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1 **Were Precambrian oceans devoid of planktonic cyanobacteria? Insights from metabolism**

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9

10 **Abstract**

11

12 Studies on biospheric evolution often assume planktonic cyanobacteria existed in Precambrian oceans,  
13 but that their productivity was limited due to various factors. However, available evidence suggests that  
14 planktonic cyanobacteria only colonized the open ocean near the Neoproterozoic-Phanerozoic boundary,  
15 close to the when a period of atmospheric oxygenation triggered the rise of animals. It is an open question  
16 if earlier planktonic marine cyanobacteria went extinct, possibly due to snowball Earth glaciations, or if  
17 cyanobacteria colonized the open ocean for the first time during this period. If the latter is true, consensus  
18 explanations are lacking for why cyanobacteria did not colonize the oceans sooner. Here I review the  
19 reconstructed metabolic evolution of marine picocyanobacteria, which perform ~25% of oceanic CO<sub>2</sub>-  
20 fixation and which emerged in the early Phanerozoic. These reconstructions provide insights into how the  
21 marine biosphere overcame earlier constraints on productivity, and how the rise of arthropods may have  
22 been key to allowing cyanobacteria to make the evolutionary leap from the benthos to the open ocean,  
23 helping transform the biosphere and Earth as a whole. The idea that photosynthesis was largely limited to  
24 shallow continental shelf waters during the Precambrian has implications for interpreting large-scale  
25 patterns of biospheric productivity and Earth history.

26

27 **Introduction**

28

29 The oxygenation of Earth's atmosphere resulted from an interplay of multiple biological and geological  
30 processes but was ultimately driven by the photosynthetic production of oxygen and the long-term  
31 sequestration of associated organic carbon in Earth's mantle, crust and oceans (1,2). Understanding large-  
32 scale patterns in biospheric productivity over Earth history (3) is key to reconstructing the rise of oxygen  
33 in Earth's atmosphere. Biospheric productivity is determined by the size and spatial extent of different  
34 ecosystems in which oxygenic photosynthesis occurs, and the rate of photosynthesis in those ecosystems.

35 Both aspects are challenging to precisely constrain in reconstructions of early Earth environments as they  
36 depend on inferences from geochemical proxies and fossil/biomarker records that become sparser and  
37 more ambiguous in deep time, especially for the deep open ocean (4-6). This in turn impacts molecular  
38 clock calculations of the evolution of major groups of photosynthetic organisms, which depend on  
39 external constraints from the geologic record (7-9). Together this leads to generally greater uncertainty  
40 regarding the scale of primary production when extrapolating further back in Earth history.

41  
42 Studies of metabolism can help address these uncertainties by suggesting linkages between the geologic  
43 and genomic records. Developing systematic frameworks of metabolic evolution identifies large-scale  
44 tradeoffs and forces that shape ecosystems and the biogeochemical cycles they mediate, informing  
45 interpretations of ecosystem change in deep time. I will illustrate these ideas using marine  
46 picocyanobacteria, consisting of the sister lineages *Prochlorococcus* (10) and marine *Synechococcus* (11).  
47 Marine picocyanobacteria perform ~20-25% of CO<sub>2</sub>-fixation in extant oceans (12,13) and their rise may  
48 have been linked to general biospheric revolutions in the early Phanerozoic (541 Mya – present) that  
49 ultimately led to the rise of macroscopic eukaryotic life (14-16). As background I will first review  
50 geologically-informed frameworks of the productivity of the marine biosphere during the Proterozoic  
51 (2500 Mya – 541 Mya), as well as evidence regarding the timelines of evolution of planktonic marine  
52 cyanobacteria in the open ocean. I will then review emerging frameworks for the metabolic evolution of  
53 marine picocyanobacteria and end by discussing their implications for large-scale patterns of biospheric  
54 productivity over Earth history.

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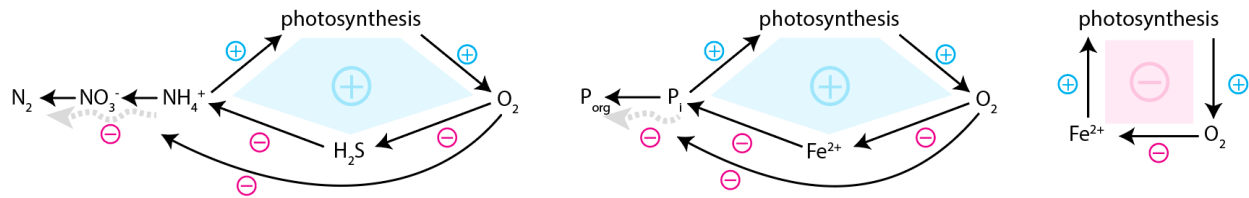
## 56 **Biospheric productivity in Proterozoic oceans**

57

58 Atmospheric oxygen underwent a dramatic rise in the early Proterozoic (17-19) and from here onward,  
59 but then remained low relative to modern levels until a second dramatic rise occurred between the late  
60 Proterozoic and early Phanerozoic (20-22). Various constraints on productivity of the Proterozoic marine  
61 biosphere have been proposed (Fig. 1), several of which involve the interplay between metals and the  
62 macronutrients nitrogen and phosphorus. For example, it has been suggested that molybdenum and other  
63 metals essential to the biological nitrogen cycle would have been scavenged by sulfide ions in sulfide-rich  
64 (i.e. euxinic) waters, leading to nitrogen-limitation of photosynthesis (23,24). Euxinic conditions were  
65 likely limited to relatively productive coastal waters, but models suggest that such environments could  
66 still have acted as an effective global sink of molybdenum due to large-scale ocean circulation processes  
67 (24). Similarly, it has been suggested that the high levels of ferrous iron (Fe<sup>2+</sup>) in the mostly anoxic

68 Proterozoic oceans (25,26) would have scavenged free phosphate, leading to phosphorus limitation of  
69 photosynthesis (27,28).

