

A Critical Review of Life Cycle Assessment in Shrimp Aquaculture: Uncovering Methodological Dominance and Analytical Blind Spots

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1 **Abstract**

2 Life Cycle Assessment (LCA) is increasingly used to evaluate the environmental impacts of shrimp
3 aquaculture, a rapidly expanding global food sector. However, existing shrimp LCA studies report widely
4 divergent results, varying by more than fiftyfold across key impact categories. This systematic review
5 identified 16 peer-reviewed shrimp LCAs and investigates the reasons for these discrepancies, revealing
6 inconsistencies across all LCA stages, such as system boundaries (e.g., inconsistent inclusion of change and
7 pond emissions), co-product allocation methods, background data sources, and impact assessment
8 methodologies. Among the 16 reviewed studies, only five provide sufficient data for reproducibility. We
9 demonstrate that methodological choices more strongly influence LCA outcomes than actual differences
10 in shrimp farming operations. Moreover, many studies neglect critical environmental concerns such as
11 biodiversity loss, land use change and antibiotic use. To enhance LCA reliability and comparability, we
12 recommend specific methodological harmonisation, suggest reporting needs for transparency, and identify
13 priority geographic and system coverage for future LCAs. Such improvements are essential for LCA results
14 to accurately inform sustainable shrimp farming practices.

15

16 **Key words**

17 Seafood; LCA; reproducibility, sustainability; global warming

18

19 **1. Introduction**

20 The global food system is responsible for roughly one quarter of global greenhouse gas (GHG) emissions
21 (Poore and Nemecek 2018) and is the leading driver of global biodiversity loss (Ritchie et al., 2022). As the
22 world's population is growing and becoming more affluent, the demand for animal-based food products is
23 increasing rapidly, with aquaculture playing an important role in meeting this demand (Salin & Ataguba,
24 2018). As animal-based foods generally require considerably more resources than plant-based alternatives,
25 the environmental burden of the food sector is expected to increase further (Godfray et al. 2018).

26 Among animal-based foods, aquaculture – the production of aquatic organisms such as fish, crustaceans,
27 and algae in controlled environments – is the fastest-growing sector in relative terms and now exceeds
28 capture fisheries in production volume (Gentry et al., 2019). In terms of value, shrimp are the second most
29 valuable aquatic export globally, following salmonids (Thorner et al., 2020). Ecuador is the largest shrimp
30 exporter, followed by India, Viet Nam, and Indonesia in 2022. Meanwhile, China is the largest importer,
31 followed by the US, Japan, and Spain (FAOFishstatJ, 2025). Shrimp are also estimated to be the most
32 consumed animal globally in terms of number of individuals (Blaxter et al., 2024).

33 Shrimp farming comprises a variety of systems operating in diverse environments (Emerenciano et al.,
34 2022). Intensive systems dominate global production, but extensive systems remain common, often
35 occupying the fragile mangrove-fringed rim of intertidal coastal zones in intertidal areas with brackish water
36 (Maiti et al., 2021). The global production of shrimp and prawns (group 45 of the International Standard
37 Statistical Classification of Aquatic Animals and Plants, FAOFishStatJ 2025) in brackish water aquaculture
38 systems amounted to an annual average of 6.7 million tonnes in 2020-2022. Whiteleg shrimp (*Litopenaeus*
39 *vannamei*) is the dominant species, accounting for 82% of global production. This prevalence can be
40 attributed to the species' rapid growth, disease resistance, and consumer popularity (Funge-Smith and
41 Briggs, 2003). China leads whiteleg shrimp production with 21% of global production by volume, followed
42 by India (17%), Ecuador (17%), Indonesia (13%), and Viet Nam (13%). The second most produced species
43 is the Asian tiger shrimp (*Penaeus monodon*), which comprises 11% of brackish water shrimp farming during
44 the same period. For this species, the top producers are Viet Nam (37% of global production by volume),
45 Indonesia (18%), China (12%), Bangladesh (9%), and Myanmar (7%).

46 Alongside growing demand, the sector faces criticism and pressure to mitigate energy use, freshwater
47 consumption, mangrove deforestation, and pollution (Serpa & Duarte, 2008). Some of these, such as
48 eutrophication, saltwater intrusion, or loss of fish habitat, negatively impact neighbouring social and
49 ecological systems, while others contribute to global environmental concerns (such as global warming)
50 (Ahmed and Ambinakudige, 2024). Additionally, the high reliance on feed resources, such as fishmeal from
51 capture fisheries and soybean meal, threatens aquatic and terrestrial biodiversity in globally telecoupled
52 locations (Majluf et al., 2024). There is therefore a need to understand a diversity of environmental impacts
53 throughout shrimp supply chains.

54 Life Cycle Assessment (LCA) is a framework used to quantify various environmental impacts of a product
55 or service and scale them to a functional unit. It is increasingly being adopted by policymakers, most notably
56 in the European Union (EU) (Sala et al., 2021), as a key tool in driving sustainability transitions. However,
57 LCA results for various food products reveal substantial discrepancies in environmental impacts on the
58 farm level; estimated impacts can vary 50-fold among producers of the same product (Poore and Nemecek,
59 2018). Many of these differences relate to methodological choices (Henriksson et al., 2012; Bohnes et al.,
60 2019) that, in theory, could be harmonised. Such ambitions have, for example, been initiated by the
61 International Organisation for Standardisation (ISO), with ISO14040 and ISO14044 providing guidelines
62 for conducting LCAs. These standards seek to promote reliable and transparent LCA results — qualities
63 essential for scientific reproducibility, informed decision-making, and credible sustainability reporting.
64 Scientific reproducibility, as defined by Popper (1959), could be translated to LCA in terms of the
65 documented methodology and data values providing sufficient information for an independent practitioner
66 to reproduce the LCA results. However, studies across multiple sectors have highlighted that poor
67 documentation on unit process data, poorly documented system boundaries, and insufficient reporting of
68 key methodological choices often compromise LCA reproducibility (Talon 2016; Dieterle et al., 2022; Philis
69 et al., 2019). In addition to generic ISO standards for LCA, the EU's Product Environmental Footprint
70 Category Rules (PEFCR) are intended to provide sector-specific guidelines on how to conduct LCAs. While
71 there is a PEFCR for Unprocessed Marine Fish Products, no PEF standards exist for crustaceans as of now
72 (The Marine Fish PEFCR project, 2025; Pedersen and Remmen, 2022).

73 This review builds on previous work by Henriksson et al. (2012) and Bohnes and Laurent (2019) to offer a
74 more pointed and evidence-based critique. We move beyond simply identifying discrepancies to
75 demonstrate that methodological inconsistency is the dominant driver of reported environmental impacts,
76 often outweighing the influence of on-farm practices. This finding has profound implications, suggesting
77 that the current body of literature is an unreliable guide for policy, certification, and improvement efforts.
78 To develop this argument, this paper is structured thematically. First, we provide an overview of the
79 fragmented landscape of shrimp LCA research. Next, we critically deconstruct how methodological choices
80 systematically shape LCA outcomes. We then turn to the field's analytical blind spots, examining the critical

81 environmental pressures that are consistently neglected. We conclude by offering a clear set of actionable
 82 recommendations for methodological harmonisation to build a more robust and representative science.

83

84 2. Methods

85 2.1 Review Protocol and Scope

86 We conducted this systematic review following the Preferred Reporting Items for Systematic Reviews and
 87 Meta-Analyses (PRISMA) guidelines (figure 1). The primary aim was to identify and critically assess all
 88 relevant peer-reviewed LCA studies concerning the aquaculture of whiteleg shrimp (*Litopenaeus vannamei*)
 89 and giant tiger prawn (*Penaeus monodon*). These two species were prioritised as they are the most widely
 90 farmed shrimp species globally and predominantly cultivated in brackish water systems within tropical
 91 coastal regions. This focus was chosen because the context and environmental consequences of these
 92 systems differ from freshwater aquaculture operations, such as those for the giant river prawn
 93 (*Macrobrachium rosenbergii*).

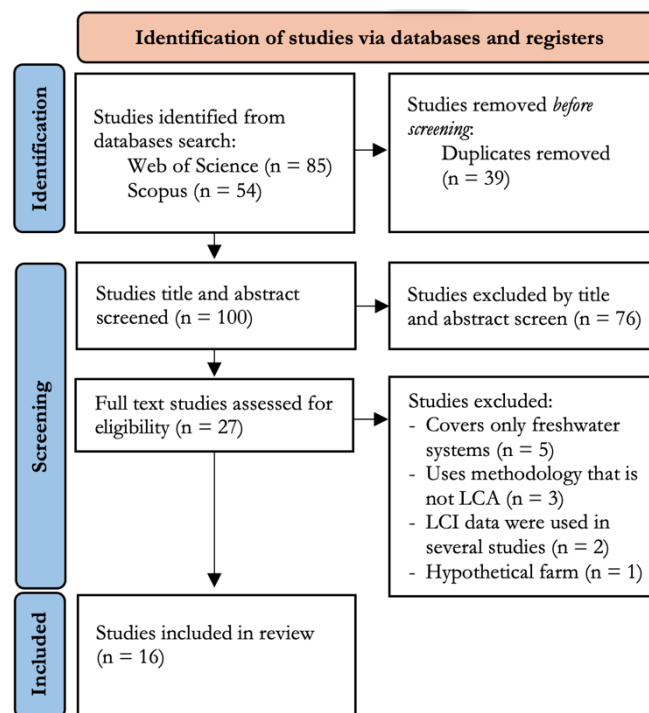


Figure 1 PRISMA flowchart, showing the criteria for inclusion in the review, and the narrowing down from 139 initial studies to 16 included in the review.

94 **2.2 Literature Search and Study Selection**

95 We employed an iterative process to develop a comprehensive literature search strategy designed to ensure
96 inclusivity while maintaining specificity. The final search string applied to the Scopus and Web of Science
97 databases was: (shrimp OR prawn) AND (aquaculture OR farming OR production) AND (LCA OR "life
98 cycle assessment" OR "life cycle analysis"). This search was conducted without filters or date restrictions
99 to maximise coverage and was finalised on January 19, 2024. Additionally, the reference lists of all identified
100 relevant articles were manually screened for further pertinent studies; this process yielded no new records.
101 Studies were included if they were peer-reviewed, applied LCA methodology to assess environmental
102 impacts, and focused on either whiteleg shrimp or giant tiger prawn aquaculture in brackish water systems.
103 Studies were excluded if they: (i) covered only freshwater systems; (ii) used a methodology that was not
104 LCA; (iii) used primary data that was also used in several other studies (to avoid duplication of datasets); or
105 (iv) assessed a purely hypothetical farm. The initial database search yielded 85 records from Web of Science
106 and 54 from Scopus, which, after removal of 39 duplicates, resulted in 100 unique records for screening
107 (figure 1). Title and abstract screening led to the exclusion of 76 records. The remaining 27 full-text articles
108 were assessed for eligibility, from which 16 studies were eligible for this review.

109 **2.3 Data Extraction and Synthesis**

110 From the 16 selected studies, a total of 41 production "cycles" were initially identified. A "cycle" is defined
111 as an LCA conducted for a unique dataset of inputs, emissions, products, and practices specific to a
112 particular farming system, intensity, species, or geographical context within a study. For example, study 15
113 examines three distinct farming cycles in China using recirculating aquaculture systems (RAS), biofloc
114 technology (BFI), and high-performance ponds (HPP).

115 For studies where primary data collection was supplemented with data from existing studies (e.g., studies 1
116 and 10), only those cycles based on the primary data collected by the respective authors were included in
117 our dataset to avoid pseudo replicates. One cycle from study 8 was excluded because it combined giant
118 tiger prawn and giant river prawn, making it difficult to isolate the impacts relevant to this review's scope.
119 After these refinements, a final dataset of 37 distinct production cycles was analysed.

120 For each included cycle, detailed information was extracted (where reported) pertaining to general context
121 and the four LCA phases outlined in ISO 14040/14044 (goal and scope definition, life cycle inventory
122 (LCI), life cycle inventory analysis (LCIA), and interpretation). All quantitative data were extracted and,
123 where necessary, harmonised to a common functional unit of one tonne of liveweight shrimp at farmgate
124 to facilitate comparison and subsequent correlation analysis. Detailed data for each of the 37 cycles are
125 provided in the Supplementary Material (SM).

126 **2.4 Analysis of Methodological Choices, Reproducibility, and Input-Impact Relationships**

127 To investigate the influence of methodological choices, data on on-farm energy use and Feed Conversion
128 Ratios (FCRs, defined as the weight of feed given divided by the weight gained; Fry et al., 2018), were
129 compared against global warming and eutrophication impact results for each cycle. Different energy inputs
130 such as electricity and diesel were standardised to megajoules, while acknowledging that this approach does
131 not account for conversion efficiency differences between energy carriers (Frischknecht et al., 2015). For
132 studies applying multiple allocation methods, we used economic allocation as this is the most common
133 allocation method (SM). Due to inconsistent reporting, other relevant factors (e.g., water consumption,
134 land occupation, chemical inputs, stocking density, and field emissions) could not be evaluated. The
135 relationships between inputs and environmental impacts of each cycle were analysed through a correlation
136 analysis.

137 To quantify the influence of methodological choices versus different farming practices, coefficients of
138 variation (CV) for global warming (GW) and eutrophication impacts were calculated for nine distinct
139 shrimp farming cycles from one study that employed a consistent methodology (Henriksson et al., 2015a).
140 These CVs were later contrasted against the percentage change in impacts observed in three identified
141 instances where identical farm-level inventory data were re-analysed using different LCA approaches (Al
142 Eissa et al., 2022; Jonell and Henriksson, 2015)).

143 **2.5 Limitations of the Review Methodology**

144 This systematic review has certain limitations primarily related to the literature search process. Firstly, the
145 search was confined to two major academic databases: Scopus and Web of Science. While these databases
146 provide extensive coverage of peer-reviewed literature, relevant studies indexed exclusively in other

147 specialised or regional databases might not have been captured. Similarly, the review was restricted to
 148 English-language publications. This means that pertinent research published in other languages would have
 149 been excluded, potentially limiting the geographical or contextual scope of the findings if significant non-
 150 English literature exists on this topic. However, an informal search of the same terms in Spanish,
 151 Portuguese, Mandarin, and Hindi did not reveal any relevant studies that fulfil the search requirements.

