tidal compensation term S^{\approx} (Table 1) to the feeding flux to allow for greater
194 rates of consumption at higher shore heights. Here S is the proportion of time submerged over
195 the entire period of the model. Without this term, the model would assume energy intake
196 declines linearly as a function of submergence times at higher shore heights. Yet many intertidal
197 species [34,67,68], including *B. glandula* [69] compensate for reduced submergence with greater

198 feeding activity and/or larger feeding structures. Initial model runs without this term severely
199 underestimated growth at higher shore heights and overestimated growth at lower shore heights.

200
$$A_{elevation,i} = A_i \times S^z$$
 eq. 10

201
202 *Observational study site*
203 Field temperatures and barnacle growth were recorded on a Southwest-facing rock wall at the
204 University of Washington's Friday Harbor Laboratories Biological Preserve (FHL), San Juan
205 Island, WA (48°32'45.1" N, 123° 0' 39.2" W) in the Salish Sea. This region is characterized by
206 mixed semi-diurnal tides, and experiences its greatest low tide temperatures in the summer, when
207 daytime low tides are most common [58]. Local water temperatures average ~10°C annually
208 [64].

209
210 *Biological observations*
211 Thirty-three 13.5cm x 13.5cm photoquadrats were established along the rock wall as part of a
212 larger study of *B. glandula* growth across multiple latitudes (Gilman et al., *unpubl. data*). At
213 each of three shore heights (1.20m, 1.55m, and 1.97m + MLLW), we spaced eleven quadrats in a
214 horizontal row approximately 1.5 m long. The three heights spanned the upper and lower limits
215 of *B. glandula* on the wall. We photographed each quadrat in February 2018, August 2018, and
216 March 2019. At each time point, a PentaxK50 digital SLR camera was placed 16 cm above the
217 quadrat. The opercular length of *B. glandula* was measured as the maximum operculum diameter
218 on the digital images using ImageJ (v.1.5) [70]. Growth was calculated as the change in
219 opercular length between each pair of time points. All analyses were restricted to the smallest

220 barnacles (1.5mm – 3mm initial opercular length) to maximize our ability to detect growth and to
221 ensure that we used an independent set of barnacles for each growth period.

222

223 *Environmental observations*

224 To estimate barnacle body temperature during both emersion and immersion, three temperature
225 dataloggers (Onset Hobo TidbiT v1 and v2) were deployed within each shore height and
226 recorded temperature every 15-min from February 2018 – March 2019. Temperature loggers
227 were attached to the rock face within individual quadrats using z-spar epoxy. We averaged the
228 replicate loggers within a shore height to reduce the effect of thermal microhabitat variation [71].
229 Tidal elevation was determined by comparing seawater temperature and 6-minute tide gauge data
230 (National Buoy Data Center Station FRDW1 - 9449880, NOAA National Ocean Service,
231 ndbc.noaa.gov/station_page.php?station=frdw1) to the 15-minute temperature logger data.
232 Weekly averages of seawater chlorophyll fluorescence were calculated from the Padilla Bay
233 Gong Site, WA (48° 33' 26.9" N, 122° 34' 19.2" W) and used to estimate food availability (YSI
234 EXO-2 sonde, Chl $\mu\text{g/L}$, NERRS CDMO PDBGSWQ, 2017 - 2020).

235

236 *Energy flux parameters*

237 We used laboratory-collected data [50,64; G. Ober *unpubl. data*] to fit the energy flux equations
238 $b_{R_AQ}(T, L)$, $b_{R_EXP}(T, L)$, and $b_{R_REC}(T, L)$ $b_I(T, L)$ (eq. 6, Supplemental Methods). Given that
239 these studies were performed over a narrow range of body sizes, the size-scaling exponents d_{AQ} ,
240 d_{AER} , and d_I were derived separately from equations reported by Wu and Levings [50] for *B.*
241 *glandula*. For energy demand, we converted the laboratory-measured respiration rates to joules
242 by multiplying by $0.457 \text{ J } \mu\text{mol O}_2 \text{ L}^{-1}$ [64,72,73].

243

244 *Model fitting*

245 We used a two-step optimization method to estimate three model parameters (p_{max} , z , F_H) on
246 independent subtidal and intertidal datasets. First, we determined the maximum assimilation rate
247 p_{max} by fitting the model to observed *B. glandula* growth over 5 weeks in subtidal conditions,
248 using data from Nishizaki and Carrington [63]. To account for a lag in growth, we used
249 chlorophyll values from 2 weeks prior to each growth timestep [74].

250

251 Second, we estimated the feeding compensation parameter (z) and the feeding half saturation
252 coefficient (F_H) by fitting the full model to the 291 intertidal barnacle growth measurements
253 across the three elevations and two-time intervals (n=10-107 growth measurements per elevation
254 and timepoint combination).

255

256 For both optimizations, the growth of individuals from the photo quadrats was compared with
257 model predictions of growth. Final length L_{final} was determined estimated from the numerical
258 model. The initial length of each replicate sample (L_{init}) was the initial length of each barnacle
259 from the photo quadrat, and the model estimated length (L_{day}) each day based on SFG, iterating
260 for each day of the 6-month dataset. The predicted growth ΔL_{pred} was calculated for each
261 replicate barnacle as the difference between the final (L_{final}) and initial lengths (L_{init}). We solved
262 for the values of p_{max} , z , and F_H that gave the lowest negative log likelihood of the data given the
263 model, optimizing for the sum of $\Delta L_{pred} = \Delta L_{obs}$ for all barnacle growth measurements.

264

265 *Statistical and numerical analysis*

266 All statistical analyses and numerical modeling were performed in R (v4.0) and RStudio (v1.3).
267 The model optimization minimized the sum of the negative log likelihoods (NLLs) of the data
268 given the model for each tidal elevations and season. NLL estimations assumed a normal
269 distribution. A bounded BFGS algorithm was used for the maximum likelihood estimation
270 (optim package in R). The hessian matrix was used to estimate approximate standard error of the
271 parameters.

272

273 *Sensitivity analysis*

274 The sensitivity analysis of the NSFG model was modified from the individual parameter
275 perturbation (IPP) approach [e.g., 5]. Rather than determining the sensitivity to a percentage
276 change in the absolute value of each parameter, we determined the sensitivity to a change of one
277 standard error (± 1 SE) in each parameter value [75] This method was used to capture the
278 sensitivity of the model relative to a realistic range of uncertainty in parameters values [76]
279 rather than a fixed percentage of the mean. The standard error of each parameter value was
280 determined from either the published literature or from our empirical parameter fits (Table 1). A
281 sensitivity of 1.5 indicates that a change in the parameter by 1 SE causes a resulting change in
282 the SFG by 50%.

283

284 *Climate change estimation*

285 The effect of ocean and aerial warming on cumulative SFG (J) was determined over each 6-
286 month interval for a barnacle of an initial size of 2.2 mm at each elevation. Seawater and/or
287 aerial warming was estimated by adding a discrete integer value from 0 to 2°C (seawater) and/or

288 0 to 4°C (aerial warming) to every 15-minute observation of estimated body temperature during
289 immersion and emersion. We chose these ranges because they encompassed the range of abiotic
290 warming expected in the Salish Sea in the year 2095 relative to 2000 under the Representative
291 Concentration Pathway 8.5 emissions scenario (RCP8.5) [36].

292 **Results**

293 *Environmental and biological observations*

294 Weekly averaged chlorophyll ranged from 0 to 18 $\mu\text{g} / \text{L}$ and was highest in June. Measured
295 water temperatures at the field site ranged from 5°C to 17°C, and low tide temperatures ranged
296 from -3°C to 31°C. Aerial exposure occurred 72% of the year at the highest elevation, 48% at
297 the mid elevation, and 33% at the lowest elevation (Fig 1b, 3a-c, 4a-c). The low shore height
298 experienced consistently cooler aerial temperatures than the other two shore heights (Fig 3d-f,
299 4d-f). However, the mid shore was often warmer at low tide than the high shore, due to a rock
300 overhang that shaded some of the high shore quadrats. Generally, the warmest aerial
301 temperatures occurred in August, while the coldest were in February.

302

303 **Figure 3.** Model inputs and predicted barnacle growth for 3 shore heights over interval 1 (Feb-
304 Aug 2018). A-C) Proportion of the day submerged and food availability (f) in the low, middle,
305 and upper intertidal zone during the first interval. D-F) Mean daily temperature during aerial
306 (solid purple line) and aquatic (solid blue line) conditions and daily 75% quantile of temperature
307 (dotted red line). G-I) Daily physiological intake (dark green), aerial cost (purple), and aquatic
308 cost (blue) of a barnacle of 2mm initial operculum length. The aerial cost includes both aerial

309 exposure and aerial recovery costs. J-L) Daily Scope for Growth of a representative barnacle of
310 ~2mm.

311

312 **Figure 4.** Model inputs and predicted barnacle growth for 3 shore heights over interval 2 (Aug
313 2018- Mar 2019). See Figure 3 for details.

314

315 Small barnacles (mean initial size 2.2 mm) grew 0.5 to 0.8 mm in operculum length over 6
316 months depending on the time interval and shore height (Figure 5, x-axis). Individuals in the low
317 intertidal tended to grow 34 and 40% more than those the upper intertidal in intervals 1 and 2,
318 respectively (Figure S2.1), but this difference was not statistically significant (Table S2.1). The
319 narrow range in mean growth rates contrasted with the large range in time submerged across the
320 three shore heights (Figure 1b).