70



71

72 **Fig. 1. Proposed constraints on productivity of the ocean biosphere during the Proterozoic.** Blue positive signs  
73 are positive, stimulatory effects, while pink negative signs are negative, inhibitory effects. Availability of nutrients  
74 (nitrogen, phosphorus, iron) stimulates photosynthesis and thereby O<sub>2</sub> production. Nitrogen and phosphorus  
75 availability are proposed to have been limited due to chemical feedbacks (23,24,27,28), as well as due to differential  
76 rates within the biological cycles of both elements, the latter represented as dashed grey arrows (29,30). Increased  
77 O<sub>2</sub> levels would have helped eliminate these negative feedbacks, leading to overall self-amplifying cycles (blue  
78 shading) that may have been stuck in local minima. In contrast, iron availability is proposed to have *become* limiting  
79 under rising O<sub>2</sub> levels due to its decreasing solubility, leading to an overall self-damping cycle (pink shading). See  
80 text for further details. Note: phytoplankton can use multiple forms of nitrogen, but the universal entry point of  
81 nitrogen into metabolism for all life is ammonia. Abbreviations: P<sub>i</sub> = inorganic phosphorus (i.e. phosphate), P<sub>org</sub> =  
82 organic phosphorus

83

84 Other proposals focus on limitations arising due to differences in the rates of processes within the  
85 biological cycles of nitrogen and phosphorus under conditions of low oxygen (Fig. 1). That is, when  
86 oxygen is low, denitrification (i.e. conversion of NO<sub>3</sub><sup>-</sup> to N<sub>2</sub> gas) rates are predicted to outpace  
87 nitrification (i.e. conversion of NH<sub>4</sub><sup>+</sup> to NO<sub>3</sub><sup>-</sup>), preventing NO<sub>3</sub><sup>-</sup> from accumulating, thereby driving  
88 nitrogen-loss (i.e. outgassing of N<sub>2</sub> from the ocean) and nitrogen limitation (29). Similarly, when oxygen  
89 is low, remineralization of organic phosphate to inorganic phosphate is predicted to have been suppressed,  
90 driving or exacerbating phosphorus-limitation (30).

91

92 These proposals share two common properties. One is that a rise in oxygen toward Phanerozoic levels  
93 ultimately acts to remove preceding nutrient limitations (Fig. 1). That is, oxygenating the ocean removes  
94 both sulfides and ferrous iron, thereby eliminating the scavenging of molybdenum (31) and phosphorus  
95 (32), respectively. Simultaneously, a rise in oxygen levels increases the rate of phosphorus  
96 remineralization (30) and the rate of nitrification relative to denitrification, thereby counteracting nitrogen  
97 loss (29) (Fig. 1). Relative to Proterozoic sediments, Phanerozoic sediments show an increase in the levels  
98 of molybdenum (31) and phosphorus (28,32), as well as the onset of a stable oceanic nitrate pool (33,34),  
99 consistent with these proposals. However, it should be noted that most of the geochemical records used to

100 make inferences of ocean nutrient inventories are largely derived from shallow continental shelf  
101 environments (28,31-34), leading to inevitable uncertainties regarding the nutrient landscape of the open  
102 ocean. Still, molecular clocks suggest that nitrogen-fixing marine planktonic cyanobacteria did not  
103 expand into the open ocean until the late-Neoproterozoic (35), consistent with nutrient-poor (or at least  
104 nitrogen-poor) oceans before this time.

105  
106 The second common feature of these proposals is they generally assume that planktonic cyanobacteria  
107 were present in the open ocean, but with their productivity suppressed due to nutrient limitations.  
108 Together these features create the potential for non-linear dynamics in the oxygenation of the ocean and  
109 atmosphere. That is, increases in oxygen will be transient so long as the marine biosphere remains in the  
110 grip of outlined nutrient limitations, but can become self-sustaining if they are significant enough to  
111 begin loosening constraints on productivity of pre-existing planktonic cyanobacteria (Fig. 1) (36). This  
112 has led the argument that a large-scale redox perturbation due to snowball Earth played a key role in  
113 triggering the transition to an oxygenated ocean, effectively by freeing cyanobacteria from constraints on  
114 their productivity (36). Proposals that seek to explain the time-course of atmospheric oxygenation through  
115 long-term changes in geologic processes rather than through changes in constraints on biospheric  
116 productivity often similarly assume – either implicitly or explicitly – that cyanobacteria were present in  
117 the open ocean (37-40). However, the validity of the assumption that planktonic cyanobacteria existed in  
118 the open ocean in the Proterozoic is not clear.

119  
120 A different dynamic emerges in a proposal that colleagues and I put forward (41,42), in which  
121 productivity of Proterozoic oceans was limited at least in part due to a negative feedback built into the  
122 molecular machinery of photosynthesis itself (Fig. 1). That is, photosynthesis depends on iron, but iron is  
123 insoluble in the presence of oxygen. Thus, as oxygenation proceeds, iron becomes less available, lowering  
124 the production of oxygen, thus creating a self-damping dynamic that contrasts with the ultimately self-  
125 amplifying dynamics as in the proposals outlined above (Fig. 1). Indeed, this proposed dynamic is  
126 independent of the ocean inventories of nitrogen and phosphorus, suggesting that external perturbations  
127 (36) or long-term changes in geologic processes (37-40) may have been insufficient to drive oxygenation  
128 of the open ocean, and that biological innovations could have been necessary to unlock the full  
129 productivity potential of the ocean (42). If the latter view is correct, it in turn raises questions on the  
130 nature of those innovations, and why they did not, or could not, happen sooner.