152 Due to the cut-off date in January 2024, the most recent studies covering novel systems, such as Arbor et
 153 al. (2024) looking at microalgae-based wastewater treatments and Sun et al. (2025) identifying tunnel
 154 greenhouse aquaculture systems, are not included in this review.

155

156 3. The Landscape of Shrimp LCA Research

157 The 16 studies under review were published between 2006 and 2023. For the twelve studies that detailed
 158 primary data collection dates, the median time between data collection and publication was four years. The
 159 average year of data collection across these eleven studies was 2013. This temporal gap draws into question
 160 the relevance of some findings to current shrimp farming practices, which have evolved significantly due
 161 to growing demand, disease outbreaks, improved farm management strategies, and technological
 162 innovations. The COVID-19 pandemic, for example, prompted a shift towards more efficient systems due
 163 to input and labour shortages (Nguyen et al., 2024).

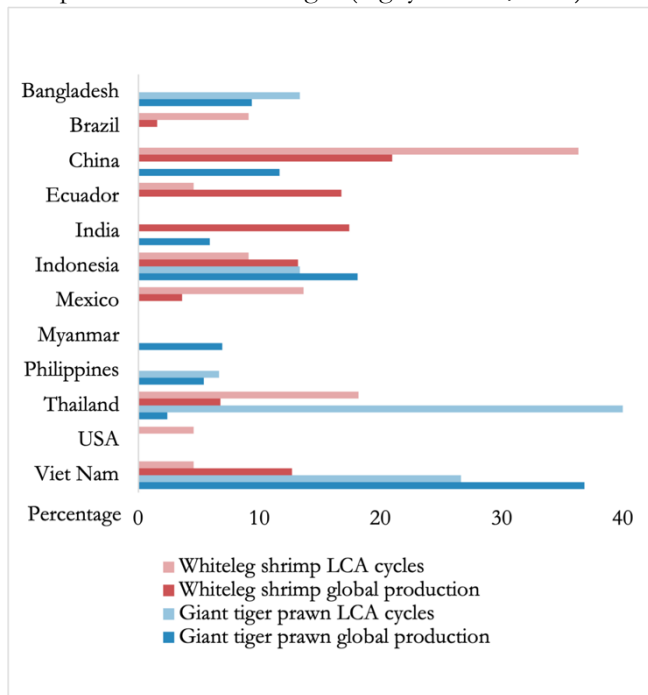


Figure 2 Percentage comparison of number of LCA cycles and global shrimp aquaculture in brackish water. Production data from FAO FishStatJ (2025), where the average of the three most recent available years (2020-2022) was used. Annual whiteleg shrimp production had an average of 5.49 million tonnes and giant tiger prawn 723 thousand tonnes.

164 The 37 analysed production cycles exhibit considerable heterogeneity. Eight cycles are based on data
 165 representing a single farm, while other cycles represent horizontally averaged data from up to 106 farms
 166 (SM). Of the 37 cycles, 30 assess monoculture systems and seven polyculture systems. Twenty-two cycles
 167 evaluate whiteleg shrimp (*Litopenaeus vannamei*) and 15 evaluate giant tiger prawn (*Penaeus monodon*).

168 A key distinction among the reviewed literature is its comparative nature. Of the 16 studies, ten are internally
 169 comparative, meaning they assess multiple distinct production cycles using a consistent internal
 170 methodology. This structure is critical as it allows for the isolation of impacts due to farming practices from
 171 those due to methodological choices. The remaining six studies each assess a single production system.

172 Eight studies define farming intensity (e.g., extensive, intensive) but fail to clarify the specific criteria for
 173 these classifications, highlighting the lack of harmonised definitions (Oddson 2020). The geographical focus
 174 of the reviewed literature is also misaligned with current global production locations, especially Ecuador
 175 (16.7% of global whiteleg shrimp production; one LCA, study 14) and India (17.4% of global whiteleg
 176 shrimp production, no shrimp LCA study, but one LCA study on shrimp feed; Ramesh et al., 2024; figure
 177 2). Thailand, on the other hand, is nowadays only responsible for 2.4% of giant tiger prawn and 6.8% of
 178 whiteleg shrimp production and had the highest representation amongst all countries in exiting LCA studies
 179 (ibid.). Among the reviewed studies, 11 claimed adherence to the ISO 14040 and 14044 standards, with
 180 study 5 self-defining as an LCA but referencing ISO 14067 for carbon footprinting. The following sections
 181 systematically evaluate how each study adheres to the stages outlined in ISO 14040/14044, and which
 182 methodological choices were made.

Study no.	Authors	Year	Title of Study
1	Al Eissa et al.	2022	Effects of feed formula and farming system on the environmental performance of shrimp production chain from a life cycle perspective
2	Aubin et al.	2014	Environmental performance of brackish water polyculture system from a life cycle perspective: A Filipino case study
3	Beletini et al.	2018	Carbon footprint in commercial cultivation of marine shrimp: A case study in southern Brazil
4	Cao et al.	2011	Life cycle assessment of Chinese shrimp farming systems targeted for export and domestic sales
5	Chang et al.	2017	Carbon footprint analysis in the aquaculture industry: Assessment of an ecological shrimp farm
6	Cortés et al.	2021	Eco-efficiency assessment of shrimp aquaculture production in Mexico
7	Flores-Pérez et al.	2023	Eco-efficiency assessment of disease-infected shrimp farming in Mexico using environmental impact assessment tools

8	Henriksson et al.	2015	Comparison of Asian Aquaculture Products by Use of Statistically Supported Life Cycle Assessment
9	Henriksson et al.	2017	Indonesian aquaculture futures – Evaluating environmental and socioeconomic potentials and limitations
10	Jonell et al.	2015	Mangrove-shrimp farms in Vietnam-Comparing organic and conventional systems using life cycle assessment
11	Koniyo et al.	2022	Role of Innovations / Interventions to Bring Sustainability in Aquaculture Growth in Indonesia: Integration of Life Cycle Assessment (LCA) Framework
12	Lebel et al.	2010	Innovation cycles, niches and sustainability in the shrimp aquaculture industry in Thailand
13	Mungkung et al.	2006	Potentials and Limitations of Life Cycle Assessment in Setting Ecolabelling Criteria: A Case Study of Thai Shrimp Aquaculture Product
14	Sanchez et al.	2023	Life Cycle Analysis of Farmed Shrimp of the Species <i>Litopenaeus Vannamei</i> in the Province of Guayas
15	Sun et al.	2023	Comparative life cycle assessment of whiteleg shrimp (<i>Penaeus vannamei</i>) cultured in recirculating aquaculture systems (RAS), biofloc technology (BFT) and higher-place ponds (HPP) farming systems in China
16	Tantipanati p et al.	2014	Life cycle assessment of pacific white shrimp (<i>penaeus vannamei</i>) farming system in trang province, Thailand

183 *Table 1: Overview of reviewed shrimp LCA studies. More information can be found in the supplementary material (SM).*

184

185 **4. Deconstruction and Reproducibility Analysis of Shrimp LCAs**

186 **4.1 Goal and Scope Definition**

187 **4.1.1 Study Goals**

188 According to the ISO 14044 standard, an LCA must begin with a clear statement of its objectives and the
189 rationale for the assessment. All reviewed studies adhere to this requirement, defining a wide array of goals.

190 In general, these objectives focused on conducting comparative assessments between different farming
191 systems, species, or geographies; identifying environmental hotspots within specific systems; or evaluating
192 the impacts of targeted scenarios such as the use of innovations, different feeds, polyculture practices, or
193 the effects of disease outbreaks (SM).

194 **4.1.2 Functional Units**

195 The functional unit (FU) is the unit of reference to which all environmental impacts are scaled. All studies
196 under review use a mass-based functional unit (FU) at farmgate, a rare point of consensus. The FU was

197 defined as either 1 kilogram (four studies) or 1 tonne (12 studies) of shrimp at the farmgate. Five studies
198 also included supplementary FUs for processed products to meet specific study goals covering the broader
199 value chain (SM). For this review, we harmonised all impact results to one tonne of liveweight shrimp at
200 the farmgate to facilitate comparison.

201 **4.1.3 System Boundaries**

202 The system boundary defines which unit processes and emissions are to be included in the LCA study. All
203 studies under review include the grow-out stage, in which post-larvae shrimp are raised to market size.
204 However, only four studies explicitly include infrastructure and 13 include transport (SM).

205 The treatment of land use and land use change (LULUC) was a major inconsistency and a key driver of
206 discrepancies. Only studies 10 and 11 quantified farm-level LULUC emissions from mangrove conversion,
207 but their differing methodologies and resulting impacts highlight the problem. In study 10, the direct
208 calculation of LULUC was responsible for 94% of the system's GW impacts, making it one of the highest
209 outlier in the dataset (SM). In contrast, study 11 incorporated LULUC by applying a pre-calculated emission
210 factor from existing literature, which resulted in GW impacts that were comparable to other studies that
211 did not include farm-level LULUC. This demonstrates how the specific methodological choice for
212 quantifying LULUC can have a more significant effect on the result than the decision to include it in the
213 first place.

214 The treatment of LULUC associated with feed ingredients was more opaque as several LCI background
215 databases (e.g., ecoinvent) used in the studies account for LULUC for some crops, while only studies 1 and
216 8 detailed the assumed origins of feed ingredients. This is particularly relevant for ingredients sourced from
217 regions like Brazil and Argentina, where soybean farming is associated with high levels of LULUC. This
218 inconsistent inclusion of LULUC represents a major driver behind discrepancies in reported impacts.

219 **4.1.4 Coproduct Allocation Methods**

220 Coproduct allocation refers to how environmental burdens are divided among multiple products originating
221 from the same unit process, or among multiple uses of one product. Only half of the studies explicitly
222 specify their coproduct allocation method, thus failing to comply with ISO 14044. Among those that did,
223 economic allocation was the most common adopted (six studies), followed by mass (three studies) and

224 energy (one study) (SM). Three studies applied multiple allocation methods, providing direct insight into
225 the influence of this choice (SM). The results showed that for the exact same farm-level inventory data, the
226 choice of allocation method could alter GW impacts by up to 58% (study 9) and eutrophication impacts by
227 up to 59% (study 8). Critically, there was no predictable pattern where one allocation method consistently
228 produced higher or lower results, demonstrating the unpredictable influence of this methodological choice.

229 **4.1.5 Assumptions**

230 LCA studies are data intensive and therefore often rely upon assumptions to fill data gaps and/or solve
231 unknown fates and origins. These assumptions are another major driver of discrepancies, yet only eleven
232 studies provide detailed documentation (SM). The profound influence of these choices is also demonstrated
233 through sensitivity analyses. For instance, study 1 assumed all its soybean meal originated from the U.S.,
234 while a sensitivity analysis reveals that sourcing from Argentina or Brazil would increase the associated GW
235 impacts by 1240% or 960%, respectively. These discrepancies are primarily due to LULUC. Similarly, study
236 4 reports that assumptions about electricity mix is highly influential, with a switch from coal to hydropower
237 or nuclear energy having the potential to reduce GW impacts of farmed shrimp by 25-50%. These examples
238 show that assumptions can have fundamental influence on LCA conclusions.

239 **4.1.6 Impact Assessment Methodologies and Categories**

240 To classify and characterise environmental emissions and resource uses towards specific environmental
241 impact categories, different impact assessment methodologies are used. CML methodology was applied in
242 nine studies, while ReCiPe was applied in three. One study applied foundational models and methods, and
243 another study ISO/TS 14067 and PAS 2050 (SM). Different impact assessment methodologies use different
244 cause-effect pathways and units to quantify how emissions and resource use contribute to specific impact
245 categories. Studies 4 and 13 compare impact results for different methodical choices. Study 4, which applies
246 the CML-IA Baseline (Guinée 2002) finds comparable outcomes for GW impacts and terrestrial
247 acidification, but lower eutrophication estimates under IMPACT 2002+. Discrepancies also arise from
248 different versions of the Intergovernmental Panel on Climate Change' (IPCC) (Kikstra et al., 2022)
249 Assessment Reports (AR). For example, the global warming potential over 100 years (GWP100) for
250 methane increased from 25 in AR4 to 27 in AR6. Meanwhile, the characterisation factors for different

251 freshwater ecotoxicity impacts can differ with orders of magnitudes depending upon the underlying data
252 (Nyberg et al. 2024). Among the reviewed studies, study 8 calculated specific freshwater ecotoxicity factors
253 using the USEtox model (Rosenbaum et al., 2008).

254 The number of impact categories assessed ranged from none to eleven, with an average of four. Most
255 assessments included global warming (13 studies), eutrophication (eleven studies), and terrestrial
256 acidification (nine studies). As shown in figure 6, ten different impact categories only appeared once,
257 suggesting a fragmented picture of the full environmental performance of shrimp across the LCA studies.
258 Noteworthy is that no study evaluated endpoint impacts (e.g., damage to human health or ecosystems)
259 (SM).

260 **4.1.7 Modelling Approaches**

261 A critical omission was the failure to declare the specific LCA modelling approach; attributional or
262 consequential. Attributional LCA is a methodology that quantifies the environmental impacts associated
263 with the lifecycle of a product or service, attributing all emissions and resource extractions directly to the
264 product or service being studied, and is typically used for reporting past impacts or comparing
265 environmental performances and identifying critical impact areas. In contrast, consequential LCA assesses
266 the environmental impacts of a decision by modelling the changes in the entire product system, including
267 market interactions and marginal effects. In this review, only two studies (2 and 10) explicitly state they use
268 an attributional methodology, while study 15 self-identifies as using a consequential LCA approach. For the
269 remaining twelve studies, the specific LCA framework is not explicitly stated, a fundamental issue as
270 attributional and consequential approaches are generally not comparable.

