1 Mercury stable isotope composition of seawater suggests important net

2 gaseous elemental mercury uptake

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14 Summary

15 Human exposure to toxic mercury (Hg) is dominated by the consumption of seafood^{1,2}. Earth system models suggest that Hg in marine ecosystems is supplied by Hg(II) deposition, with a 3x smaller 16 17 contribution from gaseous Hg(0) uptake, and that photochemical reduction of marine Hg(II) drives 18 important Hg(0) evasion to the atmosphere^{3,4}. Observations of marine Hg(II) deposition and gas exchange are sparse however⁵, leaving the suggested importance of air-sea exchange⁶ unconstrained. 19 20 Here we present the first Hg stable isotope measurements of total Hg (tHg) in surface and deep Atlantic 21 and Mediterranean seawater. We use an isotope mass balance to estimate that sea water tHg can be 22 explained by the mixing of 41% atmospheric Hg(II) deposition and 59% Hg(0) uptake. In the particulate 23 Hg (pHg) fraction, which includes phytoplankton at the base of the marine food web, and in a 24 compilation of marine fish Hg isotope data, we estimate similarly important marine Hg(0) uptake 25 fractions of 73% and 49%. We observe no photochemical odd Hg isotope anomalies in tHg, which calls 26 into question the large model Hg(0) evasion flux. Our findings indicate that direct atmospheric Hg(0) 27 uptake is important and has implications for our understanding of atmospheric Hg dispersal and 28 marine ecosystem recovery.

29 Main text

30 The consumption of seafood exposes humans to mono-methylmercury (MMHg), a toxin known to 31 affect fetal and infant neurodevelopment and cardiovascular disease in adults ^{1,2,7}. MMHg is formed from inorganic mercury (Hg) in the ocean and biomagnifies along marine food webs to high levels^{8–10}. 32 33 Anthropogenic release of Hg to the atmosphere, land and water bodies outweighs natural release at least five-fold^{11,12}, and is thought to have more than tripled the mercury content of surface ocean 34 35 waters¹³. Climate change and overfishing also affect fish MMHg levels, and require detailed models of 36 Hg cycling to assess future human exposure to Hg¹⁴. Earth system models of Hg biogeochemical cycling suggest that atmospheric Hg deposition to the open ocean is dominated by Hg(II) wet deposition (4600 37 Mg y⁻¹) compared to dissolution of gaseous Hg(0) (1700 Mg y⁻¹) in a 3:1 ratio^{3,4}. Rivers deliver similar 38 39 amounts of Hg to the oceans as atmospheric deposition (5500 Mg y⁻¹), while only 6% of river Hg is estimated to reach the open ocean¹⁵. Models also estimate that, of total deposited Hg, 4600 Mg y⁻¹ is 40 re-emitted to the atmosphere by (photo-)chemical and microbial photoreduction of Hg(II) to $Hg(0)^{3,16}$. 41 42 Dissolved Hg(0) concentrations in surface waters and atmospheric Hg(0) concentrations are sporadically measured simultaneously during oceanic cruises¹⁷, and show variable Hg(0) exchange 43 fluxes over short time periods and across different ocean basins¹⁸. The direction and magnitude of the 44 large air-sea exchange of Hg(0) depends on the parametrization of the Hg(0) exchange velocity in 45 models¹⁹. The absence of direct, long-term Hg(0) flux measurements over the ocean and limited 46

observations of Hg(II) wet and dry deposition to oceans therefore provide insufficient constraints for
model validation of sea-air Hg(0) exchange, and Hg(II) deposition fluxes. This in turn hampers our
capability to predict how Hg levels in the Ocean will respond to curbed anthropogenic Hg emissions
under the UN Minamata Convention on Mercury, and to climate change^{20–22}.

51 In terrestrial ecosystems, Hg stable isotopes have proven to be instrumental in understanding the relative importance of Hg(II) and Hg(0) deposition^{23–25}. Atmospheric Hg(0) and Hg(II) in rainfall have 52 different mass-dependent (MDF, δ^{202} Hg) and mas-independent fractionation (MIF, Δ^{199} Hg and Δ^{200} Hg) 53 signatures. Even-Hg MIF (Δ^{200} Hg) is thought to be generated exclusively by upper tropospheric 54 photochemical reactions^{26,27}, and no Hg transformations at the Earth's surface have been shown to 55 fractionate Δ^{200} Hg. Δ^{200} Hg is therefore considered as a conservative tracer for atmospheric Hg 56 deposition pathways²⁴. Apart from marine biota, the Hg stable isotope composition of total unfiltered 57 58 Hg (tHg) has only been measured in coastal seawater of the Canadian Arctic Archipelago, suggesting 59 that 50% to 80% of tHg originated from coastal erosion and river input²⁸. Analysis of particulate Hg (pHg) isotopes at the tropical Pacific station ALOHA (22°N) suggested that atmospheric deposition of 60 Hg(II) from rainfall was an important Hg source to the surface ocean²⁹. The Hg isotopic composition of 61 62 open ocean seawater, characterized by its (sub-)picomolar levels of tHg in all basins³⁰, including oceanlike seas such as the Mediterranean Sea³¹, remains unexplored. 63

In this contribution we develop and apply an ultra-clean pre-concentration method for tHg isotope analysis of seawater to understand air-sea exchange of Hg. We developed the method on oligotrophic Mediterranean seawater, and complement the tHg data with samples from the North-Atlantic Ocean, and with pHg isotope measurements for both basins. We use our new tHg isotope observations together with published atmospheric Hg(II) and Hg(0) and marine pHg, sediment and fish Hg isotope data to assess the main pathways of atmospheric Hg deposition to the ocean, and discuss the plausibility of the large global marine Hg(0) evasion flux.

71 Results and Discussion

72 Sea water Hg concentrations, speciation and isotopic composition

Seawater Hg speciation and isotope composition, and procedural standard Hg isotope data are shown in Tables S3, S4. Mediterranean station K2, 20 km off-shore, is an oligotrophic reference site that was used for the 2017 GEOTRACES Hg species intercalibration cruise because the typical water masses of the open Western Mediterranean Sea are present (Figure S1). tHg concentrations at K2 in June 2017, and Feb and May 2019 (Figure 1a, Table S1, S2) are slightly surface depleted (0.8 pM), and peak at 300m depth (1.1 pM), similar to previous observations in the Western Mediterranean Sea^{31,32}. Total 79 methylated Hg (MeHg) concentrations peak at 400-600 m, (43 % of tHg), where oxygen levels reach 80 their minimum due to microbial remineralization of particulate organic matter³². Surface tHg levels in 81 the Bay of Marseille at Endoume pier were higher, 6.0 ± 1.1 pM. In four samples from two Atlantic 82 Ocean stations (St21, St38, Figure S1), tHg increased with depth from 0.46 to 0.83 pM and represent the mixed layer (5m, 20m), intermediate water (650m), and north-east Atlantic deep water (NEADW, 83 3345m)³³. MeHg in the two Atlantic profiles were similar to Mediterranean station K2 with low levels, 84 85 0.04 pM, at the surface and elevated levels, 0.4 pM, at depth (54 % of tHg). pHg at the Mediterranean 86 stations K1, K2 and Julio were low with a mean of $0.11 \pm 0.06 \text{ pM}$ (1 σ , n=16) for the 5-800m depth 87 range. Fram Strait pHg in Atlantic Ocean waters of the West-Spitzbergen current had higher pHg levels of 0.31 ± 0.11 pM (1σ, n=9). 88

tHg isotope depth profiles for δ^{202} Hg, Δ^{199} Hg and Δ^{200} Hg signatures were replicated at station 89 90 K2 during two sampling cruises in February and May 2019 and show reproducible results (Figure 1). δ^{202} Hg is uniform with depth, with a median value of -0.09 ‰ (-0.31 ‰ to 0.00 ‰; median, IQR, n 91 = 12). Overall, Δ^{199} Hg and Δ^{200} Hg are also near-zero, with median values of 0.06 and 0.01 ‰ 92 93 respectively. The four North-Atlantic Ocean tHg samples show median δ^{202} Hg of -0.45 ‰, Δ^{199} Hg of 0.09‰, and Δ^{200} Hg of 0.06‰, and are therefore similar to Mediterranean waters, even at 3345m depth 94 95 in NEADW. Three independent lines of evidence suggest that the tHg isotope composition of seawater 96 is, to first order, a relatively constant feature: 1. The February and May replicate cruises to K2 station 97 do not show any large seasonal variation in tHg isotope profiles, 2. The eight replicate samples at the 98 Endoume pier (Table S4), covering one diurnal 24 h cycle, do not show any Hg isotope variation, and 99 3. The off-shore Mediterranean and Atlantic Ocean tHg isotope signatures are similar.