131  
132 **Reconstructed timelines for the evolution of planktonic marine cyanobacteria**

133

134 Evaluating proposals for primary production in the open ocean in deep time requires considering the  
135 evidence for the existence of planktonic marine cyanobacteria throughout Earth history. Indirect evidence  
136 comes from the geologic carbon isotope record, which shows nearshore-to-offshore gradients in the <sup>13</sup>C-  
137 content of sedimentary carbonates that could be consistent with export of organic carbon from surface  
138 waters into sediments in the Proterozoic (43) and Archaean (44). However, observed gradients are weak,  
139 and conclusions regarding export processes thereby depend on assumptions of high levels of atmospheric  
140 CO<sub>2</sub> (43,44) needed to maintain liquid oceans on the early Earth due to lower solar luminosity (45).  
141 Precise levels of CO<sub>2</sub> relative to other potential greenhouse gases (particularly CH<sub>4</sub>) remain debated  
142 (46,47), leaving uncertainty about the magnitude of potential export processes. Moreover, offshore  
143 isotopic records are limited in number and typically come from depositional environment that extend only  
144 a few tens of kilometer beyond the shelf break (43,44). In modern oceans continental shelves and near-  
145 shore open ocean waters are connected through physical transport processes driven by tides, wind stress  
146 and general ocean circulation (48,49). This leaves open the possibility that observed carbon isotopic  
147 gradients could have been generated by primary production within continental shelves followed by lateral  
148 transport of organic carbon to deeper waters.

149  
150 Direct evidence specific to the evolution of cyanobacteria ultimately comes from two sources: fossils and  
151 biomarkers available in ancient rocks (50,51), and the record of molecular evolution available in genomes  
152 of extant cyanobacteria (8,9,14). The fossil record is often less definitive for cyanobacteria than for  
153 eukaryotes due to their smaller size and simpler structural morphologies (52,53). Targeted genome  
154 sequencing efforts (e.g. 54) and biomarker analyses (e.g. 55) have increased the number of structural  
155 morphologies in rocks tied to cyanobacterial taxa, but the numbers remain limited and essentially all are  
156 derived from shallow depositional environments and assigned to taxa not exclusively associated with a  
157 planktonic lifestyle (for detailed reviews, see Refs 8,50,56). Biomarkers in turn suggest that Proterozoic  
158 marine environments were dominated by cyanobacterial photosynthesis (51), but the assignment of these  
159 molecules to specific cyanobacterial taxa remains uncertain and samples containing them were obtained  
160 from shallow depositional settings (51). In summary, Proterozoic fossil/biomarker records offer no direct  
161 evidence of planktonic cyanobacteria in the deep open ocean (50,56).

162  
163 The genomic record, in turn, is clearer. Most molecular clock calculations suggest that ancestors of  
164 marine picocyanobacteria colonized the ocean in the late Neoproterozoic (1000 – 541 Mya) or early  
165 Phanerozoic (9,14,57-59), although origins in the early Neoproterozoic have also been found (60). In  
166 general, molecular clock calculations of divergence times and associated uncertainty ranges are strongly  
167 dependent on the external constraints that are used, and as outlined above, the number of constraints

168 available from the geologic record are very limited for cyanobacteria (8,50,56). To help address this gap,  
169 a recently emerging approach is to use horizontal gene transfer (HGT) events as an additional source of  
170 constraint on molecular clock calculations (61). That is, if the topologies of both the donor and recipient  
171 trees within the phylogenies of genes undergoing HGT are sufficiently similar to those of the species trees  
172 of both groups, and either group has an absolute date constraint, this constraint can be propagated across  
173 both groups (61). Using this approach leads to a calculated mean age of 424 Mya (range 491 – 340 Mya)  
174 for crown group marine picocyanobacteria, and a calculated mean age of 576 Mya (range 666 – 492 Mya)  
175 for ‘total group’ picocyanobacteria, the latter of which includes groups from freshwater and brackish  
176 environments (9). These ages are consistent with other recent results (57,62) but reduce calculated  
177 uncertainties by several hundred million years. In other words, molecular clock calculations suggest that  
178 the cyanobacterial branch leading to crown group marine picocyanobacteria evolved over a ~150-million-  
179 year period that came after the last Neoproterozoic snowball Earth (i.e. the Marinoan glaciation), which  
180 ended ~632 Mya (63).

181  
182 In summary, the genomic record suggests that marine picocyanobacteria most likely colonized the ocean  
183 in the early Phanerozoic, while the fossil and biomarker records offer no evidence of planktonic marine  
184 cyanobacteria before then. This leaves open two basic possibilities: 1) planktonic marine cyanobacteria  
185 *did* exist in the open ocean during the Precambrian, but went extinct, possibly due to snowball Earth  
186 glaciations, with evidence of their existence erased or still to be discovered, or 2) cyanobacteria colonized  
187 the open ocean for the first time in the early Phanerozoic. If the latter is true, why did cyanobacteria not  
188 colonize the ocean sooner? For insights into these questions, I will next look at evidence from metabolic  
189 reconstructions.