271 **4.2 Life Cycle Inventory (LCI)**

272 **4.2.1 Primary Data**

273 All studies use primary data for the grow-out stage (these have been extracted and harmonised to the same
274 FU in the SM), which was a requirement for inclusion in our review, but demonstrate significant
275 inconsistencies in their primary data sampling methods and documentation, ranging from detailed case
276 studies of single farms to broader, multi-farm surveys of up to 106 farms per cycle. Ten cycles relied on
277 individual farms for data collection. Other studies employed what they described as "representative"

278 without further details. More robust approaches involved random sampling designs of farm clusters for
279 large datasets, as seen in a study collecting data from up to 100 farms per cycle across four Asian countries
280 (study 8), or using random sample size determination to select 106 farms in Thailand (study 16). Some
281 studies adopted targeted sampling strategies, such as selecting 76 commercial farms specifically affected by
282 the white spot syndrome virus in Mexico (study 7). Data collection primarily involved on-site interviews
283 and questionnaires filled out by farm owners, sometimes drawing from existing governmental databases or
284 previous studies and interviews spanning several years.

285 These diverse sampling methodologies, particularly the use of single-farm data or inadequately defined
286 "representative" samples, can significantly introduce bias and limit the generalisability of reported
287 environmental impacts across the broader shrimp aquaculture sector. Certain sampling methods potentially
288 prioritise better-managed farms with good records, which potentially result in underestimated sector-wide
289 impacts. These differences in sampling strategy mean that observed variations in environmental impacts
290 may be artefacts of the sampling method rather than true differences in farming performance. Fifteen
291 studies provided geographical specificity at least to the provincial level, with some detailing exact farm
292 locations. Three studies failed to specify data collection years, creating temporal ambiguity.

293 **4.2.2 Secondary Data**

294 The reviewed studies relied on a diverse array of secondary sources, including published literature,
295 government reports, online resources, and structured LCI databases. Details on the databases and versions
296 of these used are available in the SM. Eight studies utilised the ecoinvent database in some capacity,
297 including v2.2, v3.0, v3.01, and v3.7.1, while one study did not specify the version employed. Four studies
298 relied solely on ecoinvent, while the four others used it in combination with other LCI databases, such as
299 Agri-footprint database, LCA Food database, and national government databases. For instance, studies
300 supplemented global databases with local data for aspects like electricity mixes, local emission factors, and
301 specific farming conditions. Examples include study 3, which sourced electricity mix data from Empresa
302 de Pesquisa Energética (EPE), a Brazilian national energy research company and study 4 adapted secondary
303 data to Chinese regional conditions, referencing a publication by the Chemical Industry Press. This

304 demonstrates the necessity of integrating more specific local data to enhance the accuracy and regional
305 representativeness of life cycle assessments.

306 Three studies simply referred to the LCI databases within different versions of SimaPro (SM). Studies 12
307 and 16 did not specify background LCI databases used (SM).

308 Inconsistent data sourcing introduces discrepancies in LCA outcomes. The predominant European or
309 North American origin of many LCI databases (Henriksson et al., 2014) poses specific challenges for shrimp
310 production that is mainly conducted in Asia and Latin America, where regional conditions can differ
311 substantially from database defaults (Ossés de Eicker et al., 2010).

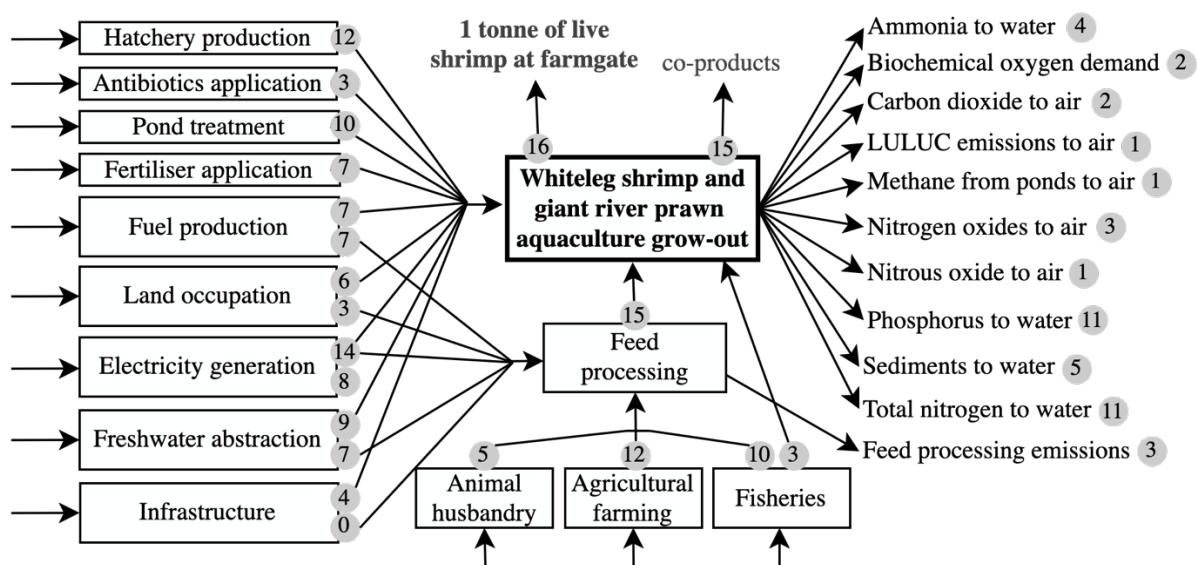
312 **4.3.3 Unit Process Data for Grow Out Cycles and Feed Mills**

313 Unit process data for the grow-out cycles represent the quantified inputs and outputs associated with one
314 cycle. While most studies detailed inputs and outputs in total units, studies 3 and 5 only reported inputs and
315 outputs in terms of associated CO₂-equivalent, making it impossible to reproduce the results of these
316 studies. Of the remaining 14 studies, all reported energy and feed use, but other critical inputs were
317 inconsistently documented (figure 4; SM). Freshwater inputs were reported in nine studies and land
318 occupation in six. Chemical treatments for water and pond soil such as chlorine, calcium carbonate, and
319 limestone were documented in ten studies, while fertilisers and productivity enhancement inputs, such as
320 urea and manure, appeared in seven studies. Notably, only one study reported types and amounts of
321 antibiotic use, while two studies explicitly stated that no antibiotics were used (SM).

322 The intensification of shrimp farming has shifted land occupation and its associated impacts from the farm-
323 level to the feed production level (Davis et al., 2021; Henriksson et al., 2018; Froehlich et al., 2018). Study
324 10, for example, where data were collected in an extensive system in 2010, reported up to 4.4 hectares of
325 land use without any external feed inputs, while study 1, which was published in 2022, documented only 84
326 m² of land use and 1.5 tonnes of feed for the same functional unit of one tonne of shrimp at the farmgate.
327 Of the 16 examined studies, 14 reported the use of feed pellets, with 13 of these quantifying the total
328 amounts used. Three studies reported supplementary feed inputs alongside pelleted feeds, including lower-
329 value fish and rice bran. In study 2, only molluscs were employed as a feed input, while no feed inputs were
330 applied in study 10. Four studies lacked documentation of feed ingredients, and two studies relied on feed

331 formulas from previously published research from different contexts. The remaining eight studies provided
 332 primary data on feed ingredients and their quantities. Within this subset of eight studies, five documented
 333 water consumption associated with pellet production and six reported energy use data (SM). Only study 8
 334 detailed the geographical origins of feed ingredients, while study 1 made assumptions about ingredient
 335 origins. Feed compositions vary, with fishmeal comprising 20-42% of pellet ingredients and soybean meal
 336 11-30%. Feed Conversion Ratios (FCRs) of monoculture cycles with pellet inputs ranged from 1.0 to 3.6.
 337 Higher FCRs are caused by the addition of less nutritious feeds, such as rice bran.

338 While all studies report product outputs at the farm gate and co-products from polyculture systems, the
 339 documentation of emissions and waste varied considerably (figure 4; SM). Total emissions of nitrogen and
 340 phosphorus were reported by eleven studies. Other emissions (figure 3) were reported by less than half of
 341 the studies, despite the impact they can have on the LCIA results, as seen in the case of the inclusion of
 342 LULUC emissions in study 10. Emissions from feed processing plants were addressed in three studies
 343 (figure 4; SM).



344

Figure 3: Inputs and outputs to and from feed processing and shrimp grow-out. Unit processes are represented by boxes and flows by arrows. Circled numbers indicate how many of the 16 studies that detail primary data, including zeroes (e. g. no feed applied and therefore zero energy use for feed production).

345

346 4.4 Life Cycle Impact Assessment (LCIA)

347 The multiplication of varying set of lifecycle inventory results with an accumulation of methodological
348 inconsistencies detailed in the preceding sections explains the divergence LCIA results. By scaling impact
349 assessment results to a functional unit of one tonne of liveweight shrimp at the farmgate, the results from
350 the different studies can be compared (figure 4, SM):

- 351 • GW impacts (reported in 33 cycles) ranged from
352 901 kg to 47,997 kg of carbon dioxide
353 equivalent (CO₂-eq) per tonne of shrimp, with a
354 standard deviation of 7,968 kg CO₂-eq t⁻¹
355 shrimp.
- 356 • Eutrophication results (reported in 28 cycles)
357 ranged from -32 to 160 kilograms of phosphate
358 equivalent (PO₄-eq.) per tonne of shrimp, with
359 a standard deviation of 52 kg PO₄-eq. t⁻¹ shrimp.
- 360 • Terrestrial acidification (reported in twenty
361 cycles) ranged from 4 to 89 kilograms of sulphur
362 dioxide equivalent (SO₂-eq.) per tonne of
363 shrimp, with a standard deviation of 27 kg SO₂-
364 eq. t⁻¹ shrimp.

365 Eutrophication showed the greatest spread, terrestrial
366 acidification displayed moderate spread, while GW
367 demonstrated the lowest spread, but with two outliers (SM,
368 figure 4). The relatively low spread in GW may be attributed
369 to more harmonised emissions models and characterisation factors (section 3.5.2).

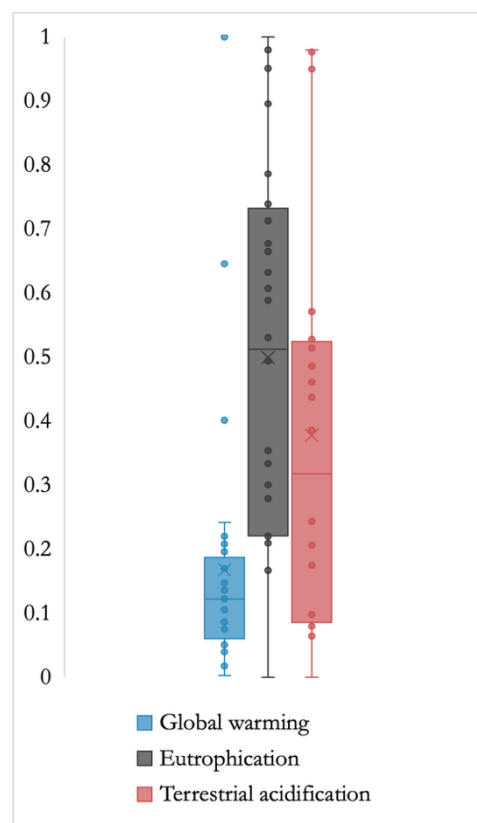


Figure 4 Normalised LCA results from all reviewed cycles, with the lowest reported value among impact results being 0 and highest being 1. Boxes represent the interquartile range with median (line) and mean ('X'). Whiskers extend to 1.5x IQR, showing individual data points and outliers.

370 4.5 Interpretation

371 4.5.1 Completeness and Consistency Analysis

372 The interpretation phase of an LCA requires checks to ensure all relevant information is included
373 (completeness) and that the methodology aligns with the study's goals (consistency). These checks,
374 mandated by ISO 14044, are essential for validating results. However, they were almost entirely absent from
375 the reviewed literature. Only studies 4 and 8 conducted a consistency check, and none of the 16 studies
376 performed an explicit completeness check as defined by ISO 14044 (Henriksen et al., 2019; Dong and Liu,
377 2022).

378 4.5.2 Uncertainty and Sensitivity Analyses

379 Incorporating uncertainty ranges enhances the robustness of LCA outcomes by accounting for error and
380 discrepancies in unit process data, emission models, and characterisation factors (Ziyadi and Al-Qadi, 2019;
381 Heijungs 2024). Six studies used Monte Carlo simulations to propagate uncertainties among parameters
382 (SM), showing that shrimp LCAs can result in very high variation in impacts: Study 8 shows that the GW
383 results of black tiger shrimp production in Eastern Bangladesh could range from a minimum of 1260 to a
384 maximum of 108,000 kg CO₂-eq. t⁻¹ frozen peeled tail-on giant tiger prawns at European ports due to
385 uncertainties in unit process data and characterisation factors.

386 Seven studies include sensitivity analyses (SM) to identify key contributing variables and improve the
387 reliability of results (Guo and Murphy, 2012). These analyses explicitly tested aspects including feed
388 compositions (such as fishmeal content and the origin of ingredients, study 1, allocation methods (studies
389 1; 8-10), FCR and impact assessment methodologies (study 4), and pond size and production-site distance
390 from the sea (study 2). For study 10, the sensitivity analysis revealed that carbon loss assumptions during
391 mangrove transformation strongly influenced results, with a 64% reduction in GW impacts when using
392 conservative estimates (25% carbon loss) and an 87% increase when assuming complete carbon loss. Study
393 4 investigated how shifting the Chinese electricity mix from coal-dominated to less CO₂-intensive
394 alternatives (such as natural gas, nuclear, or hydropower) would affect global warming, showing potential
395 reductions of 25-50%. These examples show how methodological decisions and background data can
396 overshadow actual farming practice differences in determining environmental performance outcomes.

397 **4.5.3 Conclusions, Limitations, and Recommendations**

398 Regarding study findings, studies that included a broader range of metrics found that aspects like LULUC
399 and chemical applications substantially influenced environmental profiles. The most common
400 recommendations covered changes in feed production and application (nine studies), such as lowering the
401 FCR and reducing fishmeal. Eight studies recommended the optimisation of energy consumption or use
402 of renewable energy and energy conserving technologies. Changes in wastewater and nutrient management
403 Changes in wastewater discharge and recycling of excessive nutrients were recommended by six studies.