100 pHg isotopes

At station K2, pHg Δ^{199} Hg and Δ^{200} Hg were similar in June 2017 and May 2019 at all depths, while δ^{202} Hg 101 102 was higher in June 2017 by 1.5‰ at 100-400m depth. In May 2019, the only occasion when tHg and pHg isotopes were determined simultaneously at station K2, δ^{202} Hg of pHg was depleted by -1.0 ‰ (-103 104 0.8 ‰ to -1.3‰; median and IQR, n = 4) relative to tHg (Figure 1). Light isotope enriched pHg may be explained by the preferential sorption of light Hg(II) isotopes to particulate organic matter³⁴, or by 105 106 preferential bio-uptake of light isotopes by phytoplankton that is part of the particulate pool. pHg 107 stable isotopes sampled in 2017 at two stations, Julio and K1, closer to the coast were similar to the 108 open sea station (K2) with no clear trends in water depth or distance from the shore (Figure S3). Marine sediments, analyzed for station Julio only (700m depth, Table S4), had similar δ^{202} Hg = -0.75 ‰ ± 109 0.12‰, Δ^{199} Hg = 0.04 ‰ ± 0.10‰, and Δ^{200} Hg = 0.03 ‰ ± 0.06‰ (mean ± 2 σ , n=4) to mean water 110

column pHg (K1, K2, Julio), and were similar to sediment samples taken elsewhere in the
 Mediterranean Sea^{35,36}.

113 Estimating atmospheric Hg(II) and Hg(0) deposition

114 Δ^{200} Hg has been used successfully as a conservative tracer for atmospheric Hg(0) and Hg(II) wet deposition in terrestrial ecosystems^{23,23,25}. Here we use Δ^{200} Hg to quantify the combined contribution 115 116 of Hg(II) wet and dry deposition, and Hg(0) gas exchange to marine tHg and pHg. In doing so, we 117 exclude other Hg sources such as hydrothermal or river inputs. At Mediterranean station K2, this is 118 justified by the absence of large rivers draining into the Ligurian Sea, and reflected in the open-ocean 119 type tHg levels around 1 pM. The Rhône River reaches the Mediterranean, 100 km west of K2, at the 120 continental shelf of the Gulf of Lions, and is carried further westward away from K2. Similarly, there is 121 no hydrothermal activity within 500 km from the K2 station. At the Atlantic Ocean stations St21 and St38, no influence of river¹⁵ or hydrothermal Hg inputs was found³³. 122

Modern, northern hemispheric Hg(II) wet deposition is characterized by a positive Δ^{200} Hg of 123 124 0.16 ‰ (0.11 ‰ to 0.22 ‰; median, IQR, n = 106), for background sites with Hg concentrations < 25 ng L⁻¹; Table S1). A new study shows that gaseous atmospheric Hg(II) forms, that are the precursor to 125 Hg(II) in wet deposition, have the same Δ^{200} Hg as Hg(II) wet deposition in the lower free and middle 126 127 troposphere²⁷. A recent modeling study evaluated the origin of global Hg(II) wet and dry deposition³⁷, 128 and found that Hg(II) sourced in the upper and middle troposphere constitutes 91% of the annual 129 global Hg(II) wet deposition flux. The global contribution of the upper and middle troposphere to Hg(II) dry deposition was 52%. We therefore suggest that the median northern hemisphere rainfall Δ^{200} Hg 130 131 of 0.17 ‰ is a reasonable estimate of the Δ^{200} Hg of combined wet and dry Hg(II) deposition. Atmospheric Hg(0), the larger complementary pool, exhibits slightly depleted Δ^{200} Hg of -0.05 % (-132 133 0.08 ‰ to -0.02 ‰; median, IQR, n = 126, for background samples with Hg(0) concentrations < 2 ng m⁻ 134 ³; Table S1).

All seawater tHg and pHg samples, including published Pacific Ocean pHg²⁹, show Δ^{200} Hg values 135 between those of atmospheric Hg(0) and Hg(II) (Figure 2 & Figure 4A). Based on a Δ^{200} Hg mixing model 136 (Methods, Eq. 4) we estimate the fraction of Hg(II) wet and dry deposition, $f_{Hg(II)}$, to be 41% (32% to 137 138 65%, global median, IQR, n=17) in marine tHg and 27% (13 to 51%, n=61) in pHg. This implies that the 139 majority, i.e. 59 to 73%, of marine tHg and pHg is derived from direct Hg(0) gas exchange. Current best 140 model estimates of global Hg(II) deposition and net Hg(0) uptake to the ocean surface are 4600 Mg y^{-1} 141 ¹ and 1700 Mg y⁻¹ respectively³. Our marine tHg and pHg isotope observations suggest that the Hg(II) 142 to Hg(0) net deposition balance is in fact opposite, favoring Hg(0) at the mid-latitude marine stations

we investigated. Whereas Δ^{200} Hg informs on the relative proportions of net Hg(II) and Hg(0) deposition, 143 144 it cannot independently inform on Hg fluxes in mass per time units. Additional information is therefore needed to assess which number, 4600 or 1700 Mg y⁻¹, or both are incorrect. Direct Hg(II) wet and dry 145 146 deposition measurements over the ocean are limited. Median Hg(II) wet deposition observed at six coastal monitoring sites between 2011 and 2015 is 1.6 µg m⁻² y^{-1 5}, suggesting an approximate global 147 Hg(II) wet deposition of 560 Mg y⁻¹ to oceans. Model Hg(II) wet deposition generally is on the order of 148 60% of total Hg(II) deposition, the remaining 40% being Hg(II) dry deposition³. This suggests that total 149 150 Hg(II) deposition to oceans may be closer to 1000 Mg y^{-1} rather than 4600 Mg y^{-1} . Applying our 151 calculated Hg(II)/Hg(0) net deposition ratio of 41%/59%, we suggest that net Hg(0) deposition to Oceans is then on the order of 1350 Mg y⁻¹. Our combined marine and atmospheric Hg isotope mass 152 153 balance lacks observations in the southern hemisphere. While this adds uncertainty to the global 154 estimates, we suggest that the global model 3:1 Hg(II) to Hg(0) deposition ratio over oceans is in reality 155 closer to 1:1.

We apply the same Δ^{200} Hg mass balance to published Hg isotope data in northern hemisphere marine sediments and fish (Table S2). Similar to sea water tHg and pHg, with median $f_{Hg(II)}$ of 41% and 27%, global sediments and marine fish Δ^{200} Hg suggest lower than expected $f_{Hg(II)}$ of 32% and 51% respectively. The marine sediment and fish Δ^{200} Hg data cover a much larger geographical range than our new tHg and pHg data, including South Pacific sediments, and therefore reinforce the idea that Ocean Hg(0) uptake is a globally important driver of marine ecosystem Hg levels.