190

### 191 **Metabolic evolution of marine picocyanobacteria**

192

193 The metabolic makeup of a cell reflects how it transforms environmental inputs to make additional copies  
194 of itself. Metabolic pathways are governed by various ecological tradeoffs, for example due to differences  
195 in their energetic or protein biomass requirements (64), the metals needed in enzyme cofactors (65), the  
196 nature and redox state of the inorganic nutrients that cells assimilate (66), or the relative amounts of  
197 carbon, nitrogen, or other elements, that are generated in the case of catabolic pathways. Metabolic  
198 innovations that occur over microbial evolution thus reflect changes in the ecosystems they are part of,  
199 whether due to changes or perturbations in the Earth system or due to the colonization of new  
200 environments. Relating metabolic diversity to Earth history requires developing systematic frameworks of  
201 metabolic evolution. Colleagues and I have been pursuing this through the integration of phylogenetic and

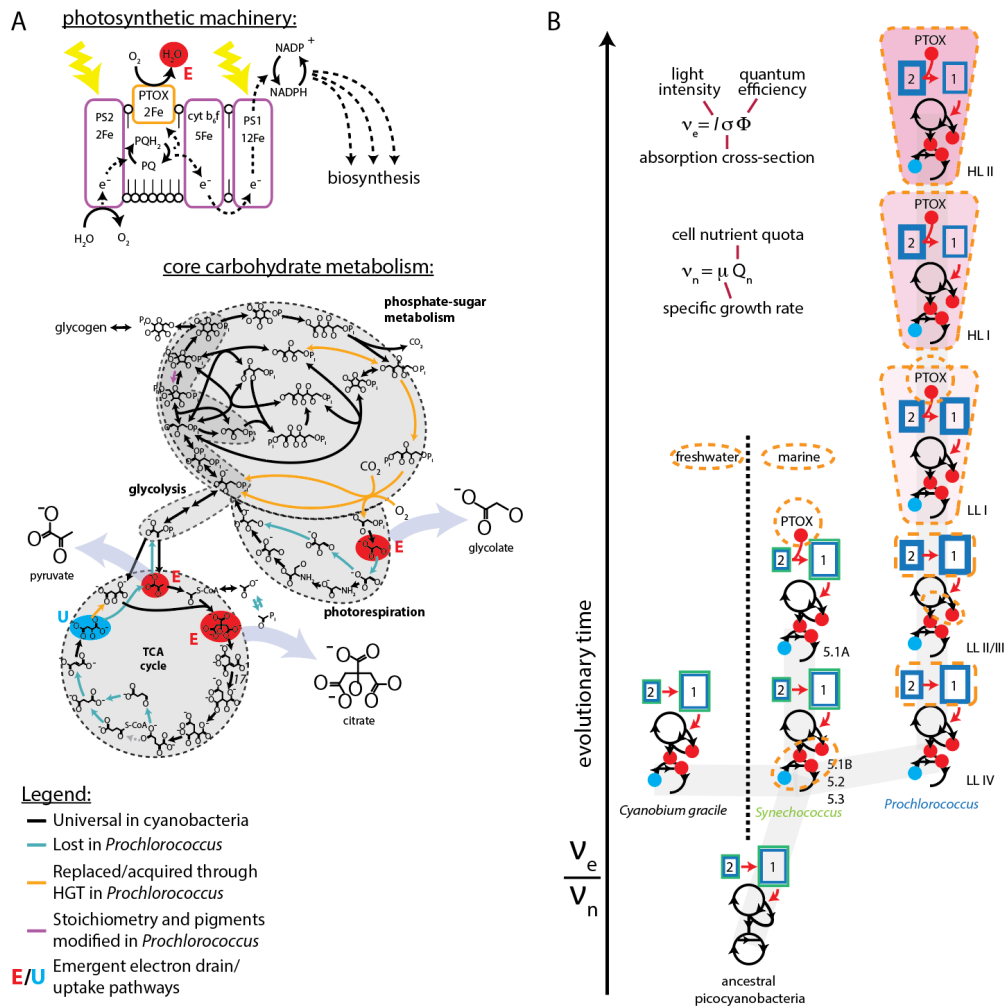
202 metabolic network reconstructions (41,67). This allows us to pinpoint specific innovations that occur  
203 along major divergences within microbial phylogenies and infer the underlying tradeoffs and driving  
204 forces acting on them. Identifying these tradeoffs and evolutionary driving forces at major branches in  
205 microbial phylogenies offers opportunities for linking the timeline of these branches as calculated using  
206 molecular clocks with records of ecosystem change observed in the rock record.

207

208 We have taken this approach to reconstruct the metabolic evolution of marine picocyanobacteria, starting  
209 with the metabolic core, i.e. the pathways involved in photosynthesis, electron transfer and carbon-  
210 fixation (Fig. 2A). The metabolic core is where light energy and CO<sub>2</sub> are combined to generate the carbon  
211 backbones of all biomolecules making up the cell. Because everything else depends on it, the evolution of  
212 the metabolic core is highly constrained and any changes that do occur reflect strong selective pressures  
213 (68). In our reconstructions, we found that the metabolic core of marine picocyanobacteria underwent a  
214 large-scale remodeling as they diverged from other ancestral cyanobacteria (Fig. 2) (41). That is, many  
215 genes involved in core carbohydrate metabolism or photosynthesis were lost, gained or replaced through  
216 horizontal gene transfer, while the pigments and stoichiometry of the photosynthetic machinery were also  
217 modified (Fig. 2). Metabolic changes include the addition of several pathways for the exudation of  
218 intermediates of core carbohydrate metabolism, suggesting that selection has favored an increase in  
219 organic carbon exudation of cells (41). This whole-sale reorganization of core pathways indicates that the  
220 metabolism of marine picocyanobacteria came under intense selection as they colonized the open ocean,  
221 reflecting the unique challenges this environment presents.