404 Only studies 8-11, 13 and 15 explicitly acknowledged limitations in their methodologies, such as the lack of
405 inadequate region-specific data (study 15). Four studies also recommended methodological improvements
406 for LCA practitioner including: combining quantitative LCA with qualitative "hurdle criteria" to address
407 impacts not captured by traditional metrics (study 13); adopting statistically supported approaches to
408 quantify data uncertainty (study 8); integrating spatiotemporal considerations (study 10); and expanding
409 data on LULUC emissions (study 9).

410 **4.6 Reproducibility**

411 This review assessed reproducibility based on the transparency of calculation methodologies,
412 acknowledging that true reproducibility would also require documentation of primary data collection
413 processes, such as providing surveys — a level of transparency lacking in all reviewed studies. While all
414 studies clearly documented FUs and system boundaries, and 13 documented primary data origins,
415 significant reporting gaps existed across other fundamental aspects. Only half of the studies reported their
416 allocation methods, nine detailed their underlying assumptions, and only seven provided emission models
417 sufficient for replication. Furthermore, two studies described themselves as LCAs without conducting any
418 impact assessment, and two only reported aggregated CO₂-eq., making independent verification impossible.

419 The quantitative assessment (table 2) reveals a stark picture: only five of the 16 studies fulfilled all eleven
420 criteria for complete reproducibility, while two studies only lacked one aspect. Lack of transparency
421 hampers the reproducibility of study 3, which reports the highest GW discrepancies, casting doubt on its
422 findings, particularly given the apparent absence of LULUC accounting. Conversely, the value of
423 transparency is highlighted by study 13. Its exemplary documentation allowed for the identification of a

424 detectable allocation error, where trawling impacts were overestimated by attributing 94.63% to broodstock
 425 instead of the correct ~57%. This error was only identifiable because of the study's transparency, proving
 426 that proper documentation enables critical evaluation and scientific self-correction. These findings align
 427 with broader challenges in LCA, where methodological inconsistencies, documentation gaps, and restricted
 428 access to proprietary data are recognised barriers to reproducibility (Dolan & Heath, 2012; Dieterle et al.,
 429 2022; Vafi and Brandt, 2014).

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
Functional units	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue
System boundaries	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue
Allocation	Blue	Blue	Red	Blue	Red	Red	Red	Blue	Blue	Blue	Blue	Red	Blue	Red	Red	Red
Primary data collection	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Red	Blue	Red	Red	Blue
Unit process data	Blue	Blue	Red	Blue	Red	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue
Background data documentation	Blue	Blue	Red	Blue	Blue	Red	Blue	Blue	Blue	Blue	Blue	Red	Blue	Blue	Blue	Red
Clearly stated assumptions	Blue	Blue	Blue	Blue	Blue	Blue	Red	Blue	Blue	Blue	Red	Red	Blue	Red	Blue	Red
Farm level emissions	Blue	Red	Blue	Blue	Red	Blue	Blue	Blue	Blue	Blue	Red	Red	Blue	Blue	Blue	Red
Emission models	Red	Blue	Red	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Red	Red	Blue	Red	Red	Red
Impact categories	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Red	Blue	Red	Blue	Red
Characterisation	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Blue	Red	Red	Blue	Red	Blue	Red
Reproducibility score (criteria met / total criteria)	$\frac{10}{11}$	$\frac{10}{11}$	$\frac{6}{11}$	$\frac{11}{11}$	$\frac{8}{11}$	$\frac{8}{11}$	$\frac{9}{11}$	$\frac{11}{11}$	$\frac{11}{11}$	$\frac{11}{11}$	$\frac{7}{11}$	$\frac{3}{11}$	$\frac{11}{11}$	$\frac{5}{11}$	$\frac{8}{11}$	$\frac{4}{11}$

430 *Table 2: Completeness, transparency and reproducibility evaluation of eleven key aspects among the reviewed shrimp LCAs. Blue means*
 431 *data are reported. Red means data are not transparently reported.*

432

433 5. The Consequences of Inconsistency

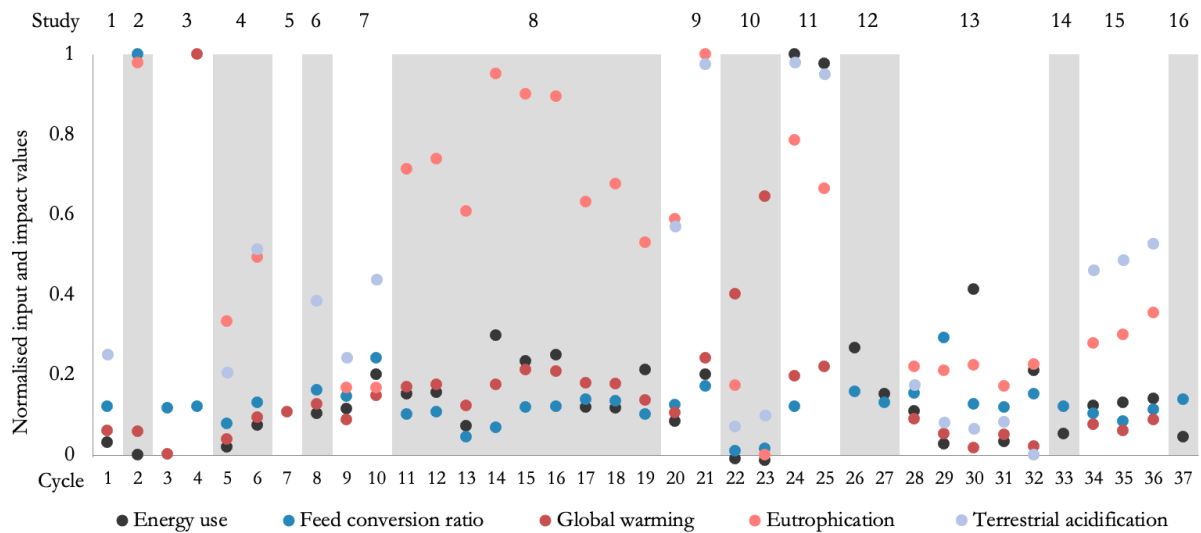
434 It is crucial to emphasise that LCA is a relative, not an absolute, measure of environmental impact
 435 (Henriksson et al., 2015b), meaning that LCA provides comparative insights rather than definitive totals,
 436 and its primary strength lies in comparing different systems or identifying relative environmental hotspots
 437 within a consistent methodological framework. This review substantiates earlier findings from broader
 438 aquaculture LCA reviews (Bohnes and Laurent, 2019; Henriksson et al., 2012), revealing that the current

439 body of shrimp LCA literature is defined by deep methodological inconsistencies that limit its utility and
440 comparability. This is not to undermine the individual strengths of certain studies under review, which may
441 have had specific aims unrelated to comparability; rather, it is an observation about the field as a whole,
442 and therefore the ability to draw generalised conclusions about the environmental impacts of this type of
443 shrimp aquaculture system.

444 445 **5.1 Correlation Analysis of Key Inputs and Environmental Impacts**

446 Shrimp LCAs consistently name feed production and on-farm energy consumption as primary sources of
447 environmental impact (Pazmiño et al., 2024). While intensification can improve resource-use efficiency
448 (Tamariska et al., 2024; Davis et al., 2021; Henriksson et al., 2018), our analysis reveals that pervasive
449 methodological inconsistencies obscure these expected relationships. We found no correlation across
450 studies between GW impacts and on-farm energy use ($r=0.04$, $R^2=0.0016$, $n=30$) and only a weak negative
451 correlation with FCR ($r=-0.21$, $R^2=0.043$, $n=33$). Excluding study 10, which included LULUC and
452 therefore had some of the highest GW results, strengthened the correlation between GW and energy use
453 tenfold (from $r=0.04$ to $r=0.40$, $n=28$), suggesting methodological noise is indeed to blame for the lack of
454 expected relationships. For eutrophication, correlations were also weak for both on-farm energy use
455 ($r=0.35$, $R^2=0.121$, $n=28$) and FCR ($r=0.24$, $R^2=0.058$, $n=27$).

456 Figure 5 visualises these counterintuitive patterns. Again, the two extensive farming cycles (23 and 24 from
457 study 10) report the highest GW impacts because of LULUC while having the lowest eutrophication
458 impacts. The stark contrast in eutrophication impacts between cycles from studies 8 and 9 and the rest of
459 the dataset highlights how different modelling approaches (sections 3.5.1 and 3.5.2) can dramatically affect
460 results.



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466 **5.2 The Dominance of Methodological Choice in Shaping LCA Outcomes**

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Figure 5 Normalised data on energy use, FCR, global warming and eutrophication impacts obtained from 37 shrimp LCA cycles detailed in 16 studies. Same background colour of adjacent cycles indicates that these cycles originate from the same study. Overlapping points have been jittered for better visibility.

The primary strength of LCA lies in comparing products or systems, a practice only valid when conducted within a consistent methodological framework as stipulated by ISO 14040. Across the ten studies that assessed and compare multiple farming cycles using a consistent internal methodology, the average CV across the farming cycles compared for GW results was 23.6%. In the most comprehensive single study, which analysed nine distinct production cycles (study 8), this variation attributable to farming practices was even lower, with a CV of 15.2%.

In contrast, when identical farm-level inventory data were re-analysed using different LCA approaches, the resulting impacts changed dramatically. Study 10 recalculated two cycles of study 8 in addition to its own cycles, and study 1 recalculated one cycle of study 10 in addition to its own cycle. In these three instances, GW results changed by an average of CV=41.6%. When study 1 recalculated data for an extensive system from study 10, the GW impact decreased by 47% (from 19,800 to 10,503 kg CO₂-eq.), the acidification impact was reduced to zero, and the eutrophication impact inverted from a positive 1.44 to a negative -11.66 kg PO₄-eq. For the exact same farm inventory, the reported environmental profile is therefore largely a function of the analyst's choices. This has profound implications for consumer-facing initiatives such as product labelling and certification. An eco-label awarded based on these LCA results may be rewarding

482 favourable methodological choices rather than genuinely superior on-farm environmental performance,
483 misleading consumers and undermining the credibility of such schemes.

484 The correlation analysis revealed surprisingly weak relationships between key inputs like on-farm energy
485 use and GW impacts ($r=0.04$) or between FCRs and GW impacts ($r=-0.21$). This does not imply that feed
486 and energy are unimportant; rather, it proves that pervasive methodological variations introduce substantial
487 statistical noise, obscuring these fundamental input-impact relationships when data are aggregated across
488 studies. Consequently, any meta-analysis that simply averages results from methodologically diverse LCAs
489 risks drawing conclusions from figures that are not fundamentally comparable. This is exemplified by how
490 directly averaging the data of study 10, which uniquely and showed LULUC emissions with studies lacking
491 such comprehensive LULUC accounting, would disproportionately skew overall findings, as could be
492 observed in analyses performed by studies like Clune et al. (2017).

493 Specific methodological decisions have different influence on this divergence. The choice of co-product
494 allocation method alone can alter reported GW and eutrophication results by up to 59%. Decisions
495 regarding system boundaries, particularly the inclusion or exclusion of LULUC, can be even more
496 influential, increasing GW results by as much as 94% in one case (study 10). This problem is compounded
497 by a critical lack of transparency and reproducibility. Only five of the 16 studies fulfilled all eleven criteria
498 deemed necessary for complete reproducibility, with one study failing to meet eight of the criteria. This
499 opacity prevents scientific scrutiny and self-correction. This lack of reproducibility undermines the
500 cumulative nature of scientific knowledge (Popper, 1959) and erodes trust in LCA as a robust tool for
501 sustainability assessment.

502 Despite these significant challenges, it is important to recognise the value of the existing body of research.
503 The reviewed LCAs have consistently identified feed composition and on-farm energy consumption as the
504 primary environmental hotspots across a wide range of production systems. This provides a crucial and
505 foundational understanding for guiding improvement efforts. Furthermore, several of the reviewed studies
506 exemplify methodological rigour with high transparency and reproducible results, offering a foundational
507 blueprint for developing more comprehensive and standardised environmental evaluation frameworks in
508 the future.

509 However, the collective utility of these studies for comparison or policy is hindered by a more fundamental
510 issue this review has quantified: the influence of methodological choice on reported impacts is greater than
511 the influence of actual on-farm practices. This analytical dominance is so profound that it can invert the
512 environmental profile of an identical farm—transforming it from a net source of eutrophication to a net
513 mitigator based solely on the modelling choice. It obscures expected biophysical relationships, making it
514 impossible to draw meaningful conclusions from cross-study comparisons. Furthermore, it means that
515 specific, often opaque, decisions, such as the inclusion of LULUC or the assumed origin of feed ingredients,
516 can single-handedly determine a product's perceived sustainability, rendering many comparative assertions
517 unreliable.