162 **Photochemical reduction of Hg(II) to Hg(0)**

163 During photoreduction of Hg(II)-organic ligand complexes the magnetic isotope effect separates even from odd Hg isotopes³⁸. Experimental photomicrobial Hg(II) photoreduction in simulated sea water 164 medium has been shown to lead to negative Δ^{199} Hg in residual aquatic Hg(II)³⁹. Here we evaluate 165 166 whether the tHg and pHg data show evidence of odd-MIF, in addition to that inherited from atmospheric deposition sources. We first calculate the excess Δ^{199} Hg_{exc} in all seawater samples, defined 167 as the difference between the measured Δ^{199} Hg in a marine sample and the expected Δ^{199} Hg, based 168 on atmospheric inputs. The expected Δ^{199} Hg was calculated by binary mixing of Δ^{199} Hg in atmospheric 169 170 Hg(II) and Hg(0) using the source contributions ($f_{Hg(II)}$) derived from Δ^{200} Hg (Methods, equations 3-4). Overall, the Δ^{199} Hg pattern in tHg and pHg of seawater and marine sediments is similar to Δ^{200} Hg and 171 172 observed Δ^{199} Hg can largely be explained by a conservative mixing of atmospheric Hg(0) and Hg(II) deposition (Figure 2B). Atlantic and Mediterranean Δ^{199} Hg_{exc} was 0.03‰ (0.02 to 0.13‰; median, IQR, 173 n = 17) in tHg and 0.01‰ (-0.07‰ to 0.08‰; median, IQR, n = 61) in pHg, which is not significant. Very 174 175 low Δ^{199} Hg_{exc} were also found in marine sediments of the Mediterranean Sea (0.09‰, 0.03‰ to

0.12‰) and the Atlantic Ocean (0.06‰, 0.02 to 0.10‰). Elevated median Δ^{199} Hg_{exc} of 1.64‰ in marine 176 fish (Table S2) can therefore be predominantly ascribed to photochemical breakdown of MMHg⁴⁰. The 177 absent diurnal tHg Δ^{199} Hg variation in coastal Mediterranean water, under fully sunlit (175 W m⁻²) and 178 179 high wind (10 m s⁻¹) conditions favorable of Hg(0) evasion, support slow marine Hg photoreduction. 180 Given that odd-MIF is one of the hallmarks of aqueous Hg(II) photoreduction, the low to absent 181 Δ^{199} Hg_{exc}, <0.09‰, in marine tHg, pHg and sediment Hg suggests that the large photoreductive marine Hg(0) evasion flux of 4600 Mg y⁻¹ in models is likely overestimated. This notion is also supported by the 182 183 absence of seasonal Hg(0) variation in the atmospheric marine boundary layer, in particular in the ocean dominated southern hemisphere⁴¹. Figure 4 illustrates (in red) changes in atmosphere-ocean Hg 184 185 exchange based on our findings. In order to maintain steady-state in atmospheric Hg inputs and 186 outputs, we adjust marine Hg emissions to 600 Mg y⁻¹, which is 8x lower than the previous estimate and compatible with the low Δ^{199} Hg_{exc} observed. Lower atmospheric Hg(II) deposition to and lower 187 188 Hg(0) evasion from oceans has profound impacts on the parameterization of Earth system Hg models, 189 the lifetime of anthropogenic Hg in atmosphere and surface ocean, and consequently the anticipated 190 recovery of ecosystems following aggressive Hg emission policy.

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207

208 Author contributions

- LEHB, JES, MJ and DP conceived the study. LEHB, MJ, DP, MP, MVP, MMD and JES performed sampling.
- 210 JES, MJ, and LEHB developed and applied the tHg isotope pre-concentration methods. JES, MJ, JM and
- 211 JC performed isotope measurements. MMD, MVP, AD, LEHB, MJ, JM, DP, and MT performed additional
- 212 laboratory work. MJ, JES and LEHB analyzed the data. JES and MJ wrote the draft paper, which was
- 213 improved by contributions from LEHB and DP, and commented by all authors.
- 214

215 Methods

The method for tHg in sea water consists of an ultra-clean shipboard sampling and post-cruise sample
 processing protocol using a standard stannous chloride (SnCl₂) reduction purge-and-trap method,
 based on the USEPA method 1631⁴².

219 Sampling. Repeated, daily cruises were undertaken on the RV Antedon II from Marseille (France) to 220 the nearby (20 km) oligotrophic, off-shore station K2 (42.98 N/5.41 E/ >1500 m-depth) between 221 14/6/2017 and 21/6/2017, on 27/02/2019 and 22/05/2019. We used an epoxy-painted trace metal 222 clean carousel, equipped with a conductivity-temperature-depth unit (CTD, Seabird SBE 911plus), and 223 oxygen sensor (Seabird SBE 43), and 8x 10 L GOFLO trace metal clean bottles (General Oceanics). 224 Samples were taken at up to 12 depths for Hg species, and at 4 depths for tHg and pHg isotopes, 225 corresponding to surface waters (5 m), the chlorophyll-maximum (20 m), the oxygen minimum zone 226 (400 m) and deep (800 m) waters. See SI for salinity, temperature, oxygen and fluorescence data. 227 Unfiltered seawater was drawn via acid-cleaned 6 mm FEP tubing into pre-cleaned 20 L Pyrex glass 228 bottles with GL45 PFA Teflon caps. Bottles were filled to the 20 L mark, and not to the brim, in order 229 to preserve a 3 L headspace for purge and trap purposes. Bottles were protected from sunlight and 230 breaking risk in 70 L plastic barrels (Kruizinga.nl Ref#53-WHV70) by using polyurethane expansive foam 231 to fit them in the barrels. We anticipate that future use of 20 L polycarbonate carboys are an alternative 232 option, for safer sampling during rough seas. Unfiltered seawater was drawn from each GOFLO bottle 233 into acid-cleaned and blank-tested 60 mL FEP Teflon bottles (Nalgene) for tHg and 250 mL PFA Teflon bottles (Savillex Purillex) for MeHg, MMHg and dissolved gaseous Hg (DGM = Hg⁰ + dimethyl-Hg 234 (DMHg)) analysis. All sampling was compliant with strict GEOTRACES trace metal clean sampling 235 236 procedures⁴³. Eight 20 L replicate samples of seawater from the Bay of Marseille were taken every 237 three hours on 12/11/2019 at the Endoume pier. Seawater is continuously pumped at 20 L min⁻¹ into 238 the Mediterranean Institute of Oceanography (MIO) seawater sensing lab (SSL@MM). Samples were 239 transported to MIO and processed within 24 h. Atlantic Ocean tHg samples were collected during the 240 GEOVIDE cruise (GEOTRACES-GA01 transect), on board the RV "Pourquoi Pas?" between 15/05/2014 241 and 30/06/2014. tHg values for GEOVIDE have been published elsewhere ³³.

242 Particulate Hg. pHg was sampled using in situ pumps (McLane LV08) at stations K1 (43.10 N, 5.49 E, 243 700 m depth, 10 km off-shore, continental slope), K2, and Julio (43.13 N, 05.36 E, 100 m-depth, 10 km 244 off-shore, shelf) in the Mediterranean Sea on 16/06/2017, and at K2 on 22/05/2019; in Fram Strait 245 Atlantic waters from 21/7/2016 to 1/9/2016 during the FS Polarstern PS100 GRIFF cruise (Geotraces-246 GN05 transect). Pre-burnt Millipore 142mm QMA, or GF/F were deployed for 1 - 3 h to collect particles 247 from 148-792 L of seawater. Filter samples were frozen onboard, shipped frozen to MIO, freeze dried 248 (Christ Gamma 1-16 LSCplus), and stored in the dark until analysis. Table S4 summarizes all sampling 249 locations.