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**Fig. 2. Reconstructed metabolic evolution of marine picocyanobacteria.** Left panel: Remodeling of the metabolic core, with genes lost highlighted in green, genes gained in orange, changes to photosystems/pigments in purple, and universal genes in black. The addition of pathways for exuding organic carbon (or water, in the case of the electron transport chain) are highlighted in red and a putative pathway for taking up organic carbon highlighted in blue. Right panel: metabolic innovations in lineages of *Synechococcus* and *Prochlorococcus* are highlighted with dashed circles and occur along an axis defined by the relative supplies of electrons and nutrients (i.e.  $v_e/v_n$ ). Little black diagrams represent core carbohydrate metabolism with direction of carbon flow indicated by arrows, green/blue boxed 1's and 2's represent photosystems I and II, and red arrows represent electron flux. Increasing pink shading high-light adapted *Prochlorococcus* ecotypes represents acquisition of UV/light stress genes that allow cells to withstand higher levels of light, thereby generating an increased photosynthetic electron flux. Figure is adapted from Refs (40,41).

238 A deeper understanding of the selective forces driving the metabolic innovations of marine  
239 picocyanobacteria comes from examining them in their physiological and ecological context. That is, the  
240 metabolic remodeling is most extensive in *Prochlorococcus*, populations of which undergo a process of  
241 depth-based niche partitioning in warm oligotrophic waters, with more recently diverging “high-light  
242 (HL) adapted” clades dominating at the surface and deeper-branching “low-light (LL) adapted” clades  
243 dominating at the bottom of the euphotic zone (69,70). The resulting higher photon fluxes received by HL  
244 adapted cells are accompanied by modifications in the stoichiometries and pigments of photosystems (71-  
245 73) that act to increase the cellular absorption cross-section (41), together driving an increase in the  
246 achievable photosynthetic electron flux ( $v_e$ ).

247  
248 This evolutionary increase in the photosynthetic electron flux ( $v_e$ ) in more recently diverging  
249 *Prochlorococcus* cells coincides with changes in their usage of inorganic nutrients. That is,  
250 *Prochlorococcus* cells are smaller and grow slower than *Synechococcus* cells (10,74), lowering the uptake  
251 flux of all resources required for growth. This is accompanied by a process of evolutionary streamlining,  
252 in which the membrane (75), genome (76), proteome (77), transcriptome (78) and metabolism (41) are  
253 modified in ways that lower the cellular contents of phosphorus, nitrogen and iron, the three main  
254 inorganic nutrients limiting phytoplankton growth in the ocean (79). Collectively these changes are  
255 inferred to drive a decrease in the assimilation flux ( $v_n$ ) of limiting nutrients, and thus an increase in the  
256 electron-to-nutrient flux ratio of cells ( $v_e/v_n$ ) over the course of evolution in marine picocyanobacteria  
257 (Fig. 2B). The addition of pathways for exudation of organic carbon in the metabolic core thus reflects a  
258 growing excess flux of electrons and fixed carbon that is released from picocyanobacterial cells into the  
259 environment.

260  
261 Selection for increases in  $v_e/v_n$  and associated removal of excess electrons through exudation of organic  
262 carbon can in turn be understood as follows. First, the increase in  $v_e/v_n$  has progressed the furthest in  
263 *Prochlorococcus*, which dominates in the most nutrient-poor regions of the open ocean (12,69),  
264 suggesting a link between metabolic evolution and nutrient assimilation. Second, photosynthetic electron  
265 flux ( $v_e$ ) carries solar energy into metabolism, suggesting metabolic evolution has increased the energy  
266 flux density, or metabolic power, of cells. Third, the free energy cost of nutrient assimilation scales with  
267 the nutrient gradient across cellular membranes (i.e.  $\Delta G_r \propto \ln ([n]_{in}/[n]_{out})$  for nutrient  $n$ ). Taken  
268 together this led us to conclude that the reconstructed evolutionary increase in  $v_e/v_n$  in marine  
269 picocyanobacteria reflects a necessary increase in metabolic power of cells to allow growth at decreasing

270 nutrient concentrations, and that added pathways for exuding organic carbon reflects the inevitable  
271 “carbon exhaust” of nutrient assimilation (41,42).

272

273 The above view of metabolic evolution in marine picocyanobacteria led us to propose a “depth-  
274 displacement” model for the evolutionary niche partitioning of *Prochlorococcus* clades. In this model,  
275 new innovations that increase  $v_e/v_n$  and allow cells to grow at lower nutrient concentrations draw  
276 nutrients down near the surface, restricting cells from deeper branching clades to regions deeper in the  
277 water column (41). This view of *Prochlorococcus* macroevolution is consistent with the distribution of  
278 nitrate assimilation genes in *Prochlorococcus* (80). That is, nitrate assimilation genes are absent in most  
279 deeply branching clades of *Prochlorococcus* but are retained in recently diverging HL-adapted clades,  
280 where genomic analyses suggest they have been inherited through a process of vertical descent that  
281 reaches back to crown group *Prochlorococcus*. Further, the frequency of nitrate assimilation genes in HL-  
282 adapted clades is enhanced under conditions of nitrogen-limitation (81). Together this suggests that  
283 conditions of nitrogen limitation in surface waters provided the selective pressure to retain nitrate  
284 assimilation genes in surface-dwelling clades, but that these genes were lost in deeper-branching clades as  
285 they were displaced further down the water column where general availability of nitrogen from other  
286 sources increases (80). This consistent with the depth-displacement model of *Prochlorococcus* evolution  
287 as emerged from metabolic reconstructions (41).