321

322 **Figure 5.** The relationship between predicted growth and observed growth in mm at the three
323 elevations (Low – blue, mid – orange, upper – grey) in interval 1 (closed circles) and interval 2
324 (open circles, means \pm SE).

325

326 *Model Fit*

327 Predictions from the NSFG model matched mean observed growth fairly well ($R^2 = 0.76$), with
328 the largest model discrepancies in the low intertidal, which had the fewest samples (Figure 5).

329 There was also a large amount of inter-individual variation in growth within each time point and
330 shore height, and individual growth was not well predicted by the SFG model ($R^2 = 0.07$, data

331 not shown). Representative trajectories of energy fluxes in aerial and aquatic conditions over the
332 course of a day for each elevation are shown in Supplemental Figure S2.2.

333

334 *Parameter estimation*

335 Based on the subtidal analysis, we found p_{max} , the size-dependent maximum assimilation rate
336 was 2.0 ± 0.1 J normalized per mg of tissue (AFDW, Table 1). The chlorophyll half-saturation
337 coefficient, fit on the intertidal growth data, was low ($F_H = 0.13 \pm 0.04$ $\mu\text{g Chl / L}$), relative to
338 the observed chlorophyll concentrations. Consequently, the model predicted feeding and
339 ingestion (f) to remain over 80% of the maximum ingestion rate throughout the study (Figure
340 S2.3, Fig 3a-c, Fig 4a-c). The feeding compensation exponent, z , was estimated as 1.16 ± 0.04 ,
341 which corresponds to a three-fold increase in feeding activity for barnacles at the high shore
342 height compared to those at the low shore height (Table 1, Figure 6).

343

344 **Figure 6.** Estimated feeding activity in the upper (square), mid (circle), low (triangle) tidal
345 elevations as a function of the proportion of time submerged over each 6-month interval (interval
346 1 – open symbols, interval 2 – closed symbols). Feeding activity at each elevation is calculated
347 as an exponential function of the proportion of time submerged (S) over the full 6-month
348 interval, $y = S^z$ ($z = 1.16 + 0.04$, mean + SE, Table 1).

349

350 *Daily energy fluxes and predicted Scope for Growth*

351 Daily energy intake and costs followed the daily patterns in submergence time. On days with
352 more time emerged aerial costs increased, while feeding and aquatic costs increased on days with
353 more time submerged. As barnacles grew over each 6-month period, all energy fluxes increased

354 (Fig 3G-F, 4G-F). Interestingly, barnacles in the low intertidal exhibited the greatest energy
355 intake even after including feeding compensation in the model (Fig. 7, Supplemental Figure
356 S2.4). For animals at higher shore heights, the decreased time in aquatic conditions yielded a
357 decrease in immersion costs and an increase in emersion costs (Figure 7). Overall, predicted SFG
358 increased over time across all elevations. Interestingly, except for the low shore (Fig 3J, 4J), SFG
359 was not monotonic with time. In the upper intertidal, the cumulative SFG initially decreased in
360 September, a period of low submergence durations and warm aerial temperatures, before
361 increasing later in the interval (Figure 4L).

362

363 **Figure 7.** Violin plots of daily energy fluxes at each tidal elevation for interval 1 (A) and interval
364 2 (B). Cumulative intake (green), aquatic cost (blue) and aerial cost (purple). Aerial cost is the
365 sum of exposure and recovery costs.

366

367 *Model sensitivity to parameter estimates*

368 The individual parameter perturbation (IPP) found that the SFG (J) was most sensitive to
369 uncertainty in the temperature parameters for feeding, followed by aerial and aquatic respiration
370 (Figure 8). Increasing the feeding coefficient (c_f) and feeding temperature exponent (a_f) by each
371 parameter's empirical standard error (SE) had the largest effect on predicted SFG. A one SE
372 increase in c_f resulted in a 300% increase in SFG, while a one SE increase in a_f led to a 260%
373 increase in SFG. Increasing the aquatic and aerial respiration coefficients ($a_{R_{aq}}$ and $a_{R_{aer}}$) by
374 one SE caused a ~ 80% decrease in SFG. In contrast, SFG was minimally sensitive to changes in
375 the energy density parameter (E_D , J mg⁻¹) and in the size scaling parameters for all energy fluxes
376 (d_f , $d_{R_{aq}}$, $d_{R_{aer}}$). SFG was also more sensitive to the aerial exposure parameter uncertainty ($a_{R_{EXP}}$,

377 $c_{R EXP}$) compared to the aerial recovery parameter uncertainty ($a_{R REC}$, $c_{R REC}$). Finally, the
378 variability of the SFG estimate due to the variability in initial body size was 40% (population SE,
379 Pop_{SE} , Figure 8).

380

381 **Figure 8.** Sensitivity analysis representing the influence of parameter uncertainty on cumulative
382 barnacle SFG (J) over 6 months (see Table 1 for parameter definitions). The sensitivities are for
383 the SFG of a representative barnacle in the mid elevations in interval 1 (Feb – Aug), and the
384 Pop_{SE} is the population uncertainty among the barnacles in the middle elevation.

385

386 *Warming body temperatures and SFG*

387 Warming body temperatures during emersion decreased predicted growth by 10-30% per 1°C
388 increase, depending on the shore height and time interval, while warming during immersion
389 increased predicted growth by 15-40% per 1°C (Fig 9). For example, in the mid shore, a 2°C
390 increase in aerial body temperatures resulted in a 30-50% reduction in SFG, while a 2° C
391 increase during immersion increased predicted SFG by 40-60%. Thus, when body temperatures
392 warmed similar amounts in air and water there was almost no change in predicted growth.

393

394 In scenarios that remained below 2°C warming, SFG remained high in the low shore sites (>70
395 J), with only a 17-23% decrease in predicted growth due to warming aerial body temperatures.
396 Both the cost of aerial warming and the benefit of warming during immersion were greater in the
397 upper intertidal, where 4°C aerial warming alone caused the SFG prediction to drop to near or
398 below 0 (-7 to 12J). However, when that aerial warming was combined with 2°C aquatic

399 warming, the high shore SFG remained above 0, and within 30-50% of the present day SFG
400 prediction.

401

402 **Figure 9.** The influence of increased body temperatures during aerial and aquatic exposure on
403 cumulative barnacle SFG (J) over 6 months in the upper (A, B), mid (C, D), and low (E, F)
404 elevations in interval 1 (Feb – Aug; A,C,E) and interval 2 (Aug – Feb; B,D,F) for a theoretical
405 barnacle of 2.2 mm initial operculum length. The SFG for the observed environmental
406 temperatures ($\Delta^{\circ}\text{C} = 0$; lower left quadrant of each 3X3 square) ranged from 40 to >100 J.

407

408 **Discussion**

409 Successfully predicting the effects of warming temperatures from climate change on species
410 requires building physiologically accurate, field-tested models of organismal responses to
411 warming temperatures [1,77]. Here we developed a Numerical Scope for Growth (NSFG) model
412 for the intertidal barnacle *Balanus glandula* and used it to predict the effects of warming body
413 temperature during high and low tide. The model explained 76% of the variability in growth
414 across the three intertidal shore heights and two time periods, after adding a new parameter for
415 feeding compensation. We found that increasing body temperatures during immersion buffered
416 the negative effects of increased air temperatures at low tide. Specifically, higher aquatic body
417 temperatures increased growth by increasing feeding rates during immersion more than they
418 increased energy costs, while warming aerial body temperatures reduced growth by increasing
419 costs without changing feeding. The NSFG model demonstrated a new method for developing
420 and testing OBMs that used high resolution timesteps, independent curve fits for different energy

421 fluxes, and estimated one parameter, rather than many, with an independent subtidal growth
422 dataset [11]. However, our findings are contingent on the accuracy of the laboratory-measured
423 parameters for feeding and energy demand. Our sensitivity analysis showed that lab-based
424 predictions of growth were greatly influenced by the uncertainty in the relationship between
425 temperature and feeding. Finally, this work highlights the importance of model testing and
426 sensitivity analyses to identify information gaps and form new hypotheses that can improve
427 future models.

428

429 *Warming body temperatures and barnacle SFG*

430 Our model suggests that *B. glandula* will respond very differently to warming body temperatures
431 at high and low tide. Warming during submersion enhanced growth while warming during
432 emersion reduced growth (Fig 9). When submerged, the energetic costs of warming were offset
433 by larger gains in energy intake. In contrast, emersion warming could only influence costs. This
434 effect was greatest on the low shore, where Scope for Growth increased with warming as long as
435 the magnitude of aerial warming did not exceed aquatic warming (Fig 9E,F). In contrast,
436 warming on the high shore only led to increased growth when aquatic warming was greater than
437 aerial warming (Fig 9A,B). Interestingly, shore height also influenced the relationship between
438 SFG and aerial warming, as the same amount of aerial warming depressed SFG more on the high
439 than the mid shore, even though the mid-shore had warmer emersion temperatures overall.

440

441 Importantly, terrestrial rates of warming are predicted to exceed marine warming, a phenomenon
442 known as the “land-sea warming contrast” [78]. In the Salish Sea, average air temperatures are
443 predicted to increase by 3.5C by 2095 under RCP 8.5, more than double the predicted sea surface

444 temperatures warming of 1.57°C [36,37]. At this 2:1 warming ratio of emersion to immersion,
445 our model predicts up to a 40-70% decrease in SFG at the higher elevation, and a 10-40%
446 decrease in SFG at the lower elevation, depending on the time interval.