250 Pre-concentration of tHg from seawater for stable isotope analysis

251 The 20L Mediterranean tHg samples were acidified and oxidized, within 12 h after sampling, at the 252 MIO laboratory, using 5 mL 0.2 N BrCl (Sigma-Aldrich KBr and KBrO₃ salts) in concentrated bi-distilled 253 9 N HCl, leading to an initial sample HCl concentration of 0.0023 N. BrCl blanks were analyzed before 254 addition, as it potentially constitutes the largest single component of the method blank. The volume 255 of BrCl was optimized by visual inspection of the sample turning slightly yellow, indicating excess BrCl 256 over reduced seawater components such as dissolved and particulate organic matter (DOC, POC, OM). 257 We anticipate that the volume of BrCl added, and the time needed to convert all Hg species to labile 258 Hg(II) forms will depend on the OM levels of seawater elsewhere. Samples were let sit for 12 h, after 259 which a 60 mL subsample was taken with an acid cleaned, 60 cm long, burette into pre-combusted 60 260 mL glass vials with acid-washed Teflon-lined caps for tHg concentration analysis. This tHg subsample is 261 compared to the shipboard 60mL tHg FEP sample and serves to verify that the 20L bottles are neither contaminated, nor subjected to tHg loss. 262

263 tHg pre-concentration was started by replacing the GL45 PFA Teflon caps (Savillex) by GL45 264 two-port PFA Teflon caps (Savillex) to guide 60 cm long 6 mm OD (3 - 4 mm ID) Pyrex bubbling post 265 with a 1 cm long P3 porosity frit (VitraPOR Micro Filter-Candle, Robuglas, Germany, custom assembled 266 by Verres Vagner, Toulouse, France). The second port on the GL45 cap hosted a 10 cm long 6 mm OD 267 FEP tube that was connected with a short piece of 10 mm OD, 4 mm ID silicone tubing to a shorter, 25 268 cm long, elbowed 6mm OD Pyrex bubbling post, with 1 cm long P2 frit (VitraPOR). The P2 bubbler post 269 was then inserted into a 60 mL, 20 cm long glass test tube, filled with 6 mL of oxidizing 40 vol% inverse 270 aqua regia (iAR) solution. The finer P3 bubbler frit inside the bottle leads to abundant small bubbles, 271 but can technically be replaced by coarser frits. The medium P2 bubbler frit inside the 40% iAR trap is 272 critical to avoid over pressure and leaks of Hg(0) from the 20 L bottle. All glassware was pre-combusted 273 at 530°C and Teflon-ware was cleaned by multiple bi-distilled HCl steps in a class 100 clean room.

274 Standard protocols for Hg(II) analysis by SnCl₂ reduction, such as EPA method 1631, use a large 275 excess of $SnCl_2$ over Hg(II). While $SnCl_2$ does not pose a blank issue (purging it removes all Hg traces), 276 it generates large volumes of toxic waste in each 20 L bottle after pre-concentration. We therefore 277 tested up to 100x lower levels of anhydrous Sn(II)Cl₂ (Sigma-Aldrich). Hg(II) reduction by Sn(II) 278 competes however with excess BrCl and natural oxygen present in seawater, i.e. Sn(II) is oxidized by 279 BrCl and O_2 before it can reduce Hg(II). We therefore neutralized excess BrCl by adding 4 mL of NH₄.HCl 280 (4.3 M) and verified disappearance of the yellow color. We then pre-purged (before adding SnCl₂) the 281 20 L sample with Hg-free argon at 300 mL min⁻¹ for 3 h to remove > 90 % of dissolved oxygen. The 40% 282 iAR solution trap was connected during pre-purging to trap any potential, though unlikely, loss of 283 gaseous Hg from the sample. Adding SnCl₂ at this point, had however shown abundant precipitation of a fine white solid, presumably SnCl_{2,s}, which we remedied by adding an additional 80 mL volume of 284 285 HCl to stabilize Sn(II) in the samples. Sn(II) addition was performed in two ways: i) 125 mL of SnCl₂ was slowly pumped with a peristaltic pump (Gilson) at 1.25 mL min⁻¹, and 1/16" PFA tubing, via the central 286 287 bubbling post into the bottle over 2 h; ii) the 125 mL was added instantly via the GL45 cap. No 288 differences in Hg recovery were found between the two. Upon addition of SnCl₂ by pump or in batch, 289 we purged the 20 L samples for 8 h at 300 mL min⁻¹, in order to quantitatively collect sample Hg into 290 the oxidizing 40% iAR solution trap. At the end of 6 h of pre-concentration, the argon flow was stopped, 291 and the 40 v% iAR traps removed and diluted with MQ water to 20 v% iAR and stored cold (4°C) in the 292 dark until analysis.

For the Atlantic Ocean tHg samples, a different pre-concentration technique was applied onboard which is described in detail in the Supporting Information. In brief, 48L samples, stored in tedlar (Teflon) bags, were pumped at 5mL min⁻¹, through an iodated activated carbon (IAC) cartridge onboard. The IAC sorbent was combusted back on land, at Geosciences Environnement Toulouse (GET), in a dual tube furnace set-up, and sample Hg(0) trapped in 6 mL 40% iAR. We abandoned this method due to higher blanks, challenging Hg recoveries from IAC, and slowness of sea water sample loading (1 week).

300 Procedural blanks and standards

Procedural Hg blanks, representing all laboratory manipulations, cleaning procedures, reagent additions, and different operators, were determined as follows. Following pre-concentration of a batch of 8 samples for tHg, the residual Hg-free seawater solution, was conserved in the 20 L bottle, and all reagents were newly added to 8 bottles as described above. The solutions were then purged, similar to samples, for 8 hours into newly prepared 6 mL 40% iAR traps. Eight procedural standards were preconcentrated in a similar fashion: following blanks, once more all reagents were added to the remaining 20 L of Hg-free seawater solution, and 4 to 8 ng of NIST SRM 3133 Hg was added as internal
 standard to each bottle.

309 Particulate Hg concentration analysis

310 Total particulate Hg (pHg) concentrations were measured on 25 mm stamp-outs, by combustion - cold 311 vapor atomic absorption spectrometry (CV-AAS, Leco AMA254) equipped with a low level optical cell 312 at MIO. Certified reference material NRC MESS-3 marine sediment (93 \pm 9 ng g⁻¹, 1 σ) was used for quality control, with good results in the low, <1 ng Hg, range (88 \pm 1 ng g⁻¹, 1 σ). Remaining filter material 313 314 was combusted whole on the dual tube furnace set-up at GET, following published protocols (Sun et 315 al., 2013). In brief, filters are rolled up, inserted in a 20 mm diameter, 15 cm long quartz tube, which is 316 plugged from both sides with quartz wool. The sample tube is then inserted in a 140 cm long quartz 317 tube, housed in two tube furnaces. The first furnace, hosting the sample tube, was heated from room 318 temperature to 900 °C over 6 h in an 80 mL min⁻¹ flow of high purity oxygen. The released Hg(0) vapor and other volatile compounds passed through the 2nd pyrolizing oven, maintained at 1000°C 319 320 continuously, and purged into a 40 vol% iAR oxidizing solution trap, that uses an elbowed, fritted, P2 321 porosity, Pyrex post. GF/F and GF/D filters melt at temperatures >530°C, but remain within the quartz 322 sample tubes, blocked by the quartz wool. Final trapping solutions were diluted to 20%v iAR and stored 323 cold in the dark until Hg concentration analysis by CV-AFS and Hg isotope analysis by MC-ICPMS. 324 Reference material NIST SRM1632d coal was used for weekly quality control. Combustion purge and 325 trapping recovery on samples, assessed by CV-AFS, was $104 \pm 28\%$, $100 \pm 28\%$, and $105 \pm 21\%$ (1 σ) for 326 the Mediterranean Sea, PS94 and PS100 cruise samples respectively.