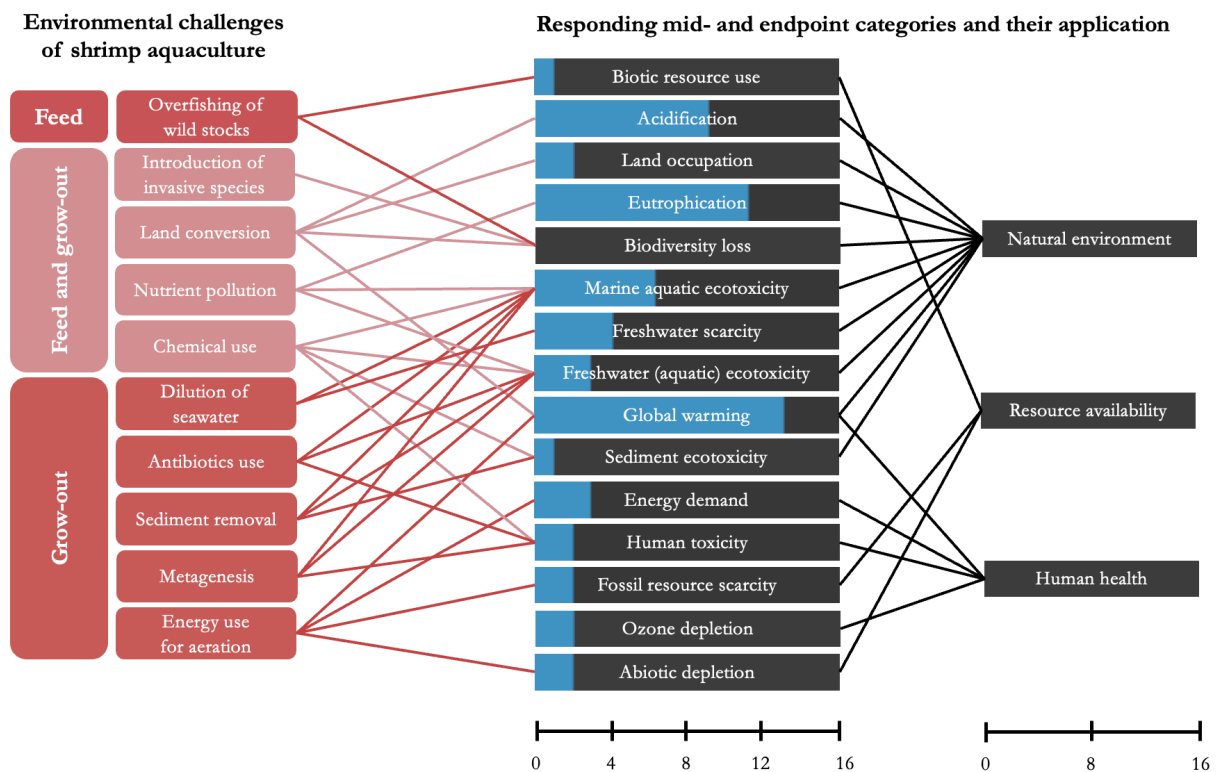
518 This review's findings demonstrate that the current, narrow focus of most shrimp LCAs is a key source of
519 this unreliability. The widespread and systematic omission of critical impact pathways creates a vacuum of
520 data and guidance. This vacuum is inevitably filled by the inconsistent assumptions and variable system
521 boundaries that have been shown to dominate the results. Therefore, for shrimp LCAs to evolve into a
522 robust tool capable of guiding policy and practice, its scope must be fundamentally expanded and
523 harmonised. Future assessments, and any prospective PEFCR for crustaceans, must move beyond a narrow
524 set of midpoints to systematically and transparently quantify the interconnected impacts of LULUC, the
525 biodiversity footprint of farms and feed, chemical and antibiotic inputs with their associated ecotoxicity
526 and human health risks, and direct GHG emissions from ponds.

527

528 **6. Analytical Blind Spots: The Neglected Environmental Dimensions**

529 The environmental critique of aquaculture extends beyond the commonly assessed impact categories of
530 global warming, eutrophication, and acidification, encompassing a wider range of environmental pressures
531 (Ahmed and Thompson, 2019; DeWeerd, 2020; Martinez-Porchas and Martinez-Cordova, 2012;
532 Mavraganis et al., 2020; Pazmiño et al., 2024; figure 6). However, the current literature systematically do
533 not allow for or omits critical environmental impacts. Biodiversity loss, for instance, is a highly relevant, yet
534 overlooked, aspect in the reviewed shrimp LCAs. Here more work needs to be done to develop biodiversity
535 impact assessment methodologies for marine environments. Furthermore, various toxicity categories were

536 calculated by only five studies, despite the documented widespread use of chemicals in shrimp aquaculture.
 537 Neither did any study evaluate endpoint impact indicators, such as effects on human health or ecosystems,
 538 thereby limiting the ability of LCA to provide a holistic assessment of shrimp aquaculture's sustainability.
 539 This selective focus creates a partial and potentially misleading picture of environmental performance of
 540 shrimp aquaculture (figure 6).



541
 542 *Figure 6 Environmental challenges of shrimp aquaculture and the responding impact categories. Blue shading of mid- and endpoint categories indicates*
 543 *the proportion of the 16 reviewed studies which address the particular category. Lines represent causal relationships between mid- and end-point categories*
 544 *and potential environment impacts of particular phases of the aquaculture cycle.*

545 6.1 The interconnected footprint of land, feed, and biodiversity

546 The environmental footprint of shrimp aquaculture is often viewed through the narrow lens of the farm
 547 boundary, yet its most profound impacts are frequently interconnected and telecoupled. This review finds
 548 that the literature systematically fails to account for these linked pressures, with three areas of particular
 549 concern:

550 **LULUC:** The conversion of coastal ecosystems, particularly carbon-rich mangrove forests, for shrimp
 551 ponds is a profound environmental transformation. Yet only studies 10 and 11 included farm-level LULUC

552 impacts, with study 10 finding that they could contribute up to 94% of a system's GW footprint. This
553 omission is critical, as emissions from mangrove conversion in Southeast Asia alone are estimated at 691.8
554 teragrams of CO₂-equivalent annually (Sasmito et al., 2025). Current approaches to land-use assessment
555 also exhibit high methodological discrepancy, with research demonstrating that the attribution of LULUC
556 emissions remains a nuanced challenge influenced by data sources, historical land-use patterns, and regional
557 dynamics (Caro et al., 2018). To address this gap, future shrimp LCAs and any prospective Product
558 Environmental Footprint Category Rules (PEFCRs) for aquaculture must mandate standardised LULUC
559 accounting. This should align with established frameworks like PAS 2050 (BSI, 2011), requiring the use of
560 region-specific carbon stock data and sensitivity analyses to address inherent uncertainties.

561 **Feed formulations and origins:** The intensification of shrimp farming has shifted this environmental
562 burden from direct land occupation at the farm site to global feed supply chains (Clawson et al., 2024; Davis
563 et al., 2021). However, the impacts of feed are poorly quantified due to inconsistent reporting of ingredients
564 and, crucially, their geographical origins. The reviewed studies demonstrate variation in feed compositions,
565 with fishmeal comprising 20–42% of pellet ingredients and soybean meal 11–30%. The sensitivity analysis
566 in study 1, which showed a potential 1240% increase in GW impacts for soybean meal sourced from Brazil
567 versus the U.S., underscores the critical importance of geographical specificity. Furthermore, minor
568 variations in reporting feed composition can lead to threefold differences in estimates of wild fish use
569 (Roberts et al., 2024), highlighting the need for high levels of detail and transparency. Therefore, future
570 LCAs and any aquaculture PEFCRs must involve transparent reporting of all feed ingredients, their
571 proportions, and their geographical origins, along with sensitivity analyses for high-impact ingredients.

572 **Biodiversity impacts:** Biodiversity loss is the ultimate consequence of these pressures, and LCA is
573 increasingly used to estimate biodiversity impacts across complex value chains (Bromwich et al., 2025), yet
574 it remains entirely unquantified in shrimp LCA studies. While several studies recognised the role of shrimp
575 farming in biodiversity loss, they excluded its quantification due to a lack of inventory data and
576 characterisation factors or lack of methods to assess these impacts. The sector drives biodiversity loss
577 through multiple pathways, including direct habitat destruction from mangrove conversion, pollution from
578 effluent, pressure on both wild fisheries for fishmeal, terrestrial ecosystems for crops like soybeans, and the
579 potential introduction of invasive species or genetic pollution from escaped stock.

580 While methods to quantify terrestrial biodiversity loss are advancing, marine biodiversity metrics is lagging
581 (Crenna et al., 2020). While the European Union's Environmental Footprint 3.1 methodology is now the
582 leading guide recommended for developing comparable PEFCRs, an examination reveals that this
583 framework is not yet equipped to address the primary biodiversity impacts of coastal aquaculture. Omitting
584 these key impact categories creates a systemic flaw in current assessments. An LCA that neglects off-farm
585 LULUC and biodiversity impacts may incorrectly favour an intensive system with a small local footprint
586 over an extensive one, even if the former's feed is sourced from recently deforested land in a global
587 biodiversity hotspot. This analytical blind spot could lead to counterproductive policy incentives that reward
588 practices that appear sustainable locally while being devastating globally. While LCA methodologies for
589 biodiversity assessment have known limitations, such as inadequate spatial differentiation (Winter et al.,
590 2017), difficulty in modelling habitat fragmentation (Kuipers et al., 2019), and gaps in addressing diverse
591 taxonomic groups (Damiani et al., 2023; Martínez-Ramón et al., 2024), researchers should begin
592 incorporating biodiversity impacts using existing frameworks like ReCiPe. Documenting key water quality
593 parameters related to biodiversity (such as biochemical oxygen demand, nitrogen, and phosphorus levels),
594 or classifying feed sources by sustainability certification would be a significant step forward.

595 **6.2 Unaccounted chemical contamination and gaseous emissions**

596 Beyond the interconnected footprint of feed and land, LCAs must also quantify critical chemical and
597 gaseous pressures originating from the farm itself.

598 **Antibiotics and ecotoxicity:** This review reveal a critical failure to assess antibiotic use in shrimp
599 aquaculture. Only one study (study 8) quantified antibiotic inputs, despite calls for more comprehensive
600 modelling of pharmaceutical emissions and their toxicity-related effects in LCA (Emara et al., 2019). This
601 is not just a matter of direct ecotoxicity, which itself was only assessed in five studies. The development of
602 antimicrobial resistance (AMR) is a profound threat to human health and may be a more severe long-term
603 impact than direct toxicity (Nyberg et al., 2021). Empirical research underscores these risks: study 8
604 highlighted toxicity inputs including pesticides, metals, and pharmaceuticals, while other studies identified
605 up to 20 different antimicrobial products in use in Viet Nam (Luu et al., 2021). Chemical residues in water,
606 sediments, and harvested shrimp potentially promote antibiotic-resistant bacteria and resistance genes

607 (Shao et al., 2021). Future LCAs should systematically incorporate these inputs. This will require
608 establishing trusted, potentially anonymised, data-sharing frameworks for sensitive farm-level data and
609 developing methods to assess not only direct toxicity but also the critical downstream impacts of AMR. If
610 primary data collection of these sensitive inputs is not feasible, assumptions should be made rather than
611 leaving out these highly influential inputs.

612 **Pond emissions:** Direct greenhouse gas emissions from the pond itself, particularly methane (CH₄) and
613 nitrous oxide (N₂O), represent a significant data gap that leads to a systematic underestimation of the
614 sector's climate impact. This review found only a fraction of studies report these emissions, despite research
615 showing they can be substantial, with shrimp ponds potentially emitting ten times more methane than the
616 coastal marsh ecosystems they often replace. To move beyond this critical omission, LCA practitioners
617 must actively incorporate predictive models to quantify these biogeochemical fluxes. While the IPCC
618 provides foundational, default methodologies for estimating these emissions from aquaculture within its
619 guidelines for wastewater, more specialised models are needed to capture the unique dynamics of these
620 systems. For instance, the Pond-NP nutrient dynamic model developed by Zhang et al. (2024) quantifies
621 the complex nitrogen cycle, estimating a significant loss to the atmosphere through processes like
622 denitrification. This work underscores a crucial point for LCA: nutrient inputs that do not end up in
623 harvested biomass are lost to the surrounding environment, partly as gaseous emissions, including potent
624 greenhouse gases.

625 Therefore, we recommend a proactive, tiered approach for practitioners to ensure these emissions are
626 accounted for. As a baseline, practitioners should use the established methodologies in the 2019 Refinement
627 to the 2006 IPCC Guidelines. This involves applying default emission factors to the nitrogen load from
628 uneaten feed and excretion to estimate N₂O, and adapting the wastewater methodology, which links CH₄
629 production to the pond's biochemical oxygen demand (BOD), to estimate methane. When more farm-
630 specific data is available, practitioners should use these parameters to apply parsimonious predictive models.
631 This practice is supported by research such as Znachor et al. (2023) which shows that GHG fluxes can be
632 estimated from a limited set of readily available data like water temperature and depth. Key data to collect

633 include feed inputs, stocking density, water exchange rates, and management practices during both the
634 culture and non-culture periods, as the latter can be a hotspot for emissions.

635 For high-quality LCAs, practitioners should leverage outputs from detailed biogeochemical process models
636 like Pond-NP, a framework that has been used to analyse complex economic and GHG relationships in
637 other aquaculture contexts. Adopting this hierarchical approach, grounded in IPCC guidance, will bridge a
638 major analytical gap and ensure that LCA can provide a more complete and accurate assessment of shrimp
639 aquaculture's climate footprint.

640

641 **7. Recommendations for More Robust and Representative LCAs**

642 To transition shrimp LCAs from a collection of disparate studies into a robust evidence base for
643 sustainability, expansions in methodology and scope are necessary.

644 **7.1 Methodological Harmonisation**

645 The EU's PEF methodology offers a promising pathway towards standardisation. It has been applied in
646 other food sectors (Hietala et al., 2023) and marine fish, providing comprehensive guidelines on system
647 boundaries, allocation, and data quality. Its requirement to assess 16 impact categories would also
648 significantly expand the scope beyond the narrow focus of current shrimp LCAs. For a future PEF standard
649 for crustaceans, we suggest that the system boundaries should include all flows in figure 3 and other
650 recommendations herein. Nonetheless, PEF's European origin may present challenges in its direct
651 applicability in major non-EU shrimp-producing regions where production conditions and data availability
652 differ substantially.

653 Complementary open-access platforms, like HESTIA (www.hestia.earth; Poore 2021), provide a
654 harmonised data and modelling platform that can further help structured unit process data and associated
655 meta-data. It enables researchers to analyse their own farm-level data using consistently using pre-defined
656 emission models and gap filling. Moreover, it also allows users to compare their results against other studies
657 and food commodities, facilitating harmonised cross-study comparisons across different systems, products,
658 and regions. Such comparisons have the potential to help identify which shrimp farming systems that are

659 most environmentally efficient, potentially influencing consumer adoption of more sustainable diets (Ran
660 et al., 2022).