447
448 These results suggest that the negative effects of climate warming are likely to be greatest on the
449 high shore. If this is the case, *B. glandula* could experience a vertical contraction of its upper
450 limit, due to negative Scope for Growth (Figs. 3,4) [79], even as individuals lower on the shore
451 continue to thrive. Indeed, Harley [30] has already documented an average 40cm drop in the
452 upper vertical limit of *B. glandula* over 50 years at 20 sites within the Salish Sea. However, the
453 relationship between survival and energetics is complex [80]. These vertical range contractions
454 could also be driven by changes in the frequency of acute lethal events [81] or changes in other
455 low tide stressors, such as desiccation [82], rather than a sustained negative Scope for Growth.
456 Further, our study animals were 2-3mm opercular length, but earlier life stages could be more
457 susceptible to abiotic stressors at high elevations [83]. Additional research is needed to determine
458 the relative importance of these different mechanisms of stress, including interactions, to
459 intertidal species distributions. It is also worth noting that, while intertidal animals will
460 experience the full amount of aquatic warming, aerial body temperature warming is often much
461 less than the full amount of abiotic aerial warming [38,84]. If barnacle body temperatures during
462 emersion warm more slowly than air temperatures, barnacles will be less negatively impacted
463 than we predict here. Accurate predictions of barnacle body temperature warming at low tide
464 will require detailed biophysical modeling [38,77].

465

466 Intertidal organisms in the Salish Sea, including *B. glandula*, occupy a unique thermal niche. In
467 this region, *B. glandula* experience some of the coldest immersion temperatures anywhere in its
468 geographic range and some of the warmest emersion body temperatures [85]. It is unclear how
469 generalizable the benefits of warming aquatic temperatures observed here are to other
470 populations of *B. glandula* or to other intertidal species. Experimental studies of other
471 Northeastern Pacific intertidal species, have found an increase in feeding rates under warming
472 water temperatures for some snails [86–88], seastars [86] and barnacles [39], but increases in
473 feeding do not always translate into increased growth [86]. Moreover, different populations of a
474 species can vary in their response to temperature [88]. The relationship between growth and
475 temperature can also be influenced by species interactions, such as non-lethal predator effects on
476 prey [39,89]. Further, species that regularly experience temperatures near and above their
477 thermal feeding optima, or that live in environments in which temperature modulates local
478 biogeochemistry may be especially vulnerable to the combined effects of aerial and water
479 warming [90]. Thus, while some intertidal species may experience the benefits of warming water
480 temperatures under specific conditions, those conditions may be idiosyncratic to specific
481 populations and/or ecological situations.

482

483 *Advantages and limitations of our Numerical Scope for Growth approach*

484 One of the central challenges of modeling organismal responses to climate change is designing
485 models that capture physiology and environmental conditions at appropriate levels of detail
486 [40,77]. A multitude of organism-level bioenergetic models have been proposed [5,6,9,46]. We
487 used a hybrid modeling approach that combined a short time-step Numerical Scope for Growth
488 model with elements of several other approaches (Fig. 2, Table 1). Our OBM model estimated

489 growth daily, which is a much shorter timestep than traditional SFG models, and relied on
490 temperature data at an even shorter timestep (15 minutes) to accurately capture the dynamic tidal
491 environment of *B. glandula*. The model also incorporated key ideas from DEB models, including
492 a nonlinear function of feeding rate relative to food concentration [6,52] and a 40% overhead
493 cost for growth [6].

494
495 One major difference among OBM frameworks is the number of distinct thermal sensitivities
496 incorporated into the model for different aspects of an organism's physiology, such as
497 consumption and metabolism. At one extreme, all thermal sensitivities in DEB and MTE are
498 modeled with a single equation derived from theoretical principles [6,7], although some
499 applications of these models allow individual parameters to vary by physiological process [e.g.,
500 9]. In contrast, fish bioenergetics and SFG models typically incorporate unique thermal
501 sensitivity equations for each process and fit these equations from empirical data [5,e.g., 47].
502 We chose the latter approach, because of a growing consensus that energy consumption and
503 energy demand exhibit different thermal sensitivities [11,20,55]. We fit four distinct curves:
504 feeding, aquatic energy demand, aerial energy demand, and aerial recovery (oxygen debt). In
505 fitting thermal sensitivity equations to empirical data, there are a wide range of curves to choose
506 from [8,91,92]. We used an information theory approach to compare 5 representative curve
507 formulations (Supplemental Methods). In fitting the curves, we drew on empirical datasets that
508 contained a wide range of temperatures, well beyond what this species currently experiences
509 (aerial: 5-38°C, aquatic: 5-26°C), to ensure the model would be relevant for novel environmental
510 conditions [11].

511

512 Divide and conquer strategies integrate sub-model simulations with top-level simulation
513 experiments with the aim of assessing research questions with more valid and realistic
514 simulations [11,93]. Many bioenergetics models include sub-model growth simulations within
515 the preparatory stage [26,94,95]. Importantly, we used a sub-model growth simulation to
516 estimate two parameters, only one of which was then integrated into the top-level model.
517 Specifically, this sub-model simulation was an independent dataset of barnacle growth in a
518 subtidal environment (no immergence). This sub-model simulation was used to estimate
519 maximum assimilation rate p_{max} . Then, the top-level experiment investigated whether the model
520 predicted growth across three tidal elevations and two time periods. When this model failed to
521 converge, we tested a new quantitative hypothesis, that feeding varied exponentially with tidal
522 elevation, and we then tested this against all three tidal elevations and two time periods. This
523 strategy allowed us to use one parameter (z) to account for the discrepancy between predicted
524 growth based on the model vs. observed growth.

525
526 Within a statistical model, only some parameters have scientific interest. In such models,
527 “nuisance parameters” address random mechanisms, and variability resulting from them, but
528 have no intrinsic value in themselves [96]. The half-saturation coefficient is used in NSFG and
529 DEB models [6,52], and allows for saturation of feeding at high food densities which is more
530 realistic [97,98]. Here, the half saturation coefficient for feeding (F_H) was estimated once,
531 independently for each of the sub model and top level model. In DEB models, this coefficient is
532 calculated for each location separately [6]. This method of validation in place of independent
533 model testing is difficult to avoid since this variable is associated with food quality and is
534 difficult to characterize empirically [6], and may vary by location [24,25]. In our study, the three

535 shore heights were within a meter of one another. While it's plausible that food quality may have
536 varied across the 6-month experiment, this work makes strategic use of a system in which food
537 quality across space is a reasonable assumption. The half-saturation coefficient value (Table 1)
538 constrained feeding to ~80-100% of the maximum feeding rate throughout the entire year (Fig
539 3a-c). Rather than considering this value as corresponding to some intrinsic value of food
540 quality, perhaps this number is best considered a nuisance parameter that constrains the influence
541 of the food metric on intake. In this study, a low F_H limited the influence of monthly chlorophyll
542 on food availability (Fig S2.3).

543

544 *Tidal compensation and sensitivity of the SFG estimate to feeding*

545 One surprising result of our model was that it failed to capture growth across the tidal gradient
546 unless we allowed feeding rate to vary with shore height. Most intertidal animals can only feed
547 while submerged so that a shorter submergence time should shorten the time for feeding. Yet,
548 growth differences across tidal elevation were not directly proportional to submergence time. *B.*
549 *glandula* at the higher elevations were submerged ~60% less than those in the lower elevation,
550 but their growth was only 30% less than that in the low elevation and this difference was not
551 statistically significant (Figure S2.2, Supplemental Table S2.1). Intertidal animals can
552 compensate for reduced submersion by either increasing feeding rates or conserving energy
553 [34,99–101]. While not all intertidal species exhibit tidal compensation [99], 1.5 to 10-fold
554 increases in feeding rates with elevation have been reported in *Balanus* species [68,69]. We
555 found a 3-fold increase in feeding activity across *B. glandula*'s vertical range, which is
556 comparable to the 6-fold increase reported by Horn et al. [69] for a central California population
557 of *B. glandula*. Other factors, such as energy conservation at high elevations [34], species

558 interactions [102], or hydrodynamics [63] could also modulate relationships between
559 submergence time and feeding activity. The need for a tidal compensation parameter highlights
560 the importance of strategic field-testing models across multiple environments or populations
561 [11]. A model based on only laboratory data or tested in only one environment would not have
562 identified the need for tidal compensation.

563
564 The sensitivity analysis also identified that the SFG was most sensitive to the uncertainty in the
565 feeding energy flux equation. Specifically, increasing the feeding coefficient by its SE increased
566 SFG by ~3-fold. Feeding rates are highly variable and challenging to characterize given effects
567 of particle selection, food quality, uptake of nutrients, current velocity [63], and the hiding
568 behaviors [68] and passive vs. active feeding behaviors [69] described above. This work suggests
569 that future lab-based growth models may be improved by focusing on more accurately
570 characterizing feeding behavior.