327 tHg and Hg species concentration analysis

328 We added 40 µL of 0.2N BrCl to the 60 FEP Teflon bottles for ambient tHg analysis. Ambient tHg 329 concentrations in 60 mL FEP Teflon bottle aliquots, in 20 L bottle aliquots before and after pre-330 concentration, and in 20% iAR solution traps were all analyzed in duplicate using a custom-made purge and trap system coupled to a cold vapor atomic fluorescence spectrometry (CV-AFS, Brooks Rand 331 332 Model III), either at MIO, at GET, or shipboard⁴⁴. The purge and trap system consists of 100 mL Teflon batch reactor (VWR), two electromagnetic valves (NResearch), a single gold trap (LECO), all connected 333 334 by 1/8 inch FEP tubing to the CV-AFS. Seawater aliquots were 35 mL, and 20% iAR aliquots 100 uL. The 335 CV-AFS was calibrated in the 1 - 20 pg range using NIST SRM 3133, and the NRC ORMS-5 certified reference material was always found within 10 % of the certified value ($26 \pm 1.3 \text{ ng L}^{-1}$, 1 σ). MeHg, 336 MMHg, and DGM were analyzed following our published protocols⁴⁴. DMHg was calculated as the 337 338 difference of tMeHg and MMHg, and dissolved Hg(0) as the difference of DGM and DMHg. Details are 339 given in the SI.

340 Hg isotope analysis

341 Hg stable isotope ratios of final 20 % iAR trap solutions were measured in duplicate during two sessions 342 by cold vapor multi-collector inductively coupled plasma mass spectrometry (CV MC-ICPMS) at the Observatoire Midi-Pyrénées, Toulouse^{45,46}. We used a CETAC ASX-520 autosampler and HGX-200 CV 343 344 system coupled to a Thermo-Scientific Neptune PLUS, equipped with a $10^{12} \Omega$ resistor, attributed to 345 the ¹⁹⁸Hg isotope in order to improve isotope ratio precision in the 20-50 mV range. Samples and standard signals at 0.25 ng g⁻¹ tHg levels were generally 180 mV on the ²⁰²Hg isotope, at a sample 346 347 introduction flow rate of 0.75 mL min⁻¹. Thallium was not used as an internal standard, and the 203 and 205 masses were monitored to survey Hg-hydride interferences (i.e. ²⁰²Hg¹H, and ²⁰²Hg¹H¹H), 348 which were found to be negligible when using standard H-cones. ¹⁹⁶Hg and ²⁰⁴Hg were not analyzed 349 350 due to low abundance, and cup configuration limitations. MDF of Hg stable isotopes is reported in 351 small delta notation (δ) in per mil (∞) deviation from to the reference NIST 3133 Hg standard:

352
$$\delta^{xxx}$$
Hg = (xxx/198Hg_{sample}/ xxx/198Hg_{NIST3133} -1) × 10³

353 where 'xxx' refers to measured isotope masses: 199, 200, 201, and 202. Mass independent 354 fractionation (MIF) is reported in capital delta notation⁴⁷, which is defined as the difference between 355 the measured δ^{199} Hg, δ^{200} Hg, and δ^{201} Hg values and those predicted for MDF relative to δ^{202} Hg using 356 the kinetic MDF law:

(1)

(2)

357
$$\Delta^{XXX}$$
Hg = δ^{XXX} Hg - SF^{XXX} × δ^{202} Hg

358 where SF^{xxx} is the mass-dependent scaling factor of 0.2520 for ¹⁹⁹Hg, 0.5024 for ²⁰⁰Hg, and 0.7520 for 359 201 Hg⁴⁷.

The long-term instrumental precision was assessed through repeated analysis of the UM-Almaden and ETH-Fluka Hg standard at 0.25 ng g⁻¹ during the two analysis sessions (SI Table S4). ETH-Fluka yielded values of $-1.45 \pm 0.20 \%$, $0.07 \pm 0.11 \%$, $0.01 \pm 0.14 \%$, $0.02 \pm 0.09 \%$, $0.00 \pm 0.18 \%$ (2σ , n = 10) for δ^{202} Hg, Δ^{199} Hg, Δ^{200} Hg, and Δ^{201} Hg respectively, in agreement with published values⁴⁸. UM-Almaden standard yielded -0.55 \pm 0.16‰, -0.03 \pm 0.11‰, -0.01 \pm 0.03 ‰, -0.05 \pm 0.23 ‰ (2σ , n = 10) respectively, in agreement with previously reported values³⁸.

366 pHg and tHg isotope method uncertainty

Mean pre-concentration recoveries for tHg isotope analysis were 90 \pm 10% (1 σ , n=12) at Mediterranean station K2, 93 \pm 18% (n=8) at Endoume coastal station, and 88 \pm 31% (n=4) for Atlantic Ocean samples. Procedural blanks were 0.23 \pm 0.08 ng Hg (1 σ , n=4) for the SnCl₂ pre-concentration method, which is <10% of tHg in 20L of seawater, e.g. typically 3 - 4 ng of tHg in this study. SnCl₂ method

replication of eight 20L coastal Mediterranean samples (6 pM, analyzed at 1.6 ng g⁻¹ by MC-ICPMS) shows good results with δ^{202} Hg, Δ^{199} Hg and Δ^{200} Hg 2 σ uncertainties of 0.23, 0.16, and 0.06 ‰. NIST SRM 3133 procedural standards returned δ^{202} Hg, Δ^{199} Hg, Δ^{200} Hg, and Δ^{201} Hg values of 0.07 ± 0.23 ‰, -0.05 ± 0.16 ‰, 0.00 ± 0.06 ‰, -0.01 ± 0.18 ‰, 0.02 ± 0.21 ‰ and a mean recovery of 86 ± 16% (2 σ , n = 8), suggesting no bias in the method.

pHg quality control results on NIST SRM 1632d yielded values of -1.75 ± 0.24 ‰, -0.04 ± 0.04 ‰, 0.00 ± 0.04 ‰, -0.02 ± 0.08 ‰ (2σ , n = 10) for δ^{202} Hg, Δ^{199} Hg, Δ^{200} Hg and Δ^{201} Hg respectively, in agreement with published values ^{45,49}. The 2σ uncertainty on pHg samples was taken to be the larger of either replicate sample analysis, procedural standard NIST SRM1632d, or secondary reference materials UM-Almaden or ETH-Fluka.

Most seawater pHg and tHg samples were analyzed in duplicate on different days. The final 2σ uncertainties reported for Mediterranean Sea and Atlantic Ocean pHg and tHg samples are the larger of the duplicate sample analysis, the 2σ on the eight NIST procedural standards (tHg), the 2σ on the ten NIST SRM 1632d standards, or the 2σ of the ETH Fluka or UM-ALmaden standards.

385 Stable isotope data analysis

The fraction of Hg in marine samples derived from atmospheric Hg(II) wet and dry deposition ($f_{Hg(II)}$) was calculated using a conservative mixing model as follows:

388
$$\Delta^{200} Hg_{sample} = f_{Hg(II)} \times \Delta^{200} Hg_{Hg(II)} + (1 - f_{Hg(II)}) \times \Delta^{200} Hg_{Hg(0)}$$
 (3)

where Δ^{200} Hg_{sample} represents the median observed value of different marine Hg pools (THg, pHg, sediment THg, fish THg), Δ^{200} Hg_{Hg(II)} and Δ^{200} Hg_{Hg(0)} the median values of previously published northern hemisphere samples for gaseous and rainfall Hg(II), and atmospheric Hg(0), respectively. The excess Δ^{199} Hg in the different marine Hg pools (Δ^{199} Hg_{exc}), representing the Δ^{199} Hg which was produced within the marine system, was calculated as follows:

394
$$\Delta^{199}Hg_{exc} = \Delta^{199}Hg_{exc} - (f_{Hg(II)} \times \Delta^{199}Hg_{Hg(II)} + (1 - f_{Hg(II)}) \times \Delta^{199}Hg_{Hg(0)})$$
(4)

395 where $f_{Hg(II)}$ was derived from the \mathbb{P}^{200} Hg of the sample according to eq. 3.