661 7.2 Representativeness in Geography and Practice

662 The current body of shrimp LCA research provides a picture of where the industry was, but is largely blind
663 to where it is today and where it is going, while farmers are facing increasing environmental and
664 socioeconomic pressures (Macusi et al., 2022). This makes our current knowledge a poor tool for guiding
665 sustainable development of the sector. Using outdated and not geographically adapted literature to guide
666 the current industry can even have highly counter-productive consequences. With an average data collection
667 year of 2013, the literature largely fails to capture the modern, intensified industry. It does not adequately
668 address newer, super-intensive systems (e.g RAS and BFT), integrated multi-trophic aquaculture (IMTA),
669 or the impacts of certification schemes. Several emerging farming techniques, such as hybrid BioRAS
670 systems, offshore shrimp farming, and various closed-loop land-based production systems, remain entirely
671 absent from current LCA literature. Similarly, the impacts of improved feed formulations aimed at reducing
672 antibiotic use are insufficiently evaluated, despite their growing adoption, particularly as economic shocks
673 like the COVID-19 pandemic have driven farmers toward greater resource efficiency (Nguyen et al. 2024).
674 The finding that infrastructure can be a major driver of GHG emissions, accounting for up to 14% of total
675 emissions in super-intensive systems (Huang et al., 2024), an element often downplayed in older studies,
676 underscores the need for updated assessments.

677 There is a significant misalignment between the regions covered by existing LCAs and the world's major
678 shrimp producers. For example, data from Thailand is overrepresented but dated, reflecting a pre-2012
679 industry structure before a major disease outbreak reshaped its production (Prompatanapak & Lopetcharat,
680 2020). Meanwhile, major producers like India and Ecuador are almost entirely absent from the peer-
681 reviewed LCA literature. This geographic imbalance is highly problematic, as production practices,
682 regulatory environments, and ecosystem sensitivities vary dramatically by region. For instance, farming in
683 India is shaped by diverse regional regulations (Kumar et al., 2023), while Ecuador's industry operates within
684 unique coastal ecosystem dynamics (Viera-Roma et al., 2024). The environmental consequences of
685 expanding shrimp farms into Egyptian deserts, utilising previously non-productive land (Soliman & Yacout,

686 2016), are vastly different from converting carbon-rich mangroves in Indonesia, where a hectare of
687 converted mangrove can release thousands of tonnes of CO₂-equivalent (Sasmito et al., 2019). Similarly,
688 Viet Nam's government supports a transition to rice-shrimp farming, a model whose impacts have not been
689 analysed from an LCA lens yet, despite studies on farmers' willingness to adopt improved practices (Ngoc
690 et al., 2021). Only through a continuous cycle of updated and geographically diverse LCAs can the field
691 keep pace with this dynamic industry and provide relevant guidance for its sustainable development.

692 **7.3 Sensitivity Analyses**

693 To make the influence of critical assumptions more transparent, we recommend that future shrimp LCAs,
694 and any forthcoming PEFCR, should mandate a minimum set of sensitivity analyses. Based on the major
695 drivers of variability identified in this review, these analyses should test the influence of several key factors.
696 Practitioners should first assess the impact of the chosen co-product allocation method by comparing the
697 results against at least one alternative, such as contrasting economic with mass-based allocation, to
698 demonstrate the robustness of the conclusions. It is also essential to test assumptions regarding the
699 geographical sourcing of high-impact inputs, for instance by evaluating how sourcing key feed ingredients
700 like soybean meal from different plausible regions with varying LULUC and biodiversity risks. Similarly,
701 given the uncertainty surrounding direct farm-level emissions, the sensitivity of results to different emission
702 models or factors for pond-level greenhouse gases should be evaluated.

703 Sensitivity analyses are also paramount for complex and developing modelling areas like LULUC and
704 biodiversity. For studies including farm-level LULUC, an analysis of the core parameters of the model—
705 such as the assumed percentage of carbon loss from soil and biomass upon conversion—is critical to frame
706 the uncertainty of this high-impact factor. As methodologies for assessing biodiversity impacts are
707 incorporated for both farm and feed stages, it is crucial to test the sensitivity of results to key methodological
708 choices, which can significantly alter outcomes. This should include assessing the sensitivity to the chosen
709 reference state (e.g., a 'natural' versus a 'managed' ecosystem baseline), the choice of biodiversity metric
710 (e.g., comparing models based on species richness with those reflecting ecosystem functionality), and the
711 taxonomic scope of the assessment (e.g., comparing impacts on well-studied taxa versus broader species
712 groups). Systematically performing and reporting on these sensitivity analyses would provide a much clearer

713 picture of how methodological choices influence the results, enhancing the credibility and utility of future
714 studies.

715 **7.4 Broader Implication for Agricultural LCAs**

716 While this review focuses on shrimp, its findings on methodological inconsistency and neglected impact
717 pathways offer critical lessons that extend beyond seafood to the broader field of food system LCA,
718 particularly for animal agriculture. The problems identified are not unique to crustaceans but are symptoms
719 of systemic challenges in assessing complex biological production systems. The central conclusion—that
720 methodological choices and analytical blind spots can obscure or even outweigh the impacts of on-farm
721 practices —is a finding of profound importance for the entire LCA community. This review demonstrates
722 that for a given farm, the reported environmental profile can be transformed based solely on the analyst's
723 choices regarding system boundaries, allocation, or the inclusion of factors like LULUC. This has direct
724 relevance for other aquatic as well as terrestrial animal production. The critical role of feed as a telecoupled
725 environmental hotspot is a shared challenge across industrial animal production, both aquatic and terrestrial.
726 The sensitivity of GW impacts to the geographical origin of soymeal, a key ingredient in many shrimp,
727 poultry, and swine feeds, exemplifies this. Any LCA of a fed animal product that lacks transparency on feed
728 composition and origin risks fundamentally misrepresenting the product's true environmental burden.

729 Furthermore, while the specific nature of neglected impacts varies by system, the problem of 'analytical
730 blind spots' is universal. In shrimp, this review highlights unquantified pond emissions, mangrove
731 conversion, and antibiotic use. In terrestrial systems, analogous blind spots may include soil carbon
732 dynamics, methane from manure management, or local biodiversity impacts from grazing (Goglio et al.,
733 2024). The key lesson for all food LCA practitioners is the need to move beyond a narrow, conventional
734 set of impact categories to identify and incorporate the system-specific pressures that may dominate the
735 total environmental footprint.

736 Finally, the geographical and temporal misalignment of the shrimp LCA literature serves as a stark warning
737 for any meta-analysis or policy decision based on LCA. Just as the evidence base for shrimp over-represents
738 dated practices in Thailand and under-represents modern production in Ecuador, the literature for other

739 commodities may suffer from similar biases, leading to flawed conclusions. Therefore, the
740 recommendations for methodological harmonisation (e.g., via PEF CRs), mandatory sensitivity analyses,
741 and a radically expanded environmental scope should be viewed as a blueprint for improving the rigour,
742 transparency, and policy-relevance of all food system sustainability assessments.

743

744 **8. Conclusion**

745 LCA is an increasingly utilised tool for evaluating the environmental performance of shrimp aquaculture,
746 and a body of research now exists, identifying key areas like LULUC, feed, and energy as key impact drivers.
747 However, this literature is marked by substantial methodologically induced discrepancies in reported
748 impacts, often differing by more than fiftyfold across key categories, and does not reflect current production
749 systems and regions. A lack of transparent reporting currently limits the reliability and comparability of
750 many shrimp LCAs. While individual studies offer valuable insights, the collective picture is fragmented,
751 making it challenging to benchmark performance accurately or develop robust, evidence-based
752 sustainability strategies. To realise the full potential of LCA as a guide for sustainable shrimp farming,
753 addressing these shortcomings is crucial. We recommend a concerted effort focused on:

- 754 • Enhancing transparency: Ensuring full and explicit documentation of all assumptions, allocation
755 approaches, system boundaries, calculation methods, and unit process data.
- 756 • Adopting harmonised methodologies: Progressing towards standardised frameworks, potentially
757 leveraging the PEF guidelines, including the development of PEF CRs, or platforms like HESTIA,
758 to improve cross-study comparability.
- 759 • Broadening environmental scope: Systematically incorporating critical impacts like LULUC,
760 biodiversity, antibiotics and water treatment chemicals, and pond emissions, using standardised
761 assessment approaches.
- 762 • Expanding geographic and system coverage: Prioritising assessments in underrepresented major
763 producing regions and incorporating emerging intensive farming systems.

764 This will support the shrimp aquaculture sector in moving towards a truly sustainable future, balancing
765 global food demands with environmental responsibilities.

766

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771

772 10. References

773 Ahmed, N., & Thompson, S. (2019). The blue dimensions of aquaculture: A global synthesis. *Science of*
774 *The Total Environment*, 652, 851–861. <https://doi.org/10.1016/j.scitotenv.2018.10.163>

775 Ahmed, Z., & Ambinakudige, S. (2024). How does shrimp farming impact agricultural production and
776 food security in coastal Bangladesh? Evidence from farmer perception and remote sensing
777 approach. *Ocean & Coastal Management*, 255, 107241.

778 <https://doi.org/10.1016/j.ocecoaman.2024.107241>

779 Al Eissa, A., Chen, P., Brown, P. B., & Huang, J. Y. (2022). Effects of feed formula and farming system
780 on the environmental performance of shrimp production chain from a life cycle perspective.
781 *Journal of Industrial Ecology*, 26(6), 2006–2019. <https://doi.org/10.1111/jiec.13370>

782 Arbour, A. J., Bhatt, P., Simsek, H., Brown, P. B., & Huang, J.-Y. (2024). Life cycle assessment on
783 environmental feasibility of microalgae-based wastewater treatment for shrimp recirculating
784 aquaculture systems. *Bioresource Technology*, 399, 130578.

785 <https://doi.org/10.1016/j.biortech.2024.130578>

786 Aubin, J., Baruthio, A., Mungkung, R., & Lazard, J. (2015). Environmental performance of brackish
787 water polyculture system from a life cycle perspective: A Filipino case study. *Aquaculture*, 435, 217–
788 227. <https://doi.org/10.1016/j.aquaculture.2014.09.019>

- 789 Belettini, F., Seiffert, W. Q., Lapa, K. R., Vieira, F. do N., Santo, C. M. do E., & Arana, L. A. V. (2018).
790 Carbon footprint in commercial cultivation of marine shrimp: A case study in southern Brazil.
791 *Revista Brasileira de Zootecnia*, 47. <https://doi.org/10.1590/rbz4720160353>
- 792 Blaxter, T., Åsbjær, E., & Fraanje, W. (2024). Animal welfare and ethics in food and agriculture.
793 <https://doi.org/10.56661/f2d8f4c7>
- 794 Bohnes, F. A., & Laurent, A. (2019). LCA of aquaculture systems: methodological issues and potential
795 improvements. *The International Journal of Life Cycle Assessment*, 24(2), 324–337.
796 <https://doi.org/10.1007/s11367-018-1517-x>
- 797 Bohnes, F. A., Hauschild, M. Z., Schlundt, J., & Laurent, A. (2019). Life cycle assessments of
798 aquaculture systems: a critical review of reported findings with recommendations for policy and
799 system development. *Reviews in Aquaculture*, 11(4), 1061–1079. <https://doi.org/10.1111/raq.12280>
- 800 Bromwich, T., White, T. B., Bouchez, A., Hawkins, I., zu Ermgassen, S., Bull, J., Bartlett, H., Bennun,
801 L., Biggs, E., Booth, H., Clark, M., el Geneidy, S., Prescott, G. W., Sonter, L. J., Starkey, M., &
802 Milner-Gulland, E. J. (2025). Navigating uncertainty in life cycle assessment-based approaches to
803 biodiversity footprinting. *Methods in Ecology and Evolution*. [https://doi.org/10.1111/2041-](https://doi.org/10.1111/2041-210X.70001)
804 [210X.70001](https://doi.org/10.1111/2041-210X.70001)
- 805 BSI. (2011). *PAS 2050:2011 Specification for the assessment of the life cycle greenhouse gas emissions of goods and*
806 *services*. British Standards Institution.
- 807 Cao, L., Diana, J. S., Keoleian, G. A., & Lai, Q. (2011). Life cycle assessment of Chinese shrimp farming
808 systems targeted for export and domestic sales. *Environmental Science and Technology*, 45(15), 6531–
809 6538. <https://doi.org/10.1021/es104058z>
- 810 Caro, D., Davis, S. J., Kebreab, E., & Mitloehner, F. (2018). Land-use change emissions from soybean
811 feed embodied in Brazilian pork and poultry meat. *Journal of Cleaner Production*, 172, 2646–2654.
812 <https://doi.org/10.1016/j.jclepro.2017.11.146>
- 813 Chang, C. C., Chang, K. C., Lin, W. C., & Wu, M. H. (2017). Carbon footprint analysis in the
814 aquaculture industry: Assessment of an ecological shrimp farm. *Journal of Cleaner Production*, 168,
815 1101–1107. <https://doi.org/10.1016/j.jclepro.2017.09.109>

816 Clawson, G., Blanchard, J. L., Cormery, M., Fulton, B., Halpern, B. S., Hamilton, H. A., O'Hara, C. C.,
817 & Cottrell, R. S. (2024). Continued Transitions from Fish Meal and Oil in Aquafeeds Require Close
818 Attention to Biodiversity Trade-Offs. <https://doi.org/10.2139/ssrn.5030822>

819 Clune, S., Crossin, E., & Verghese, K. (2017). Systematic review of greenhouse gas emissions for
820 different fresh food categories. *Journal of Cleaner Production*, 140, 766–783.
821 <https://doi.org/10.1016/j.jclepro.2016.04.082>

822 Cortés, A., Casillas-Hernández, R., Cambeses-Franco, C., Bórquez-López, R., Magallón-Barajas, F.,
823 Quadros-Seiffert, W., Feijoo, G., & Moreira, M. T. (2021). Eco-efficiency assessment of shrimp
824 aquaculture production in Mexico. *Aquaculture*, 544.
825 <https://doi.org/10.1016/j.aquaculture.2021.737145>

826 Crenna, E., Marques, A., la Notte, A., & Sala, S. (2020). Biodiversity Assessment of Value Chains: State
827 of the Art and Emerging Challenges. *Environmental Science & Technology*, 54(16), 9715–9728.
828 <https://doi.org/10.1021/acs.est.9b05153>