288

289 The emerging framework for the metabolic evolution of marine picocyanobacteria has implications for  
290 understanding how the marine biosphere overcame proposed constraints on productivity during the  
291 Proterozoic (Fig. 1). First, by lowering the nutrient concentration at which cells can grow, the metabolic  
292 innovations of marine picocyanobacteria increase their potential for expanding into the ocean in the face  
293 of proposed pre-existing nutrient limitation resulting from abiotic chemical processes in ancient oceans  
294 (23,24,27,28), as well as from ecological process (29,30) in scenarios where other planktonic  
295 cyanobacteria did exist in the open ocean before the rise of marine picocyanobacteria. By increasing  
296 ocean productivity, the global ecological expansion of marine picocyanobacteria would have helped drive  
297 oxygenation of the ocean, thereby loosening nutrient controls on productivity, helping to consolidate the  
298 transition toward an oxygenated state (Fig. 1). Although molecular clocks independently suggest that the  
299 flux of nitrogen fixation into the ocean increased in the late Neoproterozoic (35), it remains possible that  
300 ocean nutrient inventories of the deep open ocean were higher than is estimated from geochemical proxies  
301 largely derived from shallow-water sediments (27,28,31-34). In that scenario, the innovations of marine  
302 picocyanobacteria could in part reflect an adaptation to the most nutrient-poor parts of the open ocean.

303

304 Second, the metabolic evolution of marine picocyanobacteria may have also helped counteract the  
305 negative feedback on productivity involving iron (Fig. 1C). That is, in today's oxygenated ocean iron is  
306 insoluble but is made bioavailable through complexation with organic carbon (82,83). The products of the  
307 exudation pathways added to the metabolic core of marine picocyanobacteria (Fig. 2) belong to the broad  
308 class of "weak" ligands that are a major source of iron in the ocean (82,83). Thus, as marine  
309 picocyanobacteria expanded into the open ocean, they would have helped gradually increase the inventory  
310 of iron ligands, thereby helping counteract the decreased availability of iron for photosynthesis due to the  
311 diminishing solubility of iron in oxygenated waters. Indeed, modeling results suggest that ocean microbes  
312 collectively tune the total inventory of accessible iron through similar feedbacks, such that it maximizes  
313 ocean productivity (84). Importantly, this interplay of positive and negative feedbacks in the availability  
314 of iron for photosynthesis is independent of the ocean inventories of nitrogen and phosphorus and is thus  
315 consistent with the late emergence of planktonic marine cyanobacteria even if there was sufficient  
316 nitrogen and phosphorus available in earlier times.

317

#### 318 **Emergence of a planktonic lifestyle in marine picocyanobacteria: the "chitin raft hypothesis"**

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320 While the reconstructed evolution of core metabolism in marine picocyanobacteria provides insight into  
321 how the marine biosphere overcame earlier constraints on productivity (Fig. 1), it does not address the  
322 questions of why these innovations happened when they did, or why cyanobacteria may not have  
323 colonized the open ocean sooner. Here another metabolic trait that we recently discovered in marine  
324 picocyanobacteria provides relevant context: chitin degradation (85). Chitin is a carbon polymer that is  
325 highly abundant in the ocean and is largely derived from the exoskeletons of arthropods (86). The  
326 existence of genes for chitin degradation in marine picocyanobacteria is unexpected, as chitin exists  
327 mostly as particles, and marine picocyanobacteria are generally considered to live a planktonic lifestyle  
328 (14,87). However, we showed that marine *Synechococcus* and deep-branching *Prochlorococcus* attach to  
329 chitin particles and display enzymatic activity of chitinase, the main extracellular enzyme involved in  
330 cleaving chitin polymers into smaller units that cells can take up (85). We also showed that expression of  
331 chitin degradation genes and other attachment-associated genes increases when cells are exposed to  
332 chitosan, a solubilized form of chitin, and that *Prochlorococcus* obtains a growth boost from exposure to  
333 chitin when grown at low light levels. These results demonstrate that chitin degradation is an active  
334 functional trait in marine picocyanobacteria, changing our view of their ecology.

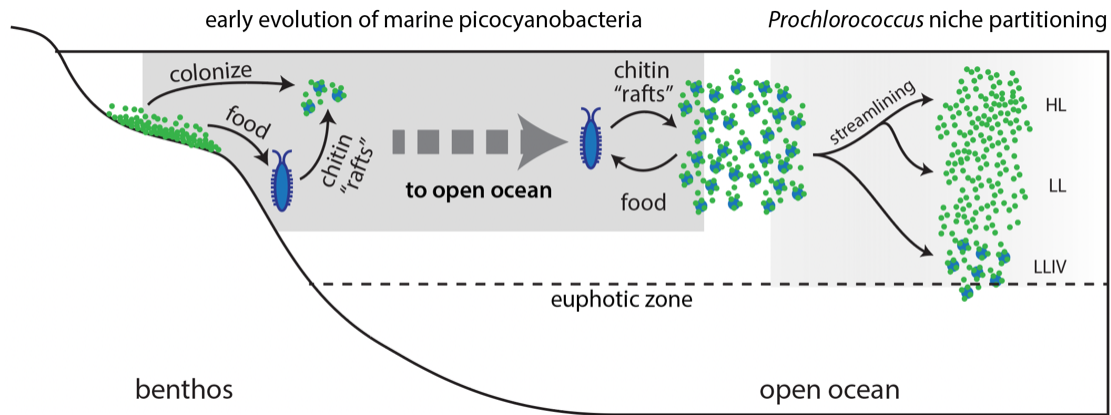
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336 The existence of chitin attachment and degradation capabilities in marine picocyanobacteria informs  
337 macroevolutionary frameworks. First, combined phylogenetic and phylogenomic analyses indicate genes