397 Figures

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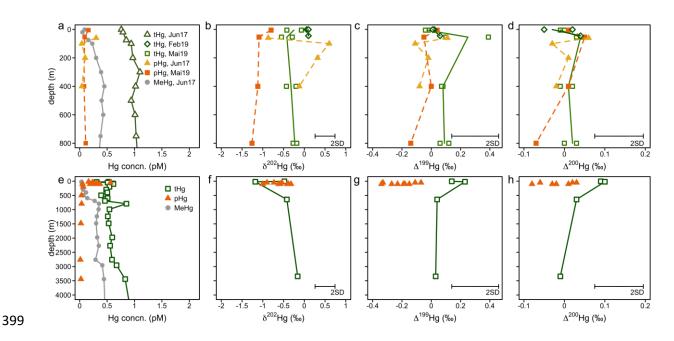
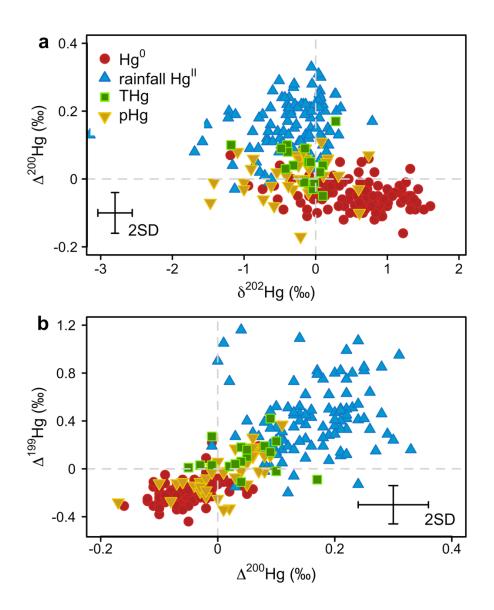


Figure 1a-h. Depth profiles of seawater Hg species concentrations and total and particulate Hg stable isotope composition at station K2 in the Mediterranean Sea (a-d) and the North Atlantic (e-h). a and e: tHg, pHg, and MeHg concentrations in pmol L⁻¹ (pM). b and f: mass-dependent fractionation (δ^{202} Hg), c and g: odd mass-independent fractionation (Δ^{199} Hg), d and h: even mass-independent fractionation (Δ^{200} Hg). Open symbols represent values for tHg and filled symbols pHg. The lines represent averages of replicate analysis of the same sample. Two standard deviation (2SD) uncertainties on Hg concentrations are 10%.

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408



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Figure 2. Hg stable isotope composition of atmospheric sources (gaseous Hg(0) and Hg(II) in rainfall) and seawater (total (HgT) and particulate (pHg)). a) even mass independent fractionation (Δ^{200} Hg) vs. mass dependent fractionation (δ^{202} Hg), and b) odd mass independent fractionation (Δ^{200} Hg) vs. even mass independent fractionation (Δ^{200} Hg). Values of atmospheric sources are from literature (see Table S1), tHg values are from this study, pHg from this study and ref.²⁹. Error bars represent the 2SD of replicate procedural standards for tHg and pHg.

417

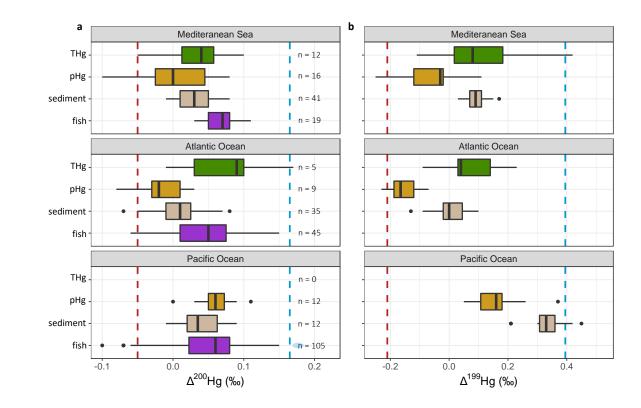
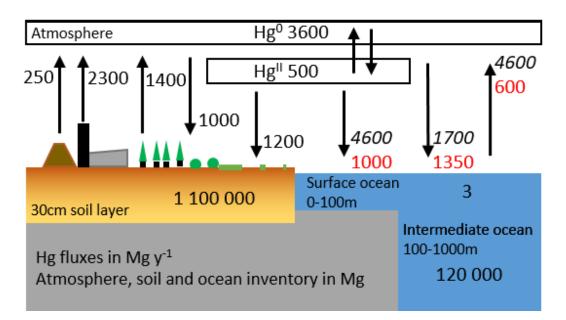




Figure 3. Hg stable isotope signatures in different ocean basins of total Hg (tHg) and particulate Hg (pHg) in seawater, marine sediments and marine fish. a: Even mass-independent signature (Δ^{200} Hg) and b: Odd mass-independent signature (Δ^{199} Hg). tHg values are from this study, pHg from this study and from Motta et al. (2019), data for sediments and biota are from literature. The atmospheric sources are shown in red for Hg(0) and blue for Hg(II) in rainfall, the dashed line represents the median and the shaded area the interquartile range. For fish, Δ^{199} Hg are not shown in this Figure as these values are heavily affected by MMHg photo-demethylation (See SI Table S2).



428

429 Figure 4. Global budget of air-sea and air-land exchange. New Hg isotope based fluxes (Mg y⁻¹) are

shown in red. All other Hg fluxes and atmospheric Hg0 and HgII inventory are from ref.⁵⁰. The upper
30cm soil and surface (0-100m) and intermediate (100-1000m) ocean inventories are from refs.^{11,51}.

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560

562 **Online Supporting Information to:**

563

564 Mercury stable isotope composition of seawater suggests important net 565 gaseous elemental mercury uptake

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576 Materials and Methods

577 Rationale

578 tHg concentration in seawater is the sum of several operationally defined species, including dissolved 579 gaseous Hg(0), dissolved gaseous dimethyl-Hg (DMHg), dissolved inorganic Hg(II) compounds, 580 dissolved monomethyl-Hg compounds (MMHg), and particulate-bound Hg(II) and MMHg compounds 581 (pHg). Both concentration and stable isotope analysis of tHg in seawater require the transformation of 582 these compounds to dissolved, labile Hg(II) forms by addition of a strong oxidant such as BrCl, prepared 583 in concentrated HCI. The concomitant oxidation and acidification also inhibits biological activity, 584 stabilizes Hg(II) in solution and minimizes potential losses to sampling container walls or to the 585 atmosphere. Two criteria need to be met for a robust Hg stable isotope analysis of seawater: first, the 586 low-picomolar seawater concentration has to be concentrated to levels suitable for Hg stable isotope analysis (typically 0.2 – 1 ng mL⁻¹ or 1 – 5 nM). This pre-concentration needs to be quantitative in order 587 588 to avoid Hg isotope fractionation during transfer. Second, the procedural blank of the method needs 589 to be low, ideally below 10% of the total amount of Hg in the sample. Low blank levels rely on ultra-590 clean sampling at sea, clean sampling vessels and ultra-low Hg levels of reagents used.