829 Damiani, M., Sinkko, T., Caldeira, C., Tosches, D., Robuchon, M., & Sala, S. (2023). Critical review of
830 methods and models for biodiversity impact assessment and their applicability in the LCA context.
831 *Environmental Impact Assessment Review*, 101, 107134. <https://doi.org/10.1016/j.eiar.2023.107134>

832 Davis, R. P., Boyd, C. E., & Davis, D. A. (2021). Resource sharing and resource sparing, understanding
833 the role of production intensity and farm practices in resource use in shrimp aquaculture. *Ocean &*
834 *Coastal Management*, 207, 105595. <https://doi.org/10.1016/j.ocecoaman.2021.105595>

835 DeWeerd, S. (2020). Can aquaculture overcome its sustainability challenges? *Nature*, 588(7837), S60–
836 S62. <https://doi.org/10.1038/d41586-020-03446-3>

837 Dieterle, M., Fischer, P., Pons, M.-N., Blume, N., Minke, C., & Bischi, A. (2022). Life cycle assessment
838 (LCA) for flow batteries: A review of methodological decisions. *Sustainable Energy Technologies and*
839 *Assessments*, 53, 102457. <https://doi.org/10.1016/j.seta.2022.102457>

840 Dolan, S. L., & Heath, G. A. (2012). Life Cycle Greenhouse Gas Emissions of Utility-Scale Wind
841 Power. *Journal of Industrial Ecology*, 16(s1). <https://doi.org/10.1111/j.1530-9290.2012.00464.x>

842 Dong, Y., & Liu, P. (2022). Evaluation of the completeness of LCA studies for residential buildings.
843 *Clean Technologies and Environmental Policy*, 24(1), 229–250. [https://doi.org/10.1007/s10098-021-](https://doi.org/10.1007/s10098-021-02115-x)
844 [02115-x](https://doi.org/10.1007/s10098-021-02115-x)

845 Emara, Y., Lehmann, A., Siegert, M.-W., & Finkbeiner, M. (2019). Modeling pharmaceutical emissions
846 and their toxicity-related effects in life cycle assessment (LCA): A review. *Integrated Environmental*
847 *Assessment and Management*, 15(1), 6–18. <https://doi.org/10.1002/ieam.4100>

848 Emerenciano, M. G. C., Rombenso, A. N., Vieira, F. d. N., Martins, M. A., Coman, G. J., Truong, H.
849 H., Noble, T. H., & Simon, C. J. (2022). Intensification of Penaeid Shrimp Culture: An Applied
850 Review of Advances in Production Systems, Nutrition and Breeding. *Animals*, 12(3), 236.
851 <https://doi.org/10.3390/ani12030236>

852 FishStatJ: For aquaculture production data, FishStatJ from FAO Fisheries Division, Statistics and
853 Information Branch for fishery statistical time series
854 (www.fao.org/fishery/en/statistics/software/fishstatj) was used, and data retrieved on March 10th
855 2025.

856 Flores-Pérez, M. B., Yépez, E. A., Robles-Morúa, A., Villa-Ibarra, M., Bórquez-López, R., Gil-Núñez, J.
857 C., Lares-Villa, F., & Casillas-Hernández, R. (2023). Eco-efficiency assessment of disease-infected
858 shrimp farming in Mexico using environmental impact assessment tools. *Science of the Total*
859 *Environment*, 858. <https://doi.org/10.1016/j.scitotenv.2022.159737>

860 Frischknecht, R., Wyss, F., Büsser Knöpfel, S., Lützkendorf, T., & Balouktsi, M. (2015). Cumulative
861 energy demand in LCA: the energy harvested approach. *The International Journal of Life Cycle*
862 *Assessment*, 20(7), 957–969. <https://doi.org/10.1007/s11367-015-0897-4>

863 Froehlich, H. E., Runge, C. A., Gentry, R. R., Gaines, S. D., & Halpern, B. S. (2018). Comparative
864 terrestrial feed and land use of an aquaculture-dominant world. *Proceedings of the National Academy of*
865 *Sciences*, 115(20), 5295–5300. <https://doi.org/10.1073/pnas.1801692115>

866 Fry, J. P., Mailloux, N. A., Love, D. C., Milli, M. C., & Cao, L. (2018). Feed conversion efficiency in
867 aquaculture: do we measure it correctly? *Environmental Research Letters*, 13(2), 024017.
868 <https://doi.org/10.1088/1748-9326/aaa273>

869 Funge-Smith, S., & Briggs, M. (2003). The introduction of *Penaeus vannamei* and *P. stylirostris* into the
870 Asia-Pacific region. *International Mechanisms for the Control and Responsible Use of Alien Species in Aquatic*
871 *Ecosystems*, 26–29.

872 Gentry, R. R., Ruff, E. O., & Lester, S. E. (2019). Temporal patterns of adoption of mariculture
873 innovation globally. *Nature Sustainability*, 2(10), 949–956. [https://doi.org/10.1038/s41893-019-](https://doi.org/10.1038/s41893-019-0395-y)
874 [0395-y](https://doi.org/10.1038/s41893-019-0395-y)

875 Godfray, H. C. J., Aveyard, P., Garnett, T., Hall, J. W., Key, T. J., Lorimer, J., Pierrehumbert, R. T.,
876 Scarborough, P., Springmann, M., & Jebb, S. A. (2018). Meat consumption, health, and the
877 environment. *Science*, 361(6399). <https://doi.org/10.1126/science.aam5324>

878 Goglio, P., Moakes, S., Knudsen, M. T., van Mierlo, K., Adams, N., Maxime, F., Maresca, A., Romero-
879 Huelva, M., Waqas, M. A., Smith, L. G., Grossi, G., Smith, W., de Camillis, C., Nemecek, T., Tei,
880 F., & Oudshoorn, F. W. (2024). Harmonizing methods to account for soil nitrous oxide emissions
881 in Life Cycle Assessment of agricultural systems. *Agricultural Systems*, 219, 104015.
882 <https://doi.org/10.1016/j.agsy.2024.104015>

883 Guinée, J. B. (2002). Handbook on life cycle assessment operational guide to the ISO standards. *The*
884 *International Journal of Life Cycle Assessment*, 7(5), 311. <https://doi.org/10.1007/BF02978897>

885 Guo, M., & Murphy, R. J. (2012). LCA data quality: Sensitivity and uncertainty analysis. *Science of The*
886 *Total Environment*, 435–436, 230–243. <https://doi.org/10.1016/j.scitotenv.2012.07.006>

887 Heijungs, R. (2024). Uncertainty and Sensitivity Analysis in Life Cycle Assessment. In *Encyclopedia of*
888 *Sustainable Technologies* (pp. 235–248). Elsevier. [https://doi.org/10.1016/B978-0-323-90386-](https://doi.org/10.1016/B978-0-323-90386-8.00039-5)
889 [8.00039-5](https://doi.org/10.1016/B978-0-323-90386-8.00039-5)

890 Henriksen, T., Levis, J. W., Barlaz, M. A., & Damgaard, A. (2019). Approaches to fill data gaps and
891 evaluate process completeness in LCA—perspectives from solid waste management systems. *The*
892 *International Journal of Life Cycle Assessment*, 24(9), 1587–1601. [https://doi.org/10.1007/s11367-019-](https://doi.org/10.1007/s11367-019-01592-z)
893 [01592-z](https://doi.org/10.1007/s11367-019-01592-z)

894 Henriksson, P. J. G., Guinée, J. B., Kleijn, R., & de Snoo, G. R. (2012). Life cycle assessment of
895 aquaculture systems—a review of methodologies. *The International Journal of Life Cycle Assessment*,
896 17(3), 304–313. <https://doi.org/10.1007/s11367-011-0369-4>

897 Henriksson, P. J. G., Guinée, J. B., Heijungs, R., de Koning, A., & Green, D. M. (2014). A protocol for
898 horizontal averaging of unit process data—including estimates for uncertainty. *The International*
899 *Journal of Life Cycle Assessment*, 19(2), 429–436. <https://doi.org/10.1007/s11367-013-0647-4>

900 Henriksson, P. J. G., Rico, A., Zhang, W., Ahmad-Al-Nahid, S., Newton, R., Phan, L. T., Zhang, Z.,
901 Jaithiang, J., Dao, H. M., Phu, T. M., Little, D. C., Murray, F. J., Satapornvanit, K., Liu, L., Liu, Q.,
902 Haque, M. M., Kruijssen, F., de Snoo, G. R., Heijungs, R., ... Guinée, J. B. (2015a). Comparison of
903 Asian Aquaculture Products by Use of Statistically Supported Life Cycle Assessment. *Environmental*
904 *Science and Technology*, 49(24), 14176–14183. <https://doi.org/10.1021/acs.est.5b04634>

905 Henriksson, P. J. G., Heijungs, R., Dao, H. M., Phan, L. T., de Snoo, G. R., & Guinée, J. B. (2015b).
906 Product Carbon Footprints and Their Uncertainties in Comparative Decision Contexts. *PLoS*
907 *ONE*, 10(3), e0121221. <https://doi.org/10.1371/journal.pone.0121221>

908 Henriksson, P. J. G., Tran, N., Mohan, C. V., Chan, C. Y., Rodriguez, U. P., Suri, S., Mateos, L. D.,
909 Utomo, N. B. P., Hall, S., & Phillips, M. J. (2017). Indonesian aquaculture futures – Evaluating
910 environmental and socioeconomic potentials and limitations. *Journal of Cleaner Production*, 162, 1482–
911 1490. <https://doi.org/10.1016/j.jclepro.2017.06.133>

912 Henriksson, P. J. G., Belton, B., Jahan, K. M., & Rico, A. (2018). Measuring the potential for
913 sustainable intensification of aquaculture in Bangladesh using life cycle assessment. *Proceedings of the*
914 *National Academy of Sciences*, 115(12), 2958–2963. <https://doi.org/10.1073/pnas.1716530115>

915 HESTIA team. HESTIA: A harmonised way to represent, share, and analyse agri-environmental data
916 (2023). <https://www.hestia.earth/>. Accessed 03 November 2024.

917 Hietala, S., Troels, K., Woodhouse, A., Ahlgren, S., & Mogensen, L. (2023). Applicability of PEF
918 methodologies in comparison of LCAs of different food products in Nordic countries.

919 Huang, M., Zhou, Y., Tian, H., Pan, S., Yang, X., Gao, Q., & Dong, S. (2024). Rapidly increased
920 greenhouse gas emissions by Pacific white shrimp aquacultural intensification and potential
921 solutions for mitigation in China. *Aquaculture*, 587.
922 <https://doi.org/10.1016/j.aquaculture.2024.740825>

923 International Organization for Standardization. (2020a). Environmental management — Life cycle
924 assessment — Principles and framework (ISO 14040:2006). <https://doi.org/10.3403/30152732U>

925 International Organization for Standardization. (2020b). Environmental management — Life cycle
926 assessment — Requirements and guidelines (ISO 14044:2006).

927 Jonell, M., & Henriksson, P. J. G. (2015). Mangrove-shrimp farms in Vietnam—Comparing organic and
928 conventional systems using life cycle assessment. In *Aquaculture* (Vol. 447, pp. 66–75). Elsevier.
929 <https://doi.org/10.1016/j.aquaculture.2014.11.001>

930 Kikstra, J. S., Nicholls, Z. R. J., Smith, C. J., Lewis, J., Lamboll, R. D., Byers, E., Sandstad, M.,
931 Meinshausen, M., Gidden, M. J., Rogelj, J., Kriegler, E., Peters, G. P., Fuglestvedt, J. S., Skeie, R.
932 B., Samset, B. H., Wienpahl, L., van Vuuren, D. P., van der Wijst, K.-I., al Khourdajie, A., ... Riahi,
933 K. (2022). The IPCC Sixth Assessment Report WGIII climate assessment of mitigation pathways:
934 from emissions to global temperatures. *Geoscientific Model Development*, 15(24), 9075–9109.
935 <https://doi.org/10.5194/gmd-15-9075-2022>

936 Koniyo, Y. (2022). *Role of Innovations / Interventions to Bring Sustainability in Aquaculture Growth in Indonesia:
937 Integration of Life Cycle Assessment (LCA) Framework* (Vol. 26).