571

572 *Model sensitivity to parameter uncertainty*

573 One advantage of Scope for Growth models over other OBMs is that they have a simpler
574 structure so model uncertainty is easier to evaluate and interpret [27,28]. We used a variation on
575 Individual Parameter Perturbation [5,76], in which we tested the model sensitivity to parameter
576 uncertainty. This method allowed for realistic estimates of uncertainty that are scaled based on
577 parameter uncertainty rather than the magnitude of the parameter. When parameter sensitivity is
578 calculated in bioenergetics models, it is often based a nominal 10% change in each parameter
579 value to investigate sensitivity to each value [5,103]. A nominal 10% change in a parameter
580 value is sensitive to the units (e.g. °C vs. K), and may not be biologically relevant. Here, we

581 estimated the error around each parameter (SE) or obtained its SE from the literature, and
582 perturbed each parameter with this uncertainty [75]. This method may be most appropriate for
583 NSFG models and other models with simple structures that meet two criteria; First, there is little
584 to no information about the shape of the priors necessary for Bayesian analyses [27, Personal
585 communication Boersch-Supan], and second, the uncertainty of individual parameters is
586 estimated separately from independent datasets such that newer multi-tier parameter estimation
587 methods are not required to characterize parameter uncertainty [104].

588

589 *Conclusions*

590 This study presents an OBM to describe physiological consequences of changing aerial and
591 atmospheric temperatures on a rocky intertidal crustacean. We found that warming of aerial and
592 aquatic temperatures had opposite effects on the growth of the intertidal barnacle *Balanus*
593 *glandula*. The high sensitivity of the model to the relationship between temperature and feeding
594 suggests that factors affecting clearance rates, feeding, and assimilation may have a large impact
595 on growth and distribution. Our OBM applied standardized statistical methods to physiological
596 energy budgets, incorporating high-resolution environmental data and a suite of physiological
597 lab-derived data. The process of field testing models is necessary to build effective predictive
598 models that forecast the effect of climate change on marine species [11]. Such models are needed
599 to adjust marine species management and conservation efforts to dynamic and changing
600 environmental conditions [10,12].

601

602

603 **Acknowledgements**

604 We thank R. Rognstad, E. Hazelton, E. Carrington and M. Nishizaki for generously sharing data
605 and providing other help. We also thank A. Gleekel, S. Malik, E. Ueland, S. Ueland, K. Roberts,
606 E. Kovachevich, and S. Martin for their contributions to the photoquadrat dataset. Finally,
607 special thanks to E. Pousse, E. Munroe, W. King, and R. Cerrato for feedback and insightful
608 discussions that improved the quality of this manuscript. This work was supported by the
609 National Science Foundation (grant IOS-1351445 to S.E.G.) with additional support from the
610 Keck Science Department of Claremont McKenna, Pitzer, and Scripps Colleges. Any opinions,
611 findings, and conclusions or recommendations expressed in this material are those of the authors
612 and do not necessarily reflect the views of the National Science Foundation.

613 **Full data citations**

614 NOAA National Estuarine Research Reserve System (NERRS). System-wide Monitoring
615 Program. Data accessed from the NOAA NERRS Centralized Data Management Office website:
616 *accessed 22 October 2020.*

617

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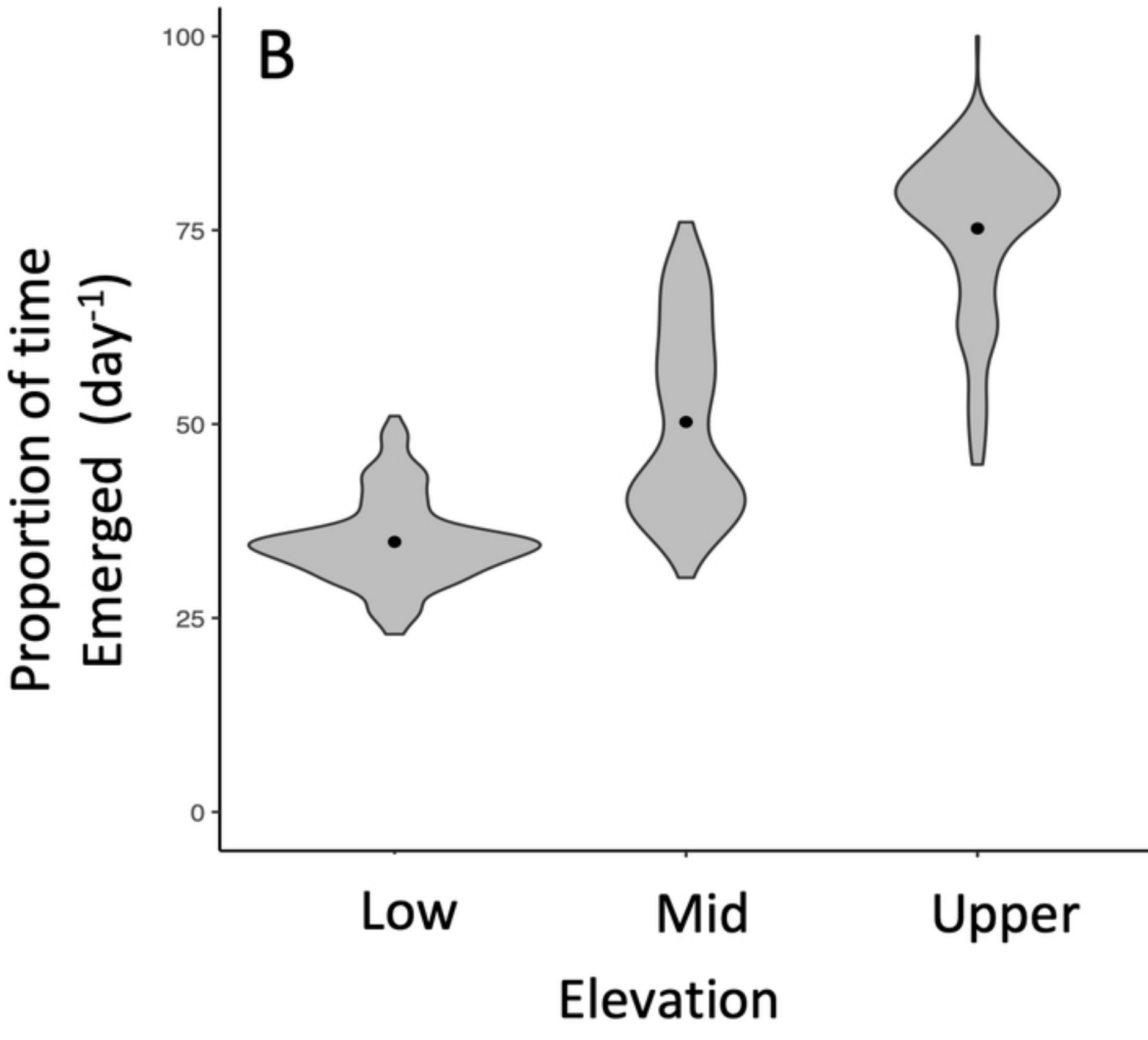
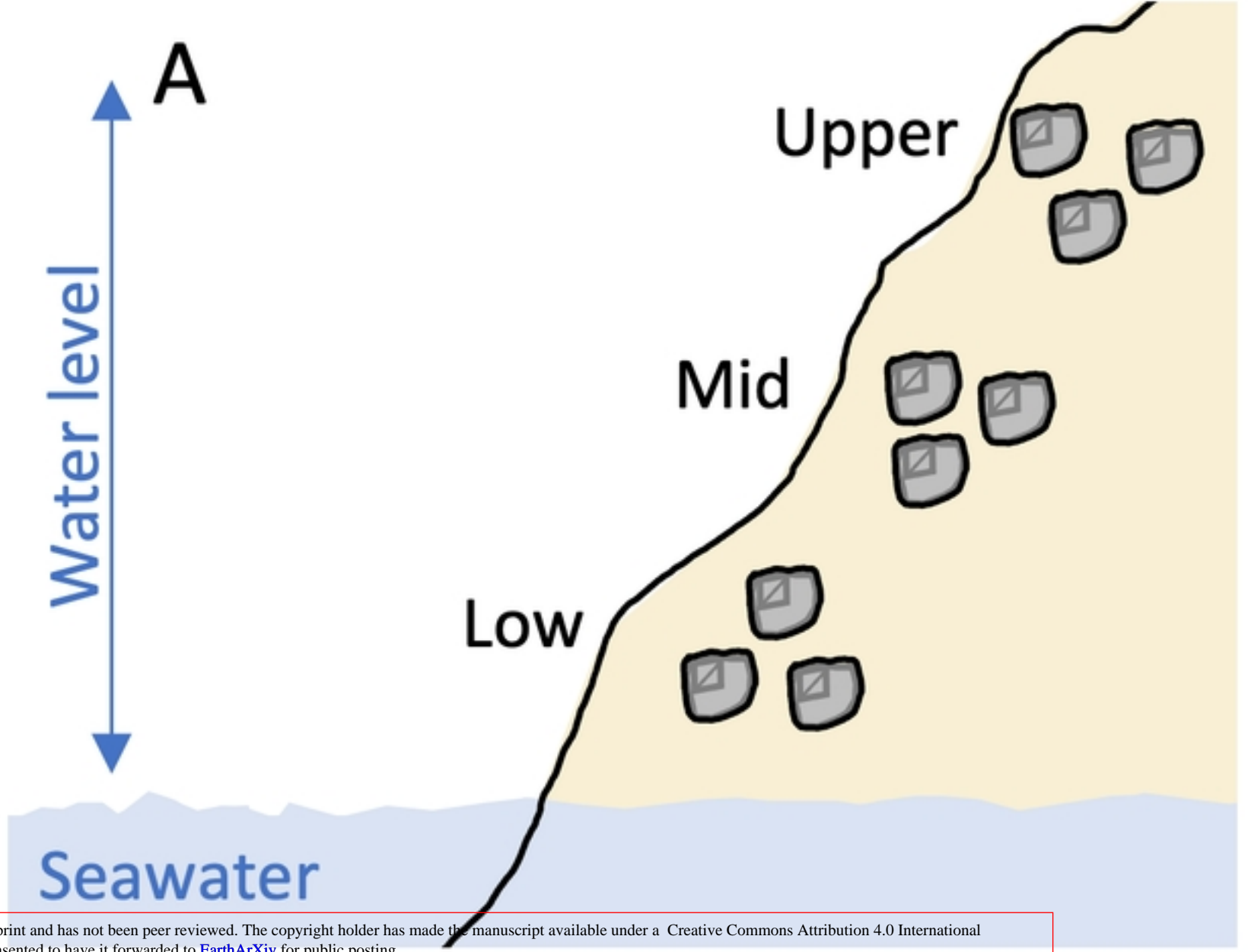


Fig1

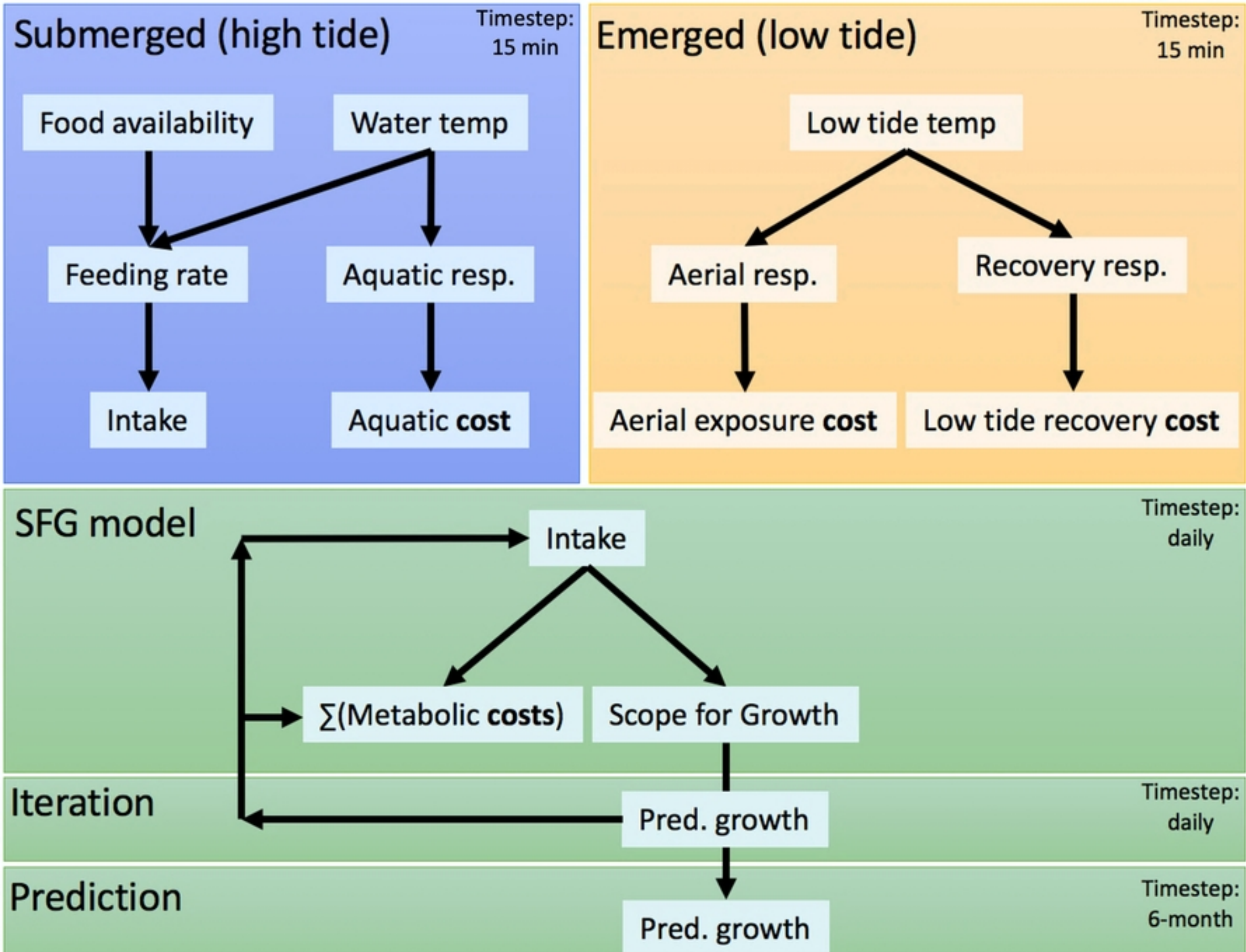


Fig2

Interval 1

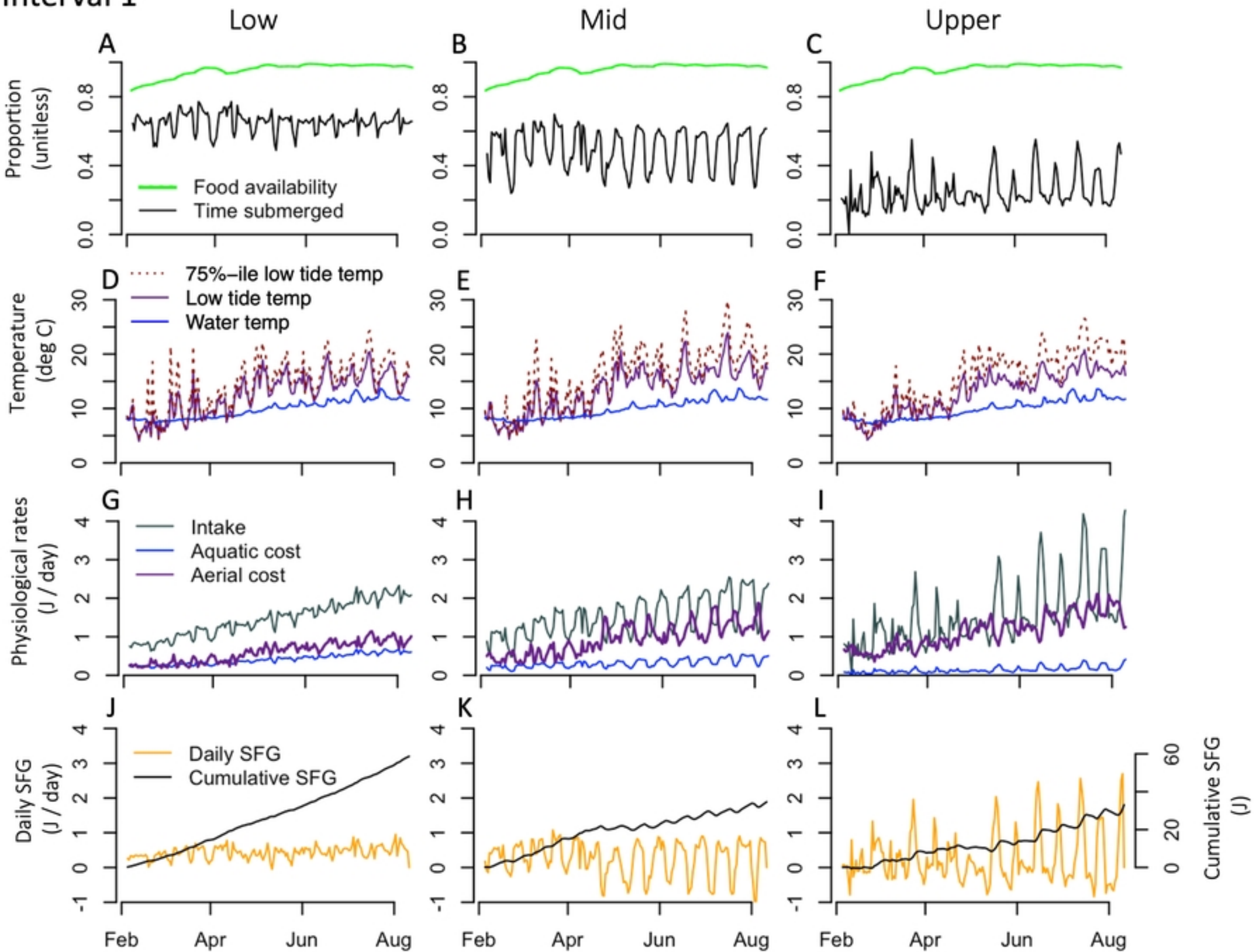


Fig3

Interval 2