591 Previous studies have successfully developed methods for large volume (1 - 20 L) pre-592 concentration of tHg from natural waters, including rain, snow, ice, lake and coastal seawater. Gratz 593 et al. (2010) and Sherman et al. (2010) first used continuous cold vapor generation, with SnCl₂ as a 594 reductant, to pre-concentrate 1 - 4 L of snow and rainfall samples into small volume (25 mL) 2 % (w/w) oxidizing KMnO₄/H₂SO₄ solution trap ^{1,2}. This method was later modified to a batch-reactor set-up to 595 596 pre-concentrate 1 L of rainfall in a 2 L Pyrex bottle, by slow, pumped addition of SnCl₂, and identical 597 $KMnO_4/H_2SO_4$ solution trap³ and trapping solutions of 40 vol% inverse agua regia (iAR, 4.2 N HNO₃, 598 1.2 N HCl; ⁴). Finally, in Obrist et al. (2017) we scaled up the batch purge and trap method to handle up to 20 L of snow and rain sample ⁵. An alternative method was developed by Chen et al. (2016) using 599 the AG-1-X4 anion exchange resin to pre-concentrate >40 ng of tHg from freshwater lakes ⁶. HgCl₄²⁻ in 600 601 the acidified, 0.1 N HCl, sample is pumped at 3.5 mL min⁻¹ over 0.5 mL of AG-1-X4 resin. Sorbed Hg is 602 then eluted with 10 mL of 0.05 % L-cysteine in 0.5 N HNO₃, oxidized by BrCl, neutralized with NH₄OH 603 and analyzed by MC-ICPMS. Štrok et al. (2015) optimized the anion exchange resin method for 604 seawater and published the first Hg stable isotope data of coastal seawater (1.5-3.1 pM) from the 605 Canadian Arctic Archipelago ⁷. The latter two methods by Štrok et al. (2015) and Obrist et al. (2017) 606 are capable of handling sample volumes of 20 L or more that are necessary for background level tHg 607 pre-concentration from seawater (0.4 to 1.5 pM). Our objective was therefore to adapt our existing large volume pre-concentration protocol, based on SnCl₂ reduction, for rain and snow to seawater. We
 optimized the protocol for quantitative pre-concentration yields and low blank levels.

610 Pre-concentration troubleshooting, Sn waste disposal and bottle cleaning

611 During pre-concentration the 20L sample bottle is under argon over-pressure in order to generate a 612 fine stream of bubbles. It is therefore important at the end of pre-concentration, or during 613 troubleshooting, not to vent the argon supply tube to the 20L sample bottle and release argon 614 pressure. The reason is that with the 20 L bottle under over-pressure, venting the argon supply tube 615 results in a sudden pressure drop, and acidified sample solution with SnCl₂ moving from the 20 L bottle 616 into the tall bubbler post, possibly all the way to the argon flow regulator, damaging the latter. Worse, 617 the 40 vol% iAR trap solution is aspirated up into its short bubbler post and into the 20 L bottle, 618 resulting in the loss of the sample. Using our final optimized Sn addition protocol, each 20 L bottle 619 contains at the end of a pre-concentration run, 2.5 g of Sn(II), and should be disposed of following local 620 environmental regulations. If needed, the dissolved Sn(II) concentration can be lowered by hydrogen 621 peroxide addition, which quantitatively precipitates Sn(IV)O₂, that can then be filtered or decanted. 622 Quantitative oxidation of Sn(II) by hydrogen peroxide also has the advantage that no traces of Sn(II) 623 remain in the 20 L bottle, which can be acid-cleaned and rinsed with abundant MQ water to receive 624 the next batch of seawater samples. In this study both sampling and pre-concentration were done in 625 the same bottle. We expect however that the use of separate bottles, possibly using plastic 20 L 626 carboys at sea for safety, does not increase blanks.

627 Alternative, activated carbon based pre-concentration method for sea water

628 We have, over the past years, tested an alternative method for low-level Hg pre-concentration in order 629 to measure Hg isotopes in sea water. The method uses small 200 mg iodated activated carbon powder 630 (Brooks Rand) cartridges, over which large volumes (50L) of sea water are pumped at a slow flow rate 631 of 5 ml min⁻¹. Following laboratory recovery and blank testing, we applied the method during the 2014 632 GEOTRACES Geovides cruise in the North-Atlantic Ocean. Unfiltered seawater was transferred using 633 clean 6mm FEP tubing from GOFLO bottles into acid washed 50L tedlar bags. A peristaltic pump, with 634 1.14mm ID (red/red) tygon pump tubing was used to load samples on the IAC traps during 1 week. 635 Traps were dried for 5 min in a dry high purity argon stream, sealed with silicone stoppers and stored 636 and transported dry and in the dark to the GET laboratory. The IAC powder was combusted using a dual tube furnace combustion method⁸. IAC blanks were 0.5 ng Hg per 200mg of IAC traps, and sample 637 638 recoveries were 88 ± 31%. In hindsight, it has been difficult to fully recover Hg from IAC traps due to 639 abundant volatile iodine release and transfer to oxidizing solution traps. We recommend that further 640 development of activated carbon based methods use commercial sulfur impregnated activated carbon

641 (Calgon HGR), which does not have this issue.

642 MMHg and DMHg analysis

643 MMHg and DMHg were determined by isotope dilution (ID)- gas chromatography - sector field ICP-644 MS (ID-GC-SF-ICPMS) method at the MIO laboratory, following previously published protocols⁹. First 645 the sum of both species (tMeHg) was quantified on an acidified sample that converts DMHg to MMHg. 646 A second replicate sample was purged to remove DMHg before acidification, allowing direct 647 measurement of MMHg. DMHg was calculated by difference, as tMeHg – MMHg. In both sample 648 aliquots MMHg and inorganic Hg species were extracted after derivatization. In brief, enriched spikes of ¹⁹⁹iHg and ²⁰¹MeHg (ISC Science, Spain) were added to a 115 mL aliquot of the sea water samples. 649 650 After 24h of equilibration, pH was adjusted to 3.9 with NH₃ (ULTREX[®] II Ultrapure Reagent, J.T. Baker, 651 USA) and a buffer solution made up with acetic acid (glacial, ULTREX® II Ultrapure Reagent, J.T. Baker, 652 USA) / sodium acetate (J.T. Baker, USA). A solution of 1 % (v:v) sodium tetra propyl borate 653 (Merseburger Spezialchemikalien, Germany) was made up freshly, under cold conditions and avoiding 654 contact with atmospheric oxygen. 1 mL of this solution was then added together with 200 µL hexane (Sigma Aldrich, USA). The glass bottles were hermetically sealed with Teflon-lined caps and vigorously 655 656 shaken for 15 minutes. The organic phase was recovered and injected in the GC (Thermo Trace Ultra), 657 coupled to a sector field ICPMS (Thermo Element XR). Detection limits were 0.005 pM for MMHg, and 658 0.01 pM for DMHg.