938 Kumar, S., Verma, A. K., Singh, S. P., & Awasthi, A. (2022). Immunostimulants for shrimp aquaculture:
939 paving pathway towards shrimp sustainability. *Environmental Science and Pollution Research*, 30(10),
940 25325–25343. <https://doi.org/10.1007/s11356-021-18433-y>

941 Kuipers, K. J. J., May, R. F., Graae, B. J., & Veronesi, F. (2019). Reviewing the potential for including
942 habitat fragmentation to improve life cycle impact assessments for land use impacts on
943 biodiversity. In *International Journal of Life Cycle Assessment* (Vol. 24, Issue 12, pp. 2206–2219).
944 Springer Verlag. <https://doi.org/10.1007/s11367-019-01647-1>

945 Lebel, L., Mungkung, R., Gheewala, S. H., & Lebel, P. (2010). Innovation cycles, niches and
946 sustainability in the shrimp aquaculture industry in Thailand. *Environmental Science and Policy*, 13(4),
947 291–302. <https://doi.org/10.1016/j.envsci.2010.03.005>

948 Luu, Q. H., Nguyen, T. B. T., Nguyen, T. L. A., Do, T. T. T., Dao, T. H. T., & Padungtod, P. (2021).
949 Antibiotics use in fish and shrimp farms in Vietnam. *Aquaculture Reports*, 20, 100711.
950 <https://doi.org/10.1016/j.aqrep.2021.100711>

951 Macusi, E. D., Estor, D. E. P., Borazon, E. Q., Clapano, M. B., & Santos, M. D. (2022). Environmental
952 and Socioeconomic Impacts of Shrimp Farming in the Philippines: A Critical Analysis Using
953 PRISMA. *Sustainability*, 14(5), 2977. <https://doi.org/10.3390/su14052977>

954 Maiti, P., Panda, A. K., & Ghorai, S. K. (2021). Mapping the wider impact of brackish water shrimp
955 culture on the local environment and economy: A brief review. *International Journal of Fisheries and*
956 *Aquatic Studies*, 9(2), 185–188. <https://doi.org/10.22271/fish.2021.v9.i2c.2449>

957 Majluf, P., Matthews, K., Pauly, D., Skerritt, D. J., & Palomares, M. L. D. (2024). A review of the global
958 use of fishmeal and fish oil and the Fish In:Fish Out metric. *Science Advances*, 10(42).
959 <https://doi.org/10.1126/sciadv.adn5650>

960 Martínez-Porchas, M., & Martínez-Cordova, L. R. (2012). World aquaculture: Environmental impacts
961 and troubleshooting alternatives. In *The Scientific World Journal* (Vol. 2012).
962 <https://doi.org/10.1100/2012/389623>

963 Martínez-Ramón, V., Bromwich, T., Modernel, P., Poore, J., & Bull, J. W. (2024). Alternative Life Cycle
964 Impact Assessment Methods for Biodiversity Footprinting Could Motivate Different Strategic
965 Priorities: A Case Study for a Dutch Dairy Multinational. *Business Strategy and the Environment*.
966 <https://doi.org/10.1002/bse.4072>

967 Mavraganis, T., Constantina, C., Kolygas, M., Vidalis, K., & Nathanailides, C. (2020). Environmental
968 issues of Aquaculture development (Vol. 24, Issue 2). www.ejabf.journals.ekb.eg

969 Mungkung, R., Udo de Haes, H., & Clift, R. (2006). Potentials and Limitations of Life Cycle Assessment
970 in Setting Ecolabelling Criteria: A Case Study of Thai Shrimp Aquaculture Product (5 pp). *The*
971 *International Journal of Life Cycle Assessment*, 11(1), 55–59. <https://doi.org/10.1065/lca2006.01.238>

972 Ngoc, Q. T. K., Xuan, B. B., Sandorf, E. D., Phong, T. N., Trung, L. C., & Hien, T. T. (2021).
973 Willingness to adopt improved shrimp aquaculture practices in Vietnam. *Aquaculture Economics &*
974 *Management*, 25(4), 430–449. <https://doi.org/10.1080/13657305.2021.1880492>

975 Nguyen, T. T., Huynh, H. H., Luu, D. D., Tran, C. T. H., Tsai, W.-P., & Sammut, J. (2024). Unveiling
976 the pandemic's ripples: a study of COVID-19's effects on catfish and shrimp farmers and export
977 enterprises in Vietnam. *Aquaculture International*, 32(7), 9457–9478.
978 <https://doi.org/10.1007/s10499-024-01623-z>

979 Nyberg, O., Rico, A., Guinée, J. B., & Henriksson, P. J. G. (2021). Characterizing antibiotics in LCA—a
980 review of current practices and proposed novel approaches for including resistance. *The International*
981 *Journal of Life Cycle Assessment*, 26(9), 1816–1831. <https://doi.org/10.1007/s11367-021-01908-y>

982 Oddson, G. V. (2020). A Definition of Aquaculture Intensity Based on Production Functions—The
983 Aquaculture Production Intensity Scale (APIS). *Water*, 12(3), 765.
984 <https://doi.org/10.3390/w12030765>

985 Ossés de Eicker, M., Hischer, R., Kulay, L. A., Lehmann, M., Zah, R., & Hurni, H. (2010). The
986 applicability of non-local LCI data for LCA. *Environmental Impact Assessment Review*, 30(3), 192–199.
987 <https://doi.org/10.1016/j.eiar.2009.08.007>

988 Pazmiño, M. L., Chico-Santamarta, L., Boero, A., & Ramirez, A. D. (2024). Environmental life cycle
989 assessment and potential improvement measures in the shrimp and prawn aquaculture sector: A
990 literature review. In *Aquaculture and Fisheries*. KeAi Communications Co.
991 <https://doi.org/10.1016/j.aaf.2024.06.003>

992 Pedersen, E., & Remmen, A. (2022). Challenges with product environmental footprint: a systematic
993 review. *The International Journal of Life Cycle Assessment*, 27(2), 342–352.
994 <https://doi.org/10.1007/s11367-022-02022-3>

995 Philis, G., Ziegler, F., Gansel, L. C., Jansen, M. D., Gracey, E. O., & Stene, A. (2019). Comparing Life
996 Cycle Assessment (LCA) of Salmonid Aquaculture Production Systems: Status and Perspectives.
997 *Sustainability*, 11(9), 2517. <https://doi.org/10.3390/su11092517>

998 Poore, J. (2021). *Reducing Agriculture's Environmental Impacts through Diverse Producers*. University of Oxford
999 (United Kingdom).

1000 Poore, J., & Nemecek, T. (2018). Reducing food's environmental impacts through producers and
1001 consumers. *Science*, 360(6392), 987–992. <https://doi.org/10.1126/science.aag0216>

1002 Popper, K. R. (1959). *The Logic of Scientific Discovery*. Hutchinson.

1003 Prompatanapak, A., & Lopetcharat, K. (2020). Managing changes and risk in seafood supply chain: A
1004 case study from Thailand. *Aquaculture*, 525. <https://doi.org/10.1016/j.aquaculture.2020.735318>

1005 Ran, Y., Nilsson Lewis, A., Dawkins, E., Grah, R., Vanhuyse, F., Engström, E., & Lambe, F. (2022).
1006 Information as an enabler of sustainable food choices: A behavioural approach to understanding

1007 consumer decision-making. *Sustainable Production and Consumption*, 31, 642–656.
1008 <https://doi.org/10.1016/j.spc.2022.03.026>

1009 Ritchie, H., Rosado, P., & Roser, M. (2022). Environmental Impacts of Food Production.
1010 <https://ourworldindata.org/environmental-impacts-of-food>

1011 Roberts, S., Jacquet, J., Majluf, P., & Hayek, M. N. (2024). Feeding global aquaculture. *Science Advances*,
1012 10(42). <https://doi.org/10.1126/sciadv.adn9698>

1013 Rosenbaum, R. K., Bachmann, T. M., Gold, L. S., Huijbregts, M. A. J., Jolliet, O., Juraske, R., Koehler,
1014 A., Larsen, H. F., MacLeod, M., Margni, M., McKone, T. E., Payet, J., Schuhmacher, M., van de
1015 Meent, D., & Hauschild, M. Z. (2008). USEtox—the UNEP-SETAC toxicity model:
1016 recommended characterisation factors for human toxicity and freshwater ecotoxicity in life cycle
1017 impact assessment. *The International Journal of Life Cycle Assessment*, 13(7), 532–546.
1018 <https://doi.org/10.1007/s11367-008-0038-4>

1019 Sala, S., Amadei, A. M., Beylot, A., & Ardente, F. (2021). The evolution of life cycle assessment in
1020 European policies over three decades. *The International Journal of Life Cycle Assessment*, 26(12), 2295–
1021 2314. <https://doi.org/10.1007/s11367-021-01893-2>

1022 Salin, K. R., & Arome Ataguba, G. (2018). Aquaculture and the Environment: Towards Sustainability.
1023 In *Sustainable Aquaculture* (pp. 1–62). Springer International Publishing.
1024 https://doi.org/10.1007/978-3-319-73257-2_1

1025 Sanchez, J. B., Zambrano, G. G. M., Briones, J. R. L. J., Coronel, C. A. V. (2023). *Life Cycle Analysis of*
1026 *Farmed Shrimp of the Species Litopenaeus Vannamei in the Province of Guayas*. 59, 1254–1271.
1027 www.migrationletters.com

1028 Sasmito, S. D., Taillardat, P., Adinugroho, W. C., Krisnawati, H., Novita, N., Fatoyinbo, L., Friess, D.
1029 A., Page, S. E., Lovelock, C. E., Murdiyarso, D., Taylor, D., & Lupascu, M. (2025). Half of land use
1030 carbon emissions in Southeast Asia can be mitigated through peat swamp forest and mangrove
1031 conservation and restoration. *Nature Communications*, 16(1), 740. [https://doi.org/10.1038/s41467-](https://doi.org/10.1038/s41467-025-55892-0)
1032 [025-55892-0](https://doi.org/10.1038/s41467-025-55892-0)

1033 Serpa, D., & Duarte, P. (2008). Impacts of aquaculture and mitigation measures. *Dynamic Biochemistry*,
1034 *Process Biotechnology and Molecular Biology*, 2, 1-20.

- 1035 Shao, Y., Wang, Y., Yuan, Y., & Xie, Y. (2021). A systematic review on antibiotics misuse in livestock
1036 and aquaculture and regulation implications in China. *Science of The Total Environment*, 798, 149205.
1037 <https://doi.org/10.1016/j.scitotenv.2021.149205>
- 1038 Soliman, N. F., & Yacout, D. M. M. (2016). Aquaculture in Egypt: status, constraints and potentials.
1039 *Aquaculture International*, 24(5), 1201–1227. <https://doi.org/10.1007/s10499-016-9989-9>
- 1040 Sun, Y., Hou, H., Dong, D., Zhang, J., Yang, X., Li, X., & Song, X. (2023). Comparative life cycle
1041 assessment of whiteleg shrimp (*Penaeus vannamei*) cultured in recirculating aquaculture systems
1042 (RAS), biofloc technology (BFT) and higher-place ponds (HPP) farming systems in China.
1043 *Aquaculture*, 574. <https://doi.org/10.1016/j.aquaculture.2023.739625>
- 1044 Sun, Y., Yu, J., Yang, X., Jia, L., Dong, D., LI, M., Song, X., & li, xian. (2025). *A Consequential Life Cycle*
1045 *Assessment of Tunnel Greenhouse Aquaculture Systems: A Case of Whiteleg Shrimp Farming in China*.
1046 <https://doi.org/10.2139/ssrn.5240908>
- 1047 Talon, O. (2016). CAN A SCIENTIST TRUSTFULLY STAND ON THE SHOULDERS OF A LCA
1048 GIANT? *Conference 2016 – Life Cycle Thinking for leading managers*.
- 1049 Tamariska, A. Y., Priyono, S. B., Suadi, & Triyatno, B. (2024). Towards Sustainable Shrimp Farming:
1050 Life Cycle Assessment of Farming Practices at the Less Favorable Areas of Yogyakarta’s Southern
1051 Coast. *Turkish Journal of Fisheries and Aquatic Sciences*, 24(9). <https://doi.org/10.4194/TRJFAS23908>
- 1052 The Marine Fish PEFCE project (2025). *Product Environmental Footprint Category Rules for Unprocessed*
1053 *Marine Fish for Human Consumption*. European Commission.
1054 https://www.marinefishpefcr.eu/files/ugd/2c010a_921a5c3f804347a0ad08b2bfd6cc20a1.pdf
- 1055 Luu, Q. H., Nguyen, T. B. T., Nguyen, T. L. A., Do, T. T. T., Dao, T. H. T., & Padungtod, P. (2021).
1056 Antibiotics use in fish and shrimp farms in Vietnam. *Aquaculture Reports*, 20, 100711.
1057 <https://doi.org/10.1016/j.aqrep.2021.100711>
- 1058 Tantipanatip, W., Jitpukdee, S., Keeratiurai, P., Tantikamton, K., & Thaneer, N. (2014). Life cycle
1059 assessment of pacific white shrimp (*penaeus vannamei*) farming system in trang province,
1060 Thailand. *Advanced Materials Research*, 1030–1032, 679–682.
1061 <https://doi.org/10.4028/www.scientific.net/AMR.1030-1032.679>

1062 Thornber, K., Verner-Jeffreys, D., Hinchliffe, S., Rahman, M. M., Bass, D., & Tyler, C. R. (2020).
1063 Evaluating antimicrobial resistance in the global shrimp industry. *Reviews in Aquaculture*, 12(2), 966–
1064 986. <https://doi.org/10.1111/raq.12367>

1065 Vafi, K., & Brandt, A. R. (2014). Reproducibility of LCA Models of Crude Oil Production. *Environmental*
1066 *Science & Technology*, 48(21), 12978–12985. <https://doi.org/10.1021/es501847p>

1067 Viera-Romero, A. M., Diemont, S. A. W., Selfa, T. L., & Luzadis, V. A. (2024). The sustainability of
1068 shrimp aquaculture: An energy-based case study in the Gulf of Guayaquil thirty years later.
1069 *Renewable and Sustainable Energy Reviews*, 194, 114326. <https://doi.org/10.1016/j.rser.2024.114326>

1070 Winter, L., Lehmann, A., Finogenova, N., & Finkbeiner, M. (2017). Including biodiversity in life cycle
1071 assessment – State of the art, gaps and research needs. In *Environmental Impact Assessment Review*
1072 (Vol. 67, pp. 88–100). Elsevier Inc. <https://doi.org/10.1016/j.eiar.2017.08.006>

1073 Yang, P., Bastviken, D., Lai, D. Y. F., Jin, B. S., Mou, X. J., Tong, C., & Yao, Y. C. (2017). Effects of
1074 coastal marsh conversion to shrimp aquaculture ponds on CH₄ and N₂O emissions. *Estuarine,*
1075 *Coastal and Shelf Science*, 199, 125–131. <https://doi.org/10.1016/j.ecss.2017.09.023>

1076 Znachor, P., Nedoma, J., Kolar, V., & Matoušů, A. (2023). Spatial and temporal variability of methane
1077 emissions and environmental conditions in a hyper-eutrophic fishpond. *Biogeosciences*, 20(20), 4273–
1078 4288. <https://doi.org/10.5194/bg-20-4273-2023>

1079 Zhang, S., Huang, J., Ji, Y., Zhang, J., Pei, P., & Gao, J. (2024). Nitrogen and phosphorus cycling for
1080 aquaculture ponds with artificially-controlled drainage: Sources, sinks and treatment strategies.
1081 *Ecological Engineering*, 206, 107331. <https://doi.org/10.1016/j.ecoleng.2024.107331>

1082 Ziyadi, M., & Al-Qadi, I. L. (2019). Model uncertainty analysis using data analytics for life-cycle
1083 assessment (LCA) applications. *The International Journal of Life Cycle Assessment*, 24(5), 945–959.
1084 <https://doi.org/10.1007/s11367-018-1528-7>

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1086 11. Abstract Art

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*Abstract art: Different global warming results for
farmed shrimp in reviewed LCA studies*