338 necessary for chitin degradation were acquired through horizontal gene transfer (likely from unspecified  
339 planctomycetes) along the branch leading to crown group marine picocyanobacteria, and that the trait was  
340 lost during the evolution of *Prochlorococcus*, but remains universally preserved in its deepest branches  
341 and in marine *Synechococcus* (85). As mentioned above, the branch leading from other picocyanobacteria  
342 to crown group marine picocyanobacteria evolved over the period of ~576 – 424 Mya (9), which overlaps  
343 with the period during which both fossils and molecular clocks indicate that marine arthropods underwent a  
344 major ecological expansion (88-91). This suggests that the early evolution of marine arthropods and  
345 marine picocyanobacteria was intertwined and may have played a role in the inferred oxygenation of  
346 shallow continental shelf waters in the late Neoproterozoic that is inferred to have preceded eventual  
347 oxygenation of the deep ocean in the early Phanerozoic (21,22).

348

349 To further understand the relevance of the acquisition of the chitin attachment and degradation trait to the  
350 ecological rise of marine picocyanobacteria, it is instructive to consider microbial lifestyle in general.  
351 Global surveys suggest that outside of lakes and oceans, microbial biomass exists predominantly within  
352 aggregates attached to surfaces due to the collective stress buffering and nutrient recycling conditions  
353 such communities create (92). Most microbes, including cyanobacteria, can facultatively switch between  
354 planktonic and aggregated states, and while there are no single universal stressors driving aggregation  
355 across all microbial systems, common triggers include salinity, UV radiation and nutrient stress (92-96),  
356 all of which are elevated in surface waters of the open ocean. These stresses were moreover likely  
357 exacerbated in the Precambrian, when UV levels were higher due to a less-developed ozone layer (97),  
358 and oceanic nutrient inventories may have been lower (23,24,27-35). How then, did planktonic  
359 cyanobacteria colonize the open ocean in the face of stresses that favor aggregation? In response to this  
360 question, we proposed the “chitin raft hypothesis” (Fig. 4). In this hypothesis (85), the exoskeleton molts  
361 that arthropods produce as a byproduct of their growth provided surfaces where picocyanobacteria could  
362 aggregate and find shelter from environmental stresses, while picocyanobacteria in turn provided a food  
363 source for arthropods. Over millions of years some *Prochlorococcus* lineages evolved the necessary  
364 adaptations that eventually allowed them to make a secondary transition of fully planktonic lifestyle.  
365



366

367 **Fig. 3. Chitin raft hypothesis.** Colonization of the open ocean by marine picocyanobacteria is proposed to have a  
 368 collective, ecosystem-level process involving two stages. In the first stage, picocyanobacteria colonized chitin  
 369 particles resulting from the waste of co-evolving arthropods, allowing them to maintain their facultatively surface-  
 370 aggregated lifestyle and find protection from environmental stresses, while in turn providing arthropods a source of  
 371 food. This association allowed both groups to expand out into the deep open ocean. Later, after acquiring  
 372 adaptations to better withstand the stresses of life in the open ocean, some lineages of *Prochlorococcus* made a  
 373 secondary evolutionary transition to a committed planktonic lifestyle. Figure is adapted from Ref (85).

374

375 The chitin raft hypothesis for is consistent with several additional lines of evidence in marine  
 376 picocyanobacteria. First, a growing number of associations of marine picocyanobacteria to particles  
 377 and/or protist hosts have been identified in recent years (98), including a dinoflagellate symbiont that  
 378 branches between *Prochlorococcus* and *Synechococcus* (99). This suggests surface attachments play a  
 379 more significant role in the ecology and evolution of marine picocyanobacteria than historically  
 380 recognized. A second line of evidence comes from the adaptations in extant lineages, including  
 381 acquisition of genes for compatible solutes that protect against osmotic stress from elevated salinity  
 382 (100,101) and for protection from UV and light stress (102,103), as well as cellular and macromolecular  
 383 changes that lower nutrient requirements while raising nutrient affinity (41,75-78), as outlined above. By  
 384 helping to adapt to the unique challenges of microbial life in the open ocean, these adaptations also  
 385 mitigate against the stresses that are generally linked to cellular aggregation and surface attachment.  
 386 Finally, a recent study found that the region in the *Prochlorococcus* tree that separates clades with chitin  
 387 metabolism from those without it contains unique photophysiology and is linked to a massive expansion  
 388 of photosystem genes (104). This is consistent with the photosynthetic machinery of *Prochlorococcus*  
 389 coming under intense selection during the evolutionary transition to a fully planktonic lifestyle.

390

391 Given the general lack of evidence of planktonic cyanobacteria before the rise of marine  
 392 picocyanobacteria, we may posit that the associations with arthropods was a necessary precondition for

393 the emergence of planktonic photosynthesis in the open ocean. Indeed, this proposal highlights how  
394 lineages do not evolve in isolation, but rather in the context of their ecological interdependencies: marine  
395 picocyanobacteria depend on the wasteful byproducts (i.e. exoskeleton molts) of arthropod growth, while  
396 arthropods in turn depend on a steady supply of food from picocyanobacteria and other particle-attached  
397 microbes. In this view, oxygenic photosynthesis could not emerge in the deep open ocean until  
398 ecosystems were complex enough such that could sustain internal