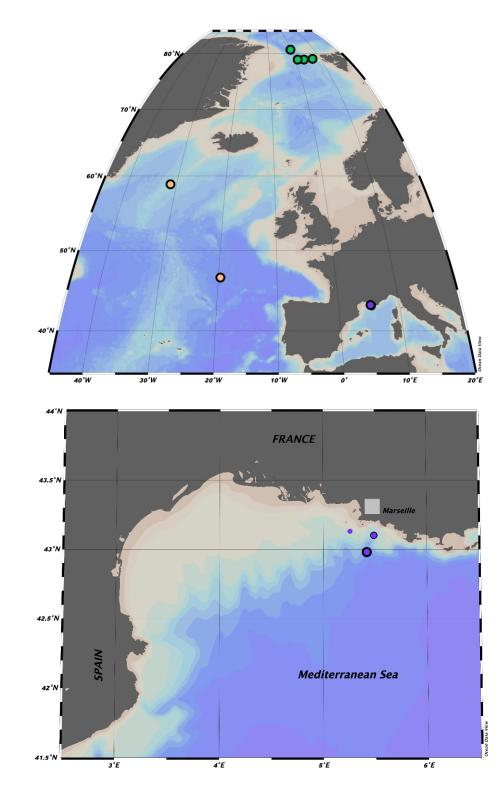
							202		100		200	
Pool	Ocean Basin	n	Hg concn.			δ^{202} Hg (‰)		Δ^{199} Hg (‰)		Δ^{200} Hg (‰)		Reference
			median	IQR	Units	median	IQR	median	IQR	median	IQR	
Hg(II) rainfall		106	13	(9 to 18)	ng/L	-0.30	(-0.63 to 0.03)	0.39	(0.21 to 0.52)	0.16	(0.11 to 0.22)	1,3,4,10–14
Hg(0)		126	1.23	(1.08 to 1.45)	ng/m ³	0.61	(0.16 to 0.90)	-0.21	(-0.24 to -0.15)	-0.05	(-0.08 to -0.02)	2,4,5,10,15–17
tHg	All samples	17				-0.15	(-0.39 to -0.02)	0.06	(0.02 to 0.18)	0.04	(0.02 to 0.09)	this study
	Med. Sea	12				-0.08	(-0.31 to 0.00)	0.08	(0.02 to 0.18)	0.04	(0.01 to 0.06)	this study
	Atlantic	5				-0.42	(-0.48 to -0.16)	0.04	(0.03 to 0.14)	0.09	(0.03 to 0.10)	this study
рНg	All samples	37				-0.22	(-0.58 to -0.10)	-0.03	(-0.14 to 0.11)	0.01	(-0.02 to 0.06)	this study, ¹¹
	Med. Sea	16				-0.14	(-0.9 to 0.19)	-0.05	(-0.12 to -0.02)	0.00	(-0.03 to 0.04)	this study
	Atlantic	9				-0.58	(-0.73 to -0.41)	-0.19	(-0.27 to -0.15)	-0.02	(-0.03 to 0.03)	this study
	Pacific	12				-0.13	(-0.22 to -0.1)	0.16	(0.1 to 0.18)	0.06	(0.05 to 0.07)	
sediment	All samples	88	55	(34to 86)	ng/g	-0.82	(-1.21 to -0.45)	0.07	(0.02 to 0.11)	0.02	(0.01 to 0.04)	
	Med. Sea	41	53	(44 to 83)	ng/g	-1.14	(-1.61 to -0.86)	0.09	(0.07 to 0.11)	0.03	(0.01 to 0.05)	This study, ^{17, 18}
	Atlantic	35	55	(33 to 137)	ng/g	-0.40	(-0.60 to 0.32)	0.00	(-0.02 to 0.05)	0.01	(-0.01 to 0.03)	18,19*
	Pacific	12	52	(20 to 74)	ng/g	-1.58	(-2.5 to -0.5)	0.33	(0.30 to 0.36)	0.04	(0.02 to 0.06)	this study
fish	All samples	169	252	(81 to 1403)	ng/g	0.49	(0.28 to 0.79)	1.72	(1.35 to 1.98)	0.06	(0.03 to 0.08)	
	Med. Sea	19	2950	(2460 to 3826)	ng/g	0.37	(0.29 to 0.47)	1.78	(1.70 to 1.81)	0.07	(0.05 to 0.08)	this study
	Atlantic	45	105	(60 to 190)	ng/g	0.34	(0.26 to 0.45)	1.32	(1.06 to 1.50)	0.05	(0.01 to 0.08)	20,21
	Pacific	105	366	(104 to 747)	ng/g	0.71	(0.35 to 0.89)	1.84	(1.48 to 2.17)	0.06	(0.02 to 0.08)	22–24

Table S1: Summary of Hg stable isotope data used for Figure 2 and calculations. *sediments considered below 1000m water depth.

660 Table S2: Modeled contribution of Hg(II) from wet and dry deposition, $f_{Hg(II)}$, and excess $\Delta^{199}Hg_{exc}$ according to Methods Equations 3, 4. Excess $\Delta^{199}Hg_{exc}$

661 represents sample Δ^{199} Hg acquired by Hg(II) photoreduction in sea water, in addition to the Δ^{199} Hg inherited from atmospheric Hg deposition.

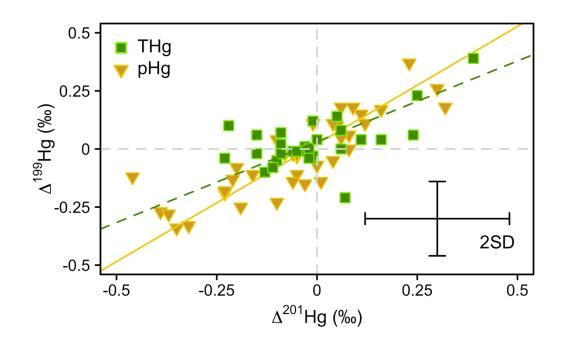
			c		Δ^{199} Hg _{exc}	
Pool	Ocean Basin	n	$f_{Hg(II)}$		(‰)	
			median	IQR	median	IQR
tHg sea water	All samples	17	0.41	(0.32 to 0.65)	0.03	(0.02 to 0.13)
	Med. Sea	12	0.41	(0.29 to 0.50)	0.06	(0.03 to 0.19)
	Atlantic	5	0.65	(0.37 to 0.69)	0.02	(-0.04 to 0.02)
pHg sea water	All samples	37	0.27	(0.13 to 0.51)	0.01	(-0.07 to 0.08)
	Med. Sea	16	0.20	(0.08 to 0.43)	0.02	(-0.08 to 0.05)
	Atlantic	9	0.13	(0.09 to 0.27)	-0.06	(-0.10 to -0.04)
	Pacific	12	0.51	(0.46 to 0.57)	0.07	(0.00 to 0.13)
marine sediment	All samples	88	0.32	(0.28 to 0.41)	0.09	(0.03 to 0.12)
	Med. Sea	41	0.37	(0.28 to 0.47)	0.09	(0.03 to 0.12)
	Atlantic	35	0.28	(0.18 to 0.35)	0.06	(0.02 to 0.10)
	Pacific	12	0.40	(0.32 to 0.52)	0.25	(0.23 to 0.33)
fish	All samples	169	0.51	(0.37 to 0.60)	1.64	(1.28 to 1.92)
	Med. Sea	19	0.55	(0.46 to 0.60)	1.65	(1.56 to 1.72)
	Atlantic	45	0.46	(0.28 to 0.58)	1.29	(1.00 to 1.40)
	Pacific	105	0.51	(0.32 to 0.60)	1.79	(1.47 to 2.08)



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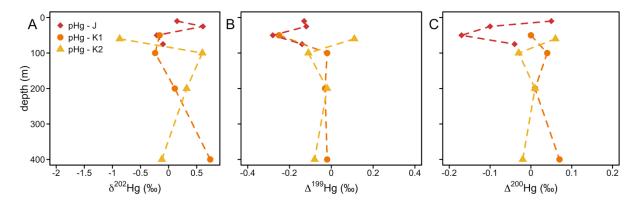
Figure S1. Top: Sampling locations K1, K2 in the Mediterranean Sea (purple), Atlantic Ocean (yellow)
and Fram Strait (green). Bottom: Zoom on the 4 Mediterranean locations, with main station K2 (large
purple circle), and pHg station K1 and Julio (small purple circles), and Endoume pier in Marseille Bay
(grey square).



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Figure S2: Scatterplot of odd Hg isotope mass-independent fractionation, Δ^{201} Hg vs. Δ^{199} Hg of all tHg and pHg seawater data, including the published data by Motta et al. 2019. The dashed line represents the York regression using IsoplotR²⁵ for tHg (Δ^{199} Hg = 0.79(±0.29) Δ^{201} Hg + (0.03±0.04), (± se), MSWD = 0.2). The solid line represents the York regression for pHg (Δ^{199} Hg = 1.01(±0.0.09) Δ^{201} Hg + (0.02±0.02), (± se), MSWD = 1.25). Error bars represent the 2 SD of replicate procedural standards for tHg and pHg.





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681Figure S3. Depth profile of particulate Hg stable isotope composition at three stations in the682Mediterranean during the June 2017 campaign. A: mass-dependent fractionation (δ^{202} Hg), B: odd683mass-independent fractionation (Δ^{199} Hg), C: even mass-independent fractionation (Δ^{200} Hg).

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