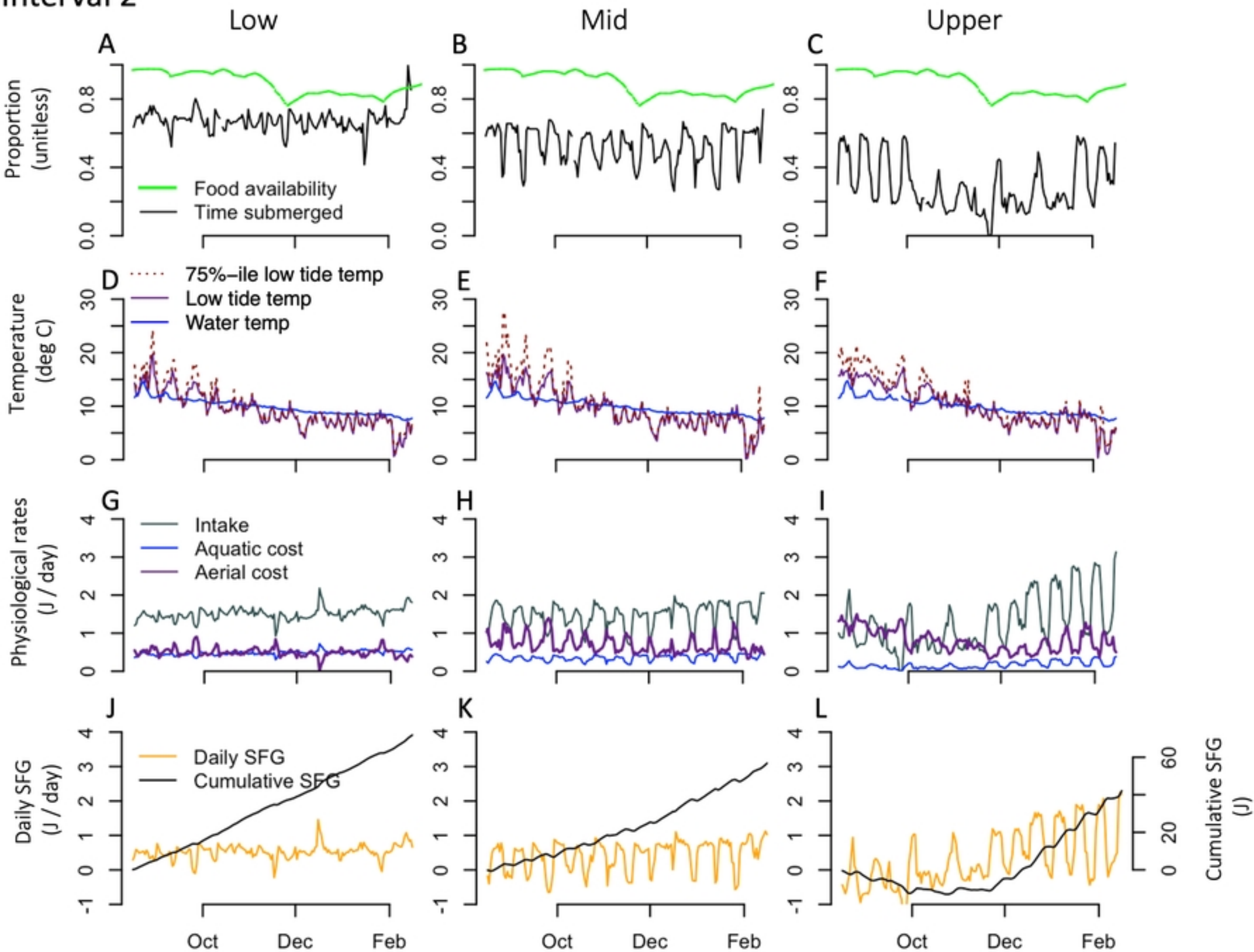


Fig4

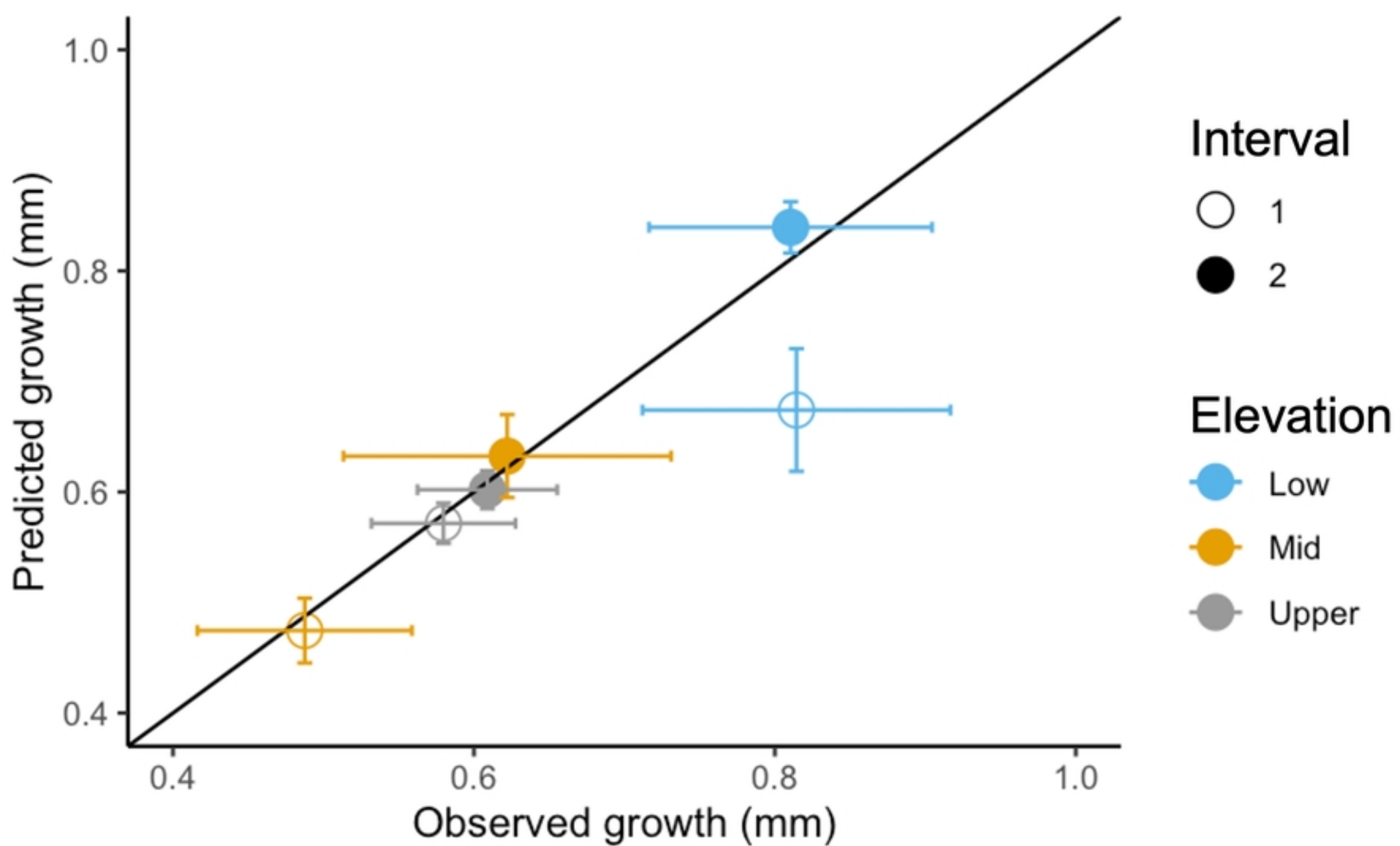


Fig5

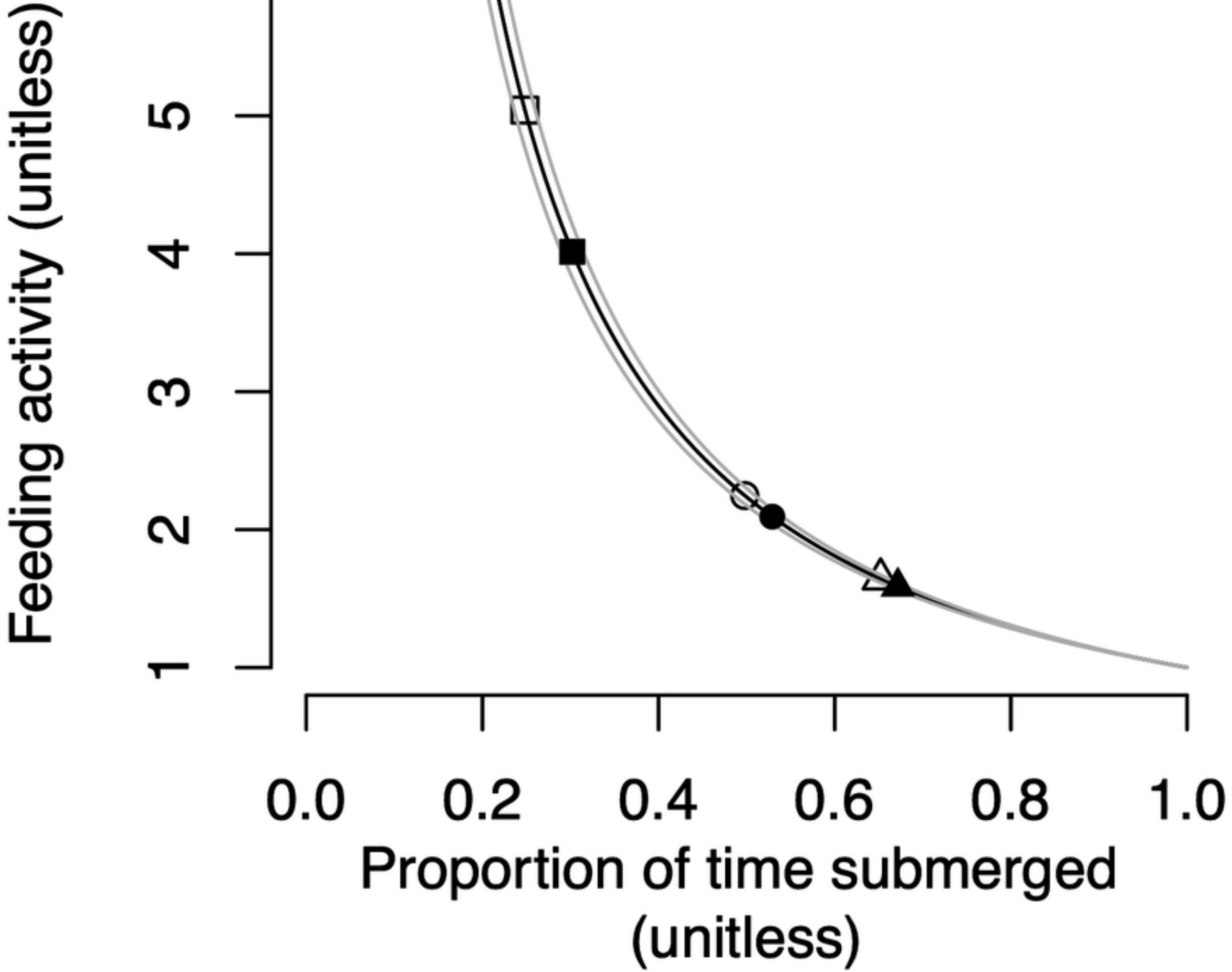


Fig6

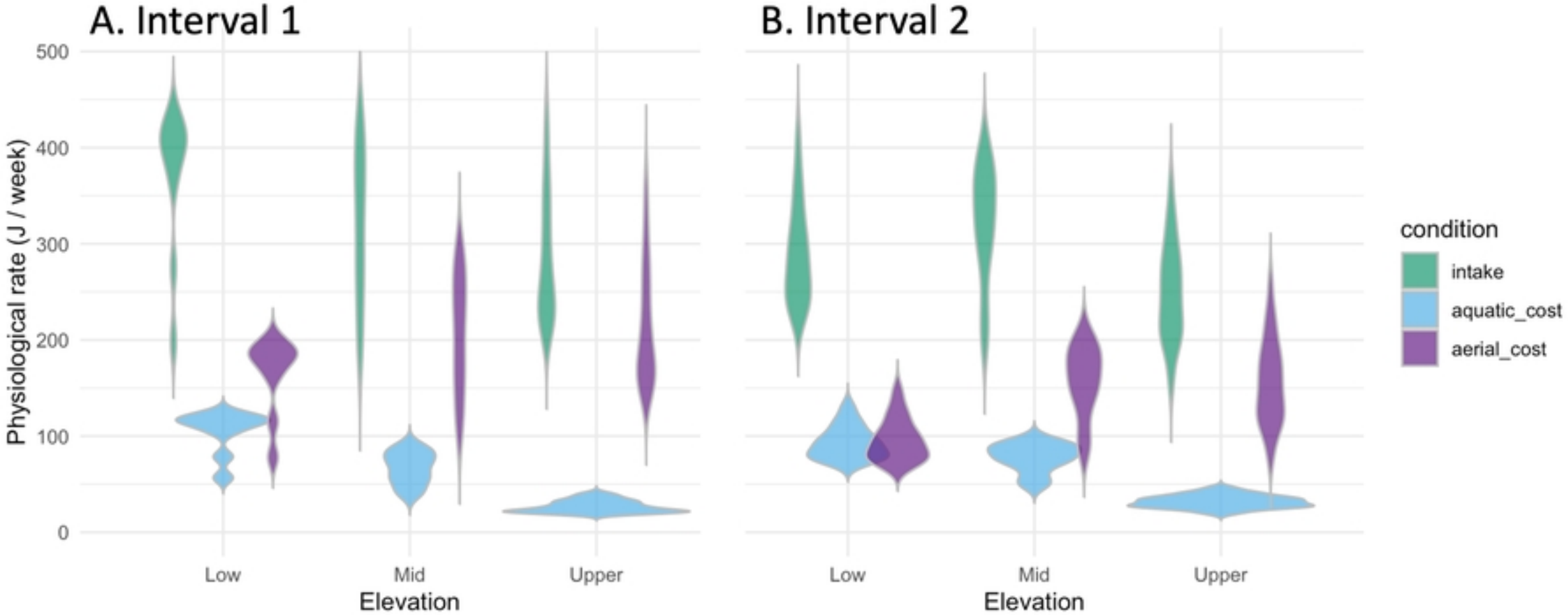


Fig7

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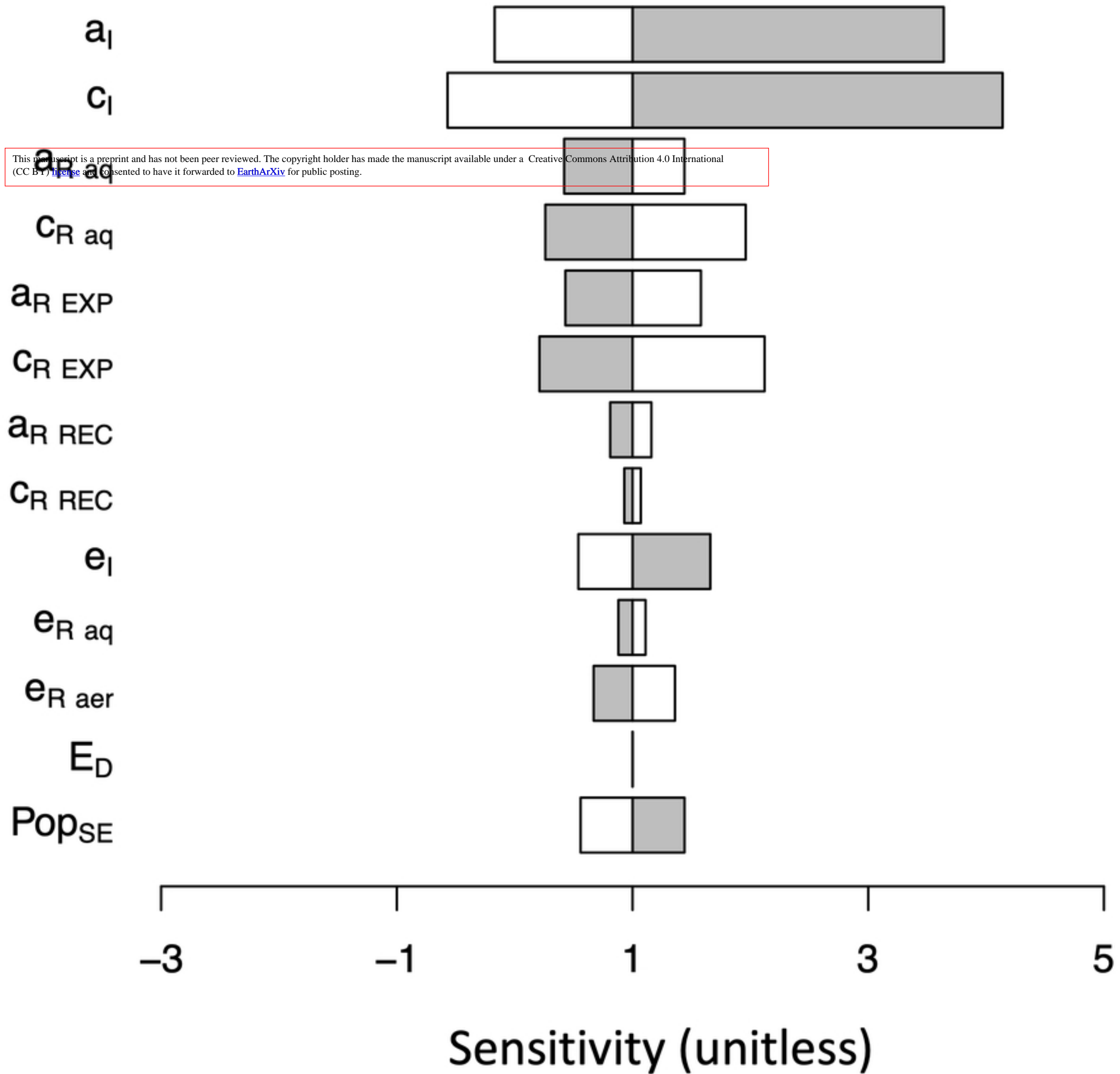


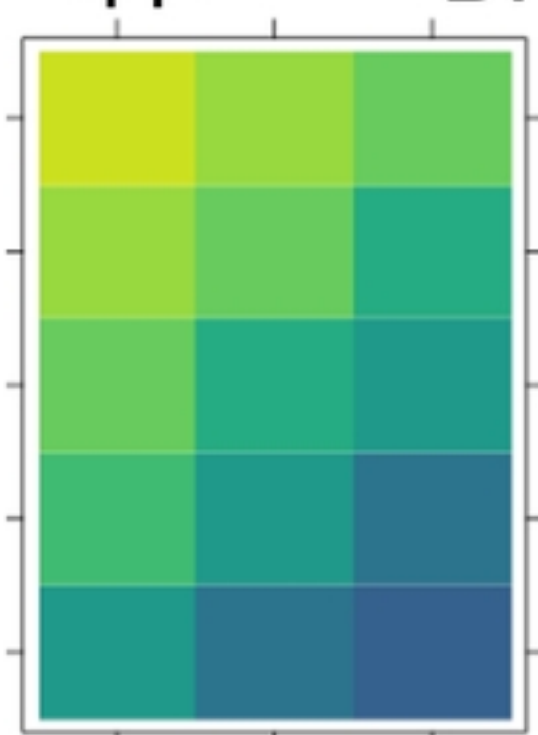
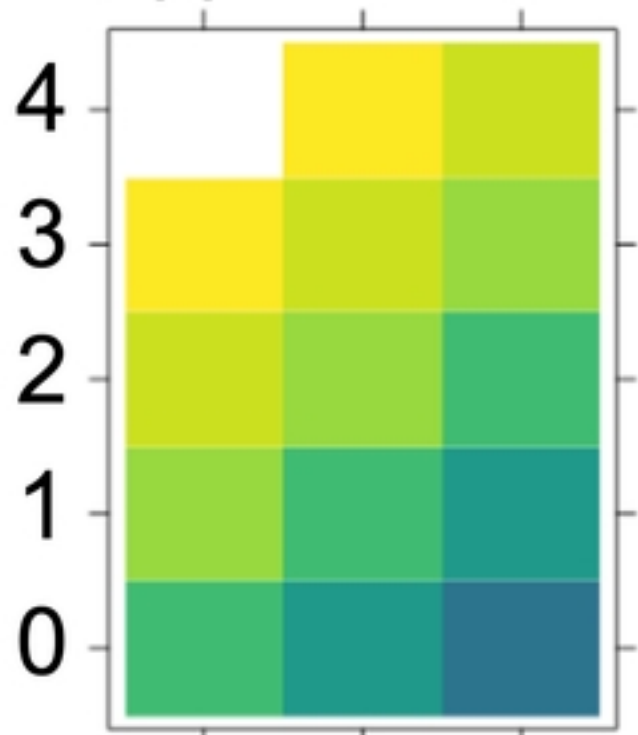
Fig8

Interval 1

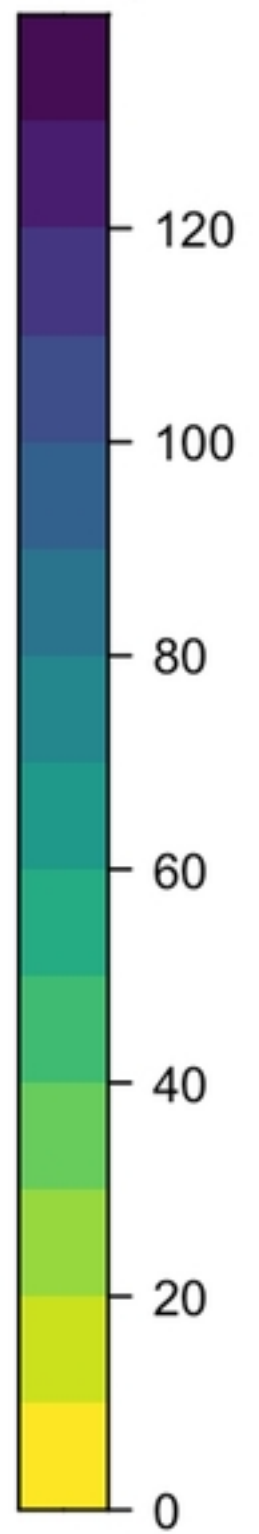
Interval 2

Upper A.

Upper B.



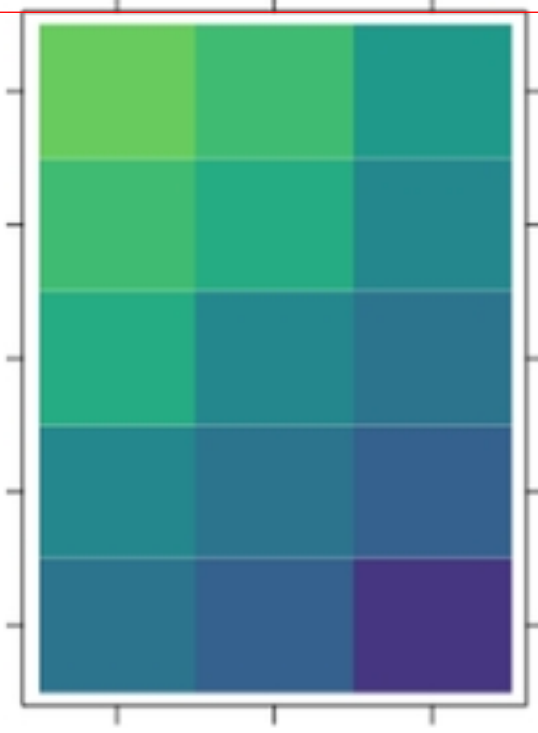
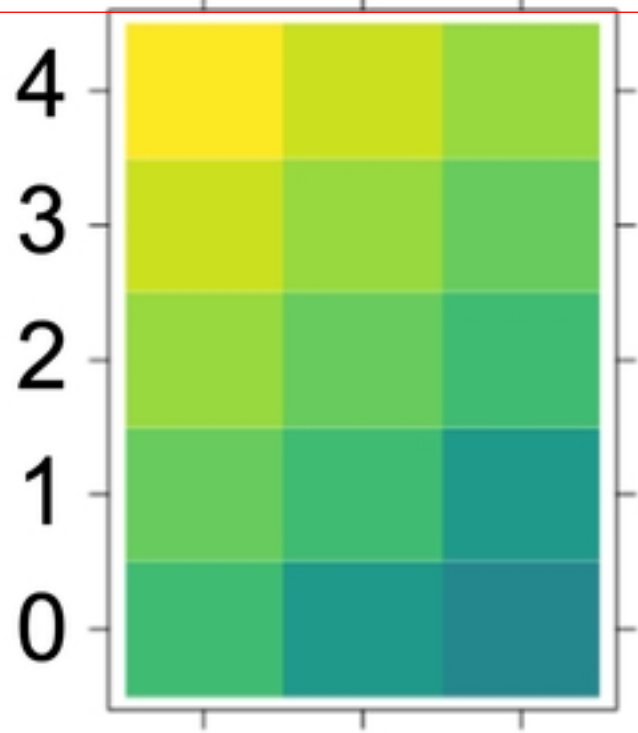
SFG (Joules)



Low tide temp ($\Delta^{\circ}\text{C}$)

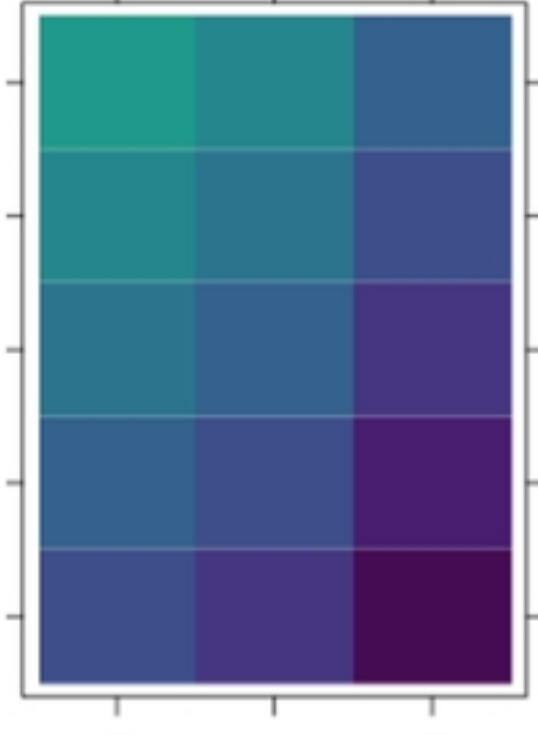
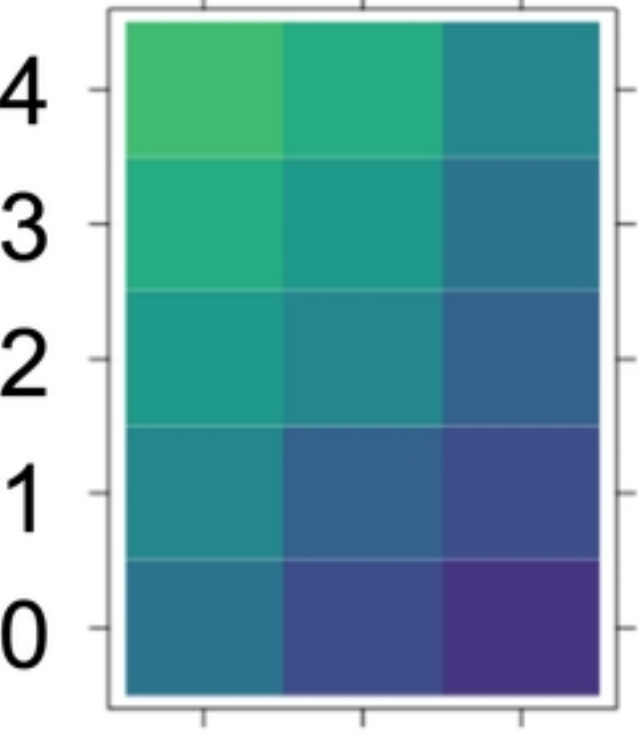
Mid C.

Mid D.



Low E.

Low F.



0 1 2

Seawater temp ($\Delta^{\circ}\text{C}$)

Fig9