399

#### 400 **Implications for biospheric productivity and Earth history**

401

402 While the chitin raft hypothesis helps resolve questions regarding both the absence of evidence for  
403 planktonic marine cyanobacteria before the rise of marine picocyanobacteria, as well as the diversity and  
404 distribution of traits within the latter, others remain unresolved. For example, planktonic nitrogen-fixing  
405 cyanobacteria also colonized the ocean sometime in the late Neoproterozoic or early Phanerozoic, and the  
406 evolution of their lifestyle has not been fully unraveled. Many marine nitrogen-fixing cyanobacteria exist  
407 as part of buoyant colonies (e.g. *Trichodesmium*) (105) or in associations with eukaryotic phytoplankton  
408 (e.g. *Richellia* or UCYN-A) (106,107), which helps address the question of how they colonized the ocean  
409 in the face of aggregation-inducing stresses. Other groups, including *Crocospaera watsonii* (108,109),  
410 have a unicellular planktonic lifestyle, but it is not yet clear if they have unrecognized abilities to switch  
411 between planktonic and attached lifestyles as we recently discovered in marine picocyanobacteria. Further  
412 studies on the diversity of lifestyles and lifestyle-associated traits among nitrogen-fixing marine  
413 cyanobacteria, and when in time different groups diverged from each other as calculated by molecular  
414 clocks will test, or at least provide nuance, to the chitin raft hypothesis.

415

416 In terms of the geologic record, additional targeted searches for evidence of planktonic cells in  
417 Proterozoic sediments, either from fossils, particularly of small cells (50), or from biomarkers (51), and  
418 from any deep-water depositional environments, would provide important tests that could invalidate our  
419 proposal. Further examination of nearshore-to-offshore gradients in carbon isotopes can in turn provide  
420 indirect evidence of the presence and/or productivity of potential planktonic cyanobacteria in open ocean  
421 waters. If high CO<sub>2</sub> levels muted such signals in the Archaean and early Proterozoic (43,44), then one  
422 might expect such gradients to become stronger in the later Proterozoic when CO<sub>2</sub> levels are expected to  
423 have been somewhat lower (45-47), as is potentially seen in some late-Mesoproterozoic sediments (110).  
424 Increasing the number of available records to improve statistics and extending included samples to  
425 include further-offshore environments less likely to be coupled to continental shelf environments through

426 physical transport processes will help understand whether there was significant organic carbon production  
427 in deep open oceans, potentially indicative of planktonic cyanobacteria.

428

429 If the chitin raft hypothesis holds up in the face of such added scrutiny, it can potentially help inform  
430 interpretations of larger patterns of biospheric productivity and atmospheric oxygenation in deep time.

431 That is, if oxygenic photosynthesis was largely restricted to shallow continental shelf environments

432 during the Precambrian, then the total productivity of the marine biosphere was limited by the total global  
433 expanse of shelf environments and the rate of nutrient supply to those environments (Fig. 4). Hence, super

434 continental cycles (111) could have had a role in modulating productivity during the Proterozoic (112), by  
435 impacting both the total size of shelf environments and the weathering supply of nutrients through

436 assembly and break-up processes (113-116). Furthermore, it has become increasingly clear from both  
437 geochemical studies and geophysical modeling results that there was a major increase in sub-aerial

438 continental surface area between the late Archaean and early Proterozoic (38,117-119). In a world of

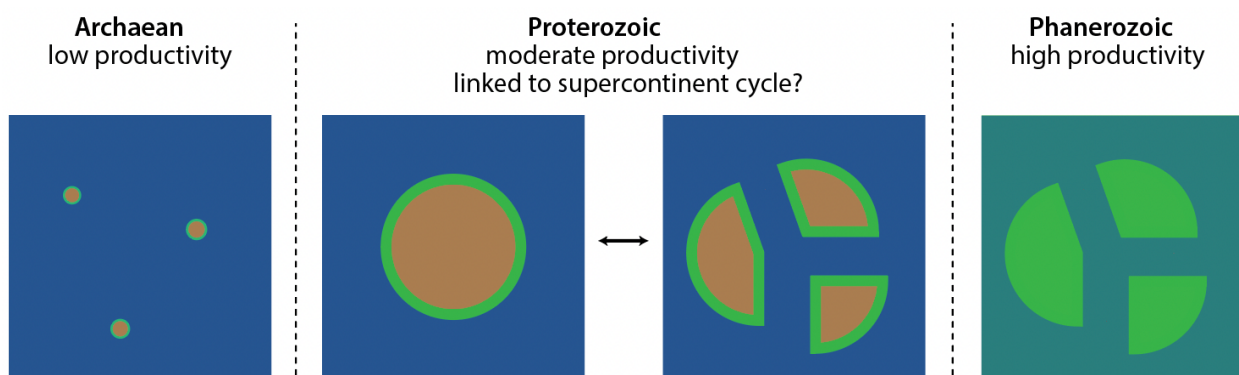
439 continental shelf-bound oxygenic photosynthesis, the transition from a ‘water world’ to a world with large  
440 continents would have led to a major expansion of shelf environments and thus of primary production,

441 possibly helping fuel the great oxidation event. These examples highlight how metabolic reconstructions  
442 of microbial evolution can provide mechanistic hypotheses for interpreting large-scale patterns of Earth

443 history.

444

445



446

447 **Fig. 4. Proposed model for the two-stage evolution of biospheric productivity over Earth history.** Shelf-bound

448 primary production during the Precambrian drove a first increase in productivity with the emergence of continents

449 near the Archaean-Proterozoic boundary. Colonization of the oceans by marine picocyanobacteria and eukaryotic

450 algae and colonization of the continents by land plants drove a second increase in productivity. Notes: green

451 circumferences of brown landmasses represent continental shelf environments where productivity was localized

452 during the Precambrian. Green oceans and continents in the Phanerozoic represent the global spread of chlorophyll

453 and thus oxygenic photosynthesis.



454

455

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457

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463

